

Step	Detailed Procedure	Simplified Procedure
	<ul style="list-style-type: none"> • Prepare Working Wash Buffer by diluting the Concentrated Wash Buffer 1:25. Pour the contents of the bottle in a graduated cylinder and fill it with freshly distilled or deionized water to 1000 mL for 96 wells/plate testing. The Working Wash Buffer is stable for 2 weeks at 15-30°C. • Note: If crystals are present in the Concentrated Wash Buffer, warm it up at 37°C until all crystals dissolve. • Remove unused strips from the microwell plate, and store in the original resealable pouch at 2-8°C. 	<ul style="list-style-type: none"> • Prepare Working Wash Buffer by diluting the Concentrated Wash Buffer 1:25 • Remove and store unused strips at 2-8°C
0	<ul style="list-style-type: none"> • Leave A1 as Blank well. 	<ul style="list-style-type: none"> • Leave A1 as Blank well
1	<ul style="list-style-type: none"> • Add 50 µL of Calibrator 1 in wells B1 and C1. • Add 50 µL of Calibrator 2 in wells D1 and E1. • Add 50 µL of Calibrator 3 in wells F1 and G1. • Add 50 µL of Calibrator 4 in wells H1 and A2. • Add 50 µL of Calibrator 5 in wells B2 and C2. <p>The colors of Calibrator 1-5 gradually change from yellow to blue.</p>	<ul style="list-style-type: none"> • B1 and C1: Add 50 µL Calibrator 1 • D1 and E1: Add 50 µL Calibrator 2 • F1 and G1: Add 50 µL Calibrator 3 • H1 and A2: Add 50 µL Calibrator 4 • B2 and C2: Add 50 µL Calibrator 5
2	<ul style="list-style-type: none"> • Add 50 µL of specimen to assigned wells starting at D2. 	<ul style="list-style-type: none"> • Starting D2: Add 50 µL specimen
3	<ul style="list-style-type: none"> • Add 50 µL of Conjugate to each well except for the Blank well. (Red Reagent). 	<ul style="list-style-type: none"> • Add 50 µL of Conjugate to each well except for the Blank well
4	<ul style="list-style-type: none"> • Mix gently by swirling the microwell plate on a flat bench for 30 seconds. • Cover the microwell plate with the Plate Sealer and incubate in a water bath or an incubator at 37°C ± 2°C for 30 minutes ± 2 minutes. 	<ul style="list-style-type: none"> • Mix gently • Cover the microwell plate with the Plate Sealer and incubate at 37°C for 30 min

5	<ul style="list-style-type: none">Remove the Plate Sealer.Wash each well 5 times with 350 µL of Working Wash Buffer per well, then remove the liquid.Turn the microwell plate upside down on absorbent tissue for a few seconds. Ensure that all wells have been completely washed and dried. Note: Improper washing may cause false positive results.	<ul style="list-style-type: none">Remove the Plate SealerWash each well 5 times with 350 µL of Working Wash BufferTurn the microwell plate upside down on absorbent tissue
6	<ul style="list-style-type: none">Add 50 µL of Substrate A to each well. (Clear Reagent)Add 50 µL of Substrate B to each well. (Clear Reagent) Then a blue color should develop in wells containing Positive specimens.	<ul style="list-style-type: none">Add 50 µL of Substrate A to each wellAdd 50 µL of Substrate B to each well
7	<ul style="list-style-type: none">Mix gently then cover microwell plate with Plate Sealer and incubate in a water bath or incubator at 37°C ± 2°C for 15 minutes ± 1 minute.	<ul style="list-style-type: none">Mix then cover microwell plate with Plate Sealer and incubate at 37°C for 15 min
8	<ul style="list-style-type: none">Remove the Plate Sealer.Add 50 µL of Stop Solution to each well. (Clear Reagent) Then a yellow color should develop in wells containing Positive specimens.	<ul style="list-style-type: none">Remove the Plate SealerAdd 50 µL of Stop Solution to each well
9	<ul style="list-style-type: none">Read at 450/630-700 nm within 30 minutes. Note: Microwell plate can also be read at 450 nm, but it is strongly recommended to read it at 450/630-700 nm for better results.	<ul style="list-style-type: none">Read at 450/630-700 nm within 30 min

AUTOMATED PROCESSING

Automatic EIA microplate processors may be used to perform the assay after validating the results to ensure they are equivalent to those obtained using the manual method for the same specimens. Incubation times may vary depending on the processors used but do not program less incubation times than the procedure listed above. When automatic EIA microplate processors are used, periodic validation is recommended to ensure proper results.

VALIDATION REQUIREMENTS AND QUALITY CONTROL

1. Calculate the Mean Absorbance of Calibrators 1-5 by referring to the table below.

Example of Calibrator 2 Calculation

Item	Absorbance
Calibrator 2: Well D1	0.165
Calibrator 2: Well E1	0.161
Total Absorbance of Calibrator 2	0.165 + 0.161 = 0.326
Mean Absorbance of Calibrator 2	0.326/2 = 0.163
Blank Absorbance: Well A1	0.005
Mean Absorbance of Calibrator 2 – Blank Absorbance	0.163 - 0.005 = 0.158

2. Check the validation requirements below to determine if the test results are valid.

Item	Validation Requirements
Blank Well	Blank Absorbance should be < 0.050 if read at 450/630-700 nm Note: It should be < 0.100 if read at 450 nm
Calibrator 1	Mean Absorbance after subtraction of Blank Absorbance should be < 0.105
Calibrator 2	Mean Absorbance after subtraction of Blank Absorbance should be > 0.105
Calibrator 4	Mean Absorbance after subtraction of Blank Absorbance should be > 0.500

NOTE: The test results are considered invalid if the above validation requirements are not met. Repeat the test or contact your local distributor.

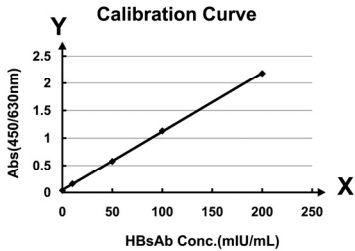
INTERPRETATION OF RESULTS

Draw the calibration curve and obtain quantitative specimen results.

1. Subtract the Blank Absorbance from the Mean Absorbance of each Calibrator, then plot them on the Y-axis against their concentration in mIU/mL on the X-axis on a linear graph paper and draw the calibration curve. Draw the best fitted line through data points to obtain a standard curve. Refer to an example of the calibration curve at right.

NOTE: Do not use the calibration curve at right to make any calculation. A calibration curve must be performed for each run.

2. Obtain quantitative specimen results from their absorbance after subtraction of Blank Absorbance



by using the calibration curve.
NOTE: Specimens that have absorbance above Calibrator 5 should be pre-diluted using Calibrator 1 and retested. The concentration must be multiplied by the dilution factor. Automated reading and calculation may be performed using linear regression function on suitable computer programs.

Results	Concentration
Negative	< 9 mIU/mL
Positive	> 11mIU/mL
Equivocal*	9 - 11mIU/mL

***NOTE:** For Equivocal results, the specimen should be retested. Specimens that are repeatedly Equivocal after retest should be confirmed using an alternate method. If the results remain Equivocal, collect a new specimen in two weeks. If the new specimen is Positive, the specimen is presumed to be Positive.

LIMITATIONS

- The HBsAb Quantitative EIA Test Kit is used for the detection of HBsAb in human serum or plasma. Diagnosis of an infectious disease should not be established based on a single test result. Further testing, including confirmatory testing, should be performed before a specimen is considered positive. A non-reactive test result does not exclude the possibility of exposure. Specimens containing precipitate may give inconsistent test results.
- As with all diagnostic tests, all results must be interpreted together with other clinical information available to the physician.
- As with other sensitive immunoassays, there is the possibility that non-repeatable reactive results may occur due to inadequate washing. The results may be affected due to procedural or instrument error.
- Erroneous result may be due to fibrin particles and microbial contamination.

PERFORMANCE CHARACTERISTICS

Clinical Sensitivity and Specificity

The HBsAb Quantitative EIA Test Kit has correctly identified specimens of a seroconversion panel and has been compared with a leading commercial HBsAb Quantitative EIA test using clinical specimens. The results show that the clinical sensitivity of the HBsAb Quantitative EIA Test Kit is 99.3%, and the clinical specificity is 99.2%.

HBsAb Quantitative EIA vs. Other EIA

HBsAb Quantitative EIA	Method	Other EIA		Total Results
	Results	Positive	Negative	
	Positive	1139	15	1154
	Negative	8	1839	1847
Total Results		1147	1854	3001

Clinical Sensitivity: 99.3% (98.6-99.7%)* Clinical Specificity: 99.2% (98.7-99.6%)*
Overall Agreement: 99.2% (98.9-99.5%) *95% Confidence Interval

Reproducibility

Intra-Assay: Within-run precision has been determined by using 20 replicates of one specimen.

Inter-Assay: Between-run precision has been determined by 3 independent assays on the same sample. Test 20 replicates of the same with three different lots of the HBsAb Quantitative EIA Test Kit.

Specimen	Intra-Assay			Inter-Assay		
	Mean Absorbance	Standard Deviation	Coefficient of Variation (%)	Mean Absorbance	Standard Deviation	Coefficient of Variation (%)
1	1.081	0.063	5.828	1.074	0.071	6.611

BIBLIOGRAPHY

- Frank Fenner and David O. White, *Medical Virology*, 4th Edition, Academic Press, 1994.
- Centers for Disease Control. Viral Hepatitis B Fact Sheet.
- Richman, D., R. Whitley, F. Hayden. *Clinical Virology*. New York: Churchill Livingstone Inc., 1997.
- World Health Organization. World Health Organization Hepatitis B Fact Sheet. N°204. Revised October 2000.
- World Health Organization. Hepatitis B. 2002.

Index of Symbols

	Consult instructions for use		Tests per kit		Manufacturer
	For <i>in vitro</i> diagnostic use only		Use by		
	Store between 2-8°C		Lot Number		Catalog #
	HBsAb		Substrate A		Substrate B
	Wash Buffer (25x)		Conjugate		Calibrator 1
	Calibrator 1		Calibrator 2		Calibrator 3
	Calibrator 4		Calibrator 5		Stop Solution
	Microwell Plate		Plate Sealer		Package Insert



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