

Specification

Solid differential medium for primary identification of enterobacteria based on the fermentation of two sugars and the hydrogen sulfide production according to ISO standard.

Presentation

20 Tubes / Slant
Tube 16 x 110mm
with: 7,5 ± 0,3 ml

Packaging Details

1 box with 20 tubes, 16x113 mm glass tubes, ink
labelled and metal cap.

Shelf Life

9 months

Storage

8-25°C

Composition

Composition (g/l):

Peptone.....	20.0
Meat extract.....	3.00
Yeast extract.....	3.00
Sodium chloride.....	5.00
Lactose.....	10.0
Glucose.....	1.00
Ferric citrate.....	0.50
Sodium thiosulfate.....	0.50
Phenol red.....	0.03
Agar.....	15.0

Description /Technique

Kligler Agar is a differential medium that has all the characteristics of the 2-Sugar Russell Agar and Lead Acetate Medium for H₂S detection. In this medium, lactose fermentation and hydrogen sulfide production can be detected, allowing a presumptive identification of most enterobacteria. Sugar fermentation is shown by acid production, which turns the indicator from red to yellow. Since there is only a small amount of sugar (dextrose) in the medium, acid production due to its fermentation is very limited and re-oxidation of the indicator occurs on the surface of the medium, causing the indicator to remain red. When lactose is fermented, a large amount of acid is produced re-oxidation does not occur and the entire medium turns yellow.

Hydrogen sulfide production is indicated by the medium turning black, due to the reaction of H₂S (liberated from thiosulfate) with the iron ions presents in the ammonium iron citrate.

Kligler Iron Agar is used in slanted tubes with short slant and a generous butt, which are inoculated on the surface and also stab inoculated. The inoculum must be copious; it has to come from a solid medium, otherwise, readings may be delayed (up to additional 2-3 days). Normal incubation is 18-24 hours at 36°C ±0,2.

A large production of H₂S may make the readings difficult, and hence early readings are strongly recommended.

To inoculate tubes follow the standard laboratory methods or the applicable norms:stab inoculation, loop inoculation.

Quality control

Physical/Chemical control

Color : Reddish

pH: 7.4 ± 0.2 at 25°C

Microbiological control

Inoculate by stabbing the butt + streak the slant

Aerobiosis. Incubation at 36 ± 2°C, reading at 18-24 h

Microorganism

Shiaella flexneri ATCC® 12022, WDCM 00126

Escherichia coli ATCC® 8739, WDCM 00012

Salmonella typhimurium ATCC® 14028, WDCM 00031

Proteus mirabilis ATCC® 43071

Growth

Good /Slant: Alk/Butt:Ac /Gas (-)/ SH2(-)

Good / Slant:Ac /Butt:Ac /Gas (+)/ SH2(-)

Good /Slant: Alk/Butt:Ac /Gas (+)/ SH2(+)

Good /Slant: Alk/Butt:Ac /Gas (+)/ SH2(+)

Sterility Control

Incubation 48 hours at 30-35°C and 48 hours at 20-25°C: NO GROWTH

Check at 7 days after incubation in same conditions

Bibliography

- ATLAS, R.M. & L.C. PARKS (1993) Handbook of Microbiological Media. CRC Press. Boca Ratón. Fla. USA.
- DOWNES, F.P. & K. ITO (2001) Compendium of Methods for the Microbiological Examination of Foods. 4th ed. APHA. Washington. DC. USA.
- ISO 6340:1995 Standard. Water Quality - Detection of *Salmonella* species. Geneva.
- ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- KLIGLER (1918) Modification of culture media used in the isolation and differentiation of typhoid, dysentery and allied bacilli. J. Exper Med. 28:319-332.
- KLIGLER (1917) A simple medium for the differentiation of members of typhoid-paratyphoid groups. Am. J. Pub. Hlth 7:1042-1044.
- MacFADDIN, J.F. (1985) Media for isolation-cultivation-identification-maintenance of medical bacteria. William & Wilkins. Baltimore. MD. USA.
- RUSELL, F.F. (1911) The isolation of typhoid bacilli from urine and feces with the description of a new double sugar tube medium. J. Med. Res. 25:217-220.