





Lublin, 26.09.2024

STATEMENT

We, BioMaxima s.a having a registered office in Poland, at Vetterow 5 street, 20-277 Lublin, Poland assign SRL Sanmedico, having a registered office at A. Corobceanu street 7A, apt. 9, Chişinău MD-2012, Moldova, as authorized representative in correspondence with the conditions of directive 93/42/EEC, 98/79/EEC and (or) 90/385/EEC.

We declare that the company mentioned above is authorized to register, notify, renew or modify the registration of medical devices on the territory of the Republic of Moldova.

atrycja Paniak-Sankowska



No. J - 2623/4/2022

This is to certify that:

BioMaxima S.A. ul. Vetterów 5, 20-277 Lublin

is in conformance with

PN-EN ISO 9001:2015-10

in the following scope of activities:

- Design, manufacturing, sales and distribution of reagents, tests, microbiological media and systems for in-vitro diagnostics and industrial applications
- Distribution of products and service of in vitro diagnostic equipment and industrial applications

The audit carried out by the Polish Centre for Testing and Certification has afforded evidence of the above.

This Certificate shall remain valid provided that above standard are respected by the Organization.

This certificate is valid:

from 16.09.2022 to 15.09.2025

Issued under the Contract No. 3009/JM/4/2022 Date of certification decision: 09.09.2022 Certificate bears a qualified signature. Warsaw, 09.09.2022







POLSKIE CENTRUM AKREDYTACJI

POLISH CENTRE FOR ACCREDITATION



Sygnatariusz EA MLA EA MLA Signatory

CERTYFIKAT AKREDYTACJI

LABORATORIUM BADAWCZEGO

ACCREDITATION CERTIFICATE OF TESTING LABORATORY

Nr AB 1863

Potwierdza się, że: / This is to confirm that:

BioMaxima S.A.

Laboratorium Kontroli Jakości Mikrobiologia

ul. Vetterów 5 20-277 Lublin

spełnia wymagania normy PN-EN ISO/IEC 17025:2018-02 meets requirements of the PN-EN ISO/IEC 17025:2018-02 standard

Akredytowana działalność jest określona w Zakresie Akredytacji Nr AB 1863

Accredited activity is defined in the Scope of Accreditation No AB 1863

Akredytacja pozostaje w mocy pod warunkiem przestrzegania wymagań jednostki akredytującej określonych w kontrakcie Nr AB 1863

This accreditation remains in force provided the Laboratory observes the requirements of Accreditation Body defined in the Contract No AB 1863



DYREKTOR
POLSKIEGO CENTRUM AKREDYTACJI

PEA

LUCYNA OLBORSKA



EU Declaration of Conformity

for In Vitro Diagnostic Medical Devices according to the Regulation (EU) 2017/746

Manufacturer: BioMaxima S.A., Vetterów 5, 20-277 Lublin, Poland

Product identification: according to the Product List Conformity route/ Certificate: Annexes I, II and III

Notified Body & Certificate: N/A

Intended purpose: microbiological media and supplements for the cultivation of microorganisms

Classification: Class A, Rule 5

The Basic UDI code - assigned to a group of products

Statement: We herewith under our sole responsibility declare that the mentioned products meet the

provisions of the Regulation (EU) 2017/746 and Standards.

The manufacturer is exclusively responsible for the declaration of conformity.

Additional information: N/A

Signed on behalf of BioMaxima S.A.:

Place and date of issue:

Lublin, 2025-01-20

Name: Henryk Lewczuk

Function: Vice President

Name: Patrycja Paniak- Sankowska

Function: Proxy

EU Declaration of Conformity Product List

| Basic UDI | Group of products | |
|---------------|---|---|
| 59026439APSYR | Dehydrated media | |
| PS 214 | Acetamide Medium | |
| PS 98 | B.G.A. LAB-AGAR™ acc. to ISO 6579 | |
| PS 88 | Bile Esculin Azide LAB-AGAR™ acc.to ISO 7899-2 | |
| PS 155 | Bile Esculin LAB-AGAR™ | |
| PS 01 | Bismuth Sulfite LAB-AGAR™ | |
| PS 159 | Blood Free Campylobacter CCDA LAB-AGAR™ Base | |
| PS 02 | Blood LAB-AGAR™ Base | |
| PS 205 | Blood No 2 LAB-AGAR™ Base | |
| PS 04 | Brain Heart Infusion Broth | |
| PS 03 | Brain Heart Infusion LAB-AGAR™ | |
| PS 137 | Brucella Broth | |
| PS 05 | Brucella LAB-AGAR™ | |
| PS 49 | Cetrimide LAB-AGAR™ | |
| PS 590 | Chromogenic Candida LAB-AGAR™ | |
| PS 589 | Chromogenic E.coli 0157 LAB-AGAR™ | |
| PS 165 | Chromogenic Listeria acc.to Ottavianii and Agostii LAB-AGAR™ Base | |
| PS 598 | Chromogenic Salmonella LAB-AGAR™ | |
| PS 371 | Chromogenic Strep B LAB-AGAR™ | |
| PS 530 | Citrate Christensen LAB-AGAR™ | |
| PS 34 | CLED LAB-AGAR™ | |
| PS 176 | Clostridium difficile LAB-AGAR™ Base | |
| PS 190 | Columbia CNA LAB-AGAR™ | |
| PS 06 | Columbia LAB-AGAR™ Base | |
| PS 715E | EMB LAB-AGAR™ | |
| PS 716E | EMB Levin LAB-AGAR™ | |
| PS 110 | Enrichment Broth | |
| PS 41 | Eosin Methylene Blue LAB-AGAR™ (EMB) | |
| PS 53 | Ewing Malonate Modified Broth | |
| PS 365 | Fastidious Anaerobe Broth | |
| PS 364 | Fastidious Anaerobe LAB-AGAR™ | |
| PS 139 | G.C. LAB-AGAR™ Base | |
| PS 182 | Haemophilus Test LAB-AGAR™ Base | |
| PS 07 | Hektoen LAB-AGAR™ | - |
| PS 188 | Kligler Iron LAB-AGAR™ | |
| PS 08 | Kligler Iron LAB-AGAR™ acc.to ISO 10273 | |
| PS 163 | Legionella CYE LAB-AGAR™ Base | |
| PS 104 | Listeria acc.to Oxford LAB-AGAR™ Base | |
| PS 104 | Listeria acc.to Palcam LAB-AGAR™ Base | |
| PS 241 | Lowenstein-Jensen Medium Base | |
| PS 150 | Lysine Decarboxylase Broth | |
| PS 10 | Mac Conkey LAB-AGAR™ | |
| PS 183 | Mac Conkey Nr 1 LAB-AGAR™ | |
| PS 183 | Mac Conkey w/o crystal violet and w/o sodium chloride LAB-AGAR™ | _ |

| PS 186 | Mannitol Salt acc.to Chapman LAB-AGAR™ |
|--------|---|
| PS 13 | Mannitol Salt LAB-AGAR™ |
| PS 525 | Motility LAB-AGAR™ |
| PS 59 | MRS LAB-AGAR™ |
| PS 92 | MRVP Medium |
| PS 79 | Mueller Hinton 2 LAB-AGAR™ |
| PS 15 | Mueller Hinton Broth |
| PS 90 | Nutrient Broth |
| PS 85 | Nutrient LAB-AGAR™ |
| PS 127 | Peptone Water |
| PS 62 | Phenylalanine LAB-AGAR™ |
| PS 583 | Ringer Solution 1/4 Strength |
| PS 233 | RPMI MOPS LAB-AGAR™ BASE |
| PS 227 | Sabouraud 2% Dex. w. Chloramhenicol LAB-AGAR™ |
| PS 547 | Sabouraud Dextrose 2% LAB -AGAR™ |
| PS 146 | Sabouraud Dextrose Broth |
| PS 16 | Sabouraud Dextrose LAB-AGAR™ |
| PS 32 | Sabouraud Dextrose w. Chloramphenicol LAB-AGAR™ |
| PS 94 | Sabouraud Dextrose w. Chloramphenicol and Cycloheximide LAB-AGAR™ |
| PS 248 | Sabouraud Dextrose w. Chloramphenicol and Gentamycine LAB-AGAR™ |
| PS 17 | Salmonella Shigella LAB-AGAR™ |
| PS 19 | Schaedler Broth |
| PS 18 | Schaedler LAB-AGAR™ |
| PS 20 | Simmons Citrate LAB-AGAR™ |
| PS 68 | Sodium Selenite Broth |
| PS 21 | Sorbitol Mac Conkey LAB-AGAR™ |
| PS 43 | Thioglycollate Fluid Medium |
| PS 72 | Todd Hewitt Broth |
| PS 44 | Triple Sugar Iron LAB-AGAR™ |
| PS 23 | Trypticasein Soy Broth (TSB) |
| PS 22 | Trypticasein Soy LAB-AGAR™ (TSA) |
| PS 132 | Tryptophan Culture Broth |
| PS 593 | Urea Broth |
| PS 506 | Urea Broth Base |
| PS 134 | Urea Indol Broth |
| PS 24 | Urea LAB-AGAR™ Base |
| PS 529 | Urea Modified Broth |
| PS 45 | XLD LAB-AGAR™ |
| PS 145 | Yersinia Selective LAB-AGAR™ Base |

| 59026439ASLYL | Supplements and Reagents | |
|---------------|--|--|
| SL 0135 | 0,5M EDTA | |
| SL 0004 | Brucella Supplement | |
| SL 0005 | Campylobacter CCDA Selective Supplement | |
| SL 0006 | Campylobacter Selective Supplement Skirrow | |

| SL 0001 | Chromogenic Listeria Set Supplement acc. to ISO 11290 |
|-----------|---|
| SL 0048 | Clostridium difficile Selective Supplement |
| SL 0160 | Defibrinated Sheep Blood |
| SL 0150 | Defibrinated Horse Blood |
| SL 0035 | Egg's Yolk Sterile Emulsion |
| SL 0054 | Haemophilus Test Supplement |
| SL 0116 | Horse Serum |
| SL 0153 | Kovac's reagent |
| SL 0046 | Legionella BCYE Supplement w/o Cysteine |
| SL 0047 | Legionella BCYE Supplement w/o Cysteine |
| SL 0017 | Legionella CYE Growth Supplement |
| SL 0053 | Legionella CYE Supplement with Cysteine |
| SL 0018 | Legionella GVPC Supplement |
| SL 0081 | Listeria Modified acc. to Oxford Supplement |
| SL 0021 | Listeria Selective acc. to Oxford Supplement |
| SL 0022 | Listeria Selective acc. to Palcam Supplement |
| SL 0136 | Phenylboronic Acid |
| SL 0112 | Ringer Tablets |
| SL 0061 | Salmonella Chromogenic Supplement |
| SL 0061 A | Salmonella Chromogenic Supplement |
| SL 0039 | Trichomonas Selective Supplement |
| SL 0044 | Yersinia CIN Supplement |

| 59026439ADPXF | Dip -slide |
|---------------|--------------------------------------|
| DP 12 | CLED LAB-AGAR™/ Mac CONKEY LAB-AGAR™ |

| 59026439APPYK | Ready to use media on plates | |
|---------------|--|--|
| PP 1111 | Azide Blood LAB-AGAR™ + 5% SB | Dett. |
| PP 1360 | BGA LAB-AGAR™ acc. to ISO 6579 | <u> </u> |
| PP 0011 | BHI LAB-AGAR™ + 6mg /l Vancomycin | 100 B |
| PP 0125 | BHI LAB-AGAR™ + Potassium Tellurite | INT I |
| PP 1075 | Bile Esculin Azide LAB-AGAR™ | vidgen a |
| PP 0019 | Bile Esculin LAB-AGAR™ | A H nee |
| PP 1040 | Bismuth Sulfite LAB-AGAR™ | NAME AND ADDRESS OF THE PARTY O |
| PP 1110 | Blood LAB-AGAR™ + 5% SB | Fel(OL.u |
| PP 0015 | Blood No 2 LAB-AGAR™ + 5% SB | U.S. 1171 & |
| PP 0007 | Brain Heart Infusion LAB-AGAR™ | 2 T |
| PP 0120 | Brucella LAB-AGAR™ + 5% HB | .0 9 11 |
| PP 0082 | Brucella LAB-AGAR™ + 5% SB + vit. K + Hemine | |
| PP 0066 | Burkholderia cepacia LAB-AGAR™ | |
| PP 0017 | CCDA LAB-AGAR™ | |
| PP 1310 | Cetrimide LAB-AGAR™ | |
| PP 0053 | Chocolate 2 LAB-AGAR™ + PV (AB) | |
| PP 0075 | Chocolate 3 LAB-AGAR™ + PV (ABV) | |
| PP 1080 | Chocolate LAB-AGAR™ + PV | |
| PP 1261 | Chocolate LAB-AGAR™ + PV + Bacitracin | |

| PP 1082 | Chocolate LAB-AGAR™ + PV + VCAT |
|----------|--|
| PP 1083 | Chocolate LAB-AGAR™ + PV + VCN |
| PP 0028 | Chromagar™ Salmonella PLUS LAB-AGAR™ |
| PP 2100 | Chromogenic Acinetobacter LAB-AGAR™ |
| PP 0139 | Chromogenic Acinetobacter MDR LAB-AGAR™ |
| PP 0137 | Chromogenic C. difficile LAB-AGAR™ |
| PP 0201 | Chromogenic C. perfringens LAB-AGAR™ |
| PP 0202 | Chromogenic C3GR LAB-AGAR™ |
| PP 0163 | Chromogenic Campylobacter LAB-AGAR™ |
| PP 0001B | Chromogenic Candida LAB-AGAR™ |
| PP 0255 | Chromogenic Candida Plus LAB-AGAR™ |
| PP 0239 | Chromogenic COL-APSE LAB-AGAR™ |
| PP 0041 | Chromogenic E. coli O157 LAB-AGAR™ |
| PP 0143 | Chromogenic E. coli STEC LAB-AGAR™ |
| PP 0155 | Chromogenic ESBL Mod. LAB-AGAR™ |
| PP 0157 | Chromogenic KPC Mod. LAB-AGAR™ |
| PP 0286 | Chromogenic LIN-R LAB-AGAR™ |
| PP 5009 | Chromogenic Listeria LAB-AGAR™ acc. ISO 11290 |
| PP 0166 | Chromogenic Listeria LAB-AGAR™ acc. ISO 11290 (140 mm) |
| PP 0043 | Chromogenic MRSA Modified LAB-AGAR™ |
| PP 0206 | Chromogenic Orientation LAB-AGAR™ |
| PP 0045 | Chromogenic S. aureus Modified LAB-AGAR™ |
| PP 0115 | Chromogenic Salmonella LAB-AGAR™ |
| PP 0104 | Chromogenic Strepto B LAB-AGAR™ |
| PP 0198 | Chromogenic Super Carba LAB-AGAR™ |
| PP 0204 | Chromogenic UTI LAB-AGAR™ |
| PP 0246 | Chromogenic VRE LAB-AGAR™ |
| PP 0074 | Chromogenic VRE LAB-AGAR™ |
| PP 0144 | Chromogenic Yersinia LAB-AGAR™ |
| PP 1030 | CLED LAB-AGAR™ |
| PP 0004 | Clostridium difficile LAB-AGAR™ |
| PP 0242 | Columbia CAP LAB-AGAR™ + 5% SB |
| PP 1191 | Columbia CNA LAB-AGAR™ + 5% SB |
| PP 0020 | Columbia LAB-AGAR™ + 5% HB |
| PP 1190 | Columbia LAB-AGAR™ + 5% SB |
| PP 0211 | Columbia LAB-AGAR™ + 5% SB + CV |
| PP 0051 | Dermatophytes LAB-AGAR™ |
| PP 1011 | Eosin Methylene Blue LAB-AGAR™ |
| PP 0270 | Fastidious Anaerobe LAB-AGAR™ +5% HB |
| PP 0271 | Fastidious Anaerobe LAB-AGAR™ +5% HB (120 mm) |
| PP 0272 | Fastidious Anaerobe LAB-AGAR™ +5% HB (140 mm) |
| PPK 0270 | Fastidious Anaerobe LAB-AGAR™ +5% HB (120 mm square) |
| PP 0005 | Gardnerella vaginalis LAB-AGAR™ |
| PP 1260 | Haemophilus Test Medium (HTM) |
| PP 1060 | Hektoen LAB-AGAR™ |
| PP 0022 | Karmali LAB-AGAR™ |

| PP 0178 | Legionella BCYE + AB LAB-AGAR™ | |
|----------|---|-----|
| PP 0026 | Legionella BCYE LAB-AGAR™ | |
| PP 0025 | Legionella GVPC LAB-AGAR™ | |
| PP 0179 | Legionella MWY LAB-AGAR™ | |
| PP 0027 | Legionella without Cysteine LAB-AGAR™ | |
| PP 1017 | Mac Conkey LAB-AGAR™ | |
| PP 1050 | Mannitol Salt LAB-AGAR™ | |
| PP 0023 | MRS LAB - AGAR™ | |
| PP 1170 | Mueller Hinton 2 LAB-AGAR™ | |
| PP 0016 | Mueller Hinton 2 LAB-AGAR™ | |
| PP 0016K | Mueller Hinton 2 LAB-AGAR™ (120 mm square) | |
| | | |
| PP 0033 | Mueller Hinton 2 LAB-AGAR™ | |
| PP 0077 | Mueller Hinton 2 LAB-AGAR™ + 2% NaCl | |
| PP 1172 | Mueller Hinton 2 LAB-AGAR™ + 5% SB | |
| PP 0018 | Mueller Hinton 2 LAB-AGAR™ + 5% SB | |
| PP 0073 | Mueller Hinton 2 LAB-AGAR™ + Cloxacillin | |
| PP 0083 | Mueller Hinton 2 LAB-AGAR™ + NAD + 5% HB | |
| PP 0092 | Mueller Hinton 2 LAB-AGAR™ + NAD + 5% HB | |
| PP 0006 | Mueller Hinton LAB-AGAR™ + 4% NaCl + Oxacillin | |
| PP 0105 | Mueller Hinton LAB-AGAR™ + Glucose + Methylene Blue | |
| PP 1503 | Nutrient LAB-AGAR™ | |
| PP 0128 | ORSIM LAB-AGAR™ | |
| PP 1292 | Oxford Listeria LAB-AGAR™ | |
| PP 1502 | Palcam Listeria LAB-AGAR™ | |
| PP 0095 | RPMI MOPS LAB-AGAR™ | |
| PP 0124 | RPMI MOPS LAB-AGAR™ | |
| PP 1230 | Sabouraud Dextrose LAB-AGAR™ | |
| PP 0021 | Sabouraud Dextrose with Chloramphenicol and Cycloheximide LAB-AGAR™ | |
| PP 1232 | Sabouraud Dextrose with Chloramphenicol and Gentamicin LAB-AGAR™ | |
| PP 1231 | Sabouraud Dextrose with Chloramphenicol LAB-AGAR™ | |
| PP 1250 | Salmonella Shigella LAB-AGAR™ | |
| PP 0002 | Schaedler Anaerobe LAB-AGAR™ | |
| PP 0012 | Schaedler Anaerobe LAB-AGAR™ + 5% SB | |
| PP 0223 | Schaedler Anaerobe LAB-AGAR™ + 5% SB + vit. K1 | 100 |
| PP 0003 | Schaedler Anaerobe LAB-AGAR™ + 5% SB + vit. K3 | |
| PP 0039 | Schaedler LAB-AGAR™ + 5% SB + CNA | |
| PP 0014 | Schaedler LAB-AGAR™ + 5% SB + K+ VA | |
| PP 0013 | Schaedler LAB-AGAR™ + 5% SB + VA + NEO | |
| PP 1021 | Sorbitol Mac Conkey LAB-AGAR™ | 4 |
| PP 1180 | Trypticasein Soy LAB-AGAR™ | |
| PP 0118 | Trypticasein Soy LAB-AGAR™ | |
| PP 1330 | XLD LAB-AGAR™ | |
| | Yersinia CIN LAB-AGAR™ | |
| PP 1090 | | |
| PD 001 | Blood LAB-AGAR™ + 5% SB / Mac Conkey LAB-AGAR™ Chromogonic Candida LAB AGAR™ / Sabouraud Doyt, LAB AGAR + Go + C | |
| PD 017 | Chromogenic Candida LAB-AGAR™/ Sabouraud Dext. LAB-AGAR + Ge + C | |
| PD 042 | Chromogenic ESBL LAB-AGAR™ / Chromogenic VRE LAB-AGAR™ | |

| PD 081 | Chromogenic KPC LAB-AGAR™ / Chromogenic VRE LAB-AGAR™ |
|--------|--|
| PD 100 | Chromogenic KPC LAB-AGAR™/ Chromogenic SUPER CARBA LAB-AGAR™ |
| PD 091 | Chromogenic KPC LAB-AGAR™/ Mac Conkey LAB-AGAR™ |
| PD 041 | Chromogenic S. aureus LAB-AGAR™ / Chromogenic MRSA LAB-AGAR™ |
| PD 023 | Chromogenic Salmonella / XLD LAB-AGAR™ |
| PD 025 | Chromogenic URI LAB-AGAR™ / Columbia CNA LAB-AGAR™ + 5% SB |
| PD 011 | Columbia CNA LAB-AGAR™ + 5% KB / Mac Conkey LAB-AGAR™ |
| PD 032 | Columbia LAB-AGAR™ + 5% SB / Columbia LAB-AGAR™ + 5% SB |
| PD 014 | Columbia LAB-AGAR™ + 5% SB / Mac Conkey LAB-AGAR™ |
| PD 094 | Mac Conkey LAB-AGAR™ / Chromogenic Orientation LAB-AGAR™ |
| PD 043 | Sabouraud Dextr. w. Chloramph. LAB-AGAR™ / Sabouraud Dextr. w. Actidione LAB-AGAR™ |
| PD 077 | Sabouraud LAB-AGAR™ / Chromogenic Candida LAB-AGAR™ |
| PD 076 | Sabouraud LAB-AGAR™ + Chloramphenicol / Fungisel LAB-AGAR™ |
| PD 021 | Schaedler LAB-AGAR™ + 5% SB / Schaedler LAB-AGAR™ + 5% SB + K + VA |
| PD 022 | Schaedler LAB-AGAR™ + 5% SB / Schaedler LAB-AGAR™ + 5% SB + NEO + VA |
| PD 006 | SS LAB-AGAR™ / Bismuth Sulfite LAB-AGAR™ |
| PD 093 | Schaedler LAB-AGAR™ + 5% SB + CNA/ Schaedler LAB-AGAR™ + 5% SB + K+ VA |
| PD 019 | SS LAB-AGAR™ / Hektoen LAB-AGAR™ |
| PD 037 | SS LAB-AGAR™ / XLD LAB-AGAR™ |

| 59026439ABTXH | Ready to use media in bottles |
|---------------|--------------------------------|
| BT 5054.01 | 0,85% Sodium Chloride |
| BT 5054.02 | 0,85% Sodium Chloride |
| BT 5054.05 | 0,85% Sodium Chloride |
| BT 5045.01 | 0,9% Sodium Chloride |
| BT 5045.02 | 0,9% Sodium Chloride |
| BT 5045.05 | 0,9% Sodium Chloride |
| BT 5052.01 | 10% Lactose Broth |
| BT 5052.02 | 10% Lactose Broth |
| BT 5052.05 | 10% Lactose Broth |
| BT 5214.01 | Acetamide Medium |
| BT 5214.02 | Acetamide Medium |
| BT 5214.05 | Acetamide Medium |
| BT 5053.01 | Arabinose Broth |
| BT 5053.05 | Arabinose Broth |
| ST 5049.01 | Arginine acc. to Falcow Broth |
| T 5049.02 | Arginine acc. to Falcow Broth |
| T 5096.05 | BGA LAB-AGAR™ acc. to ISO 6579 |
| T 5096.01 | BGA LAB-AGAR™ acc. to ISO 6579 |
| T 5096.02 | BGA LAB-AGAR™ acc. to ISO 6579 |
| T 5088.01 | Bile Esculin Azide LAB-AGAR™ |

| BT 5088.02 | Bile Esculin Azide LAB-AGAR™ |
|-------------|-------------------------------------|
| BT 5088.05 | Bile Esculin Azide LAB-AGAR™ |
| BT 5271.01 | Bile salts (2% Sodium deoxycholate) |
| BT 5003.01 | Bismuth Sulfite LAB-AGAR™ |
| BT 5003.02 | Bismuth Sulfite LAB-AGAR™ |
| BT 5034.01 | Blood LAB-AGAR™ Base |
| BT 5034.02 | Blood LAB-AGAR™ Base |
| BT 5034.05 | Blood LAB-AGAR™ Base |
| BT 5008.01 | Brain Heart Infusion Broth |
| BT 5008.02 | Brain Heart Infusion Broth |
| BT 5008.05 | Brain Heart Infusion Broth |
| BT 5122.01 | Cetrimide LAB-AGAR™ |
| BT 5122.02 | Cetrimide LAB-AGAR™ |
| BT 5122.05 | Cetrimide LAB-AGAR™ |
| BT 5017.01 | Chocolate LAB-AGAR™ |
| BT 5017.02 | Chocolate LAB-AGAR™ |
| BT 5017.05 | Chocolate LAB-AGAR™ |
| BT 5222.01 | Chromogenic Candida LAB-AGAR™ |
| BT 5222.02 | Chromogenic Candida LAB-AGAR™ |
| BT 5344.01 | Chromogenic Candida Plus LAB-AGAR™ |
| BT 5344.02 | Chromogenic Candida Plus LAB-AGAR™ |
| BT 5344.05 | Chromogenic Candida Plus LAB-AGAR™ |
| BT 5071.01 | Chromogenic E.coli O157 LAB-AGAR™ |
| BT 5071.02 | Chromogenic E.coli O157 LAB-AGAR™ |
| BT 5276.01 | Chromogenic Orientation LAB-AGAR™ |
| BT 5276.02 | Chromogenic Orientation LAB-AGAR™ |
| BT 5276.05 | Chromogenic Orientation LAB-AGAR™ |
| BT 5072.01 | Chromogenic Salmonella LAB-AGAR™ |
| BT 5072.02 | Chromogenic Salmonella LAB-AGAR™ |
| BT 5072.05 | Chromogenic Salmonella LAB-AGAR™ |
| BT 5042.01* | Citrate Christensen LAB -AGAR™ |
| BT 5042.02* | Citrate Christensen LAB -AGAR™ |
| BT 5102.01 | CLED LAB-AGAR™ |
| BT 5102.02 | CLED LAB-AGAR™ |
| BT 5102.05 | CLED LAB-AGAR™ |
| BT 5011.01 | Columbia LAB-AGAR™ Base |
| BT 5011.02 | Columbia LAB-AGAR™ Base |

| BT 5011.05 | Columbia LAB-AGAR™ Base |
|-------------|-----------------------------------|
| BT 5043.01* | Dulcytol Broth |
| BT 5043.05* | Dulcytol Broth |
| BT 5001.01 | Eosin Methylene Blue LAB-AGAR™ |
| BT 5001.02 | Eosin Methylene Blue LAB-AGAR™ |
| BT 5001.05 | Eosin Methylene Blue LAB-AGAR™ |
| BT 5018.01 | Ewing Malonate Modified Broth |
| BT 5018.02 | Ewing Malonate Modified Broth |
| BT 5018.05 | Ewing Malonate Modified Broth |
| BT 5111.01 | Hektoen LAB-AGAR™ |
| BT 5111.02 | Hektoen LAB-AGAR™ |
| BT 5111.05 | Hektoen LAB-AGAR™ |
| BT 5006.01 | Kligler LAB-AGAR™ |
| BT 5006.02 | Kligler LAB-AGAR™ |
| BT 5006.05 | Kligler LAB-AGAR™ |
| BT 5050.01 | Lysine acc. to Falkow Broth |
| BT 5050.02 | Lysine acc. to Falkow Broth |
| BT 5050.05 | Lysine acc. to Falkow Broth |
| BT 5010.01 | Mac Conkey LAB-AGAR™ |
| BT 5010.02 | Mac Conkey LAB-AGAR™ |
| BT 5010.05 | Mac Conkey LAB-AGAR™ |
| BT 5137.01 | Mac Conkey No 1 LAB-AGAR™ |
| BT 5137.02 | Mac Conkey No 1 LAB-AGAR™ |
| BT 5137.05 | Mac Conkey No 1 LAB-AGAR™ |
| BT 5104.01 | Mannitol Salt LAB-AGAR™ (Chapman) |
| BT 5104.02 | Mannitol Salt LAB-AGAR™ (Chapman) |
| BT 5104.05 | Mannitol Salt LAB-AGAR™ (Chapman) |
| BT 5026.01 | Motility LAB-AGAR™ |
| BT 5026.02 | Motility LAB-AGAR™ |
| BT 5026.05 | Motility LAB-AGAR™ |
| BT 5031.01 | MRS LAB-AGAR™ |
| BT 5031.02 | MRS LAB-AGAR™ |
| BT 5031.05 | MRS LAB-AGAR™ |
| BT 5036.01 | MRVP Medium |
| BT 5036.02 | MRVP Medium |
| BT 5036.05 | MRVP Medium |
| BT 5002.01 | Mueller Hinton 2 LAB-AGAR™ |

| BT 5002.02 | Mueller Hinton 2 LAB-AGAR™ | | | |
|------------|--|--|--|--|
| BT 5002.05 | Mueller Hinton 2 LAB-AGAR™ | | | |
| BT 5055.01 | Mueller Hinton Broth | | | |
| BT 5055.02 | Mueller Hinton Broth | | | |
| BT 5055.05 | Mueller Hinton Broth | | | |
| BT 7012.01 | Nutrient Broth | | | |
| BT 7012.02 | Nutrient Broth | | | |
| BT 7012.05 | Nutrient Broth | | | |
| BT 5126.01 | Nutrient LAB-AGAR™ | | | |
| BT 5126.02 | Nutrient LAB-AGAR™ | | | |
| BT 5126.05 | Nutrient LAB-AGAR™ | | | |
| BT 5116.01 | Nutrient S LAB-AGAR™ | | | |
| BT 5116.02 | Nutrient S LAB-AGAR™ | | | |
| BT 5116.05 | Nutrient S LAB-AGAR™ | | | |
| BT 5046.01 | Peptone Water | | | |
| BT 5046.02 | Peptone Water | | | |
| BT 5046.05 | Peptone Water | | | |
| BT 5029.01 | Phenylalanine LAB-AGAR™ | | | |
| BT 5029.02 | Phenylalanine LAB-AGAR™ | | | |
| BT 5029.05 | Phenylalanine LAB-AGAR™ | | | |
| BT 5105.01 | Phosphate Buffered Saline (PBS)(w/o lons Ca / Mg) | | | |
| BT 5105.02 | Phosphate Buffered Saline (PBS)(w/o ions Ca / Mg) | | | |
| BT 5105.05 | Phosphate Buffered Saline (PBS)(w/o lons Ca / Mg) | | | |
| BT 5140.01 | Ringer Solution | | | |
| BT 5140.02 | Ringer Solution | | | |
| BT 5140.05 | Ringer Solution | | | |
| BT 6000 | Ringer Solution | | | |
| BT 5295.01 | RPMI 1640 + NaHCO3 + L-Glutamine + Phenol Red | | | |
| BT 5295.05 | RPMI 1640 + NaHCO3 + L-Glutamine + Phenol Red | | | |
| BT 5109.01 | Sabouraud 2% Dextrose LAB-AGAR™ with Chloramphenicol | | | |
| BT 5109.02 | Sabouraud 2% Dextrose LAB-AGAR™ with Chloramphenicol | | | |
| BT 5109.05 | Sabouraud 2% Dextrose LAB-AGAR™ with Chloramphenicol | | | |
| BT 5118.01 | Sabouraud 4% Dextrose LAB-AGAR™ with Chloramphenicol | | | |
| BT 5118.02 | Sabouraud 4% Dextrose LAB-AGAR™ with Chloramphenicol | | | |
| BT 5118.05 | Sabouraud 4% Dextrose LAB-AGAR™ with Chloramphenicol | | | |
| BT 5019.01 | Sabouraud Dextrose Broth | | | |
| BT 5019.02 | Sabouraud Dextrose Broth | | | |

| BT 5019.05 | Sabouraud Dextrose Broth | | |
|------------|---|--|--|
| BT 5107.01 | Sabouraud Dextrose LAB-AGAR™ | | |
| BT 5107.02 | Sabouraud Dextrose LAB-AGAR™ | | |
| BT 5107.05 | Sabouraud Dextrose LAB-AGAR™ | | |
| BT 5108.01 | Sabouraud Dextrose LAB-AGAR™ with Chloramphenicol | | |
| BT 5108.02 | Sabouraud Dextrose LAB-AGAR™ with Chloramphenicol | | |
| BT 5108.05 | Sabouraud Dextrose LAB-AGAR™ with Chloramphenicol | | |
| BT 5025.01 | Sabouraud Dextrose with Chloramphenicol and Cycloheximide LAB-AGAR™ | | |
| BT 5025.02 | Sabouraud Dextrose with Chloramphenicol and Cycloheximide LAB-AGAR™ | | |
| BT 5025.05 | Sabouraud Dextrose with Chloramphenicol and Cycloheximide LAB-AGAR™ | | |
| BT 5142.01 | Sabouraud Dextrose with Chloramphenicol and Gentamicin LAB-AGAR™ | | |
| BT 5142.02 | Sabouraud Dextrose with Chloramphenicol and Gentamicin LAB-AGAR™ | | |
| BT 5142.05 | Sabouraud Dextrose with Chloramphenicol and Gentamicin LAB-AGAR™ | | |
| BT 5120.01 | Salmonella Shigella LAB-AGAR™ | | |
| BT 5120.02 | Salmonella Shigella LAB-AGAR™ | | |
| BT 5120.05 | Salmonella Shigella LAB-AGAR™ | | |
| BT 5014.01 | Schaedler Broth | | |
| BT 5014.02 | Schaedler Broth | | |
| BT 5014.05 | Schaedler Broth | | |
| BT 5012.01 | Schaedler Broth + vit. K3 | | |
| BT 5012.02 | Schaedler Broth + vit. K3 | | |
| BT 5012.05 | Schaedler Broth + vit. K3 | | |
| BT 5015.01 | Schaedler LAB-AGAR™ + vit. K3 | | |
| BT 5015.02 | Schaedler LAB-AGAR™ + vit. K3 | | |
| BT 5015.05 | Schaedler LAB-AGAR™ + vit. K3 | | |
| BT 5013.01 | Schaedler LAB-AGAR™ Base | | |
| BT 5013.02 | Schaedler LAB-AGAR™ Base | | |
| BT 5013.05 | Schaedler LAB-AGAR™ Base | | |
| BT 5101.01 | Simmons Citrate LAB-AGAR™ | | |
| BT 5101.02 | Simmons Citrate LAB-AGAR™ | | |
| BT 5101.05 | Simmons Citrate LAB-AGAR™ | | |
| BT 5068.01 | Sodium Selenite Broth | | |
| BT 5068.02 | Sodium Selenite Broth | | |
| BT 5068.05 | Sodium Selenite Broth | | |
| BT 5057.01 | Sorbitol Broth | | |
| BT 5057.02 | Sorbitol Broth | | |
| BT 5057.05 | Sorbitol Broth | | |

| BT 5021.01 | Sorbitol MacConkey LAB-AGAR™ | | |
|---------------|--|--|--|
| BT 5021.02 | Sorbitol MacConkey LAB-AGAR™ | | |
| BT 5021.05 | Sorbitol MacConkey LAB-AGAR™ | | |
| BT 5128.01 | Thioglycollate Fluid Medium | | |
| BT 5128.02 | Thioglycollate Fluid Medium | | |
| BT 5128.05 | Thioglycollate Fluid Medium | | |
| BT 5128.01.BS | Thioglycollate Fluid Medium - septa protective closure | | |
| BT 5005.01 | Todd Hewitt Broth | | |
| BT 5005.02 | Todd Hewitt Broth | | |
| BT 5005.05 | Todd Hewitt Broth | | |
| BT 5024.01 | Trichomonas Medium | | |
| BT 5024.02 | Trichomonas Medium | | |
| BT 5024.05 | Trichomonas Medium | | |
| BT 5123.01 | Trypticasein Soy Broth | | |
| BT 5123.02 | Trypticasein Soy Broth | | |
| BT 5123.05 | Trypticasein Soy Broth | | |
| BT 5192 | Trypticasein Soy Broth | | |
| BT 5000.01 | Trypticasein Soy LAB-AGAR™ | | |
| BT 5000.02 | Trypticasein Soy LAB-AGAR™ | | |
| BT 5000.05 | Trypticasein Soy LAB-AGAR™ | | |
| BT 5030.01 | Tryptophan Culture Broth | | |
| BT 5030.02 | Tryptophan Culture Broth | | |
| BT 5028.01 | Urea Christensen Broth | | |
| BT 5028.02 | Urea Christensen Broth | | |
| BT 5061.01 | Urea Indol Broth | | |
| BT 5061.02 | Urea Indol Broth | | |
| BT 5040.01 | Urea Broth Modified acc. to PN-A-04023 | | |
| BT 5040.02 | Urea Broth Modified acc. to PN-A-04023 | | |
| BT 5040.05 | Urea Broth Modified acc. to PN-A-04023 | | |
| | Viral Transport Medium (VTM) | | |
| BT 5340.01 | Viral Transport Medium (VTM) | | |
| BT 5340.02 | | | |
| BT 5340.05 | Viral Transport Medium (VTM) | | |
| BT 5121.01 | XLD LAB-AGAR™ | | |
| BT 5121.02 | XLD LAB-AGAR™ | | |
| BT 5121.05 | XLD LAB-AGAR™ | | |
| BT 5035.01 | Yersinia CIN LAB-AGAR™ | | |
| BT 5035.02 | Yersinia CIN LAB-AGAR™ | | |

| 59026439APWYZ | Ready to use media in tubes |
|---------------|-------------------------------------|
| PW 3151 | 0,85 % Sodium Chloride |
| PW 3240 | 0,85 % Sodium Chloride |
| PW 3145 | 0,85 % Sodium Chloride |
| PW 3053 | 0,85 % Sodium Chloride |
| PW 3054 | 0,9% Sodium Chloride |
| PW 3198 | 0,9% Sodium Chloride |
| PW 3088 | 0,9% Sodium Chloride |
| PW 3007 | 10% Lactose Broth |
| PW 3099 | 10% Lactose Broth |
| PW 4214 | Acetamide Medium |
| PW 3274 | Arabinose Broth |
| PW 3046 | Arabinose Broth |
| PW 3184 | Arginine acc. to Falkow Broth |
| PW 3048 | Arginine acc. to Falkow Broth |
| PW 3300 | Bile Esculin Azide LAB-AGAR™ |
| PW 3104 | Bile Esculin LAB-AGAR™ |
| PW 3327 | Bile Salts (2% Sodium deoxycholate) |
| PW 3015 | Brain Heart Infusion Broth |
| PW 3094 | Brain Heart Infusion Broth |
| PW 3031 | Brain Heart Infusion Broth |
| PW 4202 | Brain Heart Infusion Broth |
| PW 3155 | Brain Heart Infusion Broth |
| PW 3196 | Brain Heart Infusion Broth |
| PW 3294 | Brucella Broth + vit. K1 |
| PW 3296 | Brucella Broth + vit. K1 |
| PW 3056 | Citrate Christensen LAB-AGAR |
| PW 3059 | Dermatophytes LAB-AGAR™ |
| PW 3298 | Dermatophytes LAB-AGAR™ |
| PW 3167 | Ewing Malonate Broth Modified |
| PW 3020 | Ewing Malonate Broth Modified |
| PW 3250 | Glucose Enrichment Broth |
| PW 3082S | Glucose Enrichment Broth |
| PW 3057 | Kligler Iron LAB-AGAR™ |
| PW 3116 | Kligler Iron LAB-AGAR™ |
| PW 3021 | Ksylose Broth |
| 140/T | Lowenstein-Jensen Medium |
| 152/T | Lowenstein-Jensen Medium + PACT |
| 139/T | Lowenstein-Jensen Medium + P |
| 142/T | Lowenstein-Jensen Medium + SM 4 |
| 143/T | Lowenstein-Jensen Medium + SM 8 |
| L44/T | Lowenstein-Jensen Medium + INH 0,2 |
| 145/T | Lowenstein-Jensen Medium + INH 0,4 |
| .48/T | Lowenstein-Jensen Medium + RMP 40 |

| 149/T | Lowenstein-Jensen Medium + RMP 80 |
|---------|---|
| 146/T | Lowenstein-Jensen Medium + ETB 2 |
| 147/T | Lowenstein-Jensen Medium + ETB 4 |
| PW 3026 | Lysine acc.to Falkow Broth |
| PW 3116 | Lysine acc.to Falkow Broth |
| PW 3023 | Motility LAB-AGAR™ |
| PW 3271 | Motility LAB-AGAR™ |
| PW 3036 | MRVP Medium |
| PW 3051 | Mueller Hinton Broth with Laked Horse Blood |
| PW 3330 | Mueller Hinton II Broth |
| PW 3041 | Mueller Hinton II Broth |
| PW 3028 | Mueller Hinton Broth |
| PW 3065 | Nutrient LAB-AGAR™ |
| PW 3134 | Nutrient S LAB-AGAR™ |
| PW 3135 | Nutrient S LAB-AGAR™ |
| PW 3074 | Peptone Water |
| PW 3119 | Phenyloalanine LAB-AGAR™ |
| PW 3352 | |
| PW 3013 | Phosphate Buffered Saline (PBS) w/o ions Ca2+; Mg2+ Ramnose Broth |
| PW 3107 | |
| PW 3022 | Ringer Solution Sabouraud Dextrose Broth |
| PW 3034 | |
| PW 3064 | Sabouraud Dextrose Broth |
| PW 3270 | Sabouraud Dextrose with Chloramphenicol and Cyklohexymide LAB-AGAR™ |
| PW 3206 | Sabouraud Dextrose with Chloramphenicol and Cyklohexymide LAB-AGAR™ |
| PW 3269 | Sabouraud Dextrose with Chloramphenicol and Gentamicin LAB-AGAR™ |
| PW 3241 | Sabouraud Dextrose with Chloramphenicol LAB-AGAR™ |
| PW 3225 | Sabouraud with Chloramphenicol Broth |
| PW 3017 | Sacharose Broth |
| PW 3268 | Schaedler Broth |
| PW 3014 | Schaedler Broth + vit. K3 |
| PW 3018 | Schaedler Broth + vit. K3 |
| PW 3204 | Schaedler Broth + vit. K3 |
| PW 3058 | Schaedler Modified Broth |
| PW 3038 | Simmons Citrate LAB-AGAR™ |
| PW 3019 | Simmons Citrate LAB-AGAR™ |
| | Simmons Citrate LAB-AGAR™ |
| PW 3095 | Sodium Selenite Broth |
| PW 4209 | Sodium Selenite Broth |
| PW 3040 | Sodium Selenite Broth |
| PW 3008 | Sodium Selenite Broth |
| PW 3030 | Sodium Selenite Broth |
| PW 3045 | Sorbitol Broth |
| 137/T | Stonebrink Medium + P |
| 135/T | Stonebrink Medium + PACT |
| PW 3187 | Thioglycollate Fluid Medium |
| PW 3153 | Thioglycollate Fluid Medium |

| PW 4004 | Thioglycollate Fluid Medium | | | |
|---------|--|--|--|--|
| PW 3033 | Thioglycollate Fluid Medium | | | |
| PW 3027 | Todd Hewitt Broth | | | |
| PW 3203 | Todd Hewitt Broth with Nalidixic Acid and Colistin | | | |
| PW 3050 | Todd Hewitt Broth with Nalidixic Acid and Gentamicin | | | |
| PW 3085 | Trehalose Broth | | | |
| PW 3011 | Trichomonas Broth | | | |
| PW 4204 | Triple Sugar Iron LAB-AGAR™ | | | |
| PW 3193 | Triple Sugar Iron LAB-AGAR™ | | | |
| PW 3060 | Triple Sugar Iron LAB-AGAR™ | | | |
| PW 4019 | Trypticasein Soy Broth | | | |
| PW 4000 | Trypticasein Soy Broth | | | |
| PW 3152 | Trypticasein Soy Broth | | | |
| PW 3156 | Trypticasein Soy Broth | | | |
| PW 3062 | Trypticasein Soy LAB-AGAR™ | | | |
| PW 3226 | Tryptone Water | | | |
| PW 3012 | Tryptophan Culture Broth | | | |
| PW 3098 | Tryptophan Culture Broth | | | |
| PW 3199 | TSB Broth +20% Glicerol | | | |
| PW 3100 | Urea Broth Modified acc. to PN-A- 04023 | | | |
| PW 3304 | Urea Broth Modified acc. to PN-A- 04023 | | | |
| PW 3006 | Urea Christensen Broth | | | |
| PW 3149 | Urea Indol Broth | | | |
| PW 3267 | Urea Indol Broth | | | |
| PW 3183 | Urea LAB-AGAR™ | | | |
| PW 3120 | Urea LAB-AGAR™ | | | |

Czionek Zarządu BioMaxima S.A. Henryk ewczuk Prokurent BioMaxima S.A. Patrycja Paniak Sankowska

BACTERIOLOGICAL LAB-AGAR™

Agar is natural hydrocolloid extracted from several species of red algae, aminlz the Gelidium, Glacilaria and Pterocladia types. Bacteriological LAB-AGAR™ (European) is a gelling agent used in the preparation of culture media and other bacteriological applications. The main advantage of this agar is the absence of inhibitors, which could interfere in microorganisams growth. It has excellent transparency, high hysteresis and very reliable reproducibility. Each bath prodeced is throughly tested for biological performance against a battery of known bacterial cultures in order to ensure proper growth characteristics and absence of inhibitors. Bacteriological LAB-AGAR™ (European) type gives a higher gel strength than American version. It is used in cencentrations from 1,2% to 1,6%.

Chemical characteristics

| Appearacne | White cream powder |
|----------------------------------|----------------------------|
| Moisture | Less than 10 % |
| Ashes | ≤ 4,5 % |
| Gel strength (1,5% Nikan) | 800-1100 g/cm ² |
| pH (1,5%) before autoclaving | 7,0 ± 0,4 |
| pH (1,5%) aftere autoclaving | 6,5 ± 0,4 |
| Melting point (1,5%) | 85 ± 5°C |
| Gelling point (1,5%) | 35 ± 3°C |
| Transparency (1,5%) | ≤ 5 NTU |
| Colorimetry (absorbance) 430 nm | ≤ 0,200 |
| Particle size | 95% Over sieve 60 |

Microbiological test

| Standard plate count | Less than 3000 cfu / g |
|----------------------|------------------------|
| Yeast and molds | Less than 100 cfu / g |
| Coliforms | Less than 3 cfu / g |
| Escherichia coli | Negative |
| Salmonella | Negative |

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration. Store at 2 30°C
- ★ The expiration date is indicated on the label





CHROMOGENIC LISTERIA SET SUPPLEMENT acc. to ISO 11290

Box: 5+ 5 vials / 2 vials / 500 ml

Intended use

The selective/enrichment supplement here described is used for the preparation of Chromogenic Listeria acc. to ISO 11290 LAB-AGAR™ plating medium for the isolation of Listeria spp. from food, environmental or clinical specimens and for the detection of Listeria monocytogenes. For the details of the procedure see the technical sheet of Chromogenic Listeria LAB- AGAR™Base acc. to ISO 11290 (ref. PS 165)

Selective supplement

Vial for 500 ml of medium base

| Nalidixic acid | 10 mg | Amphotericin B | 5 mg |
|----------------|-------|----------------|----------|
| Ceftazidime | 10 mg | Polimixin B | 38350 IU |

Directions

Dissolve the contents of one vial of Selective Supplement with 5 ml of a mixture of sterile distilled water-ethanol (1:1) and add to 500 ml of Chromogenic Listeria acc.t to ISO 11290 Base (PS165) autoclaved and cooled to 50°C, together with the contents of one vial of Enrichment Supplement pre-warmed to 48-50°C. Mix well and distribute in sterile Petri dishes. Aspect of the medium: homogeneously turbid.

Enrichment supplement - ready to use

Vial for 500 ml of medium base

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label







REF. SL 0005

Box: 10 vials / 1 vial / 500 ml

Intended use

The selective supplement for the detection of enumeration of Campylobacter spp. from clinical and food samples.

Vial for 500 ml of medium base

Directions

Dissolve the contents of one vial with 4 ml of sterile distilled water and add to 500 ml of Blood Free Campylobacter CCDA LAB-AGAR™Base (ref. PS 159) autoclaved and cooled to 50°C.Mix well and distribute in sterile Petri dishes. Aspect of the medium: homogeneously turbid.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label







FRASER LISTERIA SELECTIVE SUPPLEMENT

Box: 10 vials / 1 vial / 500 ml

Intended use

Supplement for the detection of Listeria spp. from food and other samples.

Vials for 500 ml of medium base

| Ammonium iron (III) citrate | 250,0 mg |
|-----------------------------|----------|
| Nalidixic acid sodium salt | 10,0 mg |
| Acryflavine HCl | 12,5 mg |

Directions

Aseptically reconstitute 1 vial with 4 ml of 1:1 solution ethanol / sterile distilled water.

Mix gently until complete dissolution. Aseptically add vial to 500 ml Fraser Listeria Enrichment Broth Base (ref. PS 99), autoclaved and cooled to 50 °C. Mix well and distribute into sterile containers.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label







LISTERIA SELECTIVE acc. to Oxford SUPPLEMENT

Box: 10 vials / 1 vial / 500 ml

Intended use

Supplement for the detection of Listeria spp. from food and other samples.

Vials for 500 ml of medium base

| Cycloheximide | 200,00 mg | Colistin sulphate | 10,00 mg |
|---------------|-----------|-------------------|----------|
| Cefotetan | 1,00 mg | Acryflavine HCl | 2,50 mg |
| Fosfomycin | 5 00 mg | | |

Directions

Aseptically reconstitute 1 vial with 4 ml of 1:1 solution ethanol / sterile distilled water (1:1). Mix gently until complete dissolution. Aseptically add vial to 500 ml Listeria acc. to Oxford LAB-AGAR™ Base (ref. PS 104), autoclaved and cooled to 50 °C. Mix well and distribute into sterile Petri dishes.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label







NOVOBIOCIN SUPPLMENT REF. SL 0055

Box: 10 vials

Selective supplement for the isolation of Salmonella spp. and E. coli O157.

Vial for 250 ml of MKKTn Broth Base and 500 ml m-TSB Broth Base

Novobiocin10 mg

Directions

Aseptically reconstitute 1 vial with 5 ml of sterile distilled water. Mix gently until complete dissolution and add aseptically to 250 ml of MKTTn Broth Base (ref. PS 540) or 500 ml of m-TSB Broth Base (ref. PS 197), cooled to 50°C. Mix well and distribute into appropriate containers.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label



CHROMOGENIC SALMONELLA SUPPLEMENT

Box: 5+ 5 vials / 2 vials / 500 ml

Intended use

The selective/enrichment supplement here described is used for the preparation of Chromogenic Salmonella LAB-AGAR™(ref. PS 598).

Enrichment supplement - ready to use- vial A

| Vial for 500 ml of medium base |
|--|
| Liquid reagent 2 ml |
| Reagent add to 500 ml of the medium (Salmonella Chromogenic LAB-AGAR™-ref. PS 589) before autoclaving) |
| Selective supplement (vial B) Vial for 500 ml of medium base |
| Cefsulodine |

Directions

Dissolve the contents of one vial B with 2 ml of of sterile distilled water and add to 500 ml of Salmonella Chromogenic LAB-AGAR™ (ref. PS598) autoclaved and cooled to 50°C. Mix well and distribute in sterile Petri dishes.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label









HALF FRASER LISTERIA SELECTIVE SUPPLEMENT

Box: 10 vials / 1 vial / 225 ml

Intended use

Supplement for the detection of Listeria spp. from food and other samples.

Vials for 225 ml of medium base

| Ferric ammonium citrate | 112,50 mg |
|-------------------------|-----------|
| Nalidixic acid | 2,25 mg |
| Acryflavine HCl | 2,8125 mg |

Directions

Aseptically reconstitute 1 vial with 2 ml of 1:1 solution ethanol / sterile distilled water.

Mix gently until complete dissolution. Aseptically add vial to 225 ml Fraser Listeria Enrichment Broth Base (ref. PS 99), autoclaved and cooled to 50 °C. Mix well and distribute into sterile containers.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label







UREA STERILE SOLUTION 40 %

REF. SL 0111

Box: 10 vials. 1 vial /100 ml

Description

A sterile solution of 40% urea, for addition to Urea LAB-AGAR™ Base acc. to ISO 6579 (ref. PS 211) for the detection of urease production by Proteus spp.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory
- ★ Consult the material safety data sheet before the use
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label



BISMUTH SULFITE LAB-AGAR™

REF. PS 01

Highly selective, for isolation of Salmonella spp. particularly Salmonella Typhi from clinical specimens and food.

Formula in g/L

| Enzymatic digest of animal tissues 10,00 | Ferric sulphate (FeSO4)0,30 |
|---|-----------------------------|
| Beef extract5,00 | Bismuth sulfite (indicator) |
| Glucose5,00 | Brilliant green0,025 |
| Disodium hydrogen phosphate (Na ₂ HPO ₄) anh. 4,00 | Agar 20,00 |

Final pH at 25°C: 7,7 ± 0,2

Principle:

Bismuth Sulfite LAB-AGAR™ is a modification of the Wilson Blair Medium, and generally accepted as routine for the detection of most Salmonella and in particular Salmonella Typhi.

Bismuth sulfite indicator and brilliant green are inhibitors of Gram- positive bacteria and members of the coliform group. In the presence of H_2S Salmonella spp. reduced iron salts to iron sulfate, which produce a black colony, and turns the bismuth indicator to metallic bismuth surrounding the area of the colonies with a bright sheen.

Preparation: suspend 52,3 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute. DO NOT AUTOCLAVE. Cool to 45°C(very important), mix well and pour into Petri dishes. The prepared medium should be stored at 8-15°C. The colou of the prepared medium opaque white with a green tint.

The dehydrated medium should be homogenous, free-flowing and light green in color. If there are any pohysical changes, discard the medium.

Procedure:

- ★ Inoculated by streaking
- ★ Incubated at 37±1°C for 48 hours

Colony colour:

- ★ Salmonella spp bright metallic black
- ★ Escherichia coli brown green

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of $37\pm1^{\circ}$ C and observed after 40-48 hours

MicroorganismsGrowthColony colorSalmonella Enteritidis ATCC 13073GoodBlack with bright metallicSalmonella Typhi ATCC 19430GoodBlack with bright metallicEscherichia coli ATCC 25922Partial inhibitionBrown-greenShigella flexneri ATCC 12022Partial inhibitionBrown

Enrterococcus faecalis ATCC 29212 Null







BRAIN HEART INFUSION BROTH

REF. PS 04

For the growth of pathogenic cocci and other microorganisms.

Formula in g/L

| Calf brain infusion | 12,50 | Brain heart infusion | 5,00 |
|------------------------------------|-------|--|----------------------------|
| Enzymatic digest of animal tissues | 10,00 | Disodium hydrogen phosphate (Na ₂ | HPO ₄) anh2,50 |
| Sodium chloride | 5,00 | Glucose | 2,00 |

Final pH at 25°C: 7.4 ± 0.2

Principle:

Brain Herat Infusion Broth (BHIB) is a liquid medium rich in nutrients, suitable for the cultivation of several fastidious strains of bacteria, such as Streptococci, Meningococci and Pneumococci, fungi and yeasts. BHIB is recommended in Standard Methods for water testing and in antimicrobial susceptibility tests.

The nutritionally rich base of beef heart and calf brain infusion and peptone supports growth of a variety of microorganisms. Glucose is the carbon and energy source. Sodium chloride maintains the osmotic balance.

This medium is very versatile and supports the growth of many organisms

Preparation: suspend 37 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until, complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121°C for 15 minutes.

The prepared medium should be stored at 2-8°C.

Procedure:

- ★ Inoculate the medium with the sample
- ★ Incubate at 37±1°C for 24-48-72 hours

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubation at a temperature of 37±1°C and observed after 18-24 hours

| Microorganisms | Growth |
|------------------------------------|--------|
| Streptococcus pneumoniae ATCC 6303 | Good |
| Streptococcus pyogenes ATCC 19615 | Good |
| Brucella abortus ATCC 4315 | Good |
| Staphylococcus aureus ATCC 25923 | Good |







TRIPLE SUGAR IRON LAB-AGAR™

For differentiation of Enterobacterales according to ISO 6579 – 1 standard.

Formula in g/L Peptone 20,00 Glucose 1,00 Sodium thiosulphate 0,30 Sodium chloride 5,00 Lactose 10,00 Iron (III) citrate 0,30 Phenol red 0,024 Agar 12,00 Sucrose 10,00 Beef extract 3,00 Yeast extract 3,00

Final pH at 25°C: 7,4 ± 0,2

Principle:

Triple Sugar Iron LAB-AGAR $^{\text{M}}$ (T.S.I.) may be used to differential enteric Gram – negative Enterobacterales on the basis of carbohydrate fermentation and H_2S production. It used as an aid in the identification of photogenic and saprophytic Enterobacterales isolated from routine bacteriological analysis food samples. This medium is used as a key to initiate the identification of Enterobacterales in some FDA schemas.

The peptone and beef extract provide the nutrients for growth. T.S.I. contains three carbohydrates (glucose, lactose and sucrose), sources of carbon and energy. When these are fermented the acid production is indicated by the phenol red indicator, being the color changes yellow for acid production and red for alkalization. Sodium thiosulphate is reduced to hydrogen sulfide, which reacts with iron salt to give the black iron sulfide. The ferric citrate is a H₂S indicator. Sodium chloride maintains the osmotic balance of the medium.

The mode of action is similar to Kligler Iron LAB-AGAR™ (ref. PW 3016) which contains two sugars. The addition of 1% sucrose in T.S.I. agar allows to differentiate between Proteus and Salmonella. The fermentation of the sucrose by Proteus turns the color of the phenol red indicator in the slant from red to yellow. Glucose – positive, lactose – negative members of the genus Salmonella all cause a reddening of the slant and acidity the depths of the agar tubes.

Preparation: suspend 64,6 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Dispense into tubes for 10 ml of the medium and sterilize in autoclave at 121°C for 15 minutes. Allow to cool in a slanted position in order to obtain butts of 1,5-2,5 cm. depth.

The prepared medium should be stored at 2-8°C.

Procedure:

- ★ With an inoculating needle, prick the center of well isolated colonies obtained from a solid media
- \star Stab the center of the medium into the deep of the 3-5 mm the bottom
- ★ Withdraw the needle and streak the subculture the surface of the slant
- ★ Loosen closure on the tube before incubating
- ★ Incubate at 36±2°C for 24±3 hours
- ★ Read tubes for acid production of slant / butt, gas and H₂S reaction

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.









Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 36±2°C and observed after 24±3 hours

| Microorganisms | Growth | Slant | Depth | Gas | H ₂ S |
|-----------------------------------|--------|--------|--------|-----|------------------|
| Escherichia coli ATCC 25922 | Good | Yellow | Yellow | + | - |
| Proteus vulgaris ATCC 13315 | Good | Yellow | Yellow | + | + |
| Salmonella Enteritidis ATCC 13078 | Good | Red | Yellow | + | + |
| Shigella flexneri ATCC 12022 | Good | Red | Yellow | - | - |
| Pseudomonas aeruginosa ATCC 9027 | Good | Red | Red | - | - |



XLD LAB-AGAR™ REF. PS 45

Selective and differential medium for the detection of Salmonella spp. from foods acc. to ISO 6579-1 standard.

Formula in g/L

| Xylose | 3,75 | L-Lysine hydrochloride | 5,00 |
|----------------------|-------|-----------------------------|------|
| Lactose | 7,50 | Sucrose | 7,50 |
| Phenol red | 0,08 | Yeast extract | 3,00 |
| Sodium desoxycholate | 1,00 | Ammonium iron (III) citrate | 0,80 |
| Agar | 13,50 | Sodium chloride | 5,00 |
| Sodium thiosulphate | 6,80 | | |

Final pH at temp. 25° C: $7,4 \pm 0,2$

Pricniple:

The ISO 6579 recommends the XLD LAB-AGAR™ formula for the identification and presumptive identification of Salmonella after pre-enrichment in a non-selective fluid medium and enrichment in a selective fluid medium. The medium was developed principally for isolating and differentiating Gram — negative enteric bacilli, particularly Shigella and Providentia. It has been shown to be more effective than other enteric differential media.

The reactions are degradation of the three fermentable carbohydrates: xylose, lactose and sucrose, with the production of acid, manifested in the color change from red to yellow. Sodium thiosulphate serves as a reactive substance with ferric ammonium citrate as an indicator the formation of H_2S under alkaline conditions. Phenol red is the pH indicator. Yeast extract provides the carbon, protein and nutrient source required for the growth of bacteria. The bacteria that decarboxylase the L-Lysine HCl to cadaverine are identified by the presence of purple – red color around the colonies due to the elevation of the pH.

Preparation: suspend 54 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution of agar. DO NOT AUTOCLAVE. Cool to 45-50°C and pour into Petri dishes.

The prepared medium should be stored at 8-15°C

Procedure:

- ★ The specimen is seeded by streaking directly on the surface of the medium, or is first enriched in Rappaport Vassiliadis Soy Broth (ref. PW 4245) and MKTTn Broth (ref. PW 6092)
- ★ Incubate at 37°C ± 1°C for 24 h ± 3 h

Colony color:

Salmonella spp. clear red (black center)

Proteus spp. yellow, transparent (black center)

E. coli yellow

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.







Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 37±1°C and observed after 24±3 hours

Microorganisms
Escherichia coli ATCC 25922
Salmonella Typhimurium ATCC 14028
Shigella flexneri ATCC 12022
Staphylococcus aureus ATCC 25923
Packaging: 500 g

Growth Colony color
Moderate Yellow (precipitate)
Good Clear red (black center)
Good Red
Inhibited







BUFFERED PEPTONE WATER

REF. PS 52

Recommended as a diluent for the homogenization of samples in microbiological analysis of food.

Formula in g/L

| Enzymatic digest of casein 10,0 |) | Disodium hydrogen phosphate anh | 3,57* |
|------------------------------------|---|---------------------------------|-------|
| Potassium dihydrogen phosphate 1,5 | 0 | Sodium chloride | 5,00 |

^{*}Equivalent 9,0 g Disodium hydrogen phosphate dodecahydrate

Final pH at 25°C: 7,0 ± 0,2

Princple:

Buffered Peptone Water is recommended as a pre-enrichment in food samples containing suspected contaminants such as Salmonella. A feature common to all selective media that sublethally injured organisms are not generally detected and therefore a recovery step must be included in examination procedures. This is of importance, particularly in the food industry as various processes such as heat, desiccation, preservation processesses, pH changes, ect. Cause sublethal injuries to Salmonella.

The broth is a rich in nutrients and provides high resuscitation rates for sublethally injured bacteria and intense growth. Changes in pH may cause damagages to bacteria growth. Buffered Ppetone Water maintains a high pH over the enrichment period vis the phosphate buffer system and allows repair of injured cells sensitive to low pH.

Preparation: suspend 20,07 grams of the medium in one liter of distilled water. Heat with frequent agitation to completely dissolve the medium if necessary. Sterilize in autolcave at 121°C for 15 minutes. The prepared medium should be stored at 2-8°C.

Procedure:

- ★ Detection of Salmonella spp. according to standard method: Refer to standards ISO 6579-1,
- ★ Detection of Cronobacter spp. according to standard method: Refer to standards ISO 22964

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of $37\pm1^{\circ}$ C and observed 18 hours

| Growth |
|--------|
| Good |
| Good |
| Good |
| Good |
| |



LYSINE DECARBOXYLASE BROTH acc. to ISO 6579

REF. PS 57

For the biochemical confirmation of Salmonella.

| Formula in g/L | | | | |
|----------------|----------------------------|--|--|--|
| Glucose | L-Lysine hydrochloride5,00 | | | |
| Yeast extract | Bromocresol purple 0,015 | | | |

Final pH at 25°C: 6,8 ± 0,2

Principle:

Lysine Decarboxylase Broth is recommende by ISO 6579 for the biochemical confirmation of Salmonella based on lysine decarboxylation. It is also recommended by ISO 10273 for the biochemical confirmation of Yersinia. When the medium is inoculated with a bacterium that is able to ferment glucose, the acid produced lowers the pH of the medium and changes the color of the indicator from purple to yellow. The acidic condition also stimulates decarboxylase activity. The bacteria that decarboxylate the L-Lysine to cadaverine are identified by presence of a purple-red color. The production of these amines elevates the pH of the medium. A yellow color after 24 hours indicates a negative result.

Yeast extract is the source of vitamins, of particularly the B-group essential for growth. Glucoe is the fermenteable carbohydrate. Bromocresol purple is th epH indicator. Lysine is added to detect the production of the specific enzyme.

Preparation: suspend 9 of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation until completely dissolution. Dispense into test tubes and sterilize in autoclave at 121°C for 15 minutes.

The preapred medium should be stored at 2-8°C.

Procedure:

- ★ The tubes are inoculated with the microorganism samples
- ★ Incubated at 36±2°C for 24±3 hours

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 36 ± 2 °C and observed after 24 ± 3 hours

Microorganisms

Salmonella Typhi ATCC 6539

Salmonella Paratyphi ATCC 9150

Proteus vulgaris ATCC 13315

Salmonella Galinarum NCTC 9240

*Yersina enterocolitica ATCC 27729

Lysine decarboxylase

+

*Yerina decarboxylase

Lysine decarboxylase

^{*}Incubate at 30°C during 24 hours according to ISO 10273



MRS LAB-AGAR™ REF. PS 59

Selective medium for isolation of lactobacilli from foods, clinical and others samples.

Formula in g /L

| Enzymatic digest of casein | 10,00 | Beef extract | 10,00 |
|----------------------------|-------|--------------------------------|-------|
| Yeast extract | 4,00 | Glucose | 20,00 |
| Tween-80 | 1,00 | Dipotassium hydrogen phosphate | 2,00 |
| Sodium acetate | 5,00 | Tri-Ammonium citrate | 2,00 |
| Magnesium sulphate | 0,20 | Manganese sulphate | 0,05 |
| Agar | 12,00 | | |

Final pH at 25°C: 6,2 ± 0,2

Principle:

MRS LAB-AGAR™ is used for the cultivation of lactobacilli. The addition of magnesium, manganese and acetate with the Tween 80, has provided an improved medium for growth of lactobacilli, including that of very fastidious species such as Lactobacillus brevis and L. fermenti.

On the other hand, the quality enzymatic digest of casein in addition to the beef extract and yeast extract, combine together all the necessary growth factors that make the MRS LAB-AGAR™ one the best medium for the cultivation of lactobacilli.

Nevertheless, this medium selectivity is low and contaminants tend to grow in these medium, which signifies a higher selectivity is needed.

Preparation: suspend 64,2 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation to completely dissolution. Sterilize in autoclave at 121°C for 12 minutes. Cool to 45-50 °C, mix well and pour into Petri dishes.

The prepared medium should be stored at 8-15°C.

Procedure

Food samples

- ★ Plates are inoculated by spreading of 0,1 mL of the sample of its dilutions
- ★ Incubate at 30°C for 5 days in 5% CO₂ atmoshere

Clinical samples

- ★ Inoculate the plates directly from the samples
- ★ Incubate at 37°C for 72 hours in 5% CO₂ atmoshere

- ★ once opened keep powdered medium closed to avoid hydration at 2 25°C
- ★ the expiration date is indicated on the label.









MRS LAB-AGAR™ REF. PS 59

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 37° C during 3 days, or at 30° C during 5 days, in a CO_2 enriched atmosphere.

Microorganisms Lactobacillus acidophilus ATCC 4356 Lactobacillus casei ATCC 393 Escherichia coli ATCC 25922 Pseudomonas aeruginosa ATCC 27853

Packaging: 500 g

Growth Good Good

Moderate - Good

Inhibited



RAPPAPORT SOY BROTH REF. PS 65

For selective enrichment medium of Salmonella with the exception of S. Typhi and S. Paratyphi from foodstuffs and other materials. According to ISO 6579-1 standard.

Formula in g/L

| Enzymatic digest of soya 4,50 | Sodium chloride | . 7,20 |
|--|--|--------|
| Potassium dihydrogen phosphate (KH ₂ PO ₄) 1,26 | Dipotassium hydrogen phosphate(K ₂ HPO ₄) | . 0,18 |
| Magnesium chloride anh |)* Malachite green oxalate | .0,036 |

^{*}equivalent to 28,6 g * 6H₂O

Final pH at 25°C: 5,2 ± 0,2

Principle:

Rappaport Soy Broth is recommended for the selective isolation of Salmonella from food or environmental samples.

The Rappaport mdium was modified by Vassiliadis by reducing malachite green concentration and increasing incubation temperature offering a better stability of the pH of prepared medium and the optimozation of the concentration of magnesium chloride, resulting in an improved recovery of Salmonella.

The soy peptone provides essential nutrients for growth: nitrogen, vitamins, minerals and amino acids. Posassium phosphates balance the low pH of the medium, combined with the presence of magnesium chloride to raise the osmotic pressure and malachite green toinhibits other bacteria.

Peparation: suspend 26,54 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Dispense into tubes and sterilize in autoclave at 115°C for 15 minutes.

The prepared medium should be stored at 2-8°C

Procedure:

- ★ Transfer 0,1 ml pre-enrichment broth Buffered Peptone Water incubated at 37±1°C for 20 hours to 10 ml of Rappaport Vassiliadis Soy Broth
- ★ Icnubate at 41,5±0,5° for 24 hours
- **★** Subculture to selective agar media: XLD LAB-AGAR™ (ref. PP 1330), Salmonella Shigella LAB-AGAR™ (ref. PP 1250) BGA LAB-AGAR™ (ref. PP 1360), Hektoen LAB-AGAR™ (ref. PP1060); Chromogenic Salmonella LAB-AGARt (ref. PP 0115) or other solid media.

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 41,5°C and observed after 24±3 hours

Microorganisms Growth
Escherichia coli ATCC 25922 Inhibited
Salmonella Typhimurium ATCC 14028 Good

Packaging: 500 g



FRASER LISTERIA ENRICHMENT BROTH BASE

REF. PS 99

Enrichment medium for the detection of Listeria in food and environmental sapmles acc. to ISO 11290-1 standard.

| | Formul | la in g/L | |
|------------------------------------|--------|---|------|
| Sodium chloride | 20,00 | Disodium hydrogen phosphate 2 -hydrate 12 | 2,00 |
| Enzymatic digest of animal tissues | 5,00 | Tryptone (enzymatic digest of casein) | 5,00 |
| Yeast extract | 5,00 | Beef extract | 5,00 |
| Lithium chloride | 3,00 | Dipotassium hydrogen phosphate | 1,35 |
| Esculin | 1,00 | | |

Final pH at 25°C: 7.2 ± 0.2

Principle:

Fraser Listeria Enrichment Broth Base is an appropriate medium fro the selective enrichment of Listeria in the two-step method according to ISO 11290-1, for the preparation of Fraser or Half Fraser Broth by adding the respective supplements.

It is recommended for the detection of Listeria spp. in food products and in sample from the environment. All Listeria species hydrolyse the esculin to esculentin, which reacts with iron ions producing blacking of the medium. Another advantage of this medium is that the addition of ferric ammonium citrate improves the growht of Listeria monocytogenes. Lithium chloride included in the medium, along with nalidixic acid and acryflavine from the suppmelent, inhibit the growth of the accompanying flora, which can hydrolyze the esculin. The high amount of sodium chloride inhibits the growth of enterococci. Tryptone, proteose peptone nad beef extract provide nitrogen, vitamisn, minerals and amino acids essential fro growth. Yeast extract is the source of vitamins, particularly of the B-group. Phosphate salts act is a buffer.

Preparation: suspend 28,68 grams of the emdium in 500 ml of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Sterizilze in autoclave at 121°C for 15 minutes. Cool to 45-50°C and add one vial of Half Fraser Listeria Selective Supplement (ref. SL 0014) or Fraser Listeria Selective Supplement (ref. SL 0012). Mix well.

The prepared medium should be stored at 2-8°C.

Procedure:

Detection of Listeria monocytogenes according to stadnard method: Refer to standards ISO 11290-1

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of $30\pm1^{\circ}$ C and observed 24 ± 3 hours – Half Fraser Borth and after incubaction at a temperature of $37\pm1^{\circ}$ C and observed 28 ± 3 hours –Fraser Borth

MicroorganismsGrowthEsculin reationsListeria monocytogenes ATCC 19111Good+Enterococcus faecalis ATCC 29212Null-

Packaging: 500 g

Supplements: Half Fraser Listeria Selective Supplement 10 vials. 1 vial / 500 ml (ref.SL 0014)

Fraser Listeria Selective Supplement 10 vials. 1 vial / 500 ml (ref. SL 0012)





LISTERIA acc. to OXFORD LAB-AGAR™ BASE

Selective medium for the detection of Listeria monocytogenes from food and other samples. According to **ISO 11290-1** standard.

| Formula in g/L | | | |
|------------------|-------|-----------------------------|------|
| Proteose peptone | 23,00 | Ammonium iron (III) citrate | 0,50 |
| Starch | 1,00 | Sodium chloride | 5,00 |
| Agar | 10,00 | Esculin | 1,00 |
| Lithium chloride | 15,00 | | |

Final pH at 25°C: 7.0 ± 0.2

Principle:

Listeria acc. to Oxford LAB-AGAR™ is a selective medium for Listeria according to Oxford formula and is recommended for the detection of L. monocytogenes from clinical samples and foods products. It is used directly or for confirmation after using Fraser Broth (cat. no. PW4024).

All Listeria spp. hydrolyze the esculin to esculentin that reacts with the iron ions producing black colonies and a blackening of the medium. Atother advantage of this medium Columbia LAB-AGAR™ Base (cat. no. PS06) provides a rich nutrient base for growth and the addition of ferric ammonium citrate improves the growth of L. monocytogenes. Lithium chloride is an inhibiting agent, together with the other antibiotics, which inhibit the growth of Gram-negative bacteria nad a large part of Gram-positive ones. Cycloheximide inhibits yeasts.

Preparation: suspend 27,75 grams of the medium in 500 ml of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C and add one vial of Listeria Selective acc. to Oxford Supplement (ref. SL 0021). Mix well and pour into Petri dishes.

The prepared medium should be stored at 8-15°C.

Procedure

Detection of Listeria monocytogense according to stadnard method: Refer to standards ISO 11290-1

Result:

★ Listeria monocytogenes brown-grey colonies with black centerand black halo

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 37°C and observed 24-48 hours

| Microorganisms | Growth | Colony color |
|-----------------------------------|-----------|----------------------------|
| Listeria monocytogenes ATCC 13932 | Good | Brown-grey colonies with |
| | | black centerand black halo |
| Staphylococcus aureus ATCC 25923 | Inhibited | White colonies |
| Enterococcus faecalis ATCC 29212 | Null | - |
| Escherichia coli ATCC 25923 | Null | - |
| Packaging: 500 g | | |

Suplement: Listeria Selective acc. to Oxford Supplement 10 vials. 1 vial / 500 ml (ref. SL 0021) Listeria Modified acc. to Oxford Supplement 10 vials. 1 vial / 500 ml (ref. SL 0081)







BLOOD FREE CAMPYLOBACTER CCDA LAB-AGAR ™ BASE

Selective medium for the isolation of Campylobacter spp. from food and other samples. According to ISO 10272.

Formula in g/L

| Meat extract | 10,00 | Enzymatic digest of animal tissues | 10,00 |
|----------------------------|-------|------------------------------------|-------|
| Enzymatic digest of casein | 3,00 | Sodium desoxycholate | 1,00 |
| Agar | 15,00 | Ferrous sulphate (II) | 0,25 |
| Sodium pyruvate | 0,25 | Sodium chloride | 5,00 |
| Charcoal | | | |

Final pH at 25°C: $7,4 \pm 0,2$

Principle:

Blood Free Campylobacter CCDA LAB-AGAR™ is a modified formula described by Bolton et. al., replacing blood with charcoal, sodium pyruvate and ferrous sulphate. This medium supports growth of most enteric Campylobacter. It is recommended for the selective isolation of C. jejuni, C. coli, C. lari and thermophilic Campylobacter from food, clinical and non clinical specimens.

This medium contains ferrous sulphate, sodium pyruvate and charcoal to promote the growth of Campylobacter spp., as they quench the toxic forms of oxygen (hydrogen pyroxide) increasing the aero tolerance and enabling the oxygen sensitive strains to be readily isolated. Sodium desoxycholate and cephoperazone partially or completely inhibits Gram- positive and most Gram — negative bacteria. Amphotericin B inhibits yeasts.

Preparation: suspend 24,3 grams of the medium in 500 ml of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C and add one vial of Campylobacter CCDA Selective Supplement (ref. SL 0005). Mix well and pour into Petri dishes.

The prepared meiudm should be stored at 8-15°C.

Procedure:

★ Inoculate and incubate at 42°C for 24-48 hours

Results

- ★ C. jejuni produces grey, moist, flat, spreading colonies
- ★ C. coli colonies are creamy-grey, moist, slightly raised and tend to be discrete

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.









BLOOD FREE CAMPYLOBACTER CCDA LAB-AGAR ™ BASE

REF. PS 159

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction in anaerobic conditions at a temperature of 42 \pm 1°C and observed after 24-48 hours

MicroorganismsGrowthEscherichia coli ATCC 25922InhibitedCampylobacter jejuni ATCC 29428GoodCampylobacter coli ATCC 33559Good

Packaging: 500 g

Supelment: Campylobacter CCDA Selective Supplement 10 vials. 1 vial / 500 ml, Ref. SL 0005





CHROMOGENIC LISTERIA LAB-AGAR™ BASE acc. to ISO 11290

Selective medium for the detection and enumeration of Listeria monocytogenes. Medium is recommendations of the norm: ISO 11290-1 and ISO 11290-2.

Formula in g/L

| Enzymatic of digest of animal tissues 18,00 | Enzymatic digest of casein |
|--|--------------------------------------|
| Yeast extract10,00 | Sodium pyruvate2,00 |
| Glucose2,00 | Lithium chloride10,00 |
| Magnesium glycerophopshate | Magnesium sulphate anh |
| Sodium chloride 5,00 | Disodium hydrogen phosphate anh 2,50 |
| 5-bromo-4- chloro-3-indolilo-ß-D-glucopiranoside0,05 | Agar 13,50 |

Final pH at 25°C: 7,2 ± 0,2

Principle:

Chromogenic medium for the presumptive isolation, enumeration and identification of Listeria monocytogenes and Listeria spp. in food and clinical samples. It is used for confirmation after using Listeria Enrichment Broth Base (PS 99). The differential activity of the medium is due to two factors. On one hand, the presence of the chromogenic component X-glucoside, a substrate for the detection of the enzyme \(\mathbb{G}\)-glucosidase, common to all Listeria species giving the colonies their blue colour. Other organisms that possess this enzyme, for example enterococci, are inhibited by the selective agents within the medium and by the selective supplement. On the other hand, the differential activity is also obtained by the lipase substrate, upon which the specific enzyme for L. monocytogenes acts. The lipase is responsible for the opaque white halo which surrounds L. monocytogenes. The combination of both substrates allows us to differentiate the colonies of Listeria monocytogenes from the rest of Listeria spp. since, although all are blue in colour, L. monocytogenes present an opaque white halo surrounding them. It has been observed that some strains of Listeria ivanovii, mostly pathogenic to animals although some of which have caused infections in humans, also lipase activity.

Preparation: suspend 35,25 grams of the medium in 500 ml of distilled water. Mix well and dissolve by heating with frequent agitation until comlpete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C add 2 vials of Chromogenic Listeria Set Supplement acc. to ISO 11290 (ref. SL 0001). Mix well and pour into sterile Petri dishes .

Procedure:

Detection and enumeration of Listeria monocytogenes according to stadnard method: Refer to standards ISO 11290-1 and ISO 11290-2

Morphology colony:

Listeria species: blue to blue –green colonies, round reagular, without any opaque halo, diameter from 1 to 2 mm:

Listeria monocytogenes: colonies with Listeria spp. Characteristics and surrounded by an opaque halo. L. monocytogenes strains grow sa typical colonies in 24 hours

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 25°C
- ★ The expiration date is indicated on the label.

Packaging: 500 g

Supplement: Chromogenic Listeria Set Supplement acc. to ISO 11290 2 x 5 vials 1+1/500 ml Ref. SL 0001







TSYEA LAB-AGAR™ acc. to ISO 11290

REF. PS 193

For the confirmation of Listeria spp.

Formula in g/L

| Tryptone | 17,00 | Soy peptone | 3,00 |
|--------------------------------|-------|-------------|-------|
| Yeast extract | 6,00 | Glucose | 2,50 |
| Sodium chloride | 5,00 | Agar | 15,00 |
| Dipotassium hydrogen phosphate | 2.50 | | |

Final pH at 25°C: $7,3 \pm 0,2$

Principle:

TSYEA LAB-AGAR™ acc. to ISO 11290 is a general purpose medium which supports the growth of a wide variety of microorganisms.

The formula conforms to ISO 11290-1 and is used for the confirmation of Listeria monocytogenes colonies and to subculture suspected Listeria colonies.

Tryptone, Yeast extract and Soy peptone provide nitrogen, vitamins, minerals and amino acids essential for growth. Glucose is the fermentable carbohydrate providing carbon and energy. Dipotassium phosphate acts as a buffer system.

This medium is used to select colonies for the confirmation of Listeria spp. After incubation in Listeria acc. to Oxford LAB-AGAR™ (Ref. PS 104); Listeria acc. to Palcam LAB-AGAR™ (Ref. PS 102) or Chromogenic Listeria acc. LAB-AGAR™ acc. to ISO 11290 (Ref. PS 165), take 5 suspected Listeria spp colonies, and inoculate them in TSYEA LAB-AGAR.

Preparation: suspend 51 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C, mix well and dispense into Petri dishes.

The prepared medium should be stored at 8-15°C.

Procedure:

★ Inoculate and incubate at 37±1°C during 18-24 hours or until growth is satisfactory.

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubation at a temperature of 37±1° C and observed after 18-24 hours.

Microorganisms Growth
Listeria monocytogenes ATCC 19111 Good
Listeria innocua ATCC 33090 Good

Packaging: 500 g





BLOOD No2 LAB-AGAR™ BASE

REF. PS 205

For cultivation of detection of hemolytic activity of fastidious microorganisms, confirmation of Bacillus cereus (ISO 7932) and Listeria monocytogenes (ISO 11290).

| Formula in g/L | | | |
|------------------------------------|-------|---------------|-------|
| Enzymatic digest of animal tissues | 15,00 | Yeast extract | 5,00 |
| Sodium chloride | 5,00 | Agar | 15,00 |
| Liver digest | 2,50 | | |

Final pH at 25°C: 7.2 ± 0.2

Principle

Blood No 2 LAB-AGAR™ is a medium rich in nutritional properties. It is used for the isolation, cultivation and recovery of fastidious microorganisms and study of hemolysis activity. Liver extract and yeast extract provide nitrogen, vitamins, minerals and amino acids essential for growth. Sodium chloride maintains osmotic equilibrium. The blood provides growth factor for the microorganisms and is the basis for determining hemolytic reactions.

This medium has been recommended by ISO normative 7932 for the confirmation of Bacillus cereus. Incubate at 30°C for 24±2 hours and interpret the hemolysis reaction. The Bacillus cereus ha positive reaction of ß-hemolysis. The width the hemolysis zone may vary.

T is also medium recommended by ISO normative 11290 for the confirmation of Listeria monocytogenes. The normative recommends incubation at 35°C or 37°C for 18-24 hours. A zone of ß-hemolysis is considered a positive reaction

Preparation: suspend 42,5 grams of the medium in 950 ml of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C and aseptically add 50 ml defibrinated sheep blood. Mix well and pour into Petri dishes.

Procedure:

- ★ Inoculate sample onto the surface of the medium, streak for isolation with an inoculating loop
- ★ Incubate: areobic, with 5-10 % CO₂ atmosphere; at 37±1°C for 18 24 hours

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.

Microbiological test

The following results were obtained adding 5% sheep defibrinated blood in the performance of the medium from type culrure after incubation at a temperature 37±1°C and observed 24-48 hours

| Microorganisms | Growth | Hemolysis |
|--------------------------------------|--------|-----------|
| Streptococcus pneumoniae ATCC 6303 | Good | Alpha |
| Streptococcus pyogenes ATCC 19615 | Good | Beta |
| *Bacillus cereus ATCC 11778 | Good | Beta |
| ** Listeria monocytogenes ATCC 19111 | Good | Beta |

^{*} Incubate at 30°C for 24±2 hours according to ISO 7932

Packaging: 500 g





^{**} Incubate at 35 or 37°C for 18-24 hours according to ISO 11290



UREA LAB-AGAR™ BASE acc. to ISO 6579

For the differentiation of Enterobacterales, particularly Proteus spp. from Salmonella.

Formula in g/L

| Peptone | . 1,00 | Potassium dihydrogen phosphate | 2,00 |
|------------|---------|--------------------------------|-------|
| Glucose | . 1,00 | Sodium chloride | 5,00 |
| Phenol red | . 0,012 | Agar | 12,00 |

Final pH at 25°C: 6,8 ± 0,2

Principle:

Urea LAB-AGAR™Base acc. to ISO 6579 can be used for the differentiation of the urea activity of Enterobacterales as well as microogranisms of the families of Brucella, Bacillus, Micrococcus, Mycobacteria and Proteus.

Urea is a source of nitrogen for those organisms producing urease. Peptone provides nitrogen and other cofactor essential for growth. Monopotassium phosphate provides buffering capacity. Glucose is a fermentable carbohydrate carbon and enegry source. Sodium chloride maintaince the osomtic balance. Phenol red is the pH indicator.

Preparation: dissolve 2,1 grmas of the medium in 95 ml of distilled water. Mix well and dissolve by heating with frequent agitation until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 50-55°C and add aseptically 5 ml of Urea Sterile Solution 40 % (ref. SL 0041) or one vial of Urea Sterile Solution 40 % (ref. SL 00111). Mix well and dispense tubes. Allow to cool in a slanted position in order to obtain short butts. The prepared medium should be stored at 2-8°C.

Procedure:

- **★** Prepare a haevy suspension of the organism isolated from plated media and icoculate the Urea LAB-AGAR™ Base acc. to ISO 6579.
- ★ Loosen closure on the tube before incubating.
- ★ Incubate at 37°C ± 1°C or 36±2°C to 24 hours.

Result:

★ Positive urease tubes turn the phenol red indicator a deep violet red colour (alkalization)

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.







UREA LAB-AGAR™ BASE acc. to ISO 6579

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of 37±1°C; 36±2°C and observed after 24 hours

| Microorganisms | Growth | Urease |
|-----------------------------------|--------|--------------------------------------|
| Enterobacter aerogenes ATCC 13048 | Good | - / no change of color in the medium |
| Escherichia coli ATCC 25922 | Good | - / no change of color in the medium |
| Klebsiella pneumoniae ATCC 13883 | Good | + / red or purple medium |
| Proteus vulgaris ATCC 13315 | Good | + / red or purple medium |
| Salmonella Typhimurium ATCC 14028 | Good | - / no change of color in the medium |

Packaging: 500 g

Supplements: Urea Sterile Solution 40 % 100 ml ref. SL 0041

Urea Sterile Solution 40 % 10 x 5 ml ref. SL 0111



MKTTn BROTH BASE REF. PS 540

For the selective enrichment of Salmonella. Accordin to ISO 6579-1 standard.

Formula in g/L

| Beef extract | 4,30 | Enzymatic digest of casein | 8,60 |
|-------------------|--------|----------------------------|-------|
| Ox bile | 4,78 | Sodium chloride | 2,60 |
| Calcium carbonate | 38,70 | Sodium thiosulphate | 47,80 |
| Brilliant green | 0.0096 | | |

Final pH at 25°C: 8.0 ± 0.2

Principle:

MKTTn Broth is recommended by ISO 6579 to be norm to be used as a selective enrichment broth for the detection of Salmonella spp. in all food types, including milk and dairy products, moluscan, shelfish and other fish product and in environmental swabs.

Beef extract and casein peptone provide nitrogen, vitamins, minerals and amoni acids essential for growth. Calcium carbonate is neutralizer absorbs toxic metabolites. Ox bile, brilliant green and novobiocin inhibit bacteria other than Salmonella. Selectivity is also obtained by both sodium thiosulphate and tetrathionate, suppresing coliforms. Tetrathionate is formed in the medium with lodine incuded in the medium. Bacteria containing the enzyme tetrathionate reductase will thrive in this medium. Sodium chloride supplies essential electrolytes for tranport and somotic balance

Preparation: suspend 89,5 grams of the medium in one liter of distilled water by heating for 5 minutes. DO NOT AUTOCLAVE! Immediately cool to 50°C and add 4 vials of Novobiocin Supplement (ref.SL 0055).

Add 20 ml of a iodine and potassium iodide solution (20 g of iodine and 25 g of potassium iodide in 100 ml of sterile distilled water). Homogenize gently and dispense into sterile containers. The prepared medium should be stored at 2-8°C.

Procedure:

Pre-enrichment and selective enrichment

- ★ Add 25 g of the samples to 225 ml Buffered Peptone Water (Ref. BT 5020) and incubate at 34-38°C for 18±2 hours
- ★ Transfer 0,1 ml of the pre-enrichment culture to 10 ml of Rappaport Vassiliaid Soy Broth (ref. PW 4245). Incubate at 41,5°C for 24±3 hours
- ★ Transfer 1 ml of the pre-enrichment culture to 10 ml of MKTTn Broth. Incubate at 37±1°C for 24±3 hours

- ★ Once opened keep powdered medium closed to avoid hydration at 2 30°C
- ★ The expiration date is indicated on the label.





MKTTn BROTH BASE REF. PS 540

Microbiological test

The following results were obtained in the performance of the medium from type cultures after incubaction at a temperature of $37\pm1^{\circ}$ C and observed after 24 \pm 3 hours

MicroorganismsGrowthEscherichia coli ATCC 25922InhibitedSalmonella Typhimurium ATCC 13048Good

Packaging: 500 g

Novobiocin Supplement 10 vials/ 1 vial / 250 ml Ref. SL 0055



CHROMOGENIC SALMONELLA LAB-AGAR™

REF. PS 598

Chromogenic, selective medium for the isolation and presumptive identification of Salmonella spp. from environmental and other samples. According to the ISO 6579 norm.

| Formula in g/L | | | |
|---------------------|-------|---------------------|-------|
| Peptone | 10,00 | Selective compounds | 12,00 |
| Chromogenic mixture | 0,90 | Agar | 15,00 |

Final pH at 25°C: 7,2 ± 0,2

Principles:

The selectivity of the medium is improved by a cephalosporin which inhibits the growth of Pseudomonas by sodium desoxycholate which suppers the growth of Gram-positive and some Gram-negative bacteria and by Tergitol-4 which is active mainly against the growth of Proteus spp.

The differentiation between the Salmonella and non-Salmonella colonies is achieved by:

- a chromogenic substrate for a specific esterase enzyme of Salmonella, that is split with the liberation of an insoluble magenta-red dye
- a chromogenic glucopyranoside derivative which is split by β-glucosidase with the formation insoluble blue-green dye.

The chromogenic and selective compounds of the medium allow the detection also of the rare lactose positive Salmonella strains. Chromogenic Salmonella LAB-AGAR™ is useful for the detection of S. Typhi and S. paratyphi. Sometimes rare strains of Pseudomonas ana Aeromonas can cultivate with magenta-red colonies. These strains can be easy differentiated with the oxidase test.

Preparation: suspend 19 grams of the medium in 500 ml of distilled water. Mix well and add one vial (vial A) of Salmonella Chromogenic Supplement (ref. SL 0061). Heat with frequent agitation. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C, add one vial (vial B) reconstituted in 2 ml of sterile distilled water of Salmonella Chromogenic Supplement (ref. SL 0061). Mix well and pour into Petri dishes.

The prepared medium should be stored at 8-15°C.

Technique:

- ★ Chromogenic Salmonella LAB-AGAR™ can be used according to the usual laboratory practices for Salmonella isolation with direct plating or after the enrichment in non-selective liquid media
- ★ Incubate the inoculated plates at 37°C ± 1°C for 18-24 hours of observe for the presence of typical magenta-red colonies

Colour colony:

- ★ Salmonella spp magenta-red
- ★ Escherichia coli- colourless
- ★ Proteus spp. pale brown or green
- ★ Klebsiella spp. –blue –green

Storage / Shelf life

- ★ Once opened keep powdered medium closed to avoid hydration at 2 8°C
- ★ The expiration date is indicated on the label.

Packaging: 500 g

Supplement: Salmonella Chromogenic Supplement 5+5 vials. 2 vials /500 ml Ref. SL 0061







EGG`S YOLK TELLURITE STERILE EMULSION

REF. SL 0036

Flask: 100 ml

Formula:

Egg's Yolk Tellurite Sterile Emulsjon 20%......100 ml

Uses

For the detection and enumeration of coagulase positive Satphylococci.

Egg's Yolk Tellurite Emulsion, sterilized and stabilized, for use in microbiology applications, ready for use with Baird Parker LAB-AGAR™ Base acc. to ISO 6888-1 (ref. PS 33) for selection of coagulase positive Staphylococci. The addition of Egg yolk and Poatssium tellurite helps to differentiate these microorganisms from others capable of growing in the base agar, trough the detection of lecithinase and the formation of black colonies.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label



EGG`S YOLK TELLURITE STERILE EMULSION

REF. SL 0036

Flask: 100 ml

Formula:

Egg's Yolk Tellurite Sterile Emulsjon 20%......100 ml

Uses

For the detection and enumeration of coagulase positive Satphylococci.

Egg's Yolk Tellurite Emulsion, sterilized and stabilized, for use in microbiology applications, ready for use with Baird Parker LAB-AGAR™ Base acc. to ISO 6888-1 (ref. PS 33) for selection of coagulase positive Staphylococci. The addition of Egg yolk and Poatssium tellurite helps to differentiate these microorganisms from others capable of growing in the base agar, trough the detection of lecithinase and the formation of black colonies.

Precautions:

- ★ For Laboratory use only
- ★ The supplement should be used only by adequately trained personnel with knowledge of microbiological techniques in the laboratory.
- ★ Consult the material safety data sheet before the use.
- ★ Do not use beyond stated expiry date

- ★ Store at 2-8°C When stored as directed the supplement remains stable until the expiry date shown on the label
- ★ The expiration date is indicated on the label

