

**BC-760[B]/BC-760[R]/BC-780[R]**

**AUTO HEMATOLOGY ANALYZER**

**OPERATOR'S MANUAL**



# Table of Contents

<b>1 Safety Information .....</b>	<b>1 - 1</b>
1.1 Labels and Symbols on the System .....	1 - 1
1.2 Safety-related Symbols and Messages .....	1 - 8
1.2.1 General Safety Messages .....	1 - 8
1.2.2 Analyzer Transportation and Installation-related Safety Messages .....	1 - 9
1.2.3 Reagent, Control, and Calibrator-Related Safety Messages .....	1 - 9
1.2.4 Maintenance-related Safety Messages .....	1 - 10
1.2.5 Laser Warning .....	1 - 10
1.2.6 Network Security .....	1 - 11
<b>2 Using this Manual .....</b>	<b>2 - 1</b>
2.1 Overview .....	2 - 1
2.2 Who Should Read This Manual .....	2 - 1
2.3 How to Find Information .....	2 - 1
2.4 Conventions Used in This Manual .....	2 - 2
<b>3 Understanding Your System .....</b>	<b>3 - 1</b>
3.1 Intended Use .....	3 - 1
3.2 Test Parameters .....	3 - 1
3.2.1 Blood Sample Test Parameters, Histograms, and Scattergrams .....	3 - 2
3.2.2 Body Fluid Sample Test Parameters, Histograms, and Scattergrams .....	3 - 7
3.3 Device Description .....	3 - 8
3.3.1 Structure and Components .....	3 - 8
3.3.2 Modules and Components .....	3 - 8
3.3.3 Accessories .....	3 - 10
3.4 Supported Tubes, Tube Racks and Adapters .....	3 - 11
3.4.1 Supported Tubes .....	3 - 11
3.4.2 Supported Tube Racks .....	3 - 18
3.4.3 Supported Adapters .....	3 - 19
3.5 Overview of Software Interfaces .....	3 - 21
3.6 Reagents, Controls and Calibrators .....	3 - 24
3.6.1 Reagent .....	3 - 24
3.6.2 Controls and Calibrators .....	3 - 25
<b>4 Understanding the System Principles .....</b>	<b>4 - 1</b>
4.1 Overview .....	4 - 1
4.2 WBC Measurement .....	4 - 1
4.2.1 SF CUBE Cell Analysis Technology .....	4 - 1
4.2.2 Derivation of WBC-Related Parameters .....	4 - 3
4.3 Hemoglobin Concentration Measurement .....	4 - 4
4.3.1 A Test Model Using the Colorimetric Method .....	4 - 4
4.3.2 Derivation of HGB .....	4 - 5
4.4 RBC/PLT Measurement .....	4 - 5
4.4.1 Sheath Flow Impedance Method .....	4 - 5

4.4.2 Measurement Principle of Platelets in DIFF Channel (PLT-H)	4 - 5
4.4.3 SF CUBE Cell Analysis Technology	4 - 5
4.4.4 RBC-Related Parameters	4 - 6
4.4.5 PLT-Related Parameters	4 - 7
4.4.6 Reticulocyte Parameters*	4 - 8
4.5 Erythrocyte Sedimentation Rate Measurement	4 - 8
4.6 Body Fluid Parameters	4 - 8
4.7 Wash	4 - 9
<b>5 Installing and Connecting the System</b>	<b>5 - 1</b>
5.1 Notes for Analyzer Installation	5 - 1
5.1.1 Space Requirements	5 - 1
5.1.2 Power Requirements	5 - 2
5.1.3 Environment Requirements	5 - 2
5.1.4 Fuse Requirement	5 - 2
5.1.5 Moving and Installing the Analyzer	5 - 2
5.2 Connecting the Analyzer System	5 - 4
5.2.1 Connecting the Reagents	5 - 4
5.2.2 Connecting the Peripherals	5 - 6
<b>6 Customizing the Analyzer Software</b>	<b>6 - 1</b>
6.1 Introduction	6 - 1
6.2 Saving Settings after Changes	6 - 2
6.3 Analyzer Settings	6 - 2
6.3.1 System Setup ("Menu" > "Setup" > "System Setup")	6 - 2
6.3.2 User Management ("Menu" > "Setup" > "User Management")	6 - 7
6.3.3 Auxiliary Setup ("Menu" > "Setup" > "Auxiliary Setup")	6 - 8
6.3.4 Para. Setup ("Menu" > "Setup" > "Para. Setup") (Administrators)	6 - 10
6.3.5 Maintenance ("Menu" > "Setup" > "Maintenance") (Administrators)	6 - 12
6.3.6 Reagent Setup ("Menu" > "Setup" > "Reagent Setup")	6 - 13
6.3.7 Setting up Functions for the Auto-Loading of Samples (Menu > "Setup" > "Auto-loading") (administrators)	6 - 13
6.3.8 Gain Setup ("Menu" > "Setup" > "Gain Setup") (Administrators)	6 - 13
6.3.9 Setting Re-exam Rules ("Menu" > "Setup" > "Re-exam Rules Setup") (Administrators)	6 - 13
6.3.10 Setting Auto Startup/Shutdown Time ("Menu" > "Setup" > "Auto Startup/Shutdown") (Administrators)	6 - 14
<b>7 Operating Your Analyzer</b>	<b>7 - 1</b>
7.1 Overview	7 - 1
7.1.1 Operating Your Analyzer	7 - 1
7.1.2 Introduction to the Screen	7 - 2
7.2 Preparations before Operation	7 - 4
7.3 Start up and Login	7 - 5
7.3.1 Start up the Analyzer	7 - 5
7.3.2 Switching Login Account	7 - 6
7.4 Daily QC	7 - 6
7.5 Preparing Samples	7 - 6
7.5.1 Preparing Whole Blood Samples (For WB Mode)	7 - 6



7.5.2 Preparing Predilute Samples (For PD Mode)	7 - 7
7.5.3 Preparing Body Fluid Samples (For BF Mode)	7 - 8
7.5.4 Placing Barcode Labels	7 - 9
7.6 Running Samples under Closed-tube Mode	7 - 10
7.6.1 Setting up Analysis Orders	7 - 10
7.6.2 Performing Sample Analysis	7 - 12
7.7 Running Samples	7 - 17
7.7.1 Setting up Analysis Orders	7 - 17
7.7.2 Performing Sample Analysis	7 - 18
7.7.3 Stop Count	7 - 20
7.8 STAT	7 - 20
7.9 Entering/Exiting Standby Status	7 - 21
7.10 Shutting down the Analyzer	7 - 21
7.10.1 Shutting down the analyzer	7 - 21
7.11 Viewing Guidance Videos in iHelp	7 - 22
<b>8 Reviewing Sample Results</b>	<b>8 - 1</b>
8.1 Introduction	8 - 1
8.2 Reviewing Sample Results	8 - 1
8.2.1 Entering the "Table Review" Screen	8 - 1
8.2.2 Operations on the "Table Review" Screen	8 - 1
8.2.3 Searching for Sample Records	8 - 2
8.2.4 Graph Review	8 - 4
8.2.5 Communication	8 - 6
8.2.6 Exporting Sample Results	8 - 7
8.2.7 Calculating CV Values	8 - 7
8.2.8 Editing Information	8 - 7
8.2.9 Validating/Cancelling Validation (Administrators)	8 - 8
8.2.10 Deleting Sample Records	8 - 8
8.3 Flags of Analysis Results	8 - 9
8.3.1 Parameter Flags	8 - 9
8.3.2 Flags of Abnormal Blood Cell Differential or Morphology Results	8 - 9
<b>9 Using the QC Program</b>	<b>9 - 1</b>
9.1 Overview	9 - 1
9.2 L-J QC	9 - 1
9.2.1 Setting up L-J QC Files (Administrators)	9 - 2
9.2.2 Running L-J QC Tests	9 - 4
9.2.3 Reviewing L-J QC Results	9 - 7
9.2.4 Viewing Results of Single QC Count	9 - 13
9.3 X-B QC	9 - 13
9.3.1 Validity Determination for X-B QC Samples	9 - 13
9.3.2 Setting up X-B QC Rules	9 - 14
9.3.3 Reviewing X-B QC Results	9 - 14
9.4 When QC Results are Out of Range	9 - 17
<b>10 Calibrating Your Analyzer</b>	<b>10 - 1</b>

10.1 Overview .....	10 - 1
10.2 When to Calibrate .....	10 - 1
10.3 Checking before Calibration .....	10 - 1
10.4 Running the Calibration Programs .....	10 - 1
10.4.1 Notes before Calibration .....	10 - 2
10.4.2 Manual Calibration .....	10 - 2
10.4.3 Calibrating with ESR (administrators) .....	10 - 3
10.4.4 Calibrating with Calibrators (administrators) .....	10 - 4
10.4.5 Calibrating with Fresh Blood Samples (administrators) .....	10 - 5
10.4.6 Verifying Calibration Factors .....	10 - 6
10.5 Calibration History ("Menu" > "Calibrate" > "Calibration History") .....	10 - 6
<b>11 Printing .....</b>	<b>11 - 1</b>
11.1 Setting up Print Template .....	11 - 1
11.1.1 Operation Procedure .....	11 - 1
11.2 Printing Sample Result Report .....	11 - 2
11.2.1 Printing Current Sample Result Report .....	11 - 2
11.2.2 Printing from the Table Review Screen .....	11 - 2
11.2.3 Printing from the Graph Review Screen .....	11 - 3
11.2.4 Printing RUO Parameter Results .....	11 - 3
11.2.5 Printing Microscopic Parameter Results .....	11 - 4
11.3 Printing QC Result Report .....	11 - 4
11.3.1 Printing L-J QC Result from the L-J QC Table Screen .....	11 - 4
11.3.2 Printing L-J QC Graph from the L-J QC Graph Screen .....	11 - 4
11.3.3 Printing X-B QC Graph from the X-B QC Graph Screen .....	11 - 4
11.3.4 Print X-B QC Parameter Result from the X-B QC Table Screen .....	11 - 5
11.4 Printing Manual Calibration Factors .....	11 - 5
<b>12 Service .....</b>	<b>12 - 1</b>
12.1 Overview .....	12 - 1
12.2 When and Why to Perform the Maintenance .....	12 - 1
12.2.1 Maintenance of Parts and Components .....	12 - 1
12.2.2 Manual Cleaning .....	12 - 2
12.2.3 Replacing the Parts and Components .....	12 - 3
12.3 Reagent Management .....	12 - 3
12.3.1 Viewing Reagent Information ("Menu" > "Setup" > "Reagent Setup") .....	12 - 3
12.3.2 Replace the Reagents .....	12 - 3
12.3.3 Replacing the Waste Container .....	12 - 7
12.4 Probe Cleanser Maintenance .....	12 - 8
12.4.1 Daily Probe Cleanser Maintenance .....	12 - 8
12.4.2 Probe Cleanser Maintenance to Parts and Components .....	12 - 8
12.5 Auto-cleaning the Parts and Components .....	12 - 9
12.6 Manual Cleaning of Parts and Components .....	12 - 9
12.6.1 Cleaning the Probe Wipe, Floating Blood Barrier and the Blood Barrier Bracket .....	12 - 9
12.6.2 Cleaning the Analyzer Front Cover .....	12 - 11
12.6.3 Cleaning the Filter .....	12 - 12
12.7 Preparing to Ship .....	12 - 12

12.8 Screen Calibration .....	12 - 13
12.9 Viewing and Exporting Logs .....	12 - 13
12.9.1 Viewing Logs .....	12 - 13
12.9.2 Exporting Logs .....	12 - 14
12.10 Upgrading Analyzer .....	12 - 15
<b>13 Troubleshooting .....</b>	<b>13 - 1</b>
13.1 Overview .....	13 - 1
13.2 Checking Analyzer Status .....	13 - 1
13.3 Error Messages and Solutions .....	13 - 2
<b>A Index .....</b>	<b>A - I</b>
<b>B Specification .....</b>	<b>B - 1</b>
B.1 Classification .....	B - 1
B.2 Reagent .....	B - 1
B.3 Models .....	B - 4
B.4 Parameters .....	B - 4
B.5 Sampling Features .....	B - 6
B.5.1 Sample modes, test panel, and applicable model .....	B - 6
B.5.2 Sample Volumes Required for Each Analysis .....	B - 7
B.5.3 Throughput .....	B - 8
B.6 Performance Specifications .....	B - 8
B.6.1 Background/Blank Count Requirements .....	B - 8
B.6.2 Linearity Ranges .....	B - 9
B.6.3 Accuracy .....	B - 10
B.6.4 Repeatability .....	B - 10
B.6.5 Carryover .....	B - 12
B.6.6 Correlation .....	B - 13
B.6.7 Stability (Applicable to Protein Module Parameters) .....	B - 13
B.6.8 Channel Temperature Accuracy (Applicable to Protein Module Parameters) .....	B - 14
B.6.9 Dispensing Accuracy and Repeatability (Applicable to Protein Module Test Parameters) ..	B - 14
B.7 Requirements for Input/Output Devices .....	B - 14
B.7.1 Keyboard .....	B - 14
B.7.2 Mouse .....	B - 14
B.7.3 External barcode scanner .....	B - 14
B.7.4 Printer .....	B - 14
B.7.5 USB Drive .....	B - 14
B.7.6 Electronic Interface Specifications .....	B - 15
B.8 Interfaces .....	B - 15
B.9 Power supply .....	B - 15
B.10 Fuse .....	B - 15
B.11 Electromagnetic compatibility (EMC) .....	B - 15
B.12 Noise Level .....	B - 17
B.13 Normal Operating Environment .....	B - 17
B.14 Storage Environment .....	B - 17
B.15 Operating Environment .....	B - 17

B.16 Dimensions and Weight .....	B - 18
B.17 Contraindication .....	B - 18
B.18 Barcode Specifications .....	B - 18
B.19 Safety Classification .....	B - 21
B.20 Limitations .....	B - 22
B.20.1 For Blood Routine Tests .....	B - 22
B.20.2 ESR Tests .....	B - 23
<b>C Accessories and Packing List .....</b>	<b>C - 1</b>
C.1 Accessories of the Analyzer .....	C - 1
C.2 Optional Accessories of the Analyzer .....	C - 1
C.3 Packing List .....	C - 1
<b>D Communication .....</b>	<b>D - 1</b>
<b>E Radio Regulatory Compliance .....</b>	<b>E - 1</b>
<b>F References .....</b>	<b>F - 1</b>
<b>G Maintenance Logs .....</b>	<b>G - 1</b>



© 2022-2023 Shenzhen Mindray Bio-medical Electronics Co., Ltd. All rights Reserved.

For this Operator's Manual, the issued Date is 2023-04.

## Intellectual Property Statement

SHENZHEN MINDRAY BIO-MEDICAL ELECTRONICS CO., LTD. (hereinafter called Mindray) owns the intellectual property rights to this Mindray product and this manual. This manual may refer to information protected by copyright or patents and does not convey any license under the patent rights or copyright of Mindray, or of others.

Mindray intends to maintain the contents of this manual as confidential information. Disclosure of the information in this manual in any manner whatsoever without the written permission of Mindray is strictly forbidden.

Release, amendment, reproduction, distribution, rental, adaptation, translation or any other derivative work of this manual in any manner whatsoever without the written permission of Mindray is strictly forbidden.

**mindray**



**MINDRAY**

are the trademarks, registered or otherwise, of Mindray in China and other countries. All other trademarks that appear in this manual are used only for informational or editorial purposes. They are the property of their respective owners.

## Responsibility on the Manufacturer Party

Contents of this manual are subject to changes without prior notice.

All information contained in this manual is believed to be correct. Mindray shall not be liable for errors contained herein nor for incidental or consequential damages in connection with the furnishing, performance, or use of this manual.

Mindray is responsible for the effects on safety, reliability and performance of this product, only if:

- all installation operations, expansions, changes, modifications and repairs of this product are conducted by Mindray authorized personnel;
- the electrical installation of the relevant room complies with the applicable national and local requirements;
- the product is used in accordance with the instructions for use.

## NOTE

- **This equipment must be operated by skilled/trained clinical professionals.**

---

## WARNING

- **This instrument is intended to be used by clinical laboratory professionals trained by Mindray or Mindray-authorized distributors.**
  - **Be sure to use the instrument under specified environment in this manual. If not, the instrument may not work properly, the measurement may be unreliable, thus causing damage to the instrument and harm to the body.**
- 

## Warranty

THIS WARRANTY IS EXCLUSIVE AND IS IN LIEU OF ALL OTHER WARRANTIES, EXPRESSED OR IMPLIED, INCLUDING WARRANTIES OF MERCHANTABILITY OR FITNESS FOR ANY PARTICULAR PURPOSE.

## Exemptions

Mindray's obligation or liability under this warranty does not include any transportation or other charges or liability for direct, indirect or consequential damages or delay resulting from the improper use or application of the product or the use of parts or accessories not approved by Mindray or repairs by people other than Mindray authorized personnel.

This warranty shall not extend to:

- Malfunction or damage caused by improper use or man-made failure.
- Malfunction or damage caused by unstable or out-of-range power input.
- Malfunction or damage caused by force majeure such as fire and earthquake.
- Malfunction or damage caused by improper operation or repair by unqualified or unauthorized service people.
- Malfunction of the instrument or part whose serial number is not legible enough.
- Others not caused by instrument or part itself.

## Customer Service Department

Manufacturer:	Shenzhen Mindray Bio-Medical Electronics Co., Ltd.
Address	Mindray Building, Keji 12 <sup>th</sup> Road South, High-tech industrial park, Nanshan, Shenzhen 518057, P.R.China
Website	www.mindray.com
E-mail Address:	service@mindray.com.cn
Tel:	+86 755 81888998
Fax:	+86 755 26582680
EC-Representative:	Shanghai International Holding Corp. GmbH(Europe)
Address:	Eiffestraße 80 20537 Hamburg Germany
Tel:	0049-40-2513175
Fax:	0049-40-255726





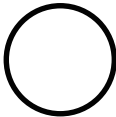




# 1 Safety Information










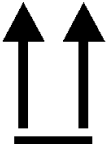


## 1.1 Labels and Symbols on the System

You may find the following symbols on package or the body of the instrument:

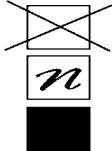



### CAUTION

- **During the daily use of the analyzer, especially the cleaning process, the operator shall ensure the intactness of the labels.**



When you see...	It means...
	Caution Note: Indicates the need for the user to consult the instructions for use for important cautionary information such as warnings and precautions that cannot, for a variety of reasons, be presented on the medical device itself.
	Biological risks
	Warning, laser beam
	PROTECTIVE CONDUCTOR TERMINAL
	Off (Power)
	On (Power)
	USB connection
	Computer Network
	Alternating current

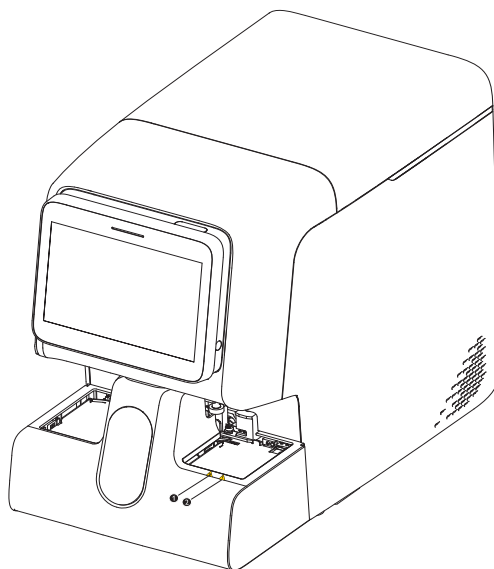
	Serial number
	In vitro diagnostic medical device
	Date of manufacture
	Temperature limit
	Humidity limitation
	Atmospheric pressure limitation
<div>  严禁在仪器指示灯闪烁状态下进行更换废液桶操作！         </div> <div>  Do not replace the waste container when the power indicator is flickering!         </div>	Biological risks (on the tube of the waste container cap assembly) Do not replace the waste container when the power indicator is flickering!
	Fragile, handle with care
	This way up
	Keep dry
	Do not roll



	Stacking limit by number
	Unique Device Identifier
	THE FOLLOWING DEFINITION OF THE WEEE LABEL APPLIES TO EU MEMBER STATES ONLY: THE USE OF THIS SYMBOL INDICATES THAT THIS PRODUCT SHOULD NOT BE TREATED AS HOUSEHOLD WASTE. BY ENSURING THAT THIS PRODUCT IS DISPOSED OF CORRECTLY, YOU WILL HELP PREVENT BRINGING POTENTIAL NEGATIVE CONSEQUENCES TO THE ENVIRONMENT AND HUMAN HEALTH. FOR MORE DETAILED INFORMATION WITH REGARD TO RETURNING AND RECYCLING THIS PRODUCT, PLEASE CONSULT THE DISTRIBUTOR FROM WHOM YOU PURCHASED THE PRODUCT.
	European Conformity

The general meaning assigned to geometric shapes, safety colors and contrast colors for safety signs are as follows:

Geometric shape	Meaning	Safety color	Contrast color	Graphical symbol color
	Warning	Yellow	Black	Black
	Warning	Yellow	Black	Black



**Figure 1-1 Warning labels on the front of main unit**



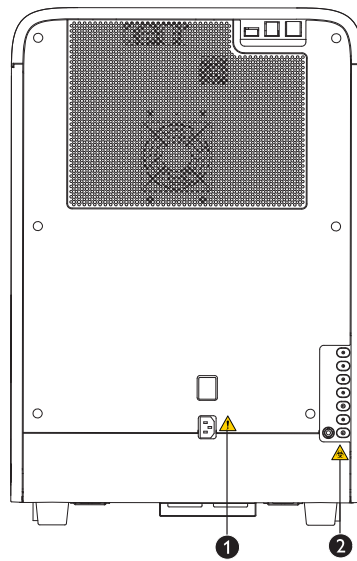
① Warning

Biohazard



② Precautions

1. Test tubes must come into the rack smoothly. If not, shape the tube by replacing the labels.
2. Make sure cap the tubes securely.
3. Make sure the left tray is not full and the detection photocoupler is not blocked, and then start analysis.



**Figure 1-2 Warning labels on the back of main unit**



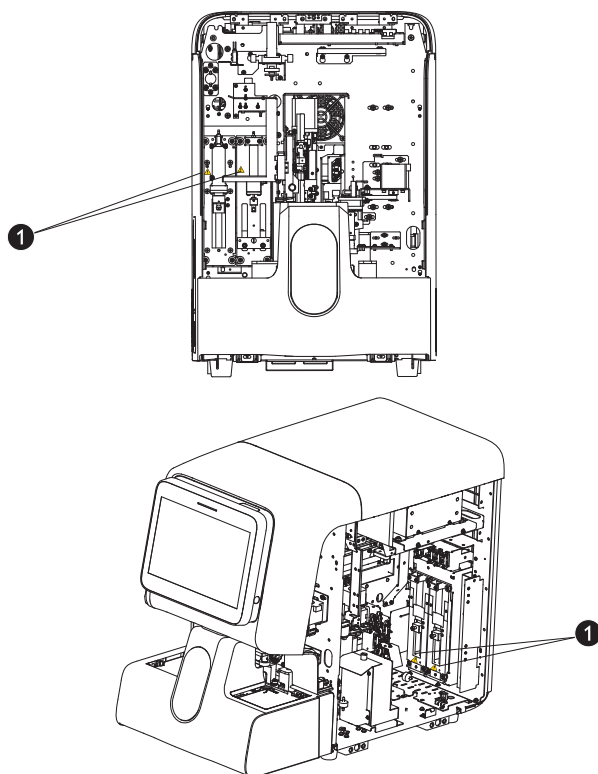
① Warning

1. Connect only to a properly earth grounded outlet.
2. To avoid electric shock, disconnect power cord prior to maintenance or servicing the analyzer.



② Warning

BIOHAZARD

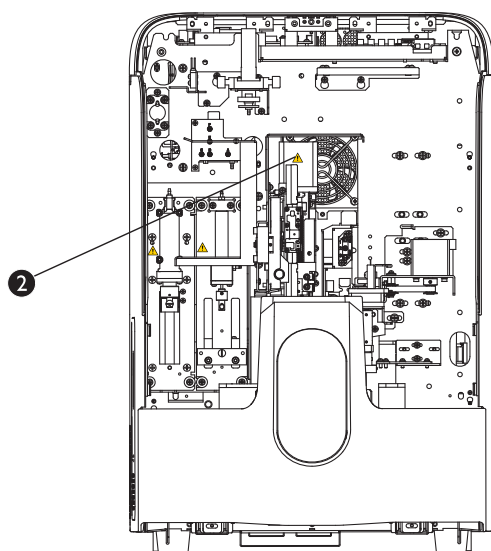


**Figure 1-3 Warning labels for moving parts 1**



① Warning

To avoid personal injury, do not put your hand under the syringe or inside the slot!



**Figure 1-4 Warning labels for moving parts 2**



## ② Warning

To avoid personal injury, do not put your hand under the pipette assembly or inside the moving track!

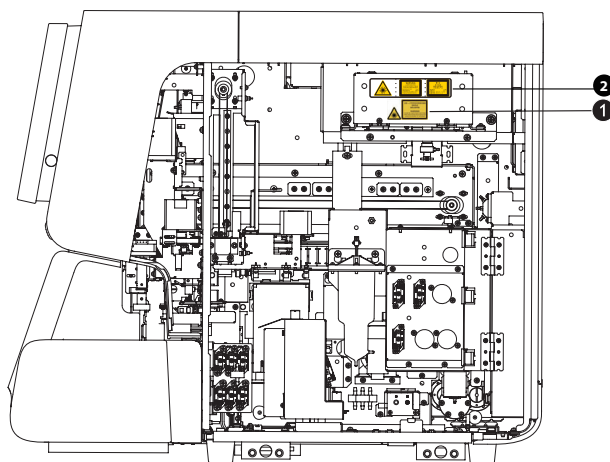


Figure 1-5 Optical assembly laser warning labels 1



## ① Warning

CLASS 3B LASER RADIATION WHEN OPEN AND INTERLOCKS DEFEATED

AVOID EXPOSURE TO THE BEAM



## ② Warning

LASER RADIATION

AVOID EXPOSURE TO BEAM

CLASS 3B LASER PRODUCT

MAXIMUM OUTPUT: 15mW

WAVELENGTH: 635nm

STANDARD NAME: IEC 60825-1:2014

PUBLICATION DATE: 2014.05

## 1.2 Safety-related Symbols and Messages

You will find the following symbols in this manual:



### **BIOLOGICAL RISK**

- Alert you to a potentially biohazardous condition.



### **WARNING**

- Alert you to an operating condition that can cause death, serious personnel injury or property damage.



### **CAUTION**

- Alert you to an operating condition that can cause minor personnel injury, system failure/ damage, and property damage.

### **NOTE**

- Alert you to information that requires your attention.

### 1.2.1 General Safety Messages



### **BIOLOGICAL RISK**

- All the samples, controls, calibrators, wastes and areas contacted them are potentially biohazardous. Wear proper personal protective equipment (e.g. gloves, lab coat, goggles, etc.) and follow safe laboratory procedures when handling them and the contacted areas in the laboratory.
- Be sure to dispose of reagents, waste, samples, consumables, etc. according to government regulations.
- Discard the system according to government regulations.



### **WARNING**

- The fuse used in the instrument is not a replaceable one. If there is any problem with the fuse, contact Mindray Customer Service Department or your local distributor your local distributor.
- Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the country in which the user and/or the patient is established.



### **CAUTION**

- Be sure to operate the instrument under the situation specified in this manual. If the instrument is used in a manner not specified by the manufacturer, the protection provided by the instrument may be impaired.
- Be sure to use the specified external devices only, and keep them away from water.
- External devices connected to the analyzer and digital interfaces must be authorized and complied with relevant safety and EMC standards (e.g. IEC 60950 Safety of Information Technology instrument Standard and CISPR 22 EMC of Information Technology instrument Standard (Class B)). Any persons who connects additional instrument to the signal input or output ports and configures an IVD system, is responsible for ensuring that the system works normally and complies within the safety and EMC requirements. If you have any questions, consult the technical service department of your local representative.

- Make sure all the safety measurements are adopted. It is prohibited to disable any safety device or sensor.
- Keep your clothes, hairs and hands away from the moving parts to avoid injury.
- Be sure to use the instrument under specified environment in this manual. If not, the instrument may not work properly, the measurement may be unreliable, thus causing damage to the instrument and harm to the body.
- This instrument is intended to be used by clinical laboratory professionals trained by Mindray or Mindray-authorized distributors.
- When power cut occurs suddenly, turn off the instrument immediately.
- Dilution is required when the analyte content in the sample exceeds the upper limit of the linear range.
- The instrument must be within the calibration validity period; otherwise, the measurement results may be inaccurate.

### 1.2.2 Analyzer Transportation and Installation-related Safety Messages

#### WARNING

- Before turning on the instrument, make sure the input voltage meets the requirements.
- When installing the instrument, ensure that the power switch is in close proximity to the instrument and within your easy access.

#### CAUTION

- Unpacking, installation or transportation by personnel not authorized or trained by Mindray may cause personal injury or damage your instrument. Do not unpack, transport or install your instrument without the presence of Mindray-authorized personnel.
- The installation, authorization, upgrade and modification of the system software must be performed by personnel authorized by Mindray. Make sure to install only Mindray-authorized software.
- Using pinboard may bring electrical interference and the analysis results may be unreliable. Place the analyzer near the electrical outlet to avoid using the pinboard.
- Use the power cord provided by the manufacturer. Using the power cord other than provided by the manufacturer may lead to system damage or unqualified smear output.
- When connecting the reagents, make sure the color of the reagent container cap assembly is the same as that of the reagent inlet to which it is connected.
- Check if the reagent tubes are properly connected before using the system. Otherwise, the results may be inaccurate.

### 1.2.3 Reagent, Control, and Calibrator-Related Safety Messages

#### CAUTION

- The reagents are irritating to eyes, skin and airway. Wear proper personal protective equipment (e.g. gloves, lab coat, goggles, etc.) and follow safe laboratory procedures when handling them and the contacted areas in the laboratory. If reagents accidentally spill on your skin or in your eyes, rinse the area with ample amount of clean water, and seek medical attention immediately.
- Use the reagents, controls and calibrators specified by the manufacturer only. Using other reagents, controls and calibrators may lead to system damage and inaccurate measurement, control and calibration results.
- Pay attention to the expiration dates and open-container stability days of all the reagents. Be sure not to use expired reagents. Otherwise, the results may be inaccurate.

- To ensure measurement accuracy, do not mix the new container of reagent with the residue in the replaced container to ensure accurate measurement, and prevent reagents from pollution following safe laboratory procedures.

## 1.2.4 Maintenance-related Safety Messages



### BIOLOGICAL RISK

- Mindray does not claim the validity of the listed chemicals in infection control. For effective control of infection, please consult the Infection Prevention Department of the hospital or the epidemic professionals.
- Remove the waste container cap and replace the waste container only when the power indicator is not flickering, in order not to make the waste overflow from the container.
- If the waste is discharged using waste container, make sure the pickup tube of the waste container cap assembly is above, and the tube is smooth and not bent. If the waste is discharged directly, make sure the waste pump is at a lower position than the waste outlet on the instrument.
- After replacing the reagent container/bag, check the tubing connected to the cap assembly and make sure it is not bent over.



### CAUTION

- Before cleaning the floating blood barrier and the blood barrier bracket, make sure the analyzer is shut down.
- Improper maintenance may damage the analyzer. Operators must follow the instruction of this Operator's Manual to perform maintenance operations. For problems not mentioned in this manual, contact Mindray customer service department for Mindray service advice.
- Only parts supplied by Mindray can be used for maintenance. For any question, contact Mindray customer service department.
- If you accidentally spill hazardous material (for example, reagents or samples) on the instrument, clean the instrument with specified disinfectant. Wear proper personal protective equipment (e.g. gloves, lab coat, etc.) and follow safe laboratory procedures when handling them and the contacted areas in the laboratory.
- The user shall perform regular cleaning and sterilization to the cover of the instrument. Use the specified materials to sterilize the instrument only. For any damage to the instrument or other accidents caused by using materials other than specified, Mindray will not provide any warranty.
- The cleaning and sterilization may damage the instrument to some extent. It is recommended to perform sterilization only when necessary according to your laboratory protocol. Remember to clean the instrument before sterilizing.
- Do not use any decontamination or cleaning agents which could cause a HAZARD as a result of a reaction with parts of the instrument or with material contained in it.
- Only use the accessories and consumables manufactured or recommended by Mindray to achieve the promised system performance and safety. For more information, contact Mindray Customer Service Department or your local distributor.
- If any of the pipes or fluidic components are worn out, stop using the analyzer and contact Mindray customer service department immediately for inspection or replacement.
- When the power indicator is flickering, it indicates that the sampling probe is lowering down. Be sure your hand is away from the sampling probe during the process, otherwise, your hand may be hurt.

## 1.2.5 Laser Warning

Class 1 laser product



**⚠ CAUTION**

- Class 3B laser radiation when open and interlocks defeated.
- Avoid exposure to the beam
- Optical density: OD4+
- Radiation exposure level: 56.77mW/cm<sup>2</sup>
- Maximum output: 15mW
- Wavelength: 635nm
- Standard name: IEC 60825-1:2014
- Publication date: 2014.05
- Use goggles when necessary

**⚠ WARNING**

- This product is a CLASS 1 embedded laser product. When opened or when the interlock defeated, there are Class 3B laser radiation. Users must not open, disassemble or damage the interlock device; otherwise may be exposed to laser exposure. For any further information or questions, please contact Mindray's Customer Service Department.

**NOTE**

- The service and maintenance of the laser product must be handled by professionals. Return the product to Mindray Customer Service for service and maintenance.

**1.2.6 Network Security****NOTE**

- The instrument software uses closed-loop operating system, meaning that the applications of the instrument works in exclusive mode and are free from other application's disturbance. Users can only operate the software interface but cannot directly access the operating system or install software. Therefore, the system is far less vulnerable to viruses, spyware, or malware attacks.
- When the instrument connects to an external computer, install anti-virus software on the computer and scan for viruses and update patches periodically. Do not use it for unintended purposes.
- Data transmission must be performed in a closed-loop network or virtual network. The network must be isolated.
- Users have the responsibility to protect the network authentication information, such as password and user information, from being obtained by unauthorized personnel.

The analyzer software is developed based in the following Ready-for-Use software.

Name	Model	Version	Title	Manufacturer
Operating system	OS Core	4.14.98	Linux embedded operating system	Linux Kernel Organization, Inc

**This page intentionally left blank.**

# 2 Using this Manual

## 2.1 Overview

This chapter explains how to use your Operator's Manual of BC-760[B]/BC-760[R]/BC-780[R] Auto Hematology Analyzer (hereby referred to as "the analyzer"). The manual is shipped with your product and describes the purpose, functions, and operations of the product. Read this manual carefully before operating your analyzer and operate your analyzer strictly as instructed in this manual.

### NOTE

- **This manual describes the use, functions and operation methods of the product based on the most complete configuration; and some of the content may not be applicable to your product. Contact Mindray if you have any questions.**

## 2.2 Who Should Read This Manual

This manual is intended to be read by clinical laboratory professionals to:

- learn about the analyzer hardware and software;
- set up system parameters;
- perform daily operating task;
- perform system maintenance and troubleshooting

## 2.3 How to Find Information

This operator's manual comprises 13 chapters and 7 appendices. Refer to the table below to find the information you need.

If you want to...	See...
learn about the safety messages of the analyzer	<b>1Safety Information</b>
learn about the intended use and parameters of the analyzer	<b>3Understanding Your System</b>
learn about the analyzer's system composition and software structure	<b>3Understanding Your System</b>
learn about how the analyzer works	<b>4Understanding the System Principles</b>
learn about the installation requirements of the analyzer	<b>5Installing and Connecting the System</b>
learn about how to define/adjust system settings	<b>6Customizing the Analyzer Software</b>
learn about how to collect, prepare and analyze the samples	<b>7Operating Your Analyzer</b>
learn about how to use the analyzer to perform daily operating tasks	<b>7Operating Your Analyzer</b>
learn about how to review the saved analysis results	<b>8Reviewing Sample Results</b>
learn about basic requirements of quality control and quality control methods of the analyzer	<b>9Using the QC Program</b>
learn about basic requirements of calibration and calibration methods of the analyzer	<b>10Calibrating Your Analyzer</b>
learn about how to use the quality control programs of the analyzer	<b>11Printing</b>
learn about how to maintain/test the analyzer	<b>12Service</b>
learn about the troubleshooting methods of the analyzer	<b>13Troubleshooting</b>

If you want to...	See...
learn about the technical specifications of the analyzer	<b><i>BSpecification</i></b>

## 2.4 Conventions Used in This Manual

This manual uses certain typographical conventions to clarify meaning in the text:

Format	Meaning
[xx]	All capital letters enclosed in [ ] indicate a key name (either on the pop-up keyboard or the external keyboard), such as [ENTER].
"xx"	Letters included in " " indicate text you can find on the screen of the analyzer.

All illustrations in this manual are provided as examples only. They may not necessarily reflect your analyzer setup or data displayed on the screen of the analyzer.

# 3 Understanding Your System

## 3.1 Intended Use

The Auto Hematology Analyzer is a quantitative, automated hematology analyzer for In Vitro Diagnostic Use in clinical laboratories. It provides Complete Blood Count, Leukocyte Differential, Hemoglobin Concentration Measurement, Reticulocyte Measurement, Nucleated Red Blood Cell Measurement and Erythrocyte Sedimentation Rate Measurement for blood samples, and provides WBC and RBC counts, and WBC Differential Measurement for body fluid samples.

Erythrocyte Sedimentation Rate (ESR) is a non-specific marker used for auxiliary diagnosis of certain pathological conditions, mainly including inflammation, injury, rheumatism, etc.

Model	Configuration Differences		
	RET	ESR	Data storage capacity
BC-760[B]		●	130,000
BC-760[R]	●	●	130,000
BC-780[R]	●	●	150,000

### NOTE

- The subtypes of body fluid supported by the analyzer include cerebrospinal fluid, pleural fluid, ascitic fluid, and synovial fluid. You may not get accurate results if you analyze body fluid samples other than specified above.
- The analyzer identifies the normal patient, with all normal system-generated parameters for In Vitro Diagnostic Use. The product flags or identifies patient results that require additional studies.

## 3.2 Test Parameters

The analyzer is used for quantitative determination of the following report parameters and Research Use Only (RUO) parameters of blood sample tests and body fluid sample tests, and provides histograms and scattergrams.

### 3.2.1 Blood Sample Test Parameters, Histograms, and Scattergrams

**Table 3-1 Blood sample test report parameters**

Group	Name	Abbreviation	Applicable Model
WBC group	White blood cell count	WBC	General
	Basophil count	Bas#	General
	Basophil percentage	Bas%	General
	Neutrophil count	Neu#	General
	Neutrophil percentage	Neu%	General
	Eosinophil count	Eos#	General
	Eosinophil percentage	Eos%	General
	Lymphocyte count	Lym#	General
	Lymphocyte percentage	Lym%	General
	Monocyte count	Mon#	General
	Monocyte percentage	Mon%	General
	Immature granulocyte count	IMG#	General
	Immature granulocyte percentage	IMG%	General
RET group	*Reticulocyte percentage	RET%	BC-760[R] BC-780[R]
	*Reticulocyte count	RET#	
	*Reticulocyte hemoglobin expression	RHE	
	*Immature reticulocyte fraction	IRF	
	*Low fluorescent ratio	LFR	
	*Middle fluorescent ratio	MFR	
	*High fluorescent ratio	HFR	
RBC group	Red blood cell count	RBC	General
	Hemoglobin concentration	HGB	General
	Mean corpuscular volume	MCV	General
	Mean corpuscular hemoglobin	MCH	General
	Mean corpuscular hemoglobin concentration	MCHC	General
	Red blood cell distribution width coefficient of variation	RDW-CV	General
	Red blood cell distribution width standard deviation	RDW-SD	General
	Hematocrit	HCT	General
	Nucleated red blood cell count	NRBC#	General
	Nucleated red blood cell percentage	NRBC%	General

Group	Name	Abbreviation	Applicable Model
Platelet group	Platelet count	PLT	General
	Mean platelet volume	MPV	General
	Platelet distribution width	PDW	General
	Plateletcrit	PCT	General
	Platelet-large cell ratio	P-LCR	General
	Platelet-large cell count	P-LCC	General
	*Immature platelet fraction	IPF	BC-760[R] BC-780[R]this parameter is optional, for more information, consult Mindray Customer Service DepartmentFor BC-760[B], this parameter is optional, for more information, consult Mindray Customer Service Department
	Platelet count- impedance	PLT-I	General (find the parameter result on the "Other Para." - "Analysis Para.")
	*Platelet count hybrid	PLT-H	BC-760[R] BC-780[R] (find the parameter result on the "Other Para." - "Analysis Para.")For BC-760[B], this parameter is optional. For more information, consult Mindray Customer Service Department
	*Optical platelet count	PLT-O	BC-760[R] BC-780[R] (find the parameter result on the "Other Para." - "Analysis Para.")
ESR	Erythrocyte sedimentation rate	ESR	General

## NOTE

- The parameters marked with \* are parameters for specified models.

**Table 3-2 Blood sample test RUO parameters**

Name	Abbreviation	Applicable Model
High fluorescent cell count	HFC#	General
High fluorescent cell percentage	HFC%	General
White blood cell count -DIFF	WBC-D	General
Total nucleated cell count-DIFF	TNC-D	General
Immature eosinophil percentage	IME%	General

Name	Abbreviation	Applicable Model
Immature eosinophil count	IME#	General
Neutrophil-to-lymphocyte ratio	NLR	General
Infected red blood cell count	InR#	General
Infected red blood cell permillage	InR‰	General
NEU# Minus IMG#	Neu#&	General
NEU% Minus IMG%	Neu%&	General
LYM# Minus HFC#	Lym#&	General
LYM% Minus HFC%	Lym%&	General
Microcyte count	Micro#	General
Microcyte percentage	Micro%	General
Macrocyte count	Macro#	General
Macrocyte percentage	Macro%	General
*Optical red blood cell count	RBC-O	BC-760[R] BC-780[R]
*Optical white blood cell count	WBC-O	
*RBC fragment count	FRC#	
*RBC fragment percentage	FRC%	
*Reticulocyte production index	RPI	
*Mean reticulocyte volume	MRV	
*RET scattergram, mean reticulocyte distribution-forward scatter intensity	RET-Y	
*RET scattergram, mean reticulocyte distribution-side fluorescent intensity	RET-X	
*RET scattergram, mean immature reticulocyte fraction distribution-forward scatter intensity	IRF-Y	
*RET scattergram, mean immature reticulocyte fraction distribution-side fluorescent intensity	IRF-X	
*RET scattergram, mean red blood cell distribution-forward scatter intensity	RET-RBC-Y	General
*RET scattergram, mean red blood cell distribution-side fluorescent intensity	RET-RBC-X	
High fluorescent immature platelet fraction	H-IPF	
Immature platelet count	IPF#	
Platelet-to-lymphocyte ratio	PLR	
Platelet distribution width standard deviation	PDW-SD	
Dimorphic population, smaller distribution RBC count	SRBC	
Dimorphic population, larger distribution RBC count	LRBC	
Dimorphic population, smaller distribution mean corpuscular volume	SMCV	
Dimorphic population, larger distribution mean corpuscular volume	LMCV	



Name	Abbreviation	Applicable Model
DIFF scattergram, mean neutrophil distribution-side scatter intensity	Neu-X	General
DIFF scattergram, mean neutrophil distribution-side fluorescent light intensity	Neu-Y	General
DIFF scattergram, mean neutrophil distribution-forward scatter intensity	Neu-Z	General
DIFF scattergram, mean lymphocyte distribution-side scatter intensity	Lym-X	General
DIFF scattergram, mean lymphocyte distribution-side fluorescent intensity	Lym-Y	General
DIFF scattergram, mean lymphocyte distribution-forward scatter intensity	Lym-Z	General
DIFF scattergram, mean monocyte distribution-side scatter intensity	Mon-X	General
DIFF scattergram, mean monocyte distribution-side fluorescent light intensity	Mon-Y	General
DIFF scattergram, mean monocyte distribution-forward scatter intensity	Mon-Z	General
DIFF scattergram, neutrophil side scatter distribution width	Neu-XW	General
DIFF scattergram, neutrophil side fluorescent light distribution width	Neu-YW	General
DIFF scattergram, neutrophil forward scatter distribution width	Neu-ZW	General
DIFF scattergram, lymphocyte side scatter distribution width	Lym-XW	General
DIFF scattergram, lymphocyte side fluorescent light distribution width	Lym-YW	General
DIFF scattergram, lymphocyte forward scatter distribution width	Lym-ZW	General
DIFF scattergram, monocyte side scatter distribution width	Mon-XW	General
DIFF scattergram, monocyte side fluorescent light distribution width	Mon-YW	General
DIFF scattergram, monocyte forward scatter distribution width	Mon-ZW	General
*Immature platelet fraction- DIFF	IPF-D	<p>These parameters are available on BC-760[R]/BC-780[R] (when the user-defined test panel includes DIFF tests, but not Ret tests.)</p> <p>For BC-760[B], this parameter is optional. For more information, consult Mindray Customer Service Department</p>
*Reticulocyte percentage- DIFF	RET%-D	
*Reticulocyte count- DIFF	RET#-D	
*Immature reticulocyte fraction- DIFF	IRF-D	
*Low fluorescent ratio- DIFF	LFR-D	
*Middle fluorescent ratio- DIFF	MFR-D	
*High fluorescent ratio- DIFF	HFR-D	
Corrected erythrocyte sedimentation rate	ESR-Corr.	General
Surface area	SA	General
Amplitude	AMP	General

Name	Abbreviation	Applicable Model
Aggregation index	AI	General
Minimum	MIN	General
Aggregation half time	T1/2	General

**NOTE**

- RUO parameters are for research purpose only. They cannot be used for diagnosis purpose.
- The parameters marked with \* are parameters for specified models.
- To print the RUO parameter results, tap "Setup"->"System Setup"->"Print Setup"->"Printing Content", and check "Print RUO Parameters".

**Table 3-3 Blood sample test histograms**

Name	Abbreviation	Applicable Model
Red blood cell histogram	RBC histogram	General
Platelet histogram	PLT histogram	General
* Platelet hybrid histogram	PLT-H histogram	BC-760[R] BC-780[R] For BC-760[B], this histogram is optional. For more information, consult Mindray Customer Service Department.
White blood cell -FSC histogram	WBC-FSC histogram	General

**NOTE**

- The parameters marked with \* are parameters for specified models.

**Table 3-4 Blood sample test scattergrams**

Name	Abbreviation	3D Scattergram	Applicable Model
Differential(FS/FL) scattergram	DIFF(FS/FL) scattergram	Available	General
Differential(FS/SS) scattergram	DIFF(FS/SS) scattergram		General
Differential(FL/SS) scattergram	DIFF(FL/SS) scattergram		General
*Reticulocyte scattergram	RET scattergram	Available	BC-760[R] BC-780[R]
*Optical platelet scattergram	PLT-O scattergram	Unavailable	
*Reticulocyte - extension scattergram	RET-EXT scattergram	Unavailable	
*Platelet hybrid scattergram	PLT-H scattergram	Available	BC-760[R] BC-780[R] For BC-760[B], this scattergram is optional. For more information, consult Mindray Customer Service Department.

**NOTE**

- The parameters marked with \* are parameters for specified models.

**3.2.2 Body Fluid Sample Test Parameters, Histograms, and Scattergrams****Table 3-5 Body fluid sample test report parameters**

Group	Name	Abbreviation	Applicable Model
WBC group	White blood cell count-body fluid	WBC-BF	General
	Total nucleated cell counts-body fluid	TC-BF#	General
	Mononuclear cell count	MN#	General
	Mononuclear cell percentage	MN%	General
	Polymorphonuclear cell count	PMN#	General
	Polymorphonuclear cell percentage	PMN%	General
RBC group	Red blood cell count-body fluid	RBC-BF	General

**Table 3-6 Body fluid sample test RUO parameters**

Name	Abbreviation	Applicable Model
Eosinophil count- body fluid	Eos-BF#	General
Eosinophil percentage- body fluid	Eos-BF%	General
Neutrophil count- body fluid	Neu-BF#	General
Neutrophil percentage- body fluid	Neu-BF%	General
Lymphocyte count- body fluid	Lym-BF#	General
Lymphocyte percentage- body fluid	Lym-BF%	General
Monocyte count- body fluid	Mon-BF#	General
Monocyte percentage- body fluid	Mon-BF%	General
High fluorescent cell count - body fluid	HFC-BF#	General
High fluorescent cell percentage- body fluid	HFC-BF%	General
Red blood cell count-body fluid	RBC-BF(R)	General

**NOTE**

- RUO parameters are for research purpose only. They cannot be used for diagnosis purpose.
- To print the RUO parameter results, tap "Setup"-"System Setup"-"Print Setup"-"Printing Content", and check "Print RUO Parameters".

**Table 3-7 Body fluid sample test histograms**

Name	Abbreviation	Applicable Model
Red blood cell histogram	RBC histogram	General
White blood cell-FSC histogram	WBC-FSC histogram	General

**Body fluid sample test scattergrams**

Name	Abbreviation	3D Scattergram	Applicable Model
Differential scattergram	DIFF scattergram	Available	General
Differential-extension scattergram	DIFF-EXT scattergram	Unavailable	General

### 3.3 Device Description

#### 3.3.1 Structure and Components

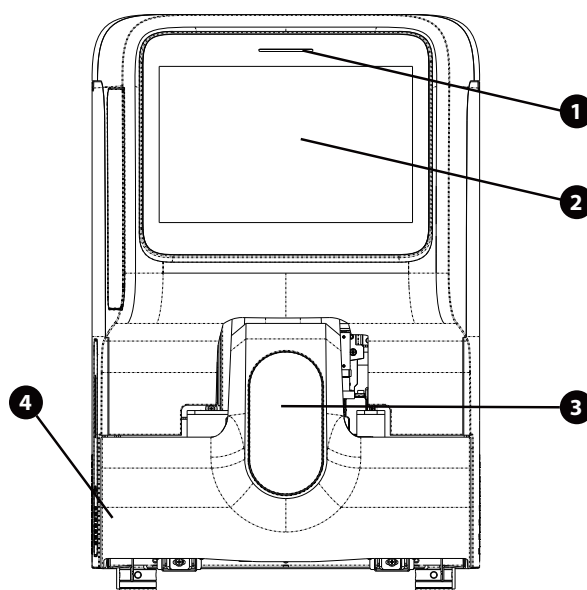
BC-760[B]/BC-760[R]/BC-780[R] Auto Hematology Analyzer consists of the sample processing unit, data management unit, result output unit, and the accessories.

#### 3.3.2 Modules and Components

**⚠ CAUTION**

- Do not use anything sharp on the touch screen or strike on it.
- Clean the touch screen with a clean and soft cloth, as well as clean water, rather than chemical solution, acid or alkali solution.

##### 3.3.2.1 Front of the analyzer

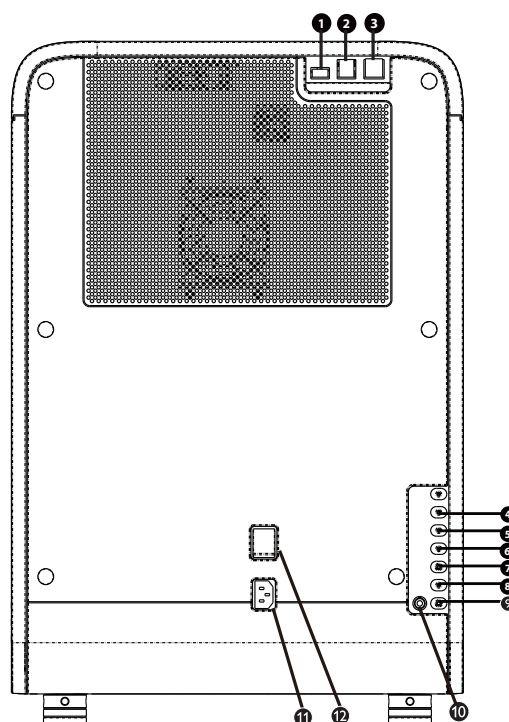


**Figure 3-1 Front of the analyzer**

- |                      |                |
|----------------------|----------------|
| ① Status indicator   | ② Touch screen |
| ③ Sample compartment | ④ Autoloader   |

①	Status Indicator	The indicator locates on the top of the touch screen; and it tells you about the status of the instrument including ready, running, error, standby and on/off, etc.	Ready: indicator stays in green
			Running: indicator flickers in green
			Sampling probe piercing: indicator flickers fast
			Error: indicator stays in red
			Sleep: indicator stays in orange
			Off: indicator off
②	Touch screen	The touch screen locates on the front of the main unit, which can be used to operate the instrument and display information.	/
③	Sample compartment	Tap closed-tube sampling mode on touch screen, and the analyzer opens the sample compartment automatically. Place applicable adapters in the compartment and sample tubes to perform analysis tests.	/
④	Autoloader	The autoloader is in the front of the analyzer. You can use it to load tubes automatically.	/

### 3.3.2.2 Back of the analyzer



**Figure 3-2 Back of the analyzer**

① USB port (protocol 3.0)

③ Network interface

⑤ LD Lyse inlet

② USB port (protocol 2.0)

④ DR Diluent inlet (applicable to the BC-760[R]/BC-780[R] model)

⑥ LH Lyse inlet

⑦ ESR Solution Reagent inlet

⑨ Waste sensor

⑪ Power inlet

⑧ DS Diluent inlet

⑩ Waste outlet

⑫ Power switch

### 3.3.2.3 Right side view of the analyzer

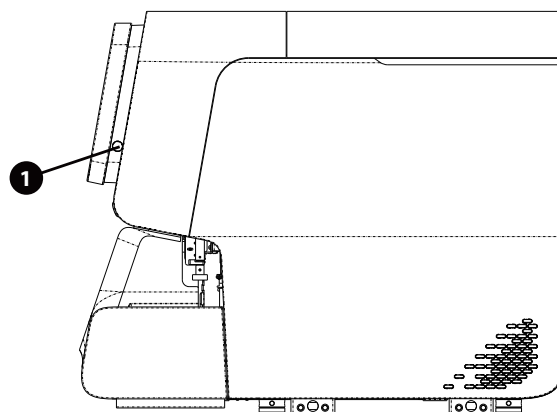


Figure 3-3 Right side view of the analyzer

① Standby

### 3.3.2.4 Left side view of the analyzer

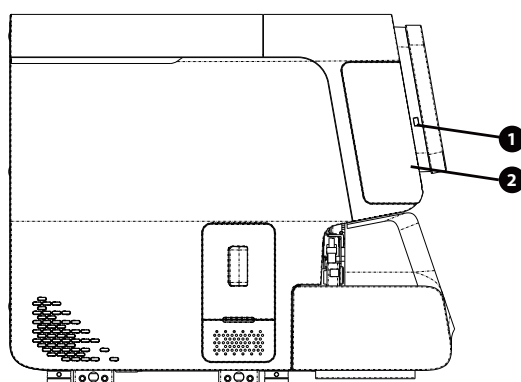


Figure 3-4 Left side of the analyzer

① USB port (protocol 2.0)

② Dye replacing compartment

## 3.3.3 Accessories

The analyzer is provided with the following configured or optional accessories.

Table 3-8 List of Accessories

	Configured	Optional
Handheld Barcode Scanner		✓
HP LaserJet Printer	✓	
Tube Rack Assembly (CAL 8000)	✓	

	Configured	Optional
Capillary blood tube rack assembly		✓

**NOTE**

- For any questions about the configured/optional accessories, consult your sales representative.
- The accessories actually attached to the product depend on your product configuration.

**CAUTION**

- Only use the accessories and consumables manufactured or recommended by Mindray to achieve the promised system performance and safety. For more information, contact Mindray Customer Service Department or your local distributor.
- Use the original power cord provided by the manufacturer. Using other electrical wire may damage the system or lead to unreliable analysis results.

## 3.4 Supported Tubes, Tube Racks and Adapters

### 3.4.1 Supported Tubes

**NOTE**

- For the use of the tubes not mentioned hereof, please contact our customer service department or the local distributor.

#### 3.4.1.1 Specifications of tubes and recommended sample volume in AL-WB mode

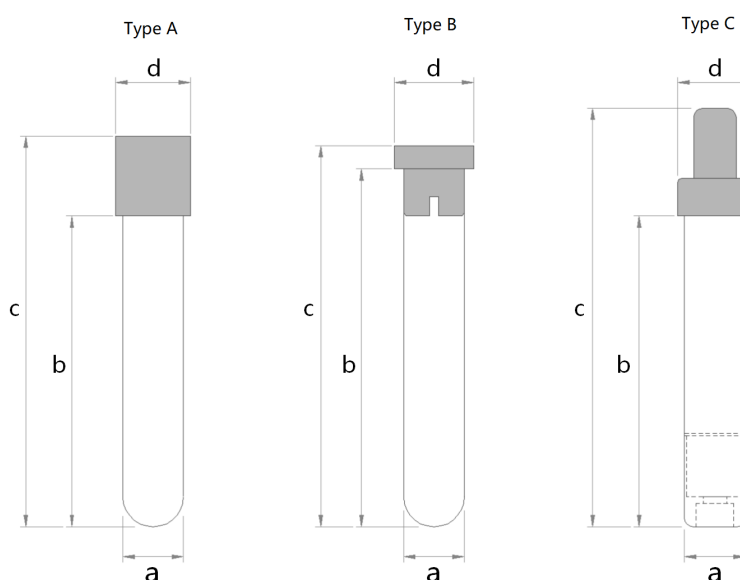


Figure 3-5 Specifications of tubes supported in AL-WB mode

Tube Diameter (a)	$11\text{ mm} \leq a \leq 15\text{ mm}$
Length Excluding Tube Cap (b)	$b \geq 58\text{ mm}$
Length Including Tube Cap (c)	$60\text{ mm} \leq c \leq 91\text{ mm}$

Tube Cap Diameter (d)	$(a+1\text{mm}) \leq d \leq 18\text{mm}$
-----------------------	--

Table 3-9 Recommended sample volume in AL-WB mode

Tube Diameter	Recommended Blood Sample Volume
$11\text{ mm} \leq a \leq 13\text{mm}$	$\geq 0.9\text{ mL}$
$13\text{ mm} < a \leq 15\text{mm}$	$\geq 2.0\text{ mL}$

Table 3-10 Tube types supported in different sampling mode

Sampling mode	Supported tube type
CT-WB	Type A and Type B
AL-WB	Type A, B and C

**NOTE**

- In CT-WB mode, do not use Sarstedt Monovette tubes for analysis.

### 3.4.1.2 Specifications of tubes and recommended sample volume supported in AL-Micro WB mode

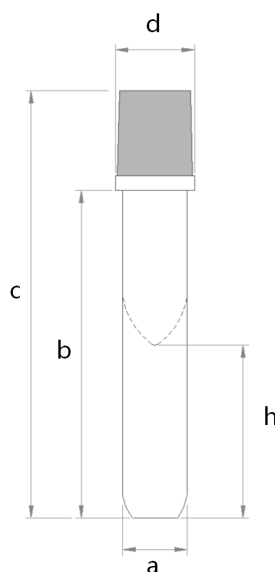


Figure 3-6 Specifications of tubes supported in AL-Micro WB mode

Tube Diameter (a)	$11.6\text{mm} \leq a \leq 12.6\text{mm}$
Length Under Tube Cap or Bulge (b)	$b \geq 58\text{mm}$
Length Including Tube Cap (c)	$60\text{ mm} \leq c \leq 85\text{mm}$
Diameter of Tube Cap or Bulge (d)	$(a+1\text{mm}) \leq d \leq 18\text{mm}$
Spacing Between Cavity Bottom and Tube Bottom (h)	$20\text{mm} \leq h \leq 36.5\text{mm}$



Mindray has performed tests on following tubes:

Manufacturer	Recommended sample volume
Kangjian KJ001-1	≥ 80μL
Xinle D type K <sub>2</sub> EDTA	≥ 80μL

#### NOTE

- The cap of the long micro WB tubes supported in AL-Micro WB mode must be pierceable and supports the pressure balancing function.

#### 3.4.1.3 Specifications of tubes and recommended sample volume supported in CT-BF mode:

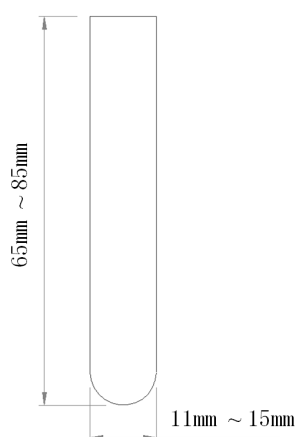
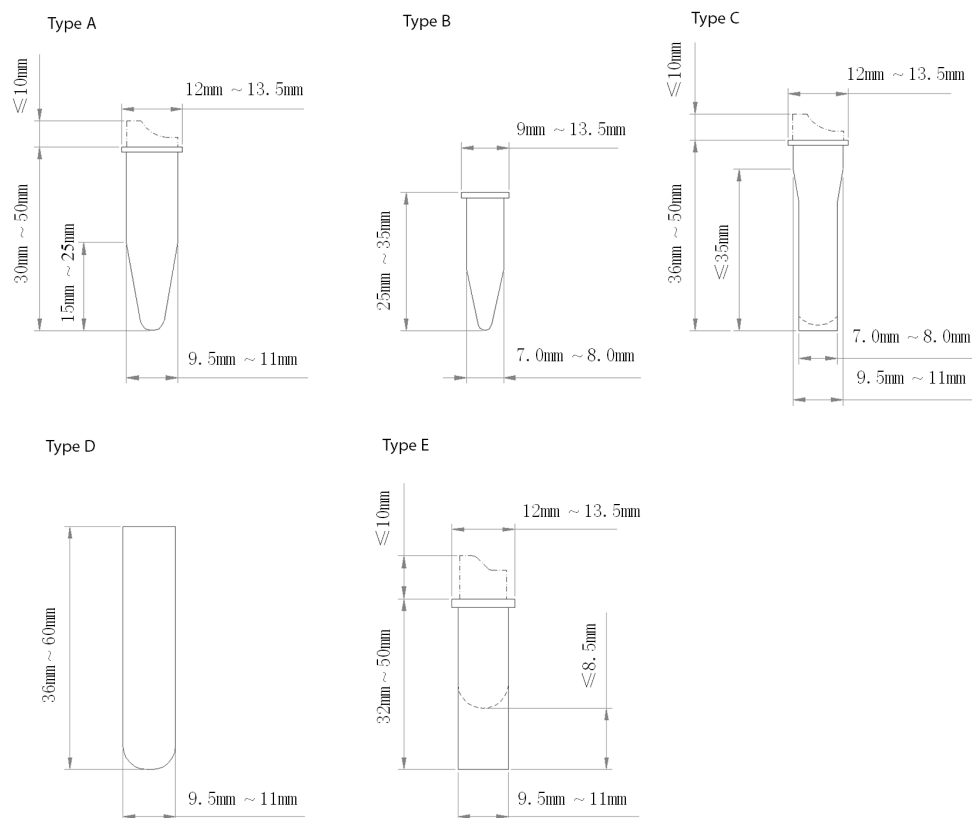


Figure 3-7 Specifications of cap-free tubes supported in CT-BF mode

Table 3-11 Recommended sample volumes supported in CT-BF mode

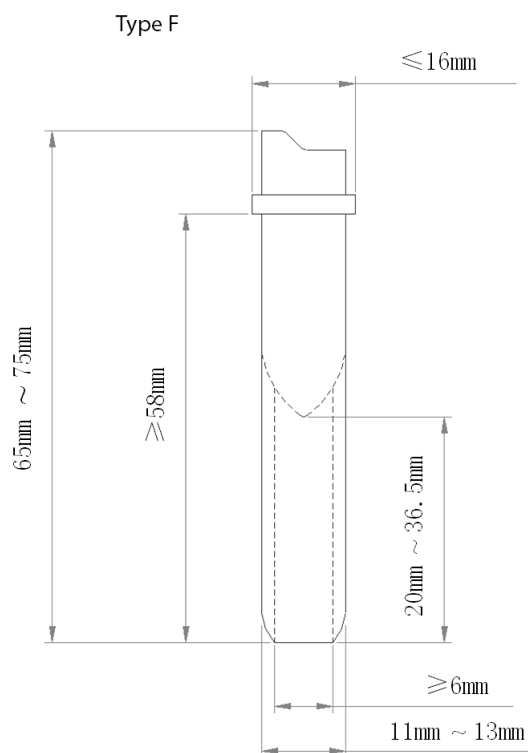
Tube Diameter	Recommended Sample Volume
11 mm~13mm	≥1.0 mL
13 mm~15mm	≥ 2.0 mL



**Figure 3-8 Specification of tubes supported in CT-Micro WB mode (type A - E)**

**Table 3-12 Recommended sample volume for tubes in CT-Micro WB mode (type A - E)**

Type	Recommended sample volume
A	$\geq 120\mu\text{L}$
B	$\geq 110\mu\text{L}$
C	$\geq 150\mu\text{L}$
D	$\geq 200\mu\text{L}$
E	$\geq 200\mu\text{L}$



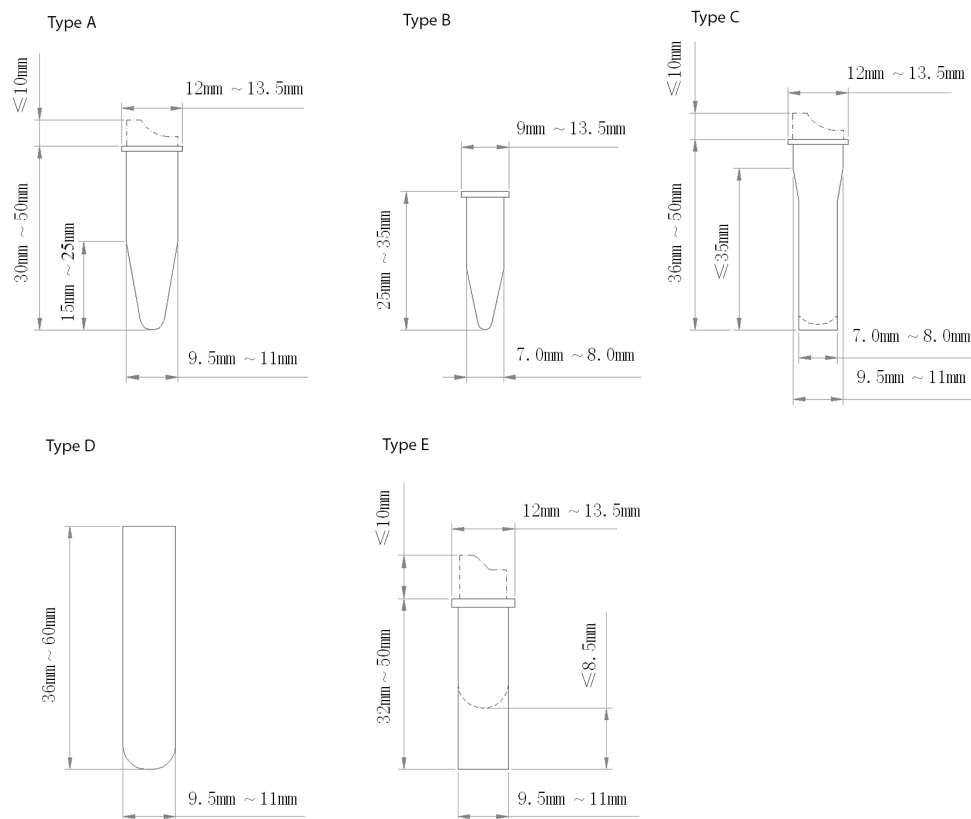
**Figure 3-9 Specification of tubes supported in CT-BF mode (type F)**

Mindray has performed tests on following type F tubes:

**Table 3-13 Recommended sample volume for long tubes in CT-Micro WB mode (type F)**

Type	Manufacture	Model	Recommended Blood Sample Volume
F	BD (the United States)	Microtainer MAP	$\geq 200\mu\text{L}$
	Guangzhou Yangpu	IMPROMINI MAP only	$\geq 200\mu\text{L}$
	Zhejiang Gongdong	GD005EK2S	$\geq 150\mu\text{L}$
	Jiangsu Kangjian	KJ001-1	$\geq 170\mu\text{L}$

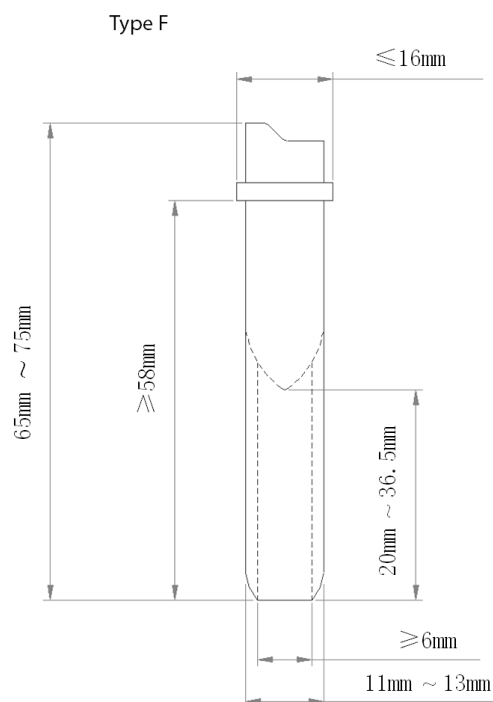
### 3.4.1.4 Specifications of tubes and recommended sample volume supported in CT-Micro WB mode



**Figure 3-10 Specifications of short tubes supported in CT-Micro WB mode (type A - E)**

**Table 3-14 Recommended sample volume for short tubes in CT-Micro WB mode (type A - E)**

Type	Recommended Blood Sample Volume
A	70 $\mu$ L
B	60 $\mu$ L
C	100 $\mu$ L
D	120 $\mu$ L
E	100 $\mu$ L



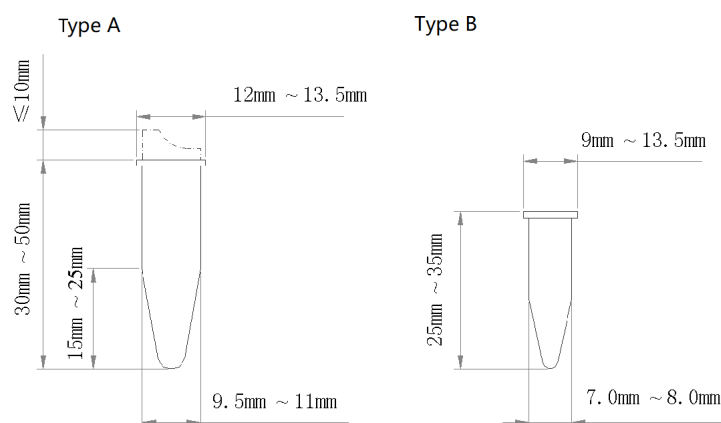
**Figure 3-11 Specifications of long tubes supported in CT-Micro WB mode (type F)**

Mindray has performed tests on following type F tubes:

**Table 3-15 Recommended blood sample volume for long micro tube in CT-Micro WB mode (type F)**

Type	Manufacture	Model	Recommended Blood Sample Volume
F	BD (the United States)	Microtainer MAP	120μL
	Guangzhou Yangpu	IMPROMINI MAP only	100μL
	Zhejiang Gongdong	GD005EK2S	80μL
	Jiangsu Kangjian	KJ001-1	80μL

### 3.4.1.5 Specifications of tubes and recommended sample volume supported in CT-PD mode:



**Figure 3-12 Specification of tubes supported in CT-PD mode**

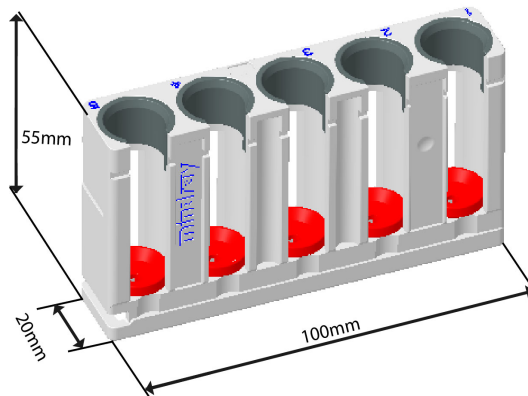
Recommended blood sample volume for short tube: at least 20 $\mu$ L.

### 3.4.2 Supported Tube Racks

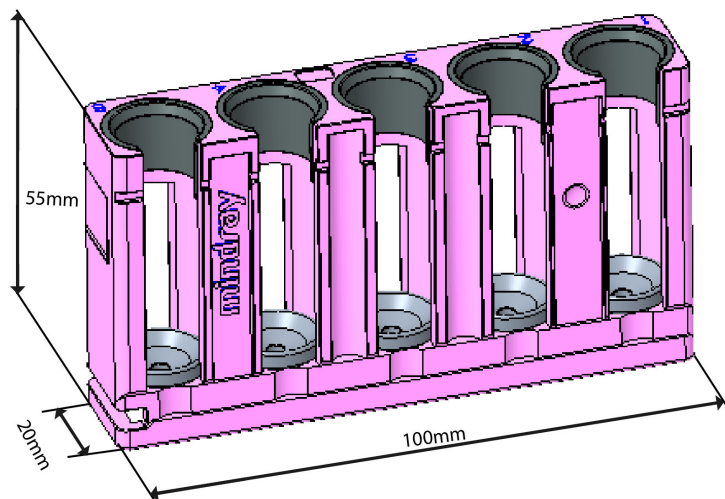
#### NOTE

- Please use the tube racks specified by the manufacturer only.

The specifications of the tube racks supported by the analyzer are shown in the figure below:



**Figure 3-13 Tube rack for regular tubes in AL-WB mode (5 positions)**



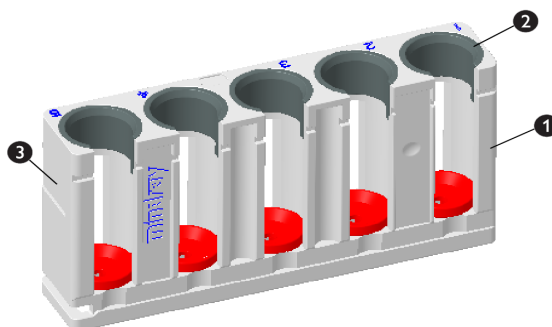
**Figure 3-14 Tube rack for AL-Micro WB mode (5 positions)**

In the auto-loading mode, the downward depth of the sampling probe may vary for the different samples in the tubes with different specifications. During installation, Mindray Service Engineer will set the downward depth of sampling probe for the tube racks with different number according to the tube types in your laboratory. Users may recognize the purposes of different tube racks based on their characteristics and number.

#### CAUTION

- The tube racks for different modes cannot be misused, otherwise, the sampling probe may fail to aspirate the sample due to insufficient downward depth, be damaged or touch the tube bottom and damage the tube due to excessive downward depth.

- To change the tube rack number, please contact Mindray Customer Service Department.



1	Tube racks
2	Tube position adapters
3	Tube rack numbers

**Table 3-16 Types of tube rack**

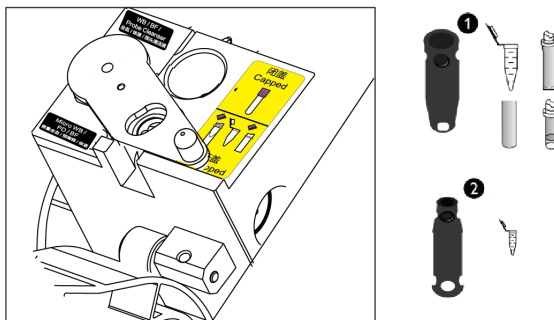
	Supported tube types	Tube rack characteristics	Other Information
Tube rack 1 for AL-WB mode	For Type A and B, refer to <b>Figure 3-5 Specifications of tubes supported in AL-WB mode</b>	White tube racks; gray inserts	Specification of tube rack depends on the analyzer model.
Tube rack 2 for AL-WB mode	Refer to <b>Figure 3-5 Specifications of tubes supported in AL-WB mode.</b>	White tube racks; colored inserts ((customization supported)	<ul style="list-style-type: none"> <li>• Confirm that the supported tubes include BD Microtainer MAP Microtube and Sarstedt Monovette tube.</li> <li>• In case of analysis with BD Microtainer MAP Microtube and Sarstedt Monovette, it is forbidden to use the different tube racks at the same time.</li> </ul>
Tube racks for AL-Micro WB mode	Refer to <b>Figure 3-6 Specifications of tubes supported in AL-Micro WB mode</b>	The tube racks are pink The inserts are white	Specification of tube rack depends on the analyzer model.

### 3.4.3 Supported Adapters

The analyzer may use the adapters for the tube positions of Micro-WB/prediluted samples and the adapters for the tube positions of calibrators.

### 3.4.3.1 Adapters for the tube positions of Micro-WB/prediluted samples

When analyzing samples under CT-Micro WB/Predilute mode, the adapters are placed in the tube positions of CT-Micro WB/prediluted samples in the sample compartment to match with the tubes of different specifications.




**Figure 3-15 Adapters for sample compartment**

1	The adapters with the larger-diameter end upward
2	The adapters with the smaller-diameter end upward


#### NOTE

- If the sample to be analyzed is micro whole blood or prediluted sample, be sure to uncap the tube before analysis.

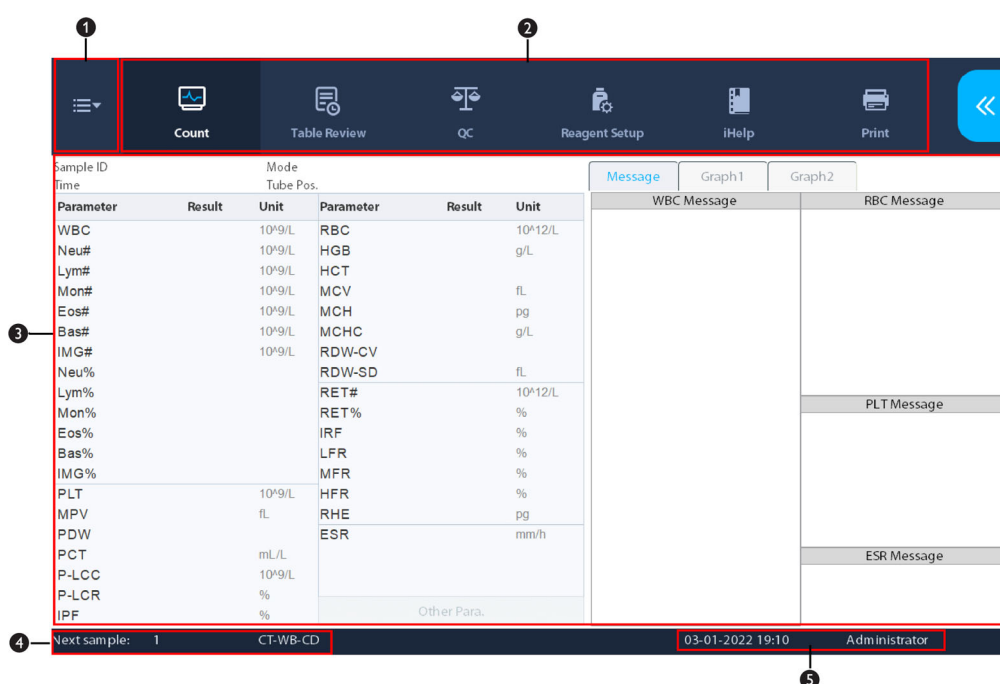
**Table 3-17 Tube position and adapters of sample compartment**

Tube Position	Adapter	Supported Mode	Applicable Tubes
WB/BF/Probe Cleanser Tube Position	/	CT-WB	Tube type A and tube type B (capped tube), as shown in <b>Figure 3-5 Specifications of tubes supported in AL-WB mode</b>
		CT-BF	As shown in <b>Figure 3-7 Specifications of cap-free tubes supported in CT-BF mode</b> .
Micro-WB/PD/BF Tube Position	/	CT-Micro WB	Tube type F, as shown in <b>Figure 3-7 Specifications of cap-free tubes supported in CT-BF mode</b> .
		CT-BF	
	 (the adapter with the larger-diameter end upward)	CT-Micro WB	Tube types A, C, D and E, as shown in <b>Figure 3-10 Specifications of short tubes supported in CT-Micro WB mode (type A - E)</b> .
		CT-PD	Tube type A, as shown in <b>Figure 3-12 Specification of tubes supported in CT-PD mode</b> .
		CT-BF	Tube types A, C, D and E, as shown in <b>Figure 3-10 Specifications of short tubes supported in CT-Micro WB mode (type A - E)</b> .




Tube Position	Adapter	Supported Mode	Applicable Tubes
	 (the adapter with the smaller-diameter end upward)	CT-Micro WB	Tube type B, as shown in <b>Figure 3-10 Specifications of short tubes supported in CT-Micro WB mode (type A - E)</b> .
		CT-PD	
		CT-BF	

### 3.5 Overview of Software Interfaces



#### ① Menu

Tap the  button at the top left of the software screen to display the system menu.

The following table lists the menu functions of the software system:

No.	Level 1	Level 2	Level 3
1	Count	/	/
2	Table Review	/	/
3	QC	L-J QC	Setup
			Count
		X-B QC	Setup
			Graph





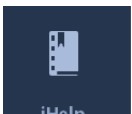

No.	Level 1	Level 2	Level 3
4	<b>Calibration</b>	<b>Manual</b>	/
		<b>ESR</b> (Administrator)	/
		<b>Calibrator</b> (Administrator)	/
		<b>Fresh Blood</b> (Administrator)	/
		<b>Calibration History</b> (Administrator)	/
5	<b>Status</b>	<b>Statistics</b>	/
		<b>Temp.&amp;Pressure</b> (Administrator)	/
		<b>Floater Status</b> (Administrator)	/
		<b>Sensor</b> (Administrator)	/
		<b>Voltage &amp; Current</b> (Administrator)	/
		<b>Version Info.</b>	/
6	<b>Setup</b>	<b>System Setup</b>	<b>Print Setup</b>
			<b>Communication</b> (Administrator)
			<b>Barcode</b> (Administrator)
			<b>Date/Time Setup</b>
			<b>Lab Info. Setup</b>
			<b>Flag Alarm Sensitivity</b> (Administrator)
			<b>Flag Rules Setup</b> (Administrator)
			<b>Tube Rack Type Setup</b>
			<b>Ext. PLT Tests</b> (Administrator)
		<b>User Management</b>	/
		<b>Auxiliary Setup</b>	/
		<b>Para. Setup</b>	<b>Parameter Unit Setup</b>
			<b>Ref. Range Setup</b>
			<b>Microsco. Para. Setup</b> (Administrator)
		<b>Maintenance</b> (Administrator)	/
		<b>Reagent Setup</b>	/
		<b>Auto-loading</b> (Administrator)	/
		<b>Gain Setup</b> (Administrator)	/
		<b>Re-exam Rules Setup</b> (Administrator)	/
		<b>Auto Startup/Shutdown</b> (Administrator)	/
7	<b>Service</b>	<b>Debug &amp; Self-Test</b>	<b>Self-Test</b>
		<b>Precise Fault Diagnosis</b> (Administrator)	/
		<b>iHelp</b>	/
		<b>Maintenance</b>	/
		<b>Screen Cal.</b>	/
		<b>Log</b>	/
8	<b>Logout</b>	/	/

No.	Level 1	Level 2	Level 3
9	Shutdown	/	/

## NOTE

- When a function is followed by "administrators", it means the function is only available to operators at administrator's level.

### ■ ② Utility Button area

Name	Icon	Functions
Count		Tap to enter the "Count" screen.
Table Review		Tap to enter the "Table Review" screen.
QC		Tap to enter to enter the "QC" screen. When the "QC" button lighted in orange, it means the analyzer is out of quality control
Reagent Setup		Tap to enter the "Reagent Setup" screen, check or set the remaining volumes and expiration date, and replace the reagent; When the "Reagent Setup" button lights in orange, it means some reagent is expired or not sufficient
iHelp		Tap to enter the "iHelp" screen.
Print		<ul style="list-style-type: none"> <li>When the analyzer is on the "Count" screen, tap the "Print" button to print the analysis results, histograms and scattergrams of the current sample in accordance with operator-customized print template</li> <li>When the analyzer is on the "Table Review" screen, tap the "Print" button to print the analysis results for all or selected samples in the table print or graph print form</li> <li>When the analyzer is on the "Graph" screen, tap the "Print" button to print the analysis results, histograms and scattergrams of the current sample in accordance with operator-customized print template</li> <li>When the analyzer is on the "QC Table" screen, tap the "Print" button to print all QC results included in the selected QC file</li> <li>When the analyzer is on the "QC Graph" screen, tap the "Print" button to print the QC graphs included in the selected QC file</li> <li>When the analyzer is on the "Manual" screen, tap "Print" to print the manual calibration factors</li> </ul>

### ■ ① Operation area

Displays contents of the screens.

For example, on the "Count" screen, the area displays function buttons related to sample analysis as well as the sample analysis results.

### ■ ② Auxiliary information area

This area displays auxiliary information of the current screen;

For example, on the "Count" screen, the area displays the ID and analysis mode of the next sample; on the "Table Review" or "Graph" screen, the area displays position of the current sample and total number of samples

### ■ ③ Other information

It displays the current system time;

When error occurs, the area displays the error message;

If you log in as an administrator, the area displays "Administrator".

## 3.6 Reagents, Controls and Calibrators

As the analyzer, reagents, controls and calibrators are components of a system, performance of the system depends on the combined integrity of all components. You must only use the Mindray-specified reagents, controls, and calibrators which are formulated specifically for the fluidic system of your analyzer in order to provide optimal system performance. Do not use the analyzer with reagents, controls, and calibrators from multiple suppliers. In such case, the analyzer may not meet the performance specified in this manual and may provide unreliable results.

All the reagents, controls, and calibrators mentioned in this manual refer to the reagents, controls, and calibrators specifically formulated for this analyzer. You must buy those reagents, controls, and calibrators from Mindray or Mindray-authorized distributors. When you need to buy reagents and consumables, please call Mindray Customer Service Department.

### 3.6.1 Reagent

All reagents used by this analyzer are specifically matched with Mindray equipment. Use for any other purpose is prohibited.

Correctly use and store reagents according to their Instructions for Use.

#### NOTE

- **For the reagent model, intended use, test principle, main components, storage conditions, expiration date, applicable models, and other information, please refer to the reagent IFU.**
- **For any questions related to reagents, controls, and calibrators, please consult your sales representatives.**

**Table 3-18 Reagents**

Applicable Channel	BC-760[B]	BC-760[R]/BC-780[R]
HGB Channel	LH Lyse	LH Lyse
DIFF Channel	LD Lyse	LD Lyse
	FD Dye	FD Dye
RET Channel	/	*DR Diluent
	/	*FR Dye
/	DS Diluent	DS Diluent
	Probe Cleanser	Probe Cleanser
ESR channel	ESR Solution Reagent	ESR Solution Reagent

### 3.6.2 Controls and Calibrators

The controls and calibrators are used to verify accurate operation and calibration of the analyzer.

The controls are suspension of simulated human blood, specifically manufactured to monitor and evaluate the analysis precision of the analyzer. The controls are prepared with three levels, namely low, normal and high. Daily use of all levels verifies the operation of the analyzer and ensures reliable results are obtained. The calibrators are commercially prepared whole-blood products used to calibrate some parameters (WBC, RBC, HGB, MCV and PLT etc.) of analyzer to build the metrological traceability of analysis results. For the use and storage of controls and calibrators, please refer to the Instruction for Use of each product.

All references related to controls in this manual refer to the "controls" and "calibrators" specifically formulated for this analyzer by Mindray. You must buy those controls and calibrators from Mindray or Mindray-authorized distributors.

The following models of controls are used with the analyzer:

**Table 3-19 Controls for complete blood count tests**

Name	Model	Level	Applicable Model	*QC Parameters
Hematology Control	BR60	High, Low, Normal	BC-760[B]/BC-760[R]/BC-780[R]	All blood sample test report parameters except ESR
Hematology Control	BC-6D	High, Low, Normal	BC-760[B]/BC-760[R]/BC-780[R]	All blood sample test report parameters except RET series
Hematology Control	BC-RET	High, Low, Normal	BC-760[R]/BC-780[R]	RET group report parameters

**Table 3-20 Controls for body fluid tests**

Name	Model	Level	Applicable Model	QC Parameters
Hematology Control	BC-BF	High, Normal, Low	BC-760[B]/BC-760[R]/BC-780[R]	Body fluid sample test report parameters

**Table 3-21 Controls for ESR tests**

Name	Model	Level	Applicable Model	QC Parameters
Hematology Control	BC-6D	Normal, Low	BC-760[B]/BC-760[R]/BC-780[R]	ESR

The following models of calibrators are used with the analyzer:

**Table 3-22 Calibrator for complete blood count tests**

Name	Model	Applicable Model
Hematology Calibrator	SC-CAL PLUS	BC-760[B]/BC-760[R]/BC-780[R]

## NOTE

- For the specific test parameters and reference values of parameters, see the reference value sheets of controls and calibrators.

**Figure 3-16 Calibrator for ESR tests**

Name	Model	Applicable Model
Hematology Calibrator	SC-CAL PLUS	BC-760[B]/BC-760[R]/BC-780[R]

**This page intentionally left blank.**

# 4 Understanding the System Principles

## 4.1 Overview

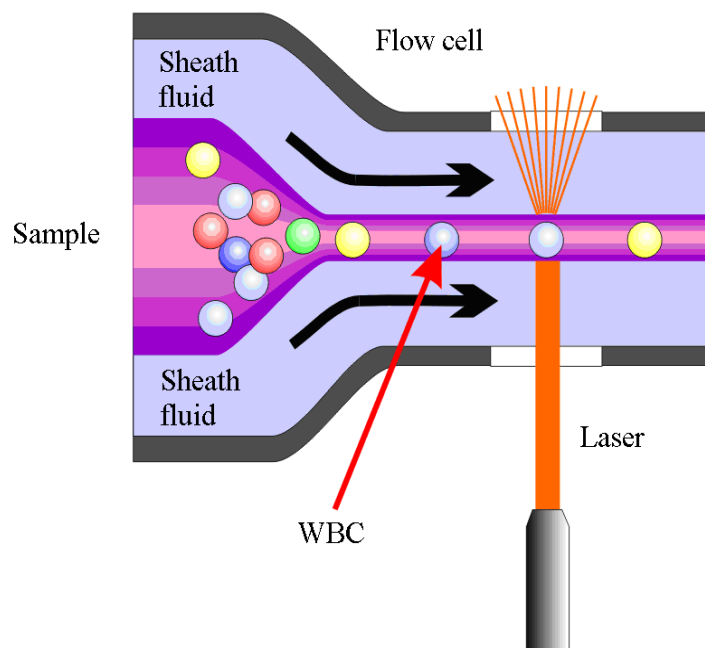
The principles used by the analyzer are:

- Sheath flow impedance method, laser scatter and SF Cube cell analysis technology (3D analysis using information from scatter of laser light at two angles and fluorescence signals) for cell differentiation and counting;
- Colorimetric method for HGB measurement.

Based on the above data, the analyzer calculates other parameters.

## 4.2 WBC Measurement

### 4.2.1 SF CUBE Cell Analysis Technology



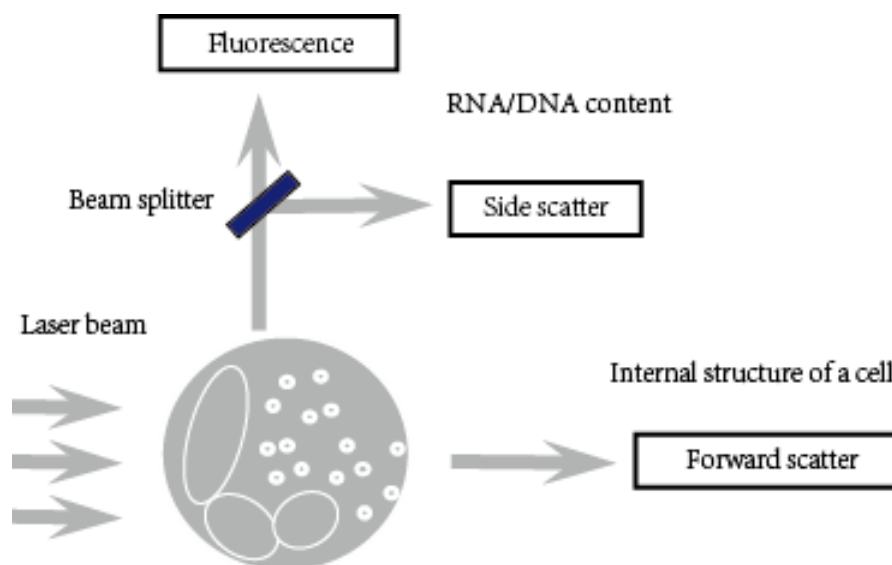
**Figure 4-1 Laser flow cytometry**

In normal peripheral blood, white blood cells can be classified into five categories: lymphocytes, monocytes, neutrophils, eosinophils and basophils. Analyzing all types of white blood cells will provide a great deal of useful information for the clinical diagnosis of diseases. Under the influence of certain diseases, the peripheral blood may contain various abnormal cells apart from the five subpopulations of normal cells, such as

atypical lymphocytes, immature cells, etc. Most of these abnormal cells are different kinds of immature cells in the cell generation process. But what they have in common is they contain a great deal of nucleic acid (DNA and RNA), the content of which decreases as the cell gets maturer. Therefore, normal cells and immature cells can be differentiated by detecting the content of nucleic acid in the cells.

Body fluid refers to the fluid in side body cavities except blood vessels. There are many sub-types of body fluid, among which the most commonly seen sub-types are cerebrospinal fluid, pleural fluid, ascitic fluid, and synovial fluid. Both cerebrospinal fluid and serous cavity fluid are colorless and transparent in normal case, but in abnormal cases, there could be increase of cells (including leukocytes and erythrocytes). Leukocytes in body fluid can be categorized into mononuclear cells (MN) and polymorphonuclear cells (PMN). The analysis of the cells in body fluid can provide useful information for clinical diagnosis.

The analyzer adopts the SF Cube cell analysis technology to recognize and detect the immature cells in blood accurately besides doing WBC 5-part differentiation, as well as identify the nucleated cells in body fluid.



**Figure 4-2 SF cube technology**

The analyzer adopts the fluorescent staining technology in its DIFF channels. The RBCs are lysed and the WBC subpopulations are made different in size and complexity by the lyse; the nucleic acid substances in WBCs are marked by the new asymmetric cyanine fluorescent substance. Due to the different content of nucleic acid in different WBC subpopulations, maturity stages or abnormal development status, the volume of fluorescent dye staining the nucleic acid substances can be different; the low-angle light scatter reflects the cell size, the high-angle light scatter reflects the intracellular granularity, and the intensity of fluorescent signal reflects the degree that the cell is stained. By sensing the difference in signal in three dimensions of the cells processed with lyse, the DIFF channel differentiates the subpopulations of WBCs (lymphocytes, monocytes, neutrophil and eosinophils), as well as identifies and flags abnormal cells like immature granulocytes, abnormal lymphocytes and blast cells.

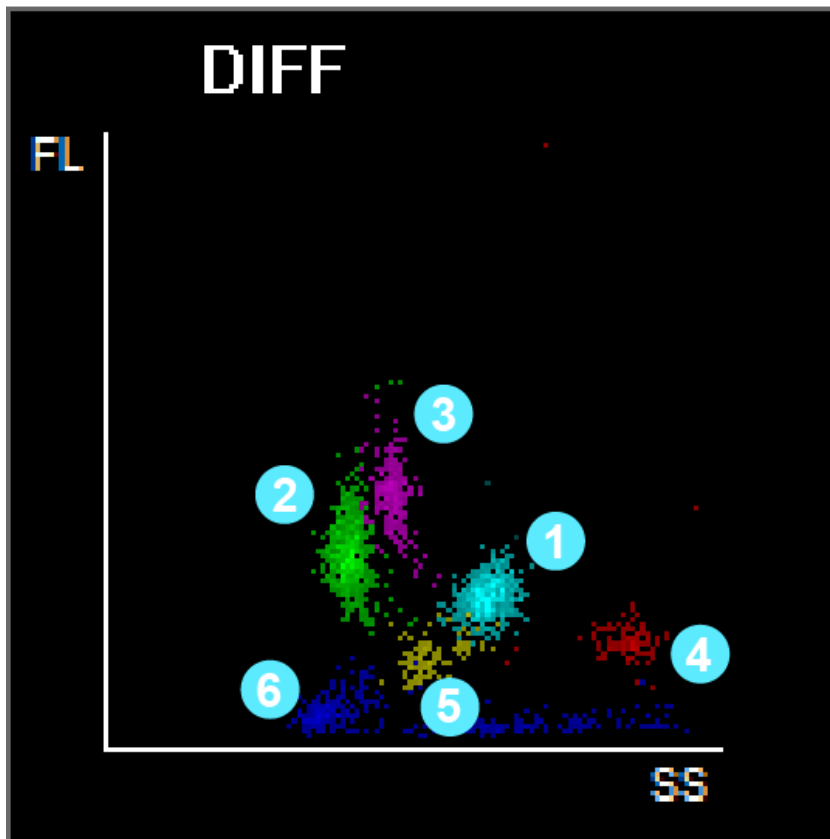
The lymphocytes are smaller in size with the nucleus taking most part of them. Lymphocytes have a high nucleus-to-cytoplasm ratio, but their nucleic acid content is low, therefore they are at a lower position in the direction of fluorescence and side scatter. The monocytes are larger in size, with high nucleus-to-cytoplasm ratio and high nucleic acid content, and less complex in structure, therefore they are at a higher position in the direction of fluorescence, and have stronger side scatter. The neutrophils and basophils are larger in size, and have medium nucleus-to-cytoplasm ratio and low nucleic acid content, therefore they are at a lower position in the direction of fluorescence, but they have stronger side scatter. The characteristics of the eosinophils are similar to those of the neutrophils, but they contain a lot of alkaline grains, so they have very strong side scatter. The blast cells, atypical lymphocytes and immature granulocytes have high nucleic acid content, so they are at a higher position in the direction of fluorescence on the scattergram.

In body fluid samples, the mononuclear cells (MN) are less complex in intracellular granularity, so the side scatter is weaker, while polymorphonuclear cells are more complex in intracellular granularity, so the side scatter is stronger.



## 4.2.2 Derivation of WBC-Related Parameters

### DIFF Scattergram



1. Neu region
2. Lym region
3. Mon region
4. Eos region
5. Baso region
6. Ghost region

Parameters	Name	Formula/Test Methods	Unit
WBC	White blood cell count	WBC=Sum of all particles in the WBC region in the DIFF channel	$10^9/L$
Bas#	Basophil count	Bas# = WBC x Bas%	$10^9/L$
Bas%	Basophil percentage	$\text{Bas}\% = \frac{\text{Particles in the Bas region in the DIFF channel}}{\text{Sum of all particles in DIFF channel except those in Ghost region}} \times 100\%$	%
Neu#	Neutrophil count	Neu# = WBC x Neu%	$10^9/L$
Neu%	Neutrophil percentage	$\text{Neu}\% = \frac{\text{Particles in the Neu region in the DIFF channel}}{\text{Sum of all particles in DIFF channel except those in Ghost region}} \times 100\%$	%
Eos#	Eosinophil count	Eos# = WBC x Eos%	$10^9/L$
Eos%	Eosinophil percentage	$\text{Eos}\% = \frac{\text{Particles in the Eos region in the DIFF channel}}{\text{Sum of all particles in DIFF channel except those in Ghost region}} \times 100\%$	%
Lym#	Lymphocyte count	Lym# = WBC x Lym%	$10^9/L$
Lym%	Lymphocyte percentage	$\text{Lym}\% = \frac{\text{Particles in the Lym region in the DIFF channel}}{\text{Sum of all particles in DIFF channel except those in Ghost region}} \times 100\%$	%
Mon#	Monocyte count	Mon# = WBC x Mon%	$10^9/L$

Parameters	Name	Formula/Test Methods	Unit
Mon%	Monocyte percentage	$\text{Mon}\% = \frac{\text{Particles in the Mon region in the DIFF channel}}{\text{Sum of all particles in DIFF channel except those in Ghost region}} \times 100\%$	%
IMG#	Immature granulocyte count	$\text{IMG}\# = \text{WBC} \times \text{IMG}\%$	$10^9/\text{L}$
IMG%	Immature granulocyte percentage	$\text{IMG}\% = \frac{\text{Particles in the IMG region in the DIFF channel}}{\text{Sum of all particles in DIFF channel except those in Ghost region}} \times 100\%$	%

## 4.3 Hemoglobin Concentration Measurement

### 4.3.1 A Test Model Using the Colorimetric Method

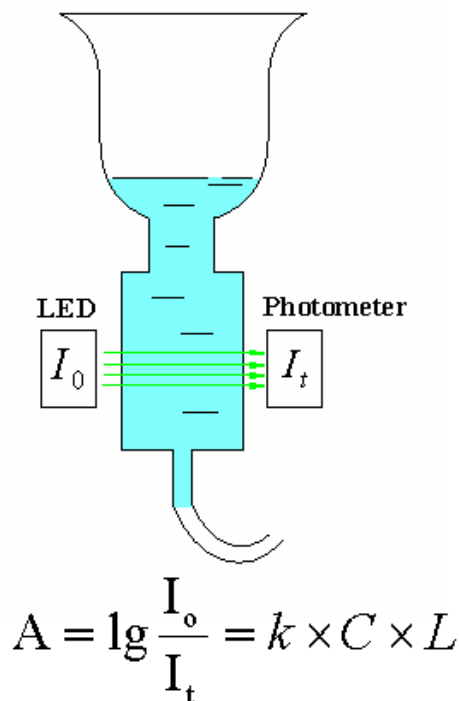


Figure 4-3 Colorimetric method

According to the Lambert-Beer Principle, when a beam of monochromatic light passes through a well-proportioned non-scattering light-absorbing solution, the absorbance  $A$  is proportional to the product of the thickness  $L$  and the concentration  $C$ . The sample in the HGB channel acts as the light absorbing substance after being treated by reagent, therefore the HGB concentration can be measured by measuring the absorbance.

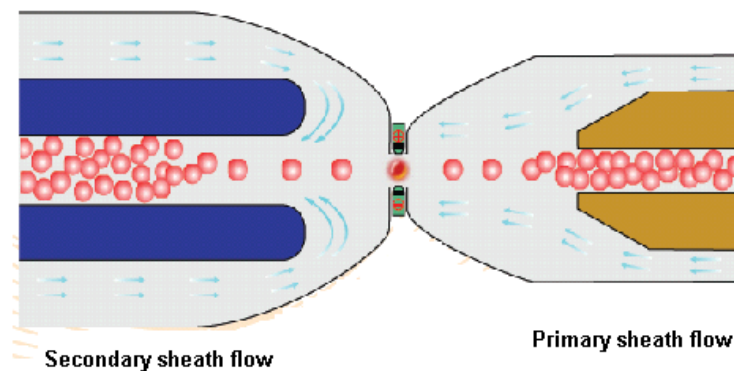
### 4.3.2 Derivation of HGB

Hemoglobin Concentration (HGB) is calculated using the following equation and expressed in g/L.

Parameters	Name	Formula/Test Methods	Unit
HGB	Hemoglobin Concentration	$\text{HGB} = \text{Constant} \times \ln \left( \frac{\text{Blank Photocurrent}}{\text{Sample Photocurrent}} \right)$	g/L

## 4.4 RBC/PLT Measurement

### 4.4.1 Sheath Flow Impedance Method



**Figure 4-4 Sheath flow impedance method**

RBCs/PLTs are counted by the sheath flow impedance method. A sensor is designed to enable the RBCs and PLTs to pass through the aperture one by one in a queue under the “focusing” effect of fluid, during which process pulses will be generated according to the Coulter Principle. The backend processor amplifies the pulses and compares them with the voltage thresholds of the RBC/PLT channel, and then the number of pulses in the RBC/PLT channel is calculated. That is to say, the pulses collected are sorted per the voltage thresholds of different channels, the number of pulses falling in the range of the RBC/PLT channel is the number of RBC/PLT. The number of cells in each channel defines the volume distribution of cells. The analyzer presents the RBC/PLT histogram, whose x-coordinate represents the cell volume (fL) and y-coordinate represents the number of the cells.

Compared with the common impedance method, the sheath flow impedance method is featured by higher efficiency, better signal quality, more accurate analysis results and lower consumption of reagents.

### 4.4.2 Measurement Principle of Platelets in DIFF Channel (PLT-H)

Traditional impedance method counts platelets by sorting and detecting the pulses fall in the PLT channel per cell size. However, as microcytes and fragments are similar in size with platelets with relatively large size, when there are microcytes/fragments present, the impedance method may deliver not so accurate PLT results. To solve the problem, the analyzer lyses the erythrocytes in the DIFF channel, so the count of platelets in relatively large sizes in DIFF channel will not be affected by erythrocytes. By combining the large-size platelet count result in DIFF channel and the small-size platelet count result in the impedance channel, the analyzer provides more accurate PLT results.

### 4.4.3 SF CUBE Cell Analysis Technology

The RET channel also adopts the SF CUBE Cell Analysis Technology. The general measurement principle in RET channel is similar to that of DIFF channel, only that in the RET channel, the RBCs are not lysed, but are spherized by RET diluent. Then the nucleic acid of the spherized RBCs and the PLTs are stained by fluorescent dyes.

**NOTE**

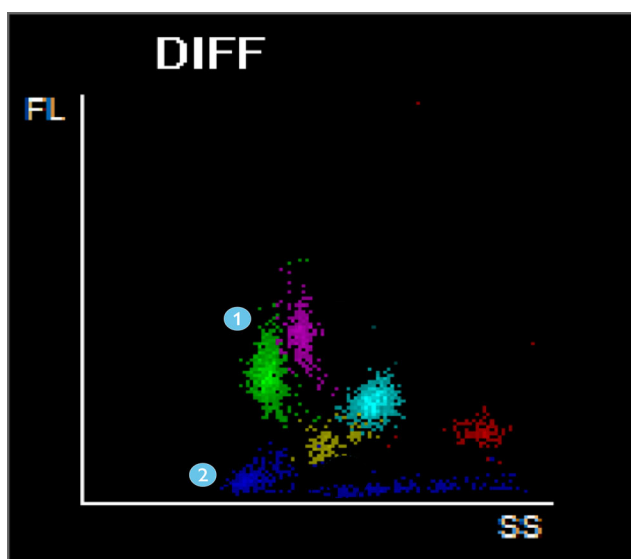
- RET channel is available only on BC-760[R]/BC-780[R] model.

**4.4.4 RBC-Related Parameters**

Parameters	Name	Formula/Test Methods	Unit
RBC	Red blood cell count	Red blood cell (RBCs) number is measured directly by counting the red blood cells passing through the aperture.	$10^{12}/L$
MCV	Mean corpuscular volume	Calculated based on the red blood cell histogram	fL
HCT	Hematocrit	$HCT = \frac{RBC \times MCV}{10}$	%
MCH	Mean corpuscular hemoglobin	$MCH = \frac{HGB}{RBC}$	pg
MCHC	Mean corpuscular hemoglobin concentration	$MCHC = \frac{HGB}{HCT} \times 100$	g/L
RDW-CV	Red blood cells distribution width coefficient of variation	RBC Histogram	%
RDW-SD	Red blood cells distribution width standard deviation	Derived based on the standard deviation of red blood cell volume distribution	fL
NRBC% <sup>1</sup>	Nucleated red blood cell percentage	Calculated based on the information derived from LYM region ① and Ghost region ② of DIFF scattergram	/100WBC
NRBC#	Nucleated red blood cell count	$NRBC\# = WBC \times NRBC\%$	$10^9/L$

Note:

1. LYM region ① and Ghost region ② of DIFF scattergram are as follow:

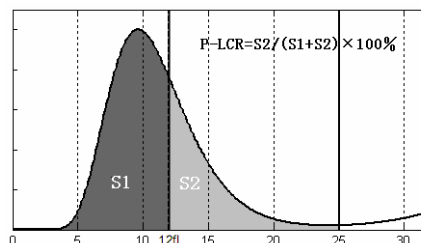


#### 4.4.5 PLT-Related Parameters

Parameters	Name	Formula/Test Methods	Unit
PLT	Platelet count	Method 1: The analyzer provides the number of platelet directly by counting the platelets passing through the aperture. Method 2: Based on the analysis result of large platelet in DIFF channel, correct the analysis result of platelet in impedance channel	$10^9/L$
MPV	Mean platelet volume	Mean platelet volume (MPV) is calculated based on the PLT histogram.	fL
PDW	Platelet distribution width	Platelet distribution width is derived from the platelet histogram, and is reported as 10 geometric standard deviation (10 GSD)	/
PCT	Plateletcrit	$PCT = \frac{PLT \times MPV}{10000}$	%
P-LCR <sup>1</sup>	Platelet-large cell ratio	P-LCR is derived from the platelet histogram, it represents the ratio of the number of platelets with a size over 12 fL to the total platelets number	%
P-LCC	Platelet-large cell count	$P-LCC = PLT \times P-LCR$	$10^9/L$
IPF <sup>2</sup>	Immature platelet fraction*	For BC-760[R]/BC-780[R], the parameter is derived based on PLT-O scattergram: $IPF = \frac{\text{Immature platelet number in the optical channel}}{\text{Sum of all platelet particles in the optical channel}} \times 100\%$ For BC-760[B], this is an optional parameter and is derived based on PLT-H scattergram. $IPF = \frac{\text{Immature platelet number in the DIFF channel}}{\text{PLT-H result}} \times 100\%$	%
PLT-I	Platelet count-Impedance	Platelet number (PLT) is measured directly by counting the platelets passing through the aperture.	$10^9/L$
PLT-H	Platelet count hybrid	PLT results derived from impedance channel, which are corrected by the DIFF channel large size platelet results.	$10^9/L$
PLT-O	Optical platelet count	PLT results derived from the RET channel	$10^9/L$

Note:

1. Platelet-Large Cell Ratio (P-LCR) is derived from the platelet histogram; it represents the ratio of the number of platelets with a size over 12fL to the total platelets number. The ratio is represented in %. In the following figure, S2 represents the number of larger platelet cells, and S1+S2 represents the total PLT count.



2. IPF parameters are applicable to the BC-760[R]/BC-780[R] model only. For BC-760[B], this is an optional parameter. For more information, contact Mindray Customer Service Department.
3. PLT-O is applicable to BC-760[R]/BC-780[R] model only.
4. PLT-H is applicable to BC-760[R]/BC-780[R] model only. For BC-760[B], this is an optional parameter. For more information, contact Mindray Customer Service Department.

#### 4.4.6 Reticulocyte Parameters\*

Parameters	Name	Formula/Test Methods	Unit
*RET%	Reticulocyte percentage	$\text{RET}\% = \frac{\text{Number of cells in the reticulocyte region}}{\text{Number of cells in mature RBC region} + \text{Number of cells in RET region}} \times 100\%$	%
*RET%	Reticulocyte count	$\text{RET}\# = \text{RBC} \times \text{RET}\%$	$10^{12}/\text{L}$
*HFR	High fluorescent ratio	$\text{HFR} = \frac{\text{Number of cells in HFR region}}{\text{Number of cells in RET region}} \times 100\%$	%
*MFR	Middle fluorescent ratio	$\text{MFR} = \frac{\text{Number of cells in MFR region}}{\text{Number of cells in RET region}} \times 100\%$	%
*LFR	Low fluorescent ratio	$\text{LFR} = \frac{\text{Number of cells in LFR region}}{\text{Number of cells in RET region}} \times 100\%$ <p>Note: Sum of all particles in the reticulocyte region = Sum of all particles in the LFR region + Sum of all particles in the MFR region + Sum of all particles in the HFR region</p>	%
*IRF	Immature reticulocyte fraction	$\text{IRF} = \text{MFR} + \text{HFR}$	%
RHE	Reticulocyte hemoglobin expression	Calculated based on the light scatter information of RET	pg

\*Note: The reticulocyte parameters are only available on the BC-760[R]/BC-780[R] model.

#### 4.5 Erythrocyte Sedimentation Rate Measurement

The analyzer uses the photometric method to measure the aggregation of erythrocytes within a specified time period and calculates the sedimentation rate. RBCs separated in the whole blood sample first aggregate and form Rouleaux formation; then the aggregation falls. The sedimentation rate depends on the degree of aggregation. As the measuring beam is subject to the RBC aggregation state, specifically manifested in that its light transmittance increases with the aggregation degree. As a result, the aggregation degree of RBCs can be obtained by measuring the change in the light transmittance of whole blood samples over time, and then the erythrocyte sedimentation rate is calculated.

#### 4.6 Body Fluid Parameters

Body fluid refers to the fluid in side body cavities except blood vessels. There are many sub-types of body fluid, among which the most commonly seen sub-types are cerebrospinal fluid, pleural fluid, ascitic fluid, and synovial fluid. Both cerebrospinal fluid and serous cavity fluid are colorless and transparent in normal case, but in abnormal cases, there could be increase of cells (including leukocytes and erythrocytes). Leukocytes in body fluid can be categorized into mononuclear cells (MN) and polymorphonuclear cells (PMN). The analysis of the cells in body fluid can provide useful information for clinical diagnosis.

Parameters	Name	Formula/Test Methods	Unit
WBC-BF	White blood cell count-body fluid	$\text{WBC-BF} = \text{Sum of all particles in the DIFF channel except those in Ghost region and HFR}$	$10^9/\text{L}$

Parameters	Name	Formula/Test Methods	Unit
TC-BF#	Total nucleated cell counts-body fluid	TC-BF# = Count of all particles in the DIFF channel except those in Ghost region	$10^9/L$
MN%	Mononuclear cell percentage	$MN\% = \frac{\text{Particles in MN region of DIFF channel}}{WBC - BF} \times 100\%$	%
PMN%	Polymorphonuclear cell percentage	$PMN\% = \frac{\text{Particles in PMN region of DIFF channel}}{WBC - BF} \times 100\%$	%
MN#	Mononuclear cell number	$MN\# = WBC - BF \times MN\%$	$10^9/L$
PMN#	Polymorphonuclear cell number	$PMN\# = WBC - BF \times PMN\%$	$10^9/L$
RBC-BF	Red Blood Cell count-body fluid	Red blood cells in body fluid (RBC-BF) are measured directly by counting the red blood cells passing through the aperture.	$10^{12}/L$

## 4.7 Wash

After each analysis cycle, all elements of the analyzer that the sample runs through are washed to ensure no residue is left.

**This page intentionally left blank.**



# 5

## Installing and Connecting the System

---

### 5.1 Notes for Analyzer Installation

---

#### CAUTION

- The installation, authorization, upgrade and modification of the system software must be performed by personnel authorized by Mindray. Make sure to install only Mindray-authorized software.
  - Unpacking, installation or transportation by personnel not authorized or trained by Mindray may cause personal injury or damage your instrument. Do not unpack, transport or install your instrument without the presence of Mindray-authorized personnel.
- 

#### NOTE

- The safety of any system incorporating the equipment is the responsibility of the assembler of the system.
  - The equipment is tested and packed with care before it is shipped from the factory. When you receive your analyzer, carefully inspect the carton. If you see any signs of mishandling or damage, contact Mindray Customer Service department or your local distributor immediately.
  - After you open the package, check the integrity of the product according to the packing list. If you find any part missing, contact Mindray Customer Service Department or your local distributor immediately.
- 

#### 5.1.1 Space Requirements

For the dimensions and weight of the analyzer, please refer to **B.16Dimensions and Weight**.

Check the site for proper space allocation. In addition to the space required for the system itself, arrange for:

- Proper height to place the analyzer;
- At least 500 mm on each side of the analyzer, which is the preferred access to perform service procedures;
- At least 600 mm above the analyzer;
- At least 250 mm behind the analyzer;
- The diluent container must be placed within 1.0 meter's reach under the main unit, the lyse container must be placed on a plane of the same level with the main unit.
- Desktop or platform must be capable to bear about the weight of 180kg.

### 5.1.2 Power Requirements

#### CAUTION

- Using pinboard may bring electrical interference and the analysis results may be unreliable. Place the analyzer near the electrical outlet to avoid using the pinboard.
- Use the original power cord provided by the manufacturer. Using other electrical wire may damage the system or lead to unreliable analysis results.

	Voltage	Frequency	Input power
Main Unit (Analyzer)	100–240 V~ (±10%)	50 Hz/60 Hz (±1 Hz)	600VA

### 5.1.3 Environment Requirements

	Normal Operation Environment	Storage and Transportation Environment	Operation Environment
Ambient temperature	10~35℃	-10~40℃	5~40℃
Relative Humidity	30%~85%	10%~90%	10%~90%
Atmospheric pressure	70.0kPa~106.0kPa <sup>Note</sup>	50.0kPa~106.0kPa	70.0kPa~106.0kPa

Note:

- The altitude requirement for normal operation is -400 m to +3000 m
- Install the instrument at a position not exposed to splashing water.
- The environment should be as free as possible from dust, mechanical vibrations, loud noises and electrical interference;
- It is advisable to evaluate the electromagnetic environment prior to operation of this analyzer.
- Do not use this instrument in close proximity to sources of strong electromagnetic radiation;
- Do not place the analyzer near brush-type motors, flickering fluorescent lights, and electrical contacts that regularly open and close;
- Do not place the analyzer in direct sunlight or in front of a source of heat or drafts;
- Do not use the instrument in a working environment with conductive or combustible gases.
- The environment shall be well ventilated;
- Do not place the analyzer on a slope;
- Connect only to a properly earth grounded outlet;
- Only use this analyzer indoors.

### 5.1.4 Fuse Requirement

#### WARNING

- The fuse used in the instrument is not a replaceable one. If there is any problem with the fuse, contact Mindray Customer Service Department or your local distributor.

### 5.1.5 Moving and Installing the Analyzer

Moving and installation of the analyzer shall be conducted by Mindray authorized personnel. Do not move or install your analyzer without the presence of Mindray-authorized personnel.

---

**⚠ CAUTION**

---

- When installing or using the analyzer, make sure at least the two inside supporters of the autoloader are on the supporting table of the analyzer.
  - After the autoloader is installed, do not lay too much pressure to it or transport the analyzer by holding the autoloader.
- 

**5.1.5.1 Analyzer transportation**

1. Insert the handle bars horizontally into the four handling holes at the bottom plate of the main unit to the limit positions respectively, as shown in the figure below.



**Figure 5-1 Inserting the handle bar**

2. Rotate the handles to the vertical position, as shown in the following figure. Pull the handles out to see whether they are firm. If yes, the main unit can be conveyed..



**Figure 5-2 Handling position of the handle bar**

**NOTE**

- The main unit must be carried by two persons with the handles to avoid tilting and other accidents. Sufficient space must be reserved for the main unit to ensure that the two inside supporters of the autoloader can be placed onto the deck panel of the main unit.
- Avoid colliding with the main unit, especially the inlets on the rear panel of the main unit.

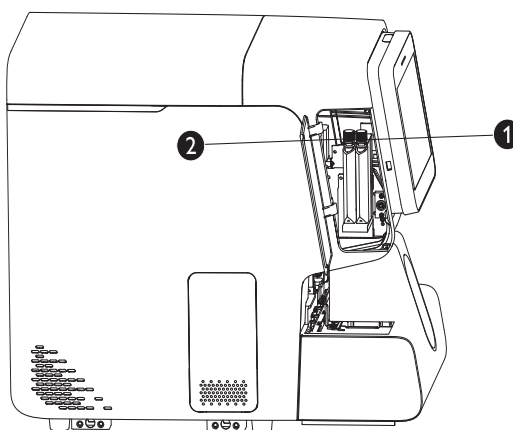
## 5.2 Connecting the Analyzer System

### 5.2.1 Connecting the Reagents

**NOTE**

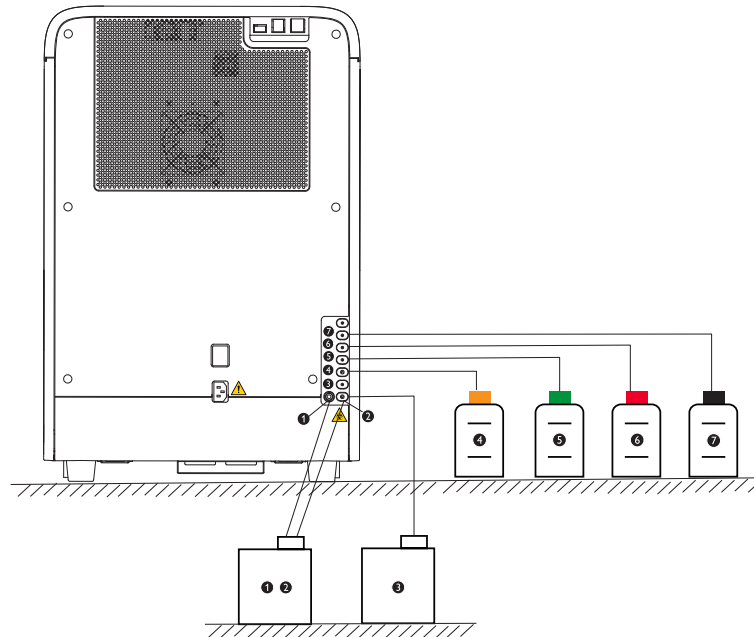
- When connecting the reagents, make sure the color of the reagent container cap assembly is the same as that of the reagent inlet to which it is connected.

Connecting the reagent containers to the analyzer as shown in the following figure



**Figure 5-3 Reagent connection - connecting fluorescent dyes**

Interface No.	Type	Description
①	FR Dye	Applicable to BC-760[R]/BC-780[R]model
②	FD Dye	/



**Figure 5-4 Connecting reagents - lyse, diluent, and waste**

Interface No.	Type of Connected Reagent	Description
①②	Waste container	Interface ② is used to connect the waste container, and interface ① connects to the waste floater sensor to detect whether the waste container is full
③	DS Diluent	/
④	ESR Solution Reagent	/
⑤	LH Lyse	/
⑥	LD Lyse	/
⑦	DR Diluent	Applicable to BC-760[R]/BC-780[R] model

## 5.2.2 Connecting the Peripherals

Make sure that the connections are correct and firm.

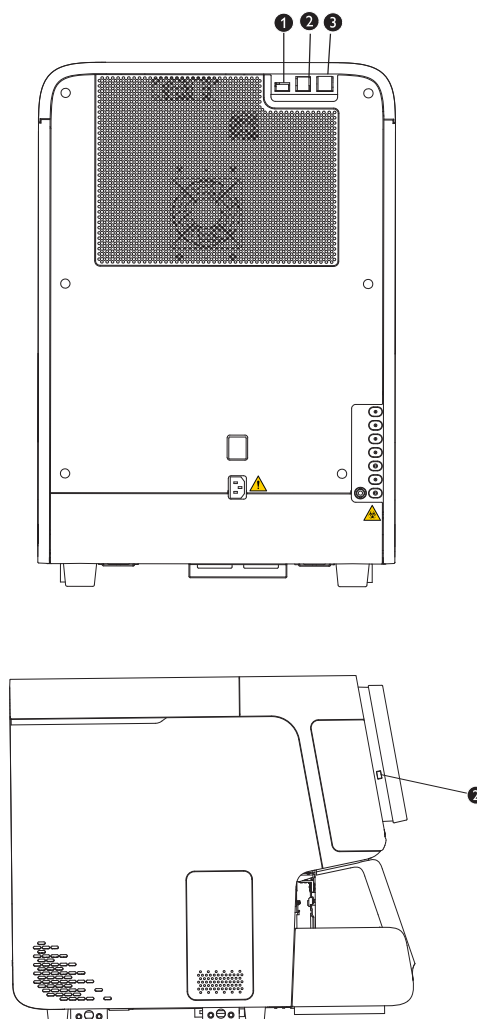


Figure 5-5 Connecting the peripherals

Interface No.	Interface	Connection
①	USB port (protocol 3.0)	Connect to optional devices such as the printer and scanner through the USB ports as needed
②	USB port (protocol 2.0)	
③	Network interface	Connect to the network port on the PC

### CAUTION

- Be sure to use the specified external devices only, and keep them away from water.
- External devices connected to the analyzer and digital interfaces must be authorized and complied with relevant safety and EMC standards (e.g. IEC 60950 Safety of Information Technology instrument Standard and CISPR 22 EMC of Information Technology instrument Standard (Class B)). Any persons who connects additional instrument to the signal input or output ports and configures an IVD system, is responsible for ensuring that the system works normally and complies within the safety and EMC requirements. If you have any questions, consult the technical service department of your local representative.

**NOTE**

- 
- The user should ensure the data safety of the USB devices connecting to the analyzer.
  - When the instrument connects to an external computer, install anti-virus software on the computer and scan for viruses and update patches periodically. Do not use it for unintended purposes.
-

**This page intentionally left blank.**



# 6 Customizing the Analyzer Software

## 6.1 Introduction

The analyzer is a flexible laboratory instrument that can be tailored to your work environment. You can customize the software options as introduced in this chapter.

For the security of the settings and data, two access levels are provided to the operator of the analyzer: “**General User**” and “**Administrator**”. The administrator access level provides the operator with access to more functions or settings, some of which can be configured to be accessible to operators.

The following tables list the users’ access by the access levels.

		Administrator's level	Operator's level
System Setup	Print Setup	√	Partly
	Communication	√	
	Barcode	√	
	Date/Time Setup	√	√
	Lab Info. Setup	√	√
	Flag Alarm Sensitivity	√	
	Flag Rules Setup	√	
	Tube Rack Type Setup	Review only	Review only
	Ext. PLT Tests	√	
User Management	New	√	
	Modify Password	√	
	Delete	√	√
Auxiliary Setup	Get Sample Information		√
	Other Settings	Predilute mode prompt	√
		Pop-up keyboard	√
		Enable fluorescent reagent detect sensor	√
		Monitor reagent expiration date	
		Flags	
		Alarm Volume	
		Enable auto exit STAT	√
		Auto replace reagent after setup	√
		De-identify health information	
		Auto log-out	√

		Administrator's level	Operator's level
<b>Para. Setup</b>	<b>Parameter Unit Setup</b>	√	Review only
	<b>Ref. Range Setup</b>	√	Review only
	<b>Microsco. Para. Setup</b>	√	
<b>Maintenance</b>	/	√	
<b>Reagent Setup</b>	/	√	√
<b>Auto-loading</b>	/	√	
<b>Gain Setup</b>	/	√	
<b>Re-exam Rules Setup</b>	/	√	
<b>Auto Startup/Shutdown</b>	/	√	

## 6.2 Saving Settings after Changes

After you have changed or modified analyzer settings, follow below steps to save the changes.

1. Tap **"Menu"** > **"Setup"** and select the setting item you want to change.
  2. Make necessary changes on the setting screen.
  3. Tap another button on the software screen.
- √ A dialog box displays asking if you want to save the change.
4. Tap **"Yes"**.
- √ The new setting is saved.

## 6.3 Analyzer Settings

### 6.3.1 System Setup ("Menu" > "Setup" > "System Setup")

#### 6.3.1.1 Print Setup ("Menu" > "Setup" > "System Setup" > "Print Setup")

For the description about print setup, refer to **11Printing**.

#### 6.3.1.2 Communication ("Menu" > "Setup" > "System Setup" > "Communication") (Administrators)

Administrators can configure the following communication settings.

- **Protocol Setup**
- **Transmission Mode**

Before you set up the communication settings, make sure that:

- you have logged in as an administrator;
- the network wire is firmly connected to the analyzer.

#### **Protocol Setup**

1. Tap **"Menu"** > **"Setup"** > **"System Setup"** > **"Communication"** to enter the **"Communication"** screen.
2. Set up the communication protocol in accordance with the real needs in your laboratory.

See below for setting descriptions

<b>IP Address</b>	Enter the correct IP address	For correct network settings, consult Mindray Customer Service Department or your network administrator.
<b>Subnet Mask</b>	Enter the correct subnet mask	
<b>Default Gateway</b>	Enter the correct gateway	
<b>ACK Synchronous Transmission</b>	Check <b>"ACK Synchronous Transmission"</b> to enable the ACK synchronous transmission function.	When <b>"ACK Synchronous Transmission"</b> is enabled, enter the ACK overtime in the <b>"ACK Overtime"</b> field (10 seconds by default)

### Transmission Mode

1. Tap **"Menu"** > **"Setup"** > **"System Setup"** > **"Communication"** to enter the **"Communication"** screen.
2. Set up the transmission mode in accordance with the real needs in your laboratory.

See below for setting descriptions

<b>Transmission Mode</b>	When you select LIS as a data channel, check to enable one or more transmission functions: <ul style="list-style-type: none"> <li>• <b>Auto Retransmit</b></li> <li>• <b>Auto Communicate</b></li> <li>• <b>Transmit as Print Bitmap Data</b></li> <li>• <b>Communicate L-J QC results as sample results</b></li> </ul>	Only when <b>"ACK Synchronous Transmission"</b> is enabled, can you enable the <b>"Auto Retransmit"</b> function.
<b>Data channel</b>	Data channel: <ul style="list-style-type: none"> <li>• <b>LIS</b></li> <li>• <b>labXpert</b></li> </ul>	Operators with user level or administrator level accounts cannot change the setting
<b>Scattergram transmitted as/ Histogram transmitted as</b>	When you select LIS as a data channel, you can select the following options from the pull-down list: <ul style="list-style-type: none"> <li>• <b>Not to be transmitted</b></li> <li>• <b>Bitmap</b></li> <li>• <b>Data</b></li> </ul>	

#### 6.3.1.3 Setting up the Barcode Type (Menu" > "Setup" > "System Setup" > "Barcode" (Administrators)

Administrators may set up barcode system as needed.

The following barcode types are supported:

- **CODE128**
- **CODE39**
- **ITF**
- **CODE93**
- **CODABAR**
- **UPC/EAN/JAN**

For the length limits and check bit of different code types, see **B.18Barcode Specifications**.

1. Tap **"Menu"** > **"Setup"** > **"System Setup"** > **"Barcode"** to enter the **"Barcode"** screen.
2. Tap the desired code type to go to the corresponding setup screen.
3. Check the **"Apply"** box to apply the code type.
4. Select the number of digits used on site in the **"Digits"** area.
5. (Optional) If you are using CODE39, ITF, and CODABAR with check bit, check the **"Check Bit"** box.

**NOTE**

- When you are using ITF, CODE39, and CODABAR with check bit, actual checked barcode digits length should be the length of barcode information plus 1 check bit.
- If you are using 9 digits barcode, and check the "Check Bit" box, you should select "10" for barcode length.

6. (Optional) Set other code types if needed.

**NOTE**

- For code types supporting check bit, use check bit in barcode labels if possible to reduce the rate of misreading.
- The code types and length limits set on the analyzer shall be those used in your laboratory.
- Do not select the code types that are not used, which may increase the rate of misreading.
- Do not use barcodes longer than 20 digits. Barcodes longer than 20 digits will not be read correctly.

**6.3.1.4 Date/Time ("Menu" > "Setup" > "System Setup" > "Date/Time")**

You can set up date and time on the "Date/Time Setup" screen.

1. Tap "Menu" > "Setup" > "System Setup" > "Date/Time Setup" to enter the "Date/Time Setup" screen.
2. Set up the date and time.

See below for setting descriptions:

<b>Date</b>	Enter the current date	/
<b>Time</b>	Enter the current time	The analyzer uses the 24-hour clock system
<b>Date Format</b>	Select a date format from the pull-down list	/

**6.3.1.5 Lab Info. Setup ("Menu" > "Setup" > "System Setup" > "Lab Info. Setup")**

Users may enter necessary laboratory information on the "Lab Info. Setup" screen.

1. Tap "Menu" > "Setup" > "System Setup" > "Lab Info. Setup" to enter the "Lab Info. Setup" screen.
2. Enter laboratory information as needed.

**6.3.1.6 Flag Alarm Sensitivity ("Menu" > "Setup" > "System Setup" > "Flag Alarm Sensitivity") (administrators)**

The analyzer provides the following flags for abnormal blood cell morphology.

Flag Message	Indication	Conditions
<b>Blasts?</b>	Possible presence of blast cells	Presence of excessive dots in blast sensitive region of the scattergram
<b>Immature Gran?</b>	Possible presence of immature granulocytes	Presence of excessive dots in immature granulocyte sensitive region of the scattergram
<b>RBC Lyse Resistance?</b>	Possible presence of RBC lyse resistance	Presence of abnormally distributed dots in the WBC sensitive region of the DIFF scattergram
<b>Abn Lymph/blast?</b>	Possible presence of abnormal lymphocytes or blasts	Presence of excessive dots in abnormal lymphocyte/blast sensitive region of the scattergram
<b>Turbidity/HGB Interfere?</b>	HGB results may be abnormal or interference may exist	Calculate and compare special parameters
<b>Fragments?</b>	Possible presence of RBC fragments	Presence of abnormally distributed dots in sensitive region of the RET channel

Flag Message	Indication	Conditions
<b>Infected RBC?</b>	Possible presence of infected RBC	Presence of excessive dots in infected RBC sensitive region of the scattergram
<b>Left Shift?</b>	Possibility of left shift	Presence of excessive dots in left shift sensitive region of the scattergram
<b>Atypical Lymph?</b>	Possible presence of atypical lymphocytes	Presence of excessive dots in atypical lymphocyte sensitive region of the scattergram
<b>RBC Agglutination?</b>	RBC results possibly inaccurate	Calculate and compare special parameters
<b>Iron Deficiency?</b>	May indicate iron deficiency anemia	Calculate and compare special parameters
<b>PLT Clump?</b>	Possibility of PLT clump	Calculate and compare special parameters
<b>Lipid Particles?</b>	Possible presence of lipid particles	Presence of excessive dots in lipid particle sensitive region of the scattergram
<b>WBC Fragments?</b>	Possible presence of WBC fragments.	Presence of abnormally distributed dots in WBC fragment sensitive region

During sample analysis, the analyzer evaluates and scores the possibility of the presence of all types of abnormal blood cell morphology. When the score for a certain type of abnormal blood cell morphology exceeds the set threshold, the analyzer reports the flag accordingly.

Administrators may tap **"Setup" > "System Setup" > "Flag Alarm Sensitivity"** to set up the flag alarm threshold values. The higher the threshold value, the lower the alarm sensitivity of the flag.

Follow below instructions:

1. Tap **"Menu" > "Setup" > "System Setup" > "Flag Alarm Sensitivity"** to enter the **"Flag Alarm Sensitivity"** screen.
2. Define the flag alarm threshold values in the **"Value (0-100)"** edit boxes as needed.

## NOTE

- The allowed range for all flag alarm threshold values is [0, 100].
- The flag alarm items vary with the models with different configurations. Please be subject to the interface of your analyzer.
- The default flag alarm threshold for **"Infected RBC?"** is 100; and the default thresholds for all other flags are all 40.

### 6.3.1.7 Flag Rules Setup ("Menu" > "Setup" > "System Setup" > "Flag Rules Setup") (administrators)

The analyzer provides the following flag messages:

Flag Message	Indication	Conditions
<b>Leukocytopenia</b>	WBC count low	$WBC < 2.50 \times 10^9/L$
<b>Leucocytosis</b>	WBC count high	$WBC > 18.00 \times 10^9/L$
<b>Neutropenia</b>	Neu# low	$Neu\# < 1.00 \times 10^9/L$
<b>Neutrophilia</b>	Neu# high	$Neu\# > 11.00 \times 10^9/L$
<b>Lymphopenia</b>	Lym# low	$Lym\# < 0.80 \times 10^9/L$
<b>Lymphocytosis</b>	Lym# high	$Lym\# > 4.00 \times 10^9/L$
<b>Monocytosis</b>	Mon# high	$Mon\# > 1.50 \times 10^9/L$

Flag Message	Indication	Conditions
<b>Eosinophilia</b>	Eos# high	Eos# > $0.70 \times 10^9/L$
<b>Basophilia</b>	Bas# high	Bas# > $0.20 \times 10^9/L$
<b>NRBC Present</b>	NRBC detected	NRBC% > 1.00% and NRBC# > $0.010 \times 10^9/L$
<b>Anisocytosis</b>	Anisocytosis	RDW-CV > 22.0% or RDW-SD > 64.0fL
<b>Hypochromia</b>	Hypochromia	Hypochromia < 290g/L
<b>Microcytosis</b>	MCV low	MCV < 70.0fL
<b>Macrocytosis</b>	MCV high	MCV > 110.0fL
<b>Erythrocytosis</b>	RBC high	RBC > $6.50 \times 10^{12}/L$
<b>Anemia</b>	Anemia	HGB < 90g/L
<b>*Reticulocytosis</b>	RET high	RET% > 5% or RET# > $0.20 \times 10^{12}/L$
<b>Thrombocytopenia</b>	PLT low	PLT < $60 \times 10^9/L$
<b>Thrombocytosis</b>	PLT high	PLT > $600 \times 10^9/L$
<b>Pancytopenia</b>	WBC, RBC and PLT low	WBC < $4.00 \times 10^9/L$ and RBC < $3.50 \times 10^{12}/L$ and PLT < $100 \times 10^9/L$

## NOTE

- The item with \* only applies to the BC-760[R]/BC-780[R] model.

Administrators may tap "Menu" > "Setup" > "System Setup" > "Flag Rules Setup" to set the flag rules for classified alarms.

Follow below instructions:

1. Tap "Menu" > "Setup" > "System Setup" > "Flag Rules Setup" to enter the "Flag Rules Setup" screen.
2. Editing a rule:
  - a. Tap in the flag rule list to select the rule to be defined and edit the rule setup below.
  - b. Tap another button, and save the setting as prompted.

Other operations:

<b>Resume</b>	To restore an edited rule to factory default, tap the "Resume" button.
<b>All to Default</b>	To restore all the edited rules to factory default, tap the "All to Default" button.
<b>Import</b>	Insert the USB device for saving the flag rules in the analyzer, and tap "Import" to import flag rules from the USB device.
<b>Export</b>	Insert the USB device for saving the flag rules in the analyzer, and tap "Export" to export the set flag rule to the USB device.

### 6.3.1.8 Tube Rack Type Setup (Menu > Setup > System Setup > Tube Rack Type Setup)

Tap "Menu" > "Setup" > "System Setup" > "Tube Rack Type Setup" to check the information of tube rack and tubes.

### 6.3.1.9 Ext. Tests ("Menu" > "Setup" > "System Setup" > "Ext. PLT Tests") (administrators)

The series of analyzers provide the "Ext. PLT Tests" function for the PLT test.

After the "Ext. PLT Tests" function is enabled, the analyzer automatically predicts the PLT results during the sample test, and automatically starts the sample rerunning analysis mode of low value when the PLT value is

lower than the set threshold. The whole process is completed at one time, not requiring re-aspirating of the sample or additional consumption of lyse and dye.

PLT 5X re-exam	During sample test procedure, the analyzer predicts PLT results. When the predicted result is lower than the set threshold, the analyzer automatically lengthens the test duration in the PLT-O channel to 5 times that of a normal test to obtain 5 times particles in the normal test mode.
----------------	---

Follow below instructions:

1. Tap **"Menu"** > **"Setup"** > **"System Setup"** > **"Ext. PLT Tests"** to enter the **"Ext. PLT Tests"** screen.

Check **"Rerun low PLT samples"** and set the threshold as needed.

## NOTE

- The Ext. PLT Tests mode only applies to the BC-760[R]BC-780[R] model.
- The Ext. Tests mode do not apply to prediluted and body fluid samples.
- The default value of low PLT sample is  $50 \times 10^9/L$ .

## 6.3.2 User Management ("Menu" > "Setup" > "User Management")

The **"User Management"** screen displays all the user accounts registered on the analyzer.

### 6.3.2.1 Adding new account (administrators)

Administrators can create new users on the "User Management" screen.

1. Tap **"Menu"** - **"Setup"** - **"User Management"** to enter **"User Management"** screen.
  2. Tap **"New"** to go to the **"Add User"** screen.
  3. Select **"Access Level"**:
    - **General User**
    - **Administrator**
  4. Enter the **"User ID"**, **"Name"**, and **"Password"** in turn.
  5. Tap **"OK"** to save the settings.
- ✓ The new user account is activated.

## NOTE

- The user ID cannot be null and up to 12 characters can be entered.
- The password can be null and up to 12 characters can be entered.
- The name can be null and up to 20 characters can be entered.

### 6.3.2.2 Changing password

## NOTE

- Users at either administrator's or operator's level can only change the passwords for the users currently logged in.

1. Tap **"Menu"** - **"Setup"** - **"User Management"** to enter **"User Management"** screen.
  2. Select the current user, and tap **"Modify Password"**.
  3. In the **"Modify Password"** dialog box, enter **"Old Password"**, and then enter the new password in the **"New Password"** and **"Confirm Password"** edit boxes.
  4. Tap **"OK"** to save the new password.
- ✓ The new password is activated.

### 6.3.2.3 Deleting account (administrators)

An administrator can delete any users registered under his/her own account.

#### NOTE

- **You cannot delete a built-in user!**

1. Tap **"Menu" - "Setup" - "User Management"** to enter **"User Management"** screen.
2. Select a user and tap **"Delete"** to delete it.
- ✓ A confirmation box displays.
3. Tap **"Yes"**.
- ✓ The selected user is deleted.

### 6.3.3 Auxiliary Setup ("Menu" > "Setup" > "Auxiliary Setup")

Tap **"Menu" - "Setup" - "Auxiliary Setup"** to enter **"Auxiliary Setup"** screen. You can set up the following contents:

- **Get Sample Information**
- **Other Settings**

#### 6.3.3.1 Getting sample information

##### Setting of the next sample

Tap **"Menu" > "Setup" > "Auxiliary Setup" > "Get Sample Information"** to enter the **"Get Sample Information"** screen. You can set the following options:

- **Entry of next sample ID**
  - ◆ **Auto increase**
  - ◆ **Manual Entry**

When **"Auto increase"** is selected for **"Entry of next sample ID"**:

- ✓ When you are running samples under the auto-loading analysis cycle and manually enter the first Sample ID, the Sample IDs for the following samples automatically increase.
- ✓ When you are running samples under closed-tube analysis, you only need to enter an ID for the first sample, and the subsequent sample IDs will automatically increase by 1 based on the previous one.

When **"Manual Entry"** is selected for **"Entry of next sample ID"**:

- ✓ When you are running samples under closed-tube analysis, you need to enter each sample ID manually.

- **Prefix Length**

When **"Auto increase"** is selected for **"Entry of next sample ID"**, this edit box will be activated.

Enter a number (n) into the edit box of **"Prefix Length"**. The first n characters in the sample ID will not be auto increased.

#### NOTE

- **Prefix Length cannot exceed 20 digits.**

##### Setting of the first sample after startup

Set the test panel for the first sample after startup in accordance with the real needs in your laboratory.

Follow below instructions:

1. Tap **"Menu" > "Setup" > "Auxiliary Setup" > "Get Sample Information"** to enter the **"Get Sample Information"** screen.
  2. In the **"Setting of the first sample after startup"** area, set the test panel for the first sample after startup.
- Define the mode for the first sample after startup:



1. In the **"First sample after startup"** pull-down list, select **"Custom"**.
2. In the **"Mode"** pull-down list, select the desired mode.
3. In the **"Sample ID"** edit box, enter the ID (1 by default) of the first sample after startup.
- ✓ After each startup, the first sample is analyzed according to the set sample ID and mode by default.
- Test the sample according to the mode of the last sample before shutdown:
  1. In the **"First sample after startup"** pull-down list, select **"Run the suspended sample after restart"**.
  - ✓ The sample ID of the first sample after each startup increases by 1 on the basis of the ID of the last sample before shutdown; the sample is tested according to the mode of the last sample before shutdown.

### Getting sample information

The setup is applicable to the analyzer with built-in rotary scanner.

This function is for "Auto-Scan rack No." setup under autoloading mode. Select "Auto-Scan rack No.," and the analyzer starts to scan rack No. automatically.

Follow below instructions:

1. Tap **"Menu"** > **"Setup"** > **"Auxiliary Setup"** > **"Get Sample Information"** to enter the **"Get Sample Information"** screen.
2. Tap **"Auto-Scan rack No."** in the **"Get Sample Information"** area.

### 6.3.3.2 Other Settings

On the **"Other Settings"** screen, you can set the following functions:

- **Predilute mode prompt**
  - **Pop-up keyboard**
  - **Enable fluorescent reagent detect sensor**
  - **Monitor reagent expiration date**
  - **Flags** (administrators)
  - **Alarm Volume** (administrators)
  - **Auto exit STAT**
  - **Auto replace reagent after setup**
  - **De-identify health information**
  - **Auto log-out**
1. Tap **"Menu"** > **"Setup"** > **"Auxiliary Setup"** > **"Other Settings"** to enter the **"Other Settings"** screen.
  2. Define the settings as needed.

See the table below for setting descriptions:

<b>Predilute mode prompt</b>	When <b>"Predilute mode prompt"</b> is checked, in predilute mode, when you start analysis, a dialog box will pop up to remind you that the current analysis mode is predilute. Uncheck <b>"Predilute mode prompt"</b> to disable the function.	/
<b>Pop-up keyboard</b>	Check <b>"Pop-up keyboard"</b> to enable the pop-up keyboard. Tap on the edit area on each screen, the pop-up keyboard will display for you to input information. When you are using an external keyboard, you can uncheck <b>"Pop-up keyboard"</b> to disable the function.	/

<b>Enable fluorescent reagent detect sensor</b>	When " <b>Enable fluorescent reagent detect sensor</b> " is checked, if the fluorescent reagents are not sufficient, the analyzer will send an alarm.	/
<b>Monitor reagent expiration date</b>	Check " <b>Monitor reagent expiration date</b> " to enable the function. When the function is enabled, the analyzer gives warning when a reagent gets expired.	/
<b>Flags</b>	Select from the pull down list to define the suspect, high and low flags (default: "R" for suspect, "H" for high and "L" for low).	Suspect: R, r
		High: H, h, ↑
		Low: L, l, ↓
<b>Alarm Volume</b>	Select the alarm volume you want	Low Medium High Max
<b>Auto exit STAT</b>	<ul style="list-style-type: none"> <li>When "<b>Auto exit STAT</b>" is selected, if you do not place STAT samples in the sample compartment during the pre-defined time range, the analyzer automatically exits STAT.</li> <li>When "<b>Auto exit STAT</b>" is not selected, manually select "<b>Exit STAT</b>" to exit STAT.</li> </ul>	The default waiting time is 2 minutes. The setup range is [2-5] minutes.
<b>Auto replace reagent after setup</b>	<ul style="list-style-type: none"> <li>When "<b>Auto replace reagent after setup</b>" is checked, after you replace the reagent with a new one and enter the barcode information of the new reagent on the "Reagent Setup" screen, the analyzer automatically replaces the reagent in the reagent container.</li> <li>If this option is not checked, you need to manually tap the "<b>Replace</b>" button to replace the reagent in the reagent container after replacing the old reagent and entering the new barcode information.</li> </ul>	The default waiting time is 30 seconds. The setup range is [0-100] seconds.
<b>De-identify health information</b>	After " <b>De-identify health information</b> " is checked, when you print, export sample information or view it on " <b>Table Review</b> " screen, identifiers (like name, gender, age and date of birth) will be shown as "*****".	/
<b>Auto log-out</b>	When " <b>Auto log-out</b> " is selected, the current account will automatically log out if the user has not performed any operation for the defined period of time.	The default log-out time is 60 minutes. The setup range is [1-120] minutes.

## 6.3.4 Para. Setup ("Menu" > "Setup" > "Para. Setup") (Administrators)

### 6.3.4.1 Parameter Unit Setup (Administrators)

Administrators may set up unit systems and parameter units.

1. Tap "**Menu**" > "**Setup**" > "**Para. Setup**" > "**Parameter Unit Setup**" to enter the "**Parameter Unit Setup**" screen.
2. (Optional) When necessary, select the unit system from the "**Unit System:**" pull-down list.
3. Tap the "**Unit**" column of the parameter for which you want to change the unit.

The available units for the parameter displays on the right side of the screen.

4. Check the desired unit for the parameter in the "**Unit option:**" area.
- ✓ The parameter unit refreshes.

## NOTE

- Users at operator's level can only review the unit system and the parameter units.
- Tap "Default" to restore the default units for all parameters.

### 6.3.4.2 Ref. Range Setup (Administrators)

The "Ref. Range Setup" screen provides 5 factory reference groups for your selection. In addition, you can set up to 10 custom reference groups. Users at administrator's level may select and customize reference ranges and reference groups.

1. Tap "Menu"-"Setup"-"Ref. Range Setup" to enter "Ref. Range Setup" screen.
2. Add a new reference group, edit or delete reference groups, or set default groups as needed.

Follow below instructions:

<b>New</b>	<ol style="list-style-type: none"> <li>1. Tap "New" to add a new reference group.</li> <li>2. On the new reference group setup screen, set up the name, lower and upper limits of ages, as well as the gender information for the new "Reference group".</li> <li>3. When necessary, tap and edit the "Upper" and "Lower" limits for the parameters.</li> </ol>	/
<b>Edit</b>	Tap to select the reference group to edit, and tap "Edit".	<ul style="list-style-type: none"> <li>• For the 5 factory reference groups, you can only edit the "Upper" and "Lower" limits of the parameters.</li> <li>• For the custom reference groups, you can edit the group name, lower and upper limits of age, gender information, and "Upper" and "Lower" of the parameters.</li> </ul>
<b>Delete</b>	Tap to select the custom reference group to delete, and tap "Delete".	You cannot delete factory reference groups.
<b>Set to Default</b>	Tap and select a reference group, and tap "Set to Default" to set the selected reference group as default reference group.	/
<b>Match customized ref. group first</b>	Check to enable "Match customized ref. group first".	<ul style="list-style-type: none"> <li>• When "Match customized ref. group first", the analyzer first finds the matching reference group based on the sample age/ gender conditions.</li> <li>• When the "Match customized ref. group first" is not enabled, the analyzer first finds the matching group in the factory reference groups based on the age/ gender.</li> </ul>

### 6.3.4.3 Microsco. Para. Setup (Administrators)

In the default mode, the "Microsco. Para. Setup" screen displays 22 microscopic parameters of hematology analysis.

Administrators may add new parameters, edit or delete existing microscopic parameters on the "Microscopic Para. Setup" screen.

1. Tap "Menu" > "Setup" > "Para. Setup" > "Microsco. Para. Setup" to enter the "Microsco. Para. Setup" screen.
2. Add new microscopic parameters, or edit or delete existing microscopic parameters as needed.

Follow below instructions:

Add a new microscopic parameter	Tap "New" to add a row in the microscopic parameter name area. Enter the new parameter name in the "Microscopic Parameter Name" column.
---------------------------------	---

Edit the microscopic parameters	Tap a parameter name in the table to edit the name.
Delete the microscopic parameters	Select a row in the table, and tap the <b>"Delete"</b> button to delete the parameter.

**NOTE**

- The analyzer can save at most 40 microscopic parameters.

## 6.3.5 Maintenance ("Menu" > "Setup" > "Maintenance") (Administrators)

### 6.3.5.1 Standby

When the time for which the analyzer is free from fluidic operations reaches that you have set on the **"Maintenance"** screen of the analyzer, the analyzer automatically enters the standby status.

On the **"Maintenance"** setup screen, administrators may set the wait time before the analyzer enters the **"Standby"** status, when the fluidic system stops working.

1. Tap **"Menu"** - **"Setup"** - **"Maintenance"** to enter **"Maintenance"** screen.
2. Set up the wait time before the analyzer entering the standby status.

**NOTE**

- The allowed range is 30 to 60 minutes. Make sure you enter the valid time and in the required format.

### 6.3.5.2 Probe cleanser maintenance

Administrators may set up the start time for daily Probe Cleanser maintenance on the **"Maintenance"** setup screen.

1. Tap **"Menu"** - **"Setup"** - **"Maintenance"** to enter **"Maintenance"** screen.
2. Set the start time and reminding interval for daily Probe Cleanser maintenance as needed.

Item	Description	Remark
Start at	When conditions for daily probe cleanser maintenance are met, the analyzer prompts you to maintain the Probe Cleanser at the set time.	<ul style="list-style-type: none"> <li>• The allowed range of time is 00: 00 to 23: 59.</li> <li>• The start time for Probe Cleanser maintenance should be earlier than that for auto shut-down, see <b>6.3.10Setting Auto Startup/Shutdown Time ("Menu" &gt; "Setup" &gt; "Auto Startup/Shutdown") (Administrators)</b>.</li> </ul>
Remind every	If you disable the daily probe cleanser, the analyzer prompts you to maintain the Probe Cleanser when the remind time is out of the set value.	The allowed range of time is 5 to 60min.

✓ Set the start time of Probe Cleanser maintenance. The system will perform Probe Cleanser maintenance for the relevant parts at the specified time according to the operating condition of the analyzer.

**NOTE**

- When the analyzer prompts you to maintain the Probe Cleanser, you can tap **"Cancel"** to ignore the prompt and up to three times is allowed to ignore. You must perform the maintenance when the prompt appears for the forth time.
- For conditions of daily probe cleanser maintenance, see **12.4.1Daily Probe Cleanser Maintenance**.

### 6.3.6 Reagent Setup ("Menu" > "Setup" > "Reagent Setup")

For the reagent replacing steps, refer to **12.3.2 Replace the Reagents**.

### 6.3.7 Setting up Functions for the Auto-Loading of Samples (Menu > "Setup" > "Auto-loading") (administrators)

Administrators may set up the following functions regarding the autoloading of samples in accordance with the real needs in your laboratory.

- How to proceed when blood/analysis mode inquiry failed
  - When there is tube vacancy, sample ID
  - Display summary after autoloading finishes
  - Auto load tube racks
1. Tap **"Menu"**-**"Setup"**-**"Auto-loading"** to enter **"Auto-loading"** screen.
  2. Set up how to proceed when blood/analysis mode inquiry failed, to set up the next sample IDs when there is tube vacancy and whether to display summary after auto-loading finishes.

See below for setting descriptions

<b>When Blood/ Analysis Mode Inquiry Failed</b>	<ul style="list-style-type: none"> <li>When <b>"Use the mode of the previous sample"</b> is enabled, the analyzer will use the analysis mode of last samples;</li> <li>Set up how to proceed when blood/analysis mode inquiry from LIS fails. If you have selected <b>"Skip after"</b>, set up the wait time (n) in seconds (default setting is 10 seconds, and n cannot be longer than 15 seconds);</li> <li>If you have selected <b>"Use specified mode"</b>, select a mode from the pull-down list</li> </ul>
<b>When there is tube vacancy, sample ID</b>	Tap the radio button to select the rule of sample ID increment when there is tube vacancy in the tube rack: <ul style="list-style-type: none"> <li><b>increases</b></li> <li><b>does not increase</b></li> </ul>
Display summary after autoloading finishes	When you have checked <b>"Display summary after autoloading finished"</b> , the screen displays the auto-loading summary, including finished blood samples, rack vacancies, ID reading errors, etc.
Auto load tube racks	<ul style="list-style-type: none"> <li>When <b>"Automatically load the tube racks on the autoloader"</b> is selected, also set the waiting time (1-30 seconds) before analyzer auto loading tube racks. When you place tube racks on the right tray of the autoloader, the analyzer will automatically load tube racks for sample analysis after the pre-defined waiting time.</li> <li>When <b>"Automatically load the tube racks on the autoloader"</b> is not selected, select <b>"Start Count"</b> to start sample analysis.</li> </ul>

### 6.3.8 Gain Setup ("Menu" > "Setup" > "Gain Setup") (Administrators)

When the analyzer reports the HGB blank voltage abnormal error, and you cannot remove the error by pressing the "Remove Error" button, adjust the HGB gains to correct the HGB blank voltage.

1. Tap **Menu** > **"Setup"** > **"Gain Setup"** > **"WB"** to enter the **"Gain Setup"** screen of whole blood.
2. Adjust the HGB default gain in the **"HGB" "Set"** text box, until the HGB blank voltage is in the range of [4.30, 4.50].

#### NOTE

- **When you modify the HGB default gain, the HGB blank voltage will change accordingly.**

3. If necessary, repeat above procedure to adjust the HGB voltages for other modes.

### 6.3.9 Setting Re-exam Rules ("Menu" > "Setup" > "Re-exam Rules Setup") (Administrators)

Administrators may enable and set up re-exam rules on the "Re-exam Rules Setup" screen of the analyzer.

Re-exam means to reexamine the analyzed samples whose results match certain conditions. The way to re-exam is determined by the laboratory procedure, which may include re-analyze on the analyzer, smear review, etc.

You can set re-exam rules for autoloading analysis on the analyzer. When the re-exam rules are enabled, the analyzer will check the auto-loading sample results and information based on the applied rules. If a sample triggers a re-exam rule, the analyzer will automatically transports the sample back for re-exam.

## NOTE

- The re-exam rules only apply to auto-loading analyses.
- If the analyzer is connected to Mindray's labXpert software, set the re-exam rules on the labXpert. For more information about the labXpert software, consult your sales representative.

Follow the following instructions to set re-exam rules for autoloading analysis on the analyzer:

1. Tap **"Menu" > "Setup" > "Re-exam Rules Setup"** to enter the **"Re-exam Rules Setup"** screen.
2. Set the re-exam rules as required.
3. Tap another button on the software screen to save the settings.

Settings	Description	Remark
Enable re-exam rules	When <b>"Re-exam Rules Setup"</b> is checked/unchecked, this function is enabled/disabled.	/
Re-exam when WBC count is low	When <b>"Re-exam when WBC count is low"</b> is checked, re-exam rules will be triggered and a related message will be prompted if measured WBC count is below the set values.	Set range: (0~4) x 10 <sup>9</sup> /L
Re-exam when PLT count is low	When <b>"Re-exam when PLT count is low"</b> is checked, re-exam rules will be triggered and a related message will be prompted if measured PLT count is below the set values.	Set range: (0~4) x 10 <sup>9</sup> /L
Re-exam when abnormal RBC/PLT result is reported	When <b>"Re-exam when abnormal RBC/PLT result is reported"</b> is checked, the analyzer will give a re-exam prompt if PLT is below 30 when the following flags occur: PLT Clump? PLT Scattergram Abn. Microcytosis	/

✓ After the re-exam rules are enabled, the analyzer checks the auto-loading sample results and information based on the applied rules.

- When the sample result triggers the re-exam rules, the "Need Re-exam" message is displayed on the graph review screen of the sample. In the meanwhile, the analyzer automatically transports the sample back for re-exam.

Sample ID	2
Time	07-28-2020 16:07 <b>Need Re-exam</b>
Mode	AL-WB-CD
Tube Pos.	1-1

### 6.3.10 Setting Auto Startup/Shutdown Time ("Menu" > "Setup" > "Auto Startup/Shutdown") (Administrators)

Administrators may set up the auto startup and shutdown time for the analyzer on the **"Auto Startup/Shutdown"** screen.

When you have set the auto startup/shutdown time, the analyzer automatically starts and shutdown at the set time.

1. Tap **"Menu" > "Setup" > "Auto Startup/Shutdown"** to enter the **"Auto Startup/Shutdown"** screen.
2. Check require dates.

	Auto Startup	Complete Time	Auto Shutdown	Time
Monday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	17 : 00
Tuesday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	17 : 00
Wednesday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	17 : 00
Thursday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	17 : 00
Friday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	17 : 00
Saturday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	12 : 00
Sunday	<input type="checkbox"/>	08 : 00	<input type="checkbox"/>	00 : 00

Note: The analyzer will begin startup procedure 20 minutes in advance of the specified startup complete time.

01-07-2022 09:48

Administrator

3. Define the auto startup complete time in the auto startup **"Complete Time"** field.  
 ✓ For example, if you set the **"Complete Time"** for auto startup to 8:00 on Monday, the analyzer automatically starts the startup procedure at 7:40, and completes the procedure at 8:00.
4. Define the auto shutdown time in the auto shutdown **"Time"** field.

### CAUTION

- For example, if you set the **"Time"** for auto shutdown to 17:00 on Monday, the analyzer automatically starts the shutdown procedure at 17:00.

### NOTE

- To use the auto startup function, do not power off the analyzer.

**This page intentionally left blank.**



# 7 Operating Your Analyzer

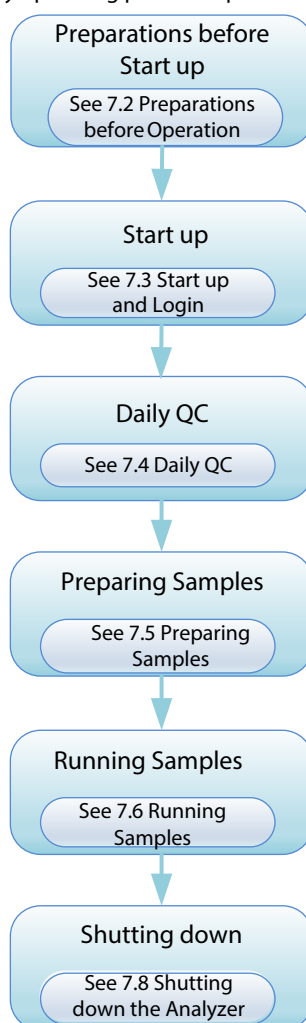
---

## 7.1 Overview

This chapter provides step-by-step procedures for operating your analyzer on a daily basis.

### 7.1.1 Operating Your Analyzer

A flow chart indicating the common daily operating process is presented below.



## 7.1.2 Introduction to the Screen

### 7.1.2.1 Introduction to the Count Screen

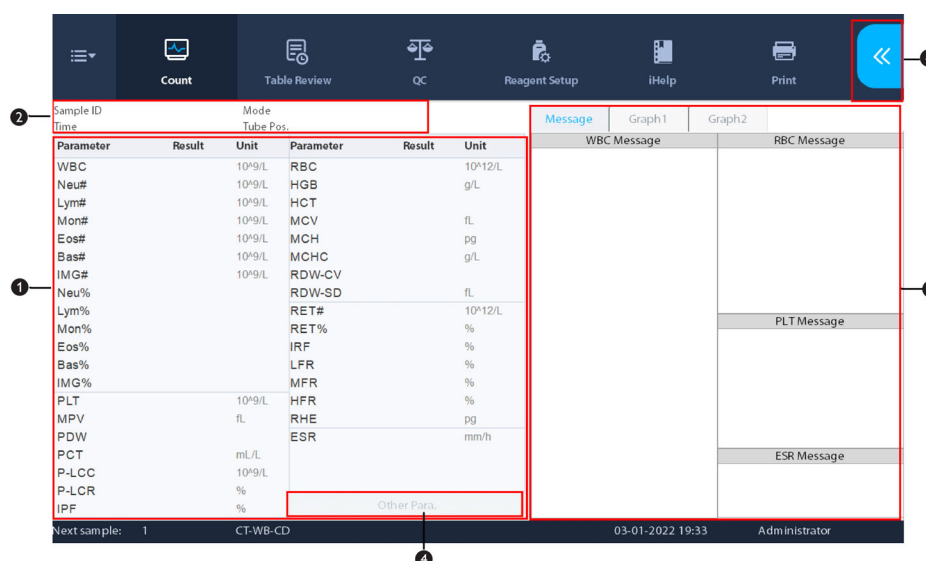


Figure 7-1 Count screen

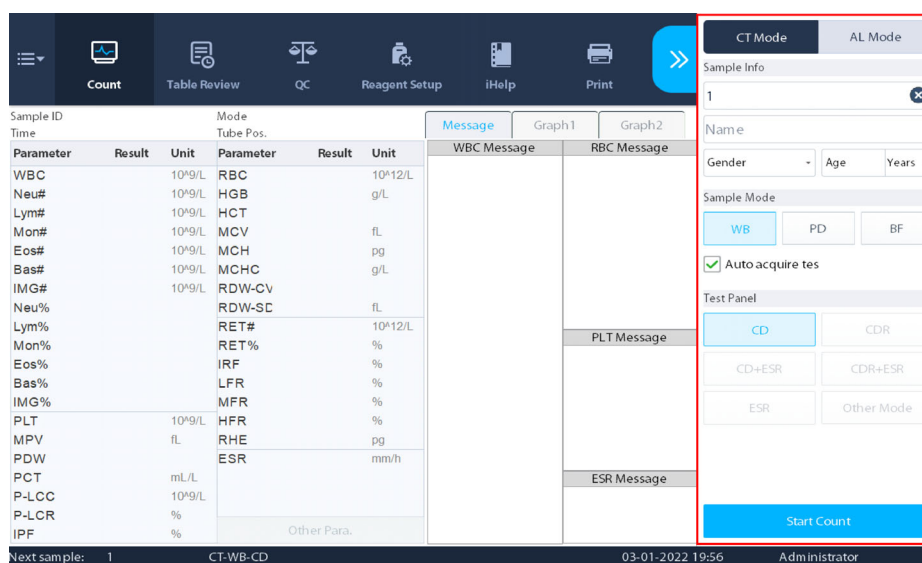



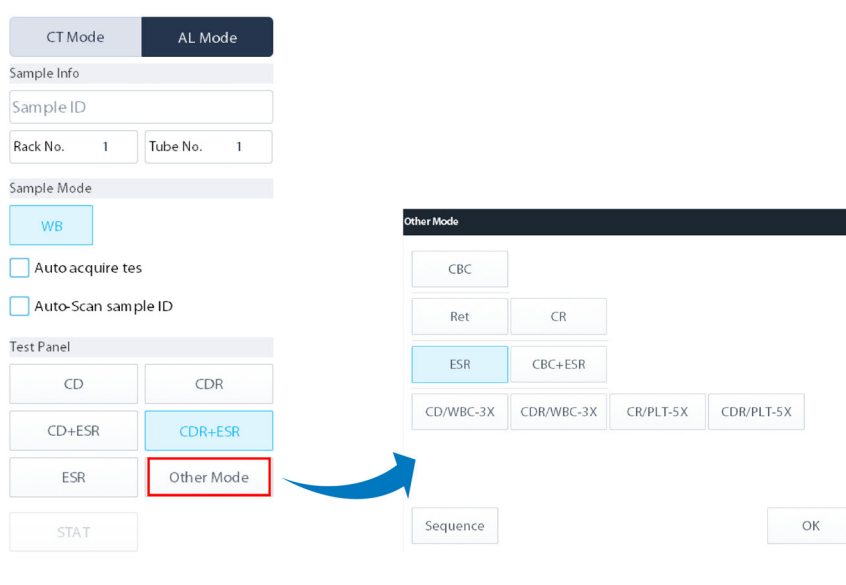
Figure 7-2 Count screen - pop-up window

①	Report parameter result area	The area displays the sample report parameter results.
②	Sample information area	The area displays sample information
③	Flags and graphs area	<ul style="list-style-type: none"> <li>Tap the <b>"Message"</b> tab to review the abnormal blood cell differential or morphology flags for current sample</li> <li>Tap <b>"Graph1"</b> and <b>"Graph2"</b> to view the scattergrams and histograms for current samples (Graph 2 only applies to blood sample analysis and the BC-760[R]/BC-780[R] model)</li> </ul>

④	Button area	<ul style="list-style-type: none"> <li>• Tap the  button to display the pop-up window and set the sample information, sample mode and test panel as needed.</li> <li>• When using the auto-loading analysis mode, tap <b>"Start Count"</b> to start analysis.</li> <li>• When auto-loading analysis is started, the <b>"Start Count"</b> button turns to <b>"Stop Count"</b>. Tap <b>"Stop Count"</b> to stop auto-loading analysis.</li> <li>• During auto-loading analysis, tap <b>"STAT"</b> to insert STAT samples.</li> <li>• When sample analysis is completed, tap the <b>"Other Para."</b> button to review the RUO parameters results of current sample, or review or edit microscopic parameter results.</li> </ul>
---	-------------	--

### 7.1.2.2 Introduction to the test panel screen

**"Test Panel"** area in the dialog box shows some common analysis modes. Tap **"Other Mode"** button to select more desired modes.

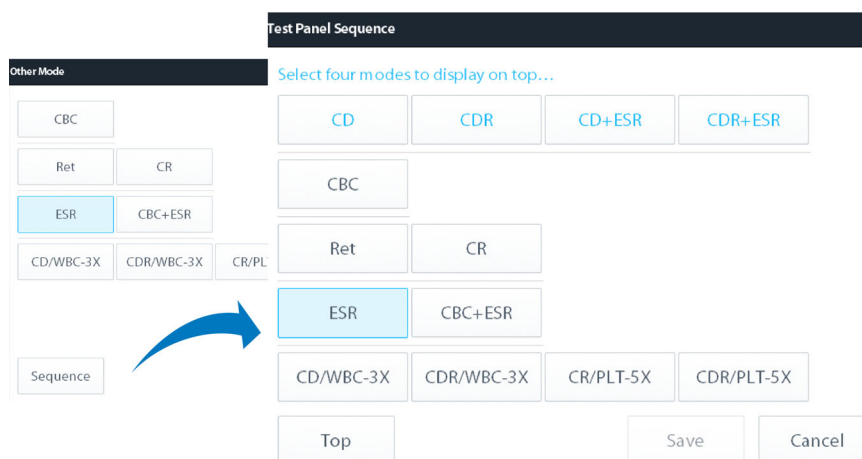


**Figure 7-3 Other mode dialog box**

#### Adjusting the sequences of analysis modes

The analyzer supports function of sequencing analysis modes. You can adjust the sequences of analysis modes in the **"Other Mode"** dialog box as needed.

1. In the **"Other Mode"** dialog box, tap **"Sequence"** button.
- ✓ The **"Test Panel Sequence"** dialog box pops up.



2. Set the sequences of modes.

Function	Description	Remark
Top	Tap <b>"Top"</b> to top up the selected mode.	A minimum of four modes can be topped up.
Save	Tap <b>"Save"</b> to save current settings.	/
Cancel	Tap <b>"Cancel"</b> to return to previous page.	/

## 7.2 Preparations before Operation

Perform the following checks before turning on the analyzer.

### ■ Checking the waste container

Provide a waste container, check and make sure the waste container is not full before startup.

To replace the waste container, refer to **✓The analyzer automatically completes the operation..**

### ■ Checking the reagents

Check whether any reagent has expired and frozen. The reagent needs to be kept still for 24 hours after long-distance transportation.

Check whether there are enough reagents for the test of the day. If the reagents run out during analysis, the analyzer will pause working automatically and prompt the operator to replace the reagents. Follow the instruction to replace the reagents. Otherwise you cannot continue the analysis.

For the safety precautions of reagents, refer to **1.2.3Reagent, Control, and Calibrator-Related Safety Messages**; for the use of reagents, controls, and calibrators, refer to the corresponding IFU; for how to connect and replace reagents, refer to **5.2.1Connecting the Reagents, 12.3.2Replace the Reagents**.

### ■ Checking for fluidic tubes and powers

Check and make sure the reagent and waste tubes are properly connected and not bent.

Check and make sure the power cord of the analyzer is properly plugged into the power outlet. For the power requirements, refer to **5.1.2Power Requirements**.

### ■ Checking the printer

Check and make sure the printer is properly installed, and there is enough paper. Check whether the printer power cord is inserted into the power socket and whether the printer cable is connected properly.

### ■ Checking the keyboard, mouse, and external computer

Check whether the network wire of the external computer is properly connected to the main unit.

## 7.3 Start up and Login

### 7.3.1 Start up the Analyzer

Start up the analyzer in accordance with the actual conditions:

Analyzer status	Operation
When the power switch is turned off	Turn on the power switch on the back of analyzer. ■ The power switch lights in green.
When the power switch is turned on	Press the [Standby] key on the side of analyzer.

√ The system automatically performs the background test and initializes the system. When the self-test and startup initialization procedures complete, the analyzer software displays the main screen.

#### WARNING

- Before turning on the instrument, make sure the input voltage meets the requirements.

#### CAUTION

- Please check the firmness of all the doors and covers before running the system, and make sure they will not open or get loose during analysis. Exercise caution when open/close, install/uninstall the doors and covers to avoid dropping to the ground.

#### NOTE

- Time needed for initializing the fluidic systems depends on how the analyzer was previously shut down. Generally the startup process takes about 10 minutes.
- Users with different accounts have different access to software functions.
- If a fault occurs during initialization (for example, the background check result exceeds the acceptable range of background/blank count results), the analyzer will send an alarm. For the troubleshooting method, refer to *13 Troubleshooting*.
- Background test refers to test on particle and electrical interference.
- If the first background test result obtained during the fluidic initialization exceeds the background range, the analyzer will automatically perform background check again.
- The sample ID of the background test is "0".
- The analyzer does not flag background test result with H/L or suspect flag.
- For the background range of each parameter, refer to *Appendix B.6.1 Background/Blank Count Requirements*.

The status indicator of the analyzer works as follows:

Status indicator status	Meaning
Stay in green	Ready
Flicker in green	Running
Flicker fast	Sampling probe piercing
In red	Error
In orange	Standby
Off	Analyzer shutdown

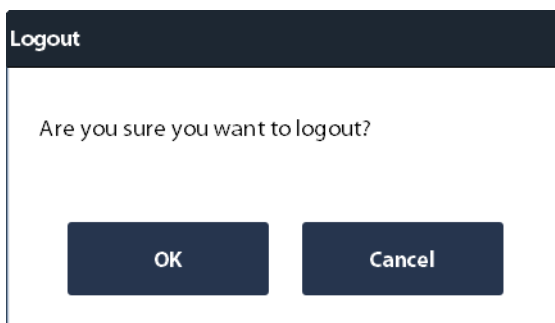
### 7.3.2 Switching Login Account

If logged in as an administrator's account, the lower right corner of the screen displays **"Administrator"**.

If necessary, perform the following steps to switch the login account. To set an account or change the password, refer to **6.3.2 User Management ("Menu" > "Setup" > "User Management")**.

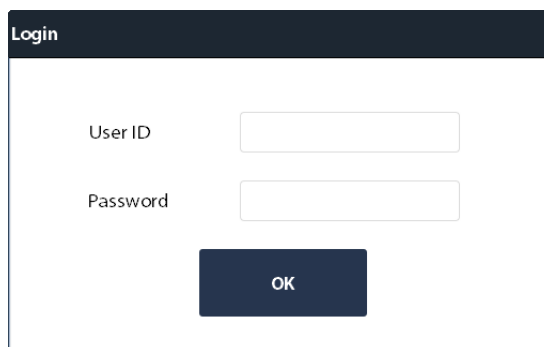
1. Tap **"Menu"** > **"Logout"** in turn.

✓ A dialog box displays

A dialog box titled "Logout" with a dark header. The main text asks "Are you sure you want to logout?". At the bottom, there are two buttons: "OK" and "Cancel".

2. Tap **"OK"**.

✓ The login dialog box displays

A login dialog box titled "Login" with a dark header. It contains two input fields: "User ID" and "Password". Below the fields is a single "OK" button.

3. Enter the new **"User ID"** and **"Password"**, and tap **"OK"** to log in.

## 7.4 Daily QC

Before running any samples, run the controls to ensure reliable results of the analyzer. Please see **9 Using the QC Program** for details.

## 7.5 Preparing Samples

### 7.5.1 Preparing Whole Blood Samples (For WB Mode)

To attain accurate analysis results, make sure the venous blood sample volume meets the following requirements:

**Table 7-1 Whole Blood Sample Volume**

Sampling and sample modes	Tube Position	Cap open?
CT-WB	Regular tube/Micro-WB tube	Either
AL-WB	Tube rack	No

1. Use evacuated blood collection tubes to collect venous blood samples, or centrifugal tubes to collect peripheral blood samples.

**NOTE**

- Use tubes matching adapter. For details, see 3.4Supported Tubes, Tube Racks and Adapters. For details about other supported tubes that are not specified here, consult your local sales representative or Mindray Customer Service Department.

2. Well mix the blood sample and EDTA K<sub>2</sub>/EDTA K<sub>3</sub> anticoagulant in the test tube rapidly.

**CAUTION**

- Be sure to use the Mindray-specified disposable products like blood collection tubes. Do not reuse disposal products, otherwise the result may be inaccurate.
- Samples may volatilize if the sample tube cap is opened for too long time, which will result in incorrect analysis results.
- To attain accurate analysis results, make sure the sample volume meets requirements. Otherwise, the results may be inaccurate.
- The samples stored under refrigeration condition (2°C to 8°C) must be kept at room temperature for at least 15 minutes before analysis.
- Be sure to mix any sample that has been prepared for a while before running it. Otherwise, the result may be inaccurate.
- For the samples to be tested for ESR, run them within 24 hours after collection if they are stored under refrigeration condition (2°C to 8°C).
- For the whole blood samples to be tested for WBC differential or MPV analysis, run them within 8 hours after collection if stored at room temperature.
- The samples not to be tested for WBC differential, MCV or PLT counts can be stored in a refrigerator at the temperature of 2°C to 8°C for 24 hours after collection.

**NOTE**

- Capillary samples cannot be tested for ESR.
- Gently squeeze when collecting capillary blood. Pressing hard will make body fluid mix into the blood, thus reducing the reliability of the measurement results.
- Before making analysis, wait for at least 5 minutes after capillary sample. You are advised to complete the test within 1 hour

**7.5.2 Preparing Predilute Samples (For PD Mode)**

Prepare the prediluted samples in the ratio of 20:100 (venous blood/capillary blood:diluent).

To attain accurate analysis results, make sure the predilute sample volume meets the following requirements:


**Table 7-2 Preparing predilute samples (for PD mode)**

Sampling and sample modes	Tube Position	Cap open?
CT-PD	Micro-WB/PD/BF Tube Position	Yes

1. Use a centrifugal tube to collect venous blood or capillary blood.


**NOTE**

- Gently squeeze when collecting capillary blood. Pressing hard will make body fluid mix into the blood, thus reducing the reliability of the measurement results.

2. On the "Count" screen, tap  button.
- √ A pop-up window appears.
3. Switch to "CT Mode" screen and select "PD".

- ✓ The analyzer switches to the predilute mode.
- ✓ When the analyzer is in the predilute mode, the software status bar lights in orange.
- 4. Tap **"Diluent"** in the pop-up window.
- 5. Make sure the tube position switch is at the regular tubes position side and proper adapter is installed. Then uncapped the centrifugal tube filled with prediluted samples and place it at Micro-WB tube position.
- 6. Tap **"OK"** button.
- ✓ The analyzer starts to dispense diluent.

## NOTE

- You can tap **"OK"** button for multiple times to dispense the diluent. The number shown on the **"Cancel"** button represents how many times you have dispensed diluent.  indicates that you have dispensed diluent twice.
  - The analyzer dispenses 100µL of diluent at a time.
7. After completing dispensing diluent, tap **"Cancel"** button.

## CAUTION

- After mixing the capillary sample with the diluent, be sure to wait at least for 3 minutes, and mix the sample again before running the sample. Otherwise, the results may be inaccurate.
- Fully mix the blood sample with the diluent for reaction. Run the prediluted samples within 30 minutes after the dilution; otherwise the measurement results may be inaccurate.
- Be sure to mix any sample that has been prepared for a while before running it. Otherwise, the result may be inaccurate.
- Samples may volatilize if the sample tube cap is opened for too long time, which will result in incorrect analysis results.
- If there is much prediluted sample adhering to the tube cap or the tube wall, the analyzer may not be able to aspirate sufficient volume of the sample.
- When preparing prediluted samples, slightly tap the bottom of the EP tubes with a finger. There should be no less than 100µL of prediluted sample settled at the tube bottom.

## NOTE

- You can also dispense diluent by pipette into the tube.
- Be sure to evaluate predilute stability based on your laboratory's sample population and sample collection techniques or methods.

### 7.5.3 Preparing Body Fluid Samples (For BF Mode)

The subtypes of body fluid supported by the current analyzer include cerebrospinal fluid, pleural fluid, ascitic fluid, and synovial fluid.

To attain accurate analysis results, make sure the sample volume meets the following requirements:

**Table 7-3 Body fluid sample volume**

Sampling and sample modes	Tube Position	Cap open?
CT-BF	WB/BF/Probe Cleanser or Micro-WB/PD/BF Tube Position	Yes

1. Use evacuated blood collection tubes, centrifugal tubes to collect body fluid samples.



### ⚠ CAUTION

- To ensure sample stability, process serous cavity fluid (pleural fluid and ascitic fluid) samples and synovial fluid samples with EDTA anticoagulant. It is not recommended to process cerebrospinal fluid samples with anticoagulant.
- To attain accurate analysis results, make sure the sample volume meets requirements. Otherwise, the results may be inaccurate.
- The samples stored under refrigeration condition (2°C-8°C) must be kept at room temperature for at least 15 minutes before analysis. Otherwise, the results may be inaccurate.
- To attain accurate analysis results, make sure the sample volume meets requirements. Otherwise, the results may be inaccurate.
- Samples may volatilize if the vial is opened for too long time, which result in incorrect analysis results.
- Misleading results may occur if the body fluid sample has floccules, clots, cryoglobulin, leukocyte clumps, bacteria, or high protein. Follow your laboratory protocol to deal with such samples.

2. Mix the sample according to your laboratory's protocol.

### NOTE

- Slightly mix the body fluid samples.

## 7.5.4 Placing Barcode Labels

To ensure good readability of the barcode, you must place the label right on the region. To ensure good readability of the barcode, you must place the label right on the region as shown in **Figure 7-4** *Where to place the barcode label*, and place the label correctly as shown in **Figure 7-5** *How to place the barcode label*.

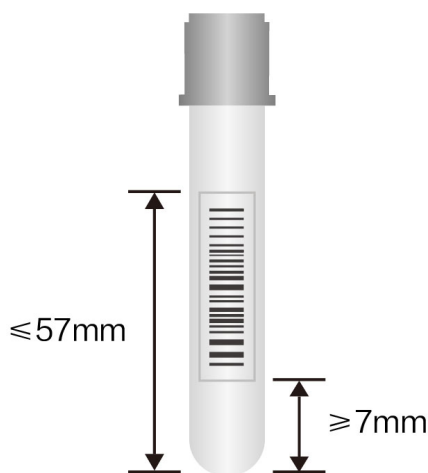


Figure 7-4 Where to place the barcode label

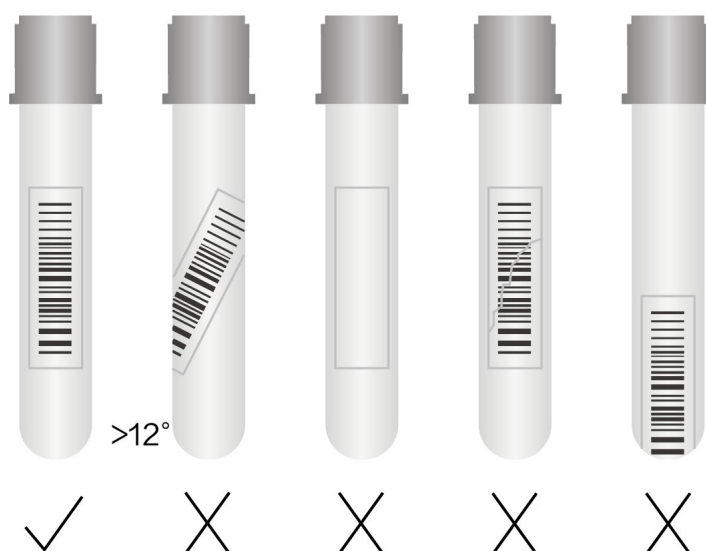



Figure 7-5 How to place the barcode label

## 7.6 Running Samples under Closed-tube Mode

### 7.6.1 Setting up Analysis Orders

You can set analysis order information on the main unit of Auto Hematology Analyzer.

1. Tap  on the "Count" screen.  
✓ The pop-up window appears.
2. Tap "CT Mode" button.  
✓ The sample compartment reaches out.

#### NOTE

- When the sample compartment is reaching out, do not block it.

3. Select the desired sampling mode and sample mode.
  - ◆ WB
  - ◆ PD
  - ◆ BF

## NOTE

- If the main unit is connected to LIS or labXpert, you can check "Auto acquire test mode". The system will automatically read the sample mode information set at the LIS or labXpert end.

4. In the "Sample ID" column, manually enter the Sample ID, or use an eternal barcode scanner to scan the barcode label on the tube to enter Sample ID into the "Sample ID" field.

## NOTE

- If you have set "Entry of next sample ID" to "Auto Increase", you only need to enter the sample ID for the first sample. The subsequent sample IDs will automatically increase by 1 based on the previous one.
- If you have set "Entry of next sample ID" to "Manual entry", you need to enter the sample ID for each sample.
- For more information about the sample ID entry method, refer to 6.3.3.1 *Getting sample information*.
- 20 characters are allowed for sample ID maximally (including prefix); the ID must end with a number, and must not be consisted of "0" only.

5. Select the desired test panel according to your model.

Test Panel	Meaning	Description
CBC	CBC	Complete blood count
CD	CBC+DIFF	Complete blood count + WBC differentiation tests
Ret	Ret	Complete blood count
CR	CBC+Ret	Complete blood count + Reticulocyte-related tests
CDR	CBC+DIFF+Ret	Complete blood count + WBC differentiation tests + Reticulocyte-related tests
ESR	ESR	Erythrocyte Sedimentation Rate
CD/WBC-3X	CBC+DIFF, WBC 3X mode	Complete blood count + WBC differentiation tests, using the extended sampling method (WBC 3X)

Test Panel	Meaning	Description
CDR/WBC-3X	CBC+DIFF+Ret, WBC 3X mode	Complete blood count + WBC differentiation tests + Reticulocyte-related tests, using the extended sampling method (WBC 3X)
CR/PLT-5X	CBC+ Ret, PLT 5X mode	Complete blood count + Reticulocyte-related tests, using the extended sampling method (PLT 5X)
CDR/PLT-5X	CBC+DIFF+Ret, PLT 5X mode	Complete blood count + WBC differentiation tests + Reticulocyte-related tests, using the extended sampling method (PLT 5X)

## NOTE

- In the "Test Panel" area of the pop-up window, only the most commonly used analysis modes are displayed. You can tap "Other Mode" to select other desired analysis modes.
- The Ret test panel only applies to the BC-760[R]/BC-780[R] model.
- Peripheral blood samples cannot be used for ESR test panel.
- The test panels vary with the models with different configurations. For details, refer to the configuration interface of your model.
- For the description of the test panels supported by the analyzer, refer to *B.5.1 Sample modes, test panel, and applicable model*.

## 7.6.2 Performing Sample Analysis

### 7.6.2.1 Analyzing samples in whole blood mode

Check the following before analysis:

- Sample(s) prepared in accordance with the laboratory protocol (refer to *7.5.1 Preparing Whole Blood Samples (For WB Mode)* and *7.5.4 Placing Barcode Labels*).
- Analysis orders are set (refer to *7.6.1 Setting up Analysis Orders*).
- The entered sample ID, sampling mode, sample mode, and the test panel are strictly in accordance with the sample to be run.
- The analyzer is ready to run samples (i.e. the analyzer indicator stays in green). Sample compartment is open.
- The bottom area of the "Count" screen displays the "Mode" as "CT-WB".

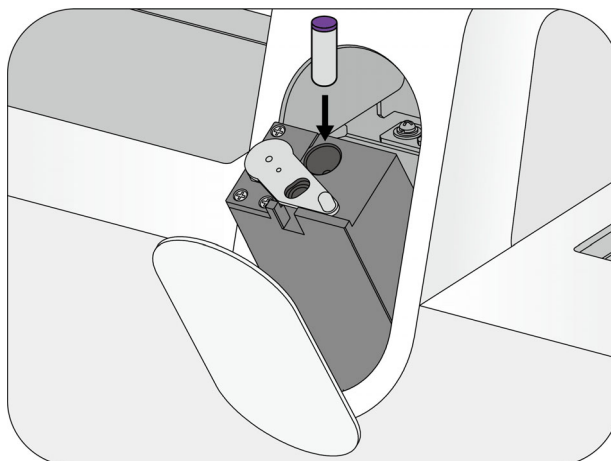
## NOTE

- If you start sample analysis immediately after selecting work mode, then the default reference range is "General". After the analysis finishes, the analyzer will flag results per the reference range "General". To change the category of a reference group, follow the instructions in *6.3.4.2 Ref. Range Setup (Administrators)*.

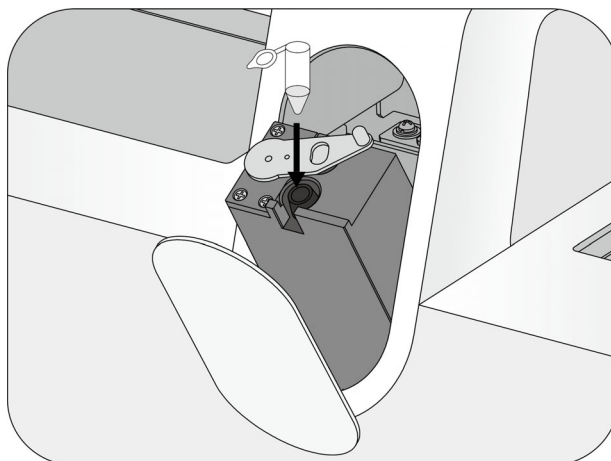
Follow below instructions:

1. Mix the sample.
  - If you are running a venous blood sample collected in an evacuated blood collection tube, shake the tube as instructed by the picture below to mix the sample thoroughly.
  - If you are running a capillary blood sample collected in a centrifugal tube, cap the tube and shake the capped tube to mix it thoroughly.
2. Place the sample tube into the appropriate tube position in the sample compartment.

- If you are running a sample collected in an evacuated collection tube, make sure the tube position switch is at the Micro WB tube position side. Then place the evacuated collection tubes into the regular position of sample compartment..



- If you are running capillary whole blood sample collected in centrifuge tubes, make sure the tube position switch is at the regular tube side and proper adapter is installed, then uncup the centrifuge tube and place it at Micro-WB tube position.



## NOTE

- **When the sample compartment is reaching out, do not block it.**

3. Press the **"Start Count"** key on screen to start sample analysis.
  - ✓ The sample compartment closes, and the sampling probe automatically aspirates sample.
4. After the analyzer finishes aspirating sample, the sample compartment opens. You can remove the sample safely.
  - ✓ The analyzer automatically analyzes the sample, the analyzer indicator is flickering in green.
  - ✓ When the analysis completes, the analyzer indicator returns to "Ready" status (stay in green).
  - ✓ The screen displays the current sample results, histograms, scattergrams and flags (if there is).

## NOTE

- **If the analyzer detects clogging or bubbles during the analysis, the corresponding error message will be displayed in the error message area and the results of all related parameters will be invalidated. See 13Troubleshooting for solutions.**
- **When the ambient temperature is out of the specified operating range, the analyzer will alarm you for abnormal ambient temperature and the analysis results may be unreliable. See 13Troubleshooting for solutions.**

### 7.6.2.2 Analyzing samples in predilute mode

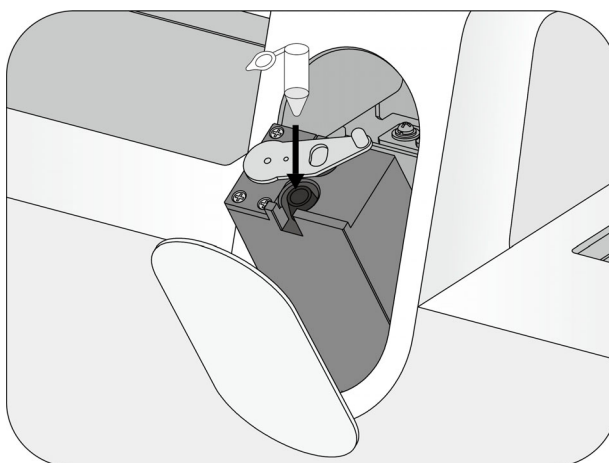
Check the following before analysis:

- Sample(s) prepared in accordance with the laboratory protocol (refer to **7.5.2Preparing Predilute Samples (For PD Mode)** and **7.5.4Placing Barcode Labels**)
- Analysis orders are set (refer to **7.6.1Setting up Analysis Orders**).
- The entered sample ID, sampling mode, sample mode, and the test panel are strictly in accordance with the sample to be run.
- The analyzer is ready to run samples (i.e. the analyzer indicator stays in green). Sample compartment is open.
- The bottom area of the "Count" screen displays the "Mode" as "CT-PD". The bottom area of the "Count" screen lights in orange.

#### NOTE

- **If you start sample analysis immediately after selecting work mode, then the default reference range is "General". After the analysis finishes, the analyzer will flag results per the reference range "General". To change the category of a reference group, follow the instructions in 6.3.4.2Ref. Range Setup (Administrators).**

1. Shake the capped tube of the predilute sample to mix it thoroughly.
2. Make sure the tube position switch is at the regular tube side and proper adapter is installed, then uncap the centrifuge tube filled with prediluted samples and place it at Micro-WB tube position.



#### NOTE

- **When the sample compartment is reaching out, do not block it.**

3. Press the "Start Count" key on screen.
- ✓ If "Predilute mode prompt" is enabled, a dialog box will pop up to remind you that the current analysis mode is PD.

#### NOTE

- **For more information about the enabling or disabling predilute mode prompt, see 6.3.3.2Other Settings.**

4. Press the "Start Count" key on screen to start sample analysis.
- ✓ The sample compartment closes, and the sampling probe automatically aspirates sample.
5. After the analyzer finishes aspirating sample, the sample compartment opens. You can remove the sample safely.
- ✓ The analyzer automatically analyzes the sample, the analyzer indicator is flickering in green.
- ✓ When the analysis completes, the analyzer indicator returns to "Ready" status (stay in green).

- ✓ The screen displays the current sample results, histograms, scattergrams and flags (if there is).

## NOTE

- If the analyzer detects clogging or bubbles during the analysis, the corresponding error message will be displayed in the error message area and the results of all related parameters will be invalidated. See *13Troubleshooting* for solutions.
- When the ambient temperature is out of the specified operating range, the analyzer will alarm you for abnormal ambient temperature and the analysis results may be unreliable. See *13Troubleshooting* for solutions.

### 7.6.2.3 Analyzing samples in body fluid mode

Check the following before analysis:

- Sample(s) prepared in accordance with the laboratory protocol (refer to *7.5.3Preparing Body Fluid Samples (For BF Mode)* and *7.5.4Placing Barcode Labels*).
- Analysis orders are set (refer to *7.6.1Setting up Analysis Orders*).
- The entered sample ID, sampling mode, sample mode, and the test panel are strictly in accordance with the sample to be run.
- The analyzer is ready to run samples (i.e. the analyzer indicator stays in green). Sample compartment is open.
- The bottom area of the "Count" screen displays the "Mode" as "OV-BF-CD".

## NOTE

- If you start sample analysis immediately after selecting work mode, then the default reference range is "General". After the analysis finishes, the analyzer will flag results per the reference range "General". To change the category of a reference group, follow the instructions in *6.3.4.2Ref. Range Setup (Administrators)*.

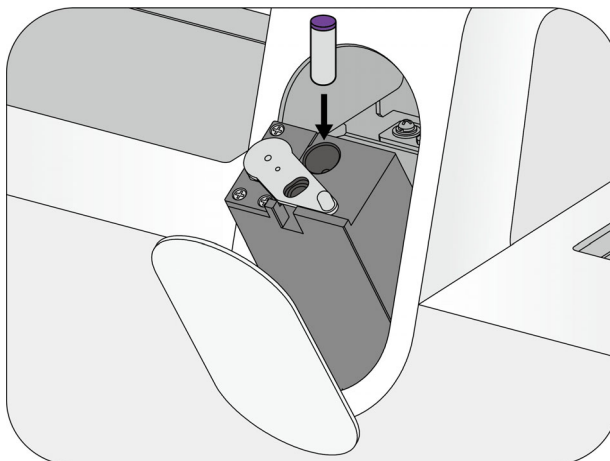
1. Mix the sample.
  - If you are running a sample collected in an evacuated blood collection tube, shake the tube as instructed by the picture below to mix the sample thoroughly.
  - If you are running a sample collected in a centrifugal tube, cap the tube and shake the capped tube to mix it thoroughly.

## CAUTION

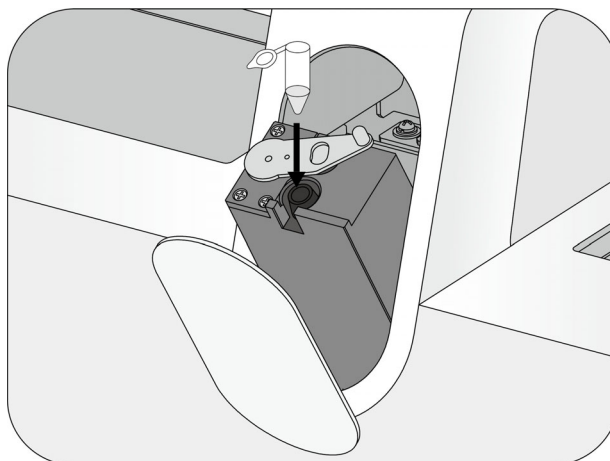
- To attain accurate analysis results, make sure the sample volume meets requirements. Otherwise, the results may be inaccurate.
- Misleading results may occur if the body fluid sample has floccules, clots, cryoglobulin, leukocyte clumps, bacteria, or high protein. Follow your laboratory protocol to deal with such samples.

2. Place the sample tube into the appropriate tube position in the sample compartment.

- If you are running a sample collected in an evacuated collection tube, make sure the tube position switch is at the Micro WB tube position side. Then place the evacuated collection tubes into the regular tube position of sample compartment.



- If you are running body fluid sample collected in centrifuge tubes, make sure the tube position switch is at the regular tube side and proper adapter is installed, then uncap the centrifuge tube and place it at Micro-WB tube position.



3. Press the "**Start Count**" key on screen to start sample analysis.
- ✓ The sample compartment closes, and the sampling probe automatically aspirates sample.
4. After the analyzer finishes aspirating sample, the sample compartment opens. You can remove the sample safely.
- ✓ The analyzer automatically analyzes the sample, the analyzer indicator is flickering in green.
- ✓ When the analysis completes, the analyzer indicator returns to "Ready" status (stay in green).
- ✓ The screen displays the current sample results, histograms, scattergrams and flags (if there is).

## NOTE

- If the analyzer detects clogging or bubbles during the analysis, the corresponding error message will be displayed in the error message area and the results of all related parameters will be invalidated. See *13Troubleshooting* for solutions.
- When the ambient temperature is out of the specified operating range, the analyzer will alarm you for abnormal ambient temperature and the analysis results may be unreliable. See *13Troubleshooting* for solutions.



## 7.7 Running Samples

### 7.7.1 Setting up Analysis Orders

You can set analysis order information on the main unit of Auto Hematology Analyzer.

1. Tap "⏪" on the "Count" screen.
- ✓ The pop-up window appears.
2. Select "AL Mode".

3. Enter the sample ID, rack No., and tube No. as needed.

#### NOTE

- If built-in barcode scanner is configured on your analyzer, the analyzer will scan the sample ID automatically after you check "Auto-Scan sample ID". Make sure that the barcode labels are intact and readable.
- If you uncheck "Auto-Scan Sample ID", manually enter the first Sample ID, and the following sample IDs will automatically increase. For more details, refer to 6.3.3.1 *Getting sample information*.
- Totally 20 characters are allowed for sample ID maximally (including prefix); the ID must end with a number, and must not be consisted of "0" only.

4. Select the desired test panel according to your model.

Test panel	Meaning	Description
CBC	CBC	Complete blood count
CD	CBC+DIFF	Complete blood count + WBC differentiation tests
Ret	Ret	Reticulocyte-related tests
CR	CBC+Ret	Complete blood count + Reticulocyte-related tests
CDR	CBC+DIFF+Ret	Complete blood count + WBC differentiation tests + Reticulocyte-related tests
ESR	ESR	Erythrocyte sedimentation rate test

Test panel	Meaning	Description
CR/PLT-5X	CBC+ Ret, PLT-5X mode	Complete blood count + Reticulocyte-related tests, using the extended sampling method (PLT-5X)
CDR/PLT-5X	CBC+DIFF+Ret/PLT-5X mode	Complete blood count + WBC differentiation tests + Reticulocyte-related tests, using the extended sampling method (PLT-5X)
CD/WBC-3X	CBC+DIFF, WBC-3X mode	Complete blood count + WBC differentiation tests, using the extended sampling method (WBC-3X)
CDR/WBC-3X	CBC+DIFF+Ret?WBC-3X mode	Complete blood count + WBC differentiation tests + Reticulocyte-related tests, using the extended sampling method (WBC-3X)

## NOTE

- Only some commonly used analysis modes are displayed on the "Test Panel" area of pop-up window. You can tap "Other Mode" button to select other desired analysis modes.
- The Ret test panel only applies to the BC-760[R]/BC-780[R] model.
- Capillary samples cannot be tested for ESR.
- The test panels vary with the models with different configurations. For details, refer to the configuration interface of your model.
- For the description of the test panels supported by the analyzer, refer to *B.5.1 Sample modes, test panel, and applicable model*.

## 7.7.2 Performing Sample Analysis

### CAUTION

- Do not run prediluted samples in autoloading mode. Otherwise the results may be inaccurate.

Check the following before analysis:

- Sample(s) prepared in accordance with the laboratory protocol (refer to *7.5.1 Preparing Whole Blood Samples (For WB Mode)* and *7.5.4 Placing Barcode Labels*).
  - Analysis orders are set (refer to *7.7.1 Setting up Analysis Orders*).
  - Make sure the entered sample ID, rack No., tube No. and the analysis mode are strictly in accordance with the sample to be run.
  - The analyzer is ready to run samples (i.e. the analyzer indicator stays in green).
  - The "Next Sample" area of the "Count" screen displays the "Mode" as "AL-WB".
  - When you have selected "Auto-Scan Sample ID" as the sample ID entry method, make sure the barcode labels are clear, complete and are properly pasted on the sample tubes.
1. Place the racks with tubes on the right tray of the autoloader, with the side opening facing the analyzer.
  2. (Optional) Tap "Start Count" on the "Count" screen.

## NOTE

- When "Auto load tube racks" is enabled, the analyzer automatically loads the tube racks on the right tray of the autoloader for sample analysis. For the instruction of enabling "Auto load tube racks", refer to *6.3.7 Setting up Functions for the Auto-Loading of Samples (Menu > "Setup" > "Auto-loading") (administrators)*.
- Repeated piercing of the vacuum blood collection tube can damage the rubber cap. The resulting debris can cause inaccurate analysis results. The recommended number of piercing actions per vacuum tube is not more than 3.
- If abnormal power interruption occurs after analysis is started, you should remove the racks manually and open the front cover to see if there are dropped tubes to be removed.

- ✓ The analyzer automatically analyzes the sample. During this process, the analyzer indicator is flickering in green.
- ✓ Save results from each sample analysis to "Table Review".
- 3. (Optional) Review the autoloading summary.

If you have enabled **"Display summary after autoloading finished"** on the **"Auto-loading"** setup screen, a statistical result dialog box will display when the analyses are finished.

Autoloading Summary		
Item	Statistics	Note
Finished blood samples	2	
Finished QC samples	0	
Rack vacancies	0	Details...
ID reading errors	0	Details...
Invalid racks	0	
Clog	0	Details...
Aspiration abnormal	2	Details...
Tap the "Details" buttons where the font color is red to check		
OK		

- a (Optional) If the results of certain items on the statistical result dialog box are not 0, tap the **"Details..."** button to check the sample ID, analysis time and sample position of the corresponding samples.

Details			
Sample ID	Time	Tube Pos.	
1	11:29	1-1	▲
2	11:29	1-2	▲
			▼
			▼
OK			

- b Tap **"OK"** to close the dialog box.
4. When the autoloading analyses are finished, all the tube racks are moved to the left tray of the autoloader automatically, and then you can remove the racks of tubes safely.

## NOTE

- When you have selected **"Auto-Scan Sample ID"** as the sample ID entry method, but the sample ID scanned is invalid, then **"Inv. test time \_ tube position"** will be the sample ID.

- If "Auto-Scan sample ID" is selected and the tube No. scanned is invalid, then "???" will be the tube No.. For the information of enabling or disabling "Auto-Scan sample ID", refer to 6.3.3.1 *Getting sample information*.
- For the information of enabling or disabling "Display summary after autoloading finished", refer to 6.3.7 *Setting up Functions for the Auto-Loading of Samples (Menu > "Setup" > "Auto-loading") (administrators)*.

### 7.7.3 Stop Count


If you need to stop auto-loading analysis during the analysis process, follow below instructions.

During the analysis process, tap the "Stop Count" button on the pop-up window.

- ✓ The analyzer will stop running after the current sample is analyzer,
- ✓ The rack of the sample will move to the left tray of the autoloader.

## 7.8 STAT

Use this function to insert STAT samples during the auto-loading analysis process.

1. On the "Count" screen, tap  and then tap the "STAT" button on the pop-up window.
  - ✓ After the current sample has been analyzed, the analyzer will stop the autoloading operation and switch from AL Mode to CT Mode.
  - ✓ The pop-up window is switched to "CT Mode" page.
  - ✓ The sample compartment reaches out.
2. Enter the sample information and select the desired sample mode and analysis mode.
3. Place STAT samples in the sample compartment.
4. Select "Start Count" to start STAT samples analysis.
5. When STAT samples analysis completes, the sample compartment reaches out. Take out STAT samples.
6. (Optional) Repeat steps 2 to 5 to complete all STAT samples analysis.
7. Exit STAT using either of the following methods.
  - Exit STAT manually.

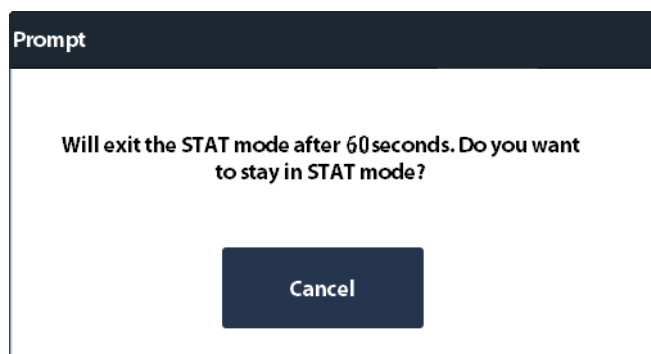
Select "Exit STAT".

- Exit STAT automatically.

### NOTE

- For the instruction of enabling "Auto exit STAT" function, refer to 6.3.3 *Auxiliary Setup ("Menu" > "Setup" > "Auxiliary Setup")*.

- a When you do not place STAT samples in the sample compartment during the pre-defined time range, a prompt as shown in the figure below is displayed.



- b (Optional) To continue STAT samples analysis, select "Cancel".
- c The analyzer automatically exits STAT when count-down ends.

- ✓ The analyzer switches back to **"AL Mode"**, then you can proceed with analysis under auto-loading mode.

## NOTE

- For the information of setting analysis order under closed-tube sampling mode, refer to **7.6.1 Setting up Analysis Orders**.
- When the sample mode or sampling mode changes, the analyzer will switch the mode automatically and a pop-up message will be displayed.

## 7.9 Entering/Exiting Standby Status

When the analyzer is free from fluidic operations or when it reaches the set standby time, the analyzer automatically enters the standby status.

When the user starts sample tests, or performs any operation that initiates fluidic system actions or moving parts actions, the analyzer automatically exits standby.

## NOTE

- Refer to **6.3.5 Maintenance ("Menu" > "Setup" > "Maintenance") (Administrators)** for how to edit waiting time before entering the standby mode.

## 7.10 Shutting down the Analyzer

### CAUTION

- Be sure to shut down the analyzer strictly as instructed. Otherwise, the tubes may be polluted, leading to wrong results.
- Do not start up the analyzer immediately after shutdown. Wait at least 10 seconds.

### 7.10.1 Shutting down the analyzer

Follow below instructions to perform the shutdown procedure.

1. Tap **"Menu"** -> **"Shutdown"** on the **"Count"** screen.
2. Tap **"OK"** to perform the shutdown procedure. The dialog box disappears automatically.
3. (Optional) When a dialog box pops up, asking you to perform Probe Cleanser maintenance, present Probe Cleanser to the sampling probe as instructed, and start Probe Cleanser maintenance.

## NOTE

- For how to perform Probe Cleanser maintenance, see **12.4 Probe Cleanser Maintenance**.

- ✓ Prompt "The analyzer will power off" displays on the screen.

## NOTE

- When the screen goes black, the analyzer exits normal working conditions and just certain modules continue working.
4. (Optional) Turn off the power switch on the back of the analyzer.
  5. Empty the waste in the waste container, and then dispose of the waste properly.

## 7.11 Viewing Guidance Videos in iHelp

Video Guidance is available in iHelp on analyzer. You can view guidance videos and know how to operate your analyzer.

Follow below instructions to view guidance videos.

1. Tap "**Menu**" > "**Service**" > "**iHelp**" on the "Count" screen or tap "**iHelp**" utility button.
2. Tap video thumbnails to play videos.

### **NOTE**

- 
- **If you have any questions about guidance videos, contact out customer service department.**
-

# 8 Reviewing Sample Results

## 8.1 Introduction

After every analysis cycle, the analyzer automatically saves the analysis results into the sample database.

Up to 130,000 analysis results can be stored in the BC-760[B]/BC-760[R] analyzers. Up to 150,000 analysis results can be stored in the BC-780[R] analyzers. When the maximum number has been reached, the newest result will overwrite the oldest.

You can review all the analysis results, scattergrams and histograms.

## 8.2 Reviewing Sample Results

### NOTE

- The screenshots in this Operator's Manual are only for reference. The actual screens are subject to the configuration of your analyzer.













### 8.2.1 Entering the "Table Review" Screen

Select "Menu" > "Table Review" or tap the "Table Review" utility button to enter the "Table Review" screen.



The sample results are sequentially displayed from left to right on the "Table Review" screen, the latest on the utmost right of the table, including the sample information, parameter results, and patient information.

### 8.2.2 Operations on the "Table Review" Screen

#### 8.2.2.1 Browsing sample record

Tap  / ,  / , and  /  to turn to the left/right column (page).  
Tap  / ,  / , and  /  to turn to the previous/next row (page).

	1	2	3*
Sample ID	1	2	3
Sample State	Not printed	Not printed	Not printed
Count Mode	WB	WB	WB
Test Panel	CD	CDR	CD+ESR
Date	12-07-2021	12-07-2021	12-07-2021
Time	19:33	19:33	19:33
Tube Pos.			
WBC	H 10.06	9.99	H 10.01
Neu#	L 1.00	L 1.00	L 1.00
Lym#	1.00	1.00	1.00
Mon#	1.00	1.00	1.00
Eos#	H 1.00	H 1.00	H 1.00
Bas#	H 1.00	H 1.00	H 1.00
IMG#	1.00	1.00	1.00
Neu%	L 0.010	L 0.010	L 0.010

SearchGraphComm.ExportCVEdit Info.Validate



Figure 8-1 Table review - parameter results

	1	2	3*	
Sample ID	1	2	3	
Sample State	Not printed	Not printed	Not printed	
Date of Birth	***	***	***	
Age	***	***	***	
Patient Type				
Ref. group	General	General	General	
Department				
Bed No.				
Draw Date				
Draw Time				
Delivery Date				
Delivery Time				
Clinician				
Operator	RD	RD	RD	
Validated By				

◀

◀◀

◀

▶

▶▶

▶

Search

Graph

Comm.

Export

CV

Edit Info.

Validate

➔

Figure 8-2 Table review - patient information



Tap  /  to switch to the second tool bar.



Figure 8-3 tool bar - 1



Figure 8-4 tool bar - 2

The position of the current sample result and the total sample results are shown in the form of "**Position/Total**" at the bottom left the screen.

### NOTE

- When the WBC result is lower than WBC DIFF threshold ( $0.1 \times 10^9$  by default), the analyzer does not display the specific result. To review the result, contact our Customer Service Department.

#### 8.2.2.2 Selecting/deselecting records

- Tap the record that you want to select to select it.
  - Tap a selected record to deselect it.
  - Tap the records that you want to select one by one to select multiple records.
- ✓ The selected sample records are highlighted.

#### 8.2.3 Searching for Sample Records

You can search sample records that match defined conditions.

- Select "**Menu**" > "**Table Review**" or tap the "**Table Review**" utility button to enter the "**Table Review**" screen.
- Tap "**Search**" on the tool bar.



✓ The following dialog box displays.

3. Define the searching conditions.

See below for setting descriptions.

Search today's samples by sample status:

<b>Not Validated Today</b>	Tap " <b>Not Validated Today</b> ", " <b>Not Printed Today</b> ", or " <b>Not Transmitted Today</b> " to search for the samples on the current day that are not validated, printed or transmitted.
<b>Not Printed Today</b>	
<b>Not Transmitted Today</b>	

Search for sample records that match defined conditions.

You can define one or more searching conditions.

When you have defined more than one searching condition, the analyzer will search for the sample records that match all defined conditions.

<b>Sample ID</b>	Enter " <b>Sample ID</b> " as a search condition, the analyzer searches for and displays all sample records whose sample ID includes the entered sample ID.	For example, if you enter "1235" into the " <b>Sample ID</b> " field, the analyzer will search for all sample records whose sample ID includes "1235".
<b>Patient ID</b>	Enter " <b>Patient ID</b> " as a search condition, the analyzer searches for and displays all sample records whose patient ID includes the entered sample ID.	For example, if you enter "1235" into the " <b>Patient ID</b> " field, the analyzer will search for all sample records whose patient ID includes "1235".
<b>Name</b>	Enter patient name as a search condition. The analyzer will search for all sample records whose patient name matches the defined name.	/
<b>Date</b>	Define the test time period in the " <b>Date</b> " fields as a search condition. The analyzer searches for all sample records tested between the defined time period of " <b>Date</b> ".	The " <b>Date</b> " range is set to the current date by default.
<b>Sample No.</b>	Enter " <b>Sample No.</b> " as a search condition, the analyzer searches for and displays all sample records whose sample ID includes the entered sample ID.	/
<b>Sample State</b>	Check " <b>Not validated</b> ", " <b>Not printed</b> ", or " <b>Not transmitted</b> ", and the analyzer searches from all sample records that match the checked state.	You can check for more than one sample state, and the analyzer searches from all sample records that match the checked states.

<b>"Auto select searched record" check box</b>	When <b>"Auto select searched record"</b> is checked, the analyzer highlights and displays all searched results.	/
--	--	---

4. Tap **"OK"**.
- ✓ All searched results display on the screen.
5. (Optional) Tap **"Cancel"** to return to the **"Table Review"** screen.

## 8.2.4 Graph Review

Follow below instructions:

1. Select **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Select one or more sample records of which you want to review the graph data.  
✓ The selected sample record is highlighted.
3. Tap **"Graph"** to go to the **"Graph"** screen.

See below for relevant operations on the **"Graph"** screen:

### 8.2.4.1 Previous/Next

Tap **"Previous"** or **"Next"** to browse the previous or next samples.

### 8.2.4.2 Table Review

Tap **"Table Review"** to enter the **"Table Review"** screen.

### 8.2.4.3 Other Parameters

The **"Other Para."** screen displays the RUO parameter results, reference results, and microscopic parameter results.

1. Select **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Select one or more sample records of which you want to review the RUO parameter results, reference results, and microscopic parameter results.  
✓ The selected sample record is highlighted.

Tap **"Graph"** to go to the **"Graph"** screen.

Tap **"Other Para."** to go to the **"Other Para."** screen.

### Research Use Only (RUO) Parameters

Tap the **"RUO Para."** tab to review the RUO parameter results.

### NOTE

- **RUO parameters and scattergrams are for research purpose only. They cannot be used for diagnosis purpose.**
- **When the user selects to use the PLT-5X test mode, or the PLT-5X test mode is automatically triggered in accordance with the re-exam settings for low PLT/WBC samples, besides the normal PLT-O result, the system will give another PLT-O result in a pair of parenthesis, and that one has one more decimal place than that of the normal one.**

### Reference Results

When the system decides a parameter result is not reliable (for example, the DIFF parameter results of certain abnormal samples), the Sample Report screen will not display the results in values, but as "\*\*\*\*\*". The estimated results will be provided on the **"Reference results"** screen. Decide if the reference results can be used for report based on a comprehensive consideration regarding information, like historical results, microscopic exam results or re-exam results.

Tap the **"Reference results"** tab to check the analysis parameter results of the corresponding samples.

### Microscopic Parameters

Before you can review microscopic parameter results, make sure you have set the microscopic parameters on the **"Setup" > "Para. Setup" > "Microsco. Para. Setup"** screen.

#### NOTE

- **For the method of defining "Microscopic Para.", refer to section 6.3.4.3Microsco. Para. Setup (Administrators).**

1. Tap the **"Microscopic Para."** tab to review the **"Microscopic Para."** results or tap the **"Result"** cell for a certain parameter to enter the microscopic parameter result.
2. (Optional) Enter the blood group information and ESR result of the sample in the **"Blood Group"** and **"ESR"** edit boxes.
3. (Optional) If necessary, tap the **"Print"** button on the **"Microscopic Para."** screen to print the microscopic parameter results.

#### NOTE

- **For the method of defining "Microscopic Para.", refer to section 6.3.4.3Microsco. Para. Setup (Administrators).**
- **You cannot edit the microscopic parameter results of a validated sample. For the method of validating samples, refer to section 8.2.9Validating/Cancelling Validation (Administrators).**

### 8.2.4.4 Editing Results (Administrators)

Administrators may edit analysis results.

1. Select **"Menu" > "Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select the sample record of which you want to edit the analysis results.  
√ The selected sample record is highlighted.

Tap **"Graph"** to go to the **"Graph"** screen.

3. Tap **"Edit Result"**.  
√ The **"Edit Result"** dialog box is displayed.
4. Tap the "Results" edit box of the parameter you want to edit and modify the parameter result.
5. Tap **"OK"** to close the dialog box.  
√ The analyzer saves the new parameter results.  
√ "E" is marked behind the parameter result data directly modified manually. If modifying a parameter result directly and manually leads to the modification of its related parameter result, "e" is marked behind its result data.

#### NOTE

- **You cannot edit the sample results of the samples that have been validated and samples for background tests. For the method of validating samples, refer to section 8.2.9Validating/Cancelling Validation (Administrators).**

Follow below instructions to restore the edited results to its original analysis results:

1. Select **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select the sample record of which you want to restore the original analysis results.  
√ The selected sample record is highlighted.
3. Tap **"Graph"** to go to the **"Graph"** screen.
4. Tap **"Edit Result"**.

- ✓ The **"Edit Result"** dialog box is displayed.
- 5. Tap **"Restore"**.
- ✓ The edited results will be changed back into the initial values.
- ✓ The "E" and "e" flags disappear.

## NOTE

- **The analyzer only saves the original measurement results of the latest 2000 samples with results modified.**

### 8.2.4.5 Reviwing Special Information

Follow below instructions:

1. Select **"Menu"** - **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select the samples of which you want to review the special information.
- ✓ The selected sample record is highlighted.
3. Tap **"Graph"** to go to the **"Graph"** screen.
4. Tap **"Special Info."**
- ✓ The screen displays the instrument-related information when analyzing the current sample.
5. (Optional) Tap the **"Error Information"** pull-down list to review the error logs (if there are) when analyzing the current sample .

### 8.2.4.6 Reviewing Traceability Information

The "Traceability" screen displays the information of the reagents and controls when analyzing the current sample.

Follow below instructions:

1. Select **"Menu"** > **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select the samples of which you want to review the traceability information.
- ✓ The selected sample record is highlighted.
3. Tap **"Graph"** to go to the **"Graph"** screen.
4. Tap **"Traceability"**.
- ✓ The "Traceability" screen displays the information of the reagents and controls when analyzing the current sample.

## 8.2.5 Communication

Before transmitting the sample records, make sure the network connection is good.

### 8.2.5.1 Transmitting selected records

1. Select samples to be transmitted on the "Table Review" screen.
2. Tap **"Comm."**, the **"Comm."** dialog box displays.
3. Select the **"Selected records"** button.
4. Tap **"OK"** to close the dialog box and start transmitting data.

### 8.2.5.2 Transmitting all records

1. Tap **"Comm."**, the **"Comm."** dialog box displays.
2. Select the **"All records"** button.

Tap **"OK"** to close the dialog box and start transmitting data.

## 8.2.6 Exporting Sample Results

Users of administrator level can export the sample records, graphic data, flags, and other parameters of the selected sample records to a USB flash drive.

Before exporting sample records, make sure that you have inserted a safe USB flash drive into the USB port on the analyzer.

### CAUTION

- **The USB port of the analyzer is only used to connect to a designated peripheral device. For details about supported devices and models, refer to *B.7 Requirements for Input/Output Devices*.**
- **The user should ensure the data safety of the USB devices connecting to the analyzer.**

### 8.2.6.1 Exporting some or all sample records

Follow below instructions to export selected or all sample records:

1. Select **"Menu"** > **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select one or more samples records that you want to export; to export all the sample records, go to the next step.
3. Tap **"Export"**.  
√ The **"Export"** dialog box displays.
4. To export some sample records, tap **"Selected records"** in the **"Export Range"** area; to export all the sample records, tap **"All records"** in the **"Export Range"** area.
5. (Optional) To export **"Flags"**, check **"Flags"**.
6. Tap **"OK"**.  
√ The analyzer exports the corresponding sample records to the USB device.

## 8.2.7 Calculating CV Values

Follow the instructions below to calculate the CV values of sample results.

1. Select **"Menu"** > **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select multiple sample records for which you want to calculate the CV values.
3. Tap **"CV"**.  
√ The screen displays the Mean value, SD value and the CV value for each parameter.

### NOTE

- **To calculate CV values, select at least 3 samples.**

## 8.2.8 Editing Information

1. Select **"Menu"** > **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Tap to select the sample records of which you want to edit the sample information.  
√ The selected sample record is highlighted.
3. Edit the sample information.
  - a Tap **"Edit Info."**

- ✓ The following dialog box displays.

**Edit Info.**

Sample ID	* 1	Patient ID	
First Name		Last Name	
Date of Birth	MM - DD - YYYY	Age	<input type="text"/> Years ▼
Gender	▼	Ref. group	General ▼
Department	▼	Bed No.	
Patient Type	▼	Draw Time	MM - DD - YYYY HH : MM
Clinician	▼	Delivery Time	MM - DD - YYYY HH : MM
Mode	OV-WB-CD	Time	12 - 07 - 2021 19 : 33
Operator	RD	Validated By	
Comments			

OK Cancel

- b Enter necessary information as needed.
4. Tap "OK" to save the entered information.

## NOTE

- For the setup of reference groups, refer to section 6.3.4.2 *Ref. Range Setup (Administrators)*.

## 8.2.9 Validating/Cancelling Validation (Administrators)

Users of administrator level can validate/cancel validate sample records.

- Select "Menu" > "Table Review" or tap the "Table Review" utility button to enter the "Table Review" screen.
  - Tap to select the sample you want to validate or cancel validation.
- ✓ The selected sample record(s) is/are highlighted.
- Tap "Validate" or "Cancel Validate".
- ✓ For the validated sample records, the "Validated By" cell displays the role of the validator.
- ✓ The "Validated By" cell for the unvalidated sample records will be empty.

## NOTE

- You cannot validate background test results or invalid sample records.

## 8.2.10 Deleting Sample Records

Administrators can delete sample records on the "Table Review" screen.

### 8.2.10.1 Deleting some or all sample records

Follow instructions below to delete some or all sample records:

- Select "Menu- "Table Review" or tap the "Table Review" utility button to enter the "Table Review" screen.
  - To delete some sample records, tap to select one or more samples to delete; to delete all the sample records, go to the next step.
- ✓ The selected record(s) is/are highlighted.

3. Tap **"Delete"**.
- ✓ The **"Delete"** dialog box displays.
4. To delete some selected sample records, tap to select **"Selected records"**. To delete all the sample records, tap to select **"All records"**.
5. Tap **"OK"**.
- ✓ The system deletes the corresponding records.

## 8.3 Flags of Analysis Results

The analyzer provides two types of flags for analysis results:

- Parameter flags;
- Flags of Abnormal Blood Cell Differential or Morphology

### 8.3.1 Parameter Flags

The analyzer provides the following parameter flags.

Flag	Message	Meaning
"H" (default) and "L" or "h" and "l" or "↑" and "↓"	High and low result flags	The analysis result exceeds the upper or lower limit of the reference range, but still within the display range
"R" (default) or "r"	Suspect flags	The analysis result is suspicious
"&"	Algorithm-rectified flag	The analysis result is rectified by the algorithm of the analyzer
@	Out of linearity range flag	The analysis result is out of the linearity range
"++++"	Out of display range flag	The analysis result is out of the display range
"*****"	Screened results	When the system decides a parameter result is not reliable (for example, the DIFF parameter results of certain abnormal samples), the Sample Report screen will not display the results in values, but as "*****".

#### NOTE

- The results of background check will not be flagged for abnormal parameters, abnormal blood cell differential or morphology.
- For the linearity range of each parameter, refer to *B.6.2Linearity Ranges*.

### 8.3.2 Flags of Abnormal Blood Cell Differential or Morphology Results

#### CAUTION

- Abnormal cells may not necessarily trigger the flags during the analysis process, it is recommended that reexamination is conducted per the operation instruction of your laboratory.

The analyzer reports the flags for the following abnormal blood cell differential or morphology.

Flag Message	Indication	Criteria
Abn. WBC scattergram	WBC scattergram abnormal	The DIFF channel scattergram is abnormal
Neutropenia	Neu# low	Neu# < 1.00×10 <sup>9</sup> /L

Flag Message	Indication	Criteria
<b>Neutrophilia</b>	Neu# high	Neu# > $11.00 \times 10^9/L$
<b>Lymphopenia</b>	Lym# low	Lym# < $0.80 \times 10^9/L$
<b>Lymphocytosis</b>	Lym# high	Lym# > $4.00 \times 10^9/L$
<b>Monocytosis</b>	Mon# high	Mon# > $1.50 \times 10^9/L$
<b>Eosinophilia</b>	Eos# high	Eos# > $0.70 \times 10^9/L$
<b>Basophilia</b>	Bas# high	Bas# > $0.20 \times 10^9/L$
<b>Leukocytopenia</b>	WBC count low	WBC < $2.50 \times 10^9/L$
<b>Leucocytosis</b>	WBC count high	WBC > $18.00 \times 10^9/L$
<b>NRBC?</b>	Possible presence of nucleated red blood cells	Presence of excessive dots in NRBC sensitive region of the scattergram
<b>Blasts?</b>	Possible presence of blast cells	Presence of excessive dots in blast sensitive region of the scattergram
<b>Abn Lymph/blast?</b>	Possible presence of abnormal lymphocytes or blasts	Presence of excessive dots in abnormal lymphocyte/blast sensitive region of the scattergram
<b>Immature Gran?</b>	Possible presence of immature granulocytes	Presence of excessive dots in immature granulocyte sensitive region of the scattergram
<b>Left Shift?</b>	Possibility of left shift	Presence of excessive dots in left shift sensitive region of the scattergram
<b>Atypical Lymph?</b>	Possible presence of atypical lymphocytes	Presence of excessive dots in atypical lymphocyte sensitive region of the scattergram
<b>RBC Histogram Abn.</b>	Abnormal distribution of RBC histogram	Abnormal distribution of RBC histogram
<b>*RET Scattergram Abn.</b>	The distribution of RET scattergram is abnormal	The distribution of RET scattergram is abnormal
<b>Dimorphic Population</b>	Dimorphic population distribution	Presence of two or more peaks on the RBC histogram
<b>Anisocytosis</b>	Anisocytosis	RDW-CV > 22 or RDW-SD > 64fL
<b>Microcytosis</b>	MCV low	MCV < 70fL
<b>Macrocytosis</b>	MCV high	MCV > 110fL
<b>Hypochromia</b>	Hypochromia	MCHC < 290
<b>Anemia</b>	Anemia	HGB < 90g/L
<b>Erythrocytosis</b>	RBC high	RBC > $6.5 \times 10^{12}/L$
<b>RBC Agglutination?</b>	RBC results possibly inaccurate	Calculate and compare special parameters
<b>Turbidity/HGB Interfere?</b>	Abnormal HGB or there may be interference	Calculate and compare special parameters
<b>Iron Deficiency?</b>	May indicate iron deficiency anemia	Calculate and compare special parameters
<b>Fragments?</b>	Possible presence of RBC fragments	Presence of abnormally distributed dots in sensitive region of the RET channel
<b>*PLT Scattergram Abn.</b>	PLT Scattergram Abn.	PLT Scattergram Abn.
<b>PLT Histogram Abn.</b>	PLT Histogram Abn.	PLT Histogram Abn.
<b>Thrombocytopenia</b>	PLT low	PLT < $60 \times 10^9/L$
<b>Thrombocytosis</b>	PLT high	PLT > $600 \times 10^9/L$



Flag Message	Indication	Criteria
<b>PLT Clump?</b>	Possibility of PLT clump	Calculate and compare special parameters
<b>Pancytopenia</b>	WBC, RBC and PLT low	WBC < 4.0 and RBC < 3.5 and PLT < 100
<b>Lipid Particles?</b>	Possible presence of lipid particles	Presence of excessive dots in lipid particle sensitive region of the scattergram
<b>Infected RBC?</b>	Possible presence of infected RBC	Presence of excessive dots in infected RBC sensitive region of the scattergram
<b>*WBC Fragments?</b>	Possible presence of WBC fragments.	Presence of abnormally distributed dots in WBC fragment sensitive region
<b>Aspiration Abnormal</b>	The sample probe is clogged or the sample volume is insufficient	Insufficient sample aspiration due to clogging of the sample probe, or insufficient sample volume
<b>*Reticulocytosis</b>	RET high	RET% > 5% or RET# > 0.20 × 10 <sup>12</sup> /L or presence of abnormally distributed dots in sensitive region of the DIFF RET channel
<b>^PLT-H Histogram Abn.</b>	The PLT-H histogram is abnormal.	The PLT-H histogram is abnormal.
<b>ESR analysis error</b>	The ESR channel may be abnormal and the ESR measurement may be inaccurate.	Analyze and monitor the measurements of the ESR channel.

## NOTE

- Items marked with \* apply only to the BC-760[R]/BC-780[R] models.
- Items marked with ^ is optional on models configured with PLT-H parameter.

**This page intentionally left blank.**

# 9

## Using the QC Program

---

### 9.1 Overview

Quality Control (QC) consists of strategies and procedures that measure the precision and stability of the analyzer. The results imply the reliability of the sample results.

QC involves measuring materials with known, stable characteristics at frequent intervals. Analysis of the results with statistical methods allows the inference that sample results are reliable.

A new lot of controls should be analyzed in parallel with the current lot prior to their expiration dates.

In order to ensure the reliability of sample analysis results, it is recommended that operators use low-level, normal-level, and high-level hematology controls. When the control of a new lot needs to be used, use the control of the new lot and the existing control in parallel for five days. Run QC twice a day. The results shall be within the reference range specified for the control.

This analyzer provides the L-J QC (including blood, BF, and protein QC) and X-B QC programs.

---

#### CAUTION

- **Use the controls and reagents specified by the manufacturer only. Store and use the controls and reagents as instructed by their instructions for use.**
- 

#### NOTE

- **For the description of reagents and controls, refer to sections 3.6.1 *Reagent* and 3.6.2 *Controls and Calibrators*.**
- 

### 9.2 L-J QC

L-J QC is named after S. Levey and E.R. Jennings, who in 1950 introduced statistical control into the clinical laboratories. Laboratories may set up allowable deviations (by standard deviations (SD) or coefficient of variation (CV%)) from the targets for the control based on their real scenario.

QC points are then plotted so the operators may easily see how far the actual QC results are from their targets. The x-axis indicates the QC date and time; and the Y-axis indicates the targets as well as the defined limits. Draw a straight line respectively at the reference value position and the upper and lower deviation limit position of control parameter along the X-axis direction on the QC graph. Lines run across the graph at the target as well as at the upper and lower limits to either side of the target value for the control. The QC points' distance from the target value is measured in SD or CV%.

You can select one of the two ways below to run controls:

- Run controls under the "QC" screen.
- Put controls together with normal samples, and run the controls on the "Count" screen

## 9.2.1 Setting up L-J QC Files (Administrators)

### 9.2.1.1 Introduction to the L-J QC file setup

You can set up L-J QC files on the QC file setup screens, as shown in **Figure 9-1 L-J QC File Setup Screen**.

Lot No.	<input type="text"/>	Level	Normal ▼	Exp. Date	MM - DD - YYYY
Mode	WB ▼	Type	Others ▼	QC Sample ID	<input type="text"/>
Test Panel	CD ▼	In Use	In Use ▼	Communication ID	<input type="text"/>

Parameter	Target	Limit (#)	Parameter	Target	Limit (#)
WBC			H-IPF		
Neu#			IPF#		
Lym#			MRV		
Mon#			FRC#		
Eos#			FRC%		
Bas#			PDW-SD		
IMG#			NRBC#		
Neu%			NRBC%		

Import File
Set Limits
Return

**Figure 9-1 L-J QC File Setup Screen**

**Table 9-1 L-J QC File**

Items	Description	Note
<b>Lot No.</b>	Find the Lot No. of controls on the vial labels of the controls	<ul style="list-style-type: none"> <li>Up to 16 digits can be entered for the lot No. You can enter characters, numbers, letters and special characters. Chinese characters are not supported.</li> <li>The lot No. is required.</li> <li>Enter the lot No. of the controls by one of the following ways: Manual entry, or using an external barcode scanner.</li> </ul>
<b>Level</b>	Levels of controls "High", "Normal", "Low"	/
<b>Exp. Date</b>	Expiration dates of the controls	The expiration date shall not be earlier than the current system date.
<b>Mode</b>	The loading and sample modes in QC tests: "AL-WB" "CT-WB" "CT-BF"	/

<b>Test Panel</b>	The analysis modes supported in QC tests: <b>CD, *CDR, *Ret, ESR, or CD+ESR</b>	<ul style="list-style-type: none"> <li>When you are using BR60 controls, select the CD test panel or *CDR test panel;</li> <li>When you are using BC-6D controls, select the CD test panel;</li> <li>When you are using the BC-RET controls, select the *RET test panel;</li> <li>When you are using the BC-BF controls, select the CD test panel;</li> </ul> <p>Note: The item with * only applies to the BC-760[R] and BC-780[R] model.</p>
<b>Type</b>	The controls are classified into two types: <b>"Others" or "Mindray"</b>	/
<b>In Use</b>	QC files in use or not: <ul style="list-style-type: none"> <li><b>In Use</b></li> <li><b>Not in Use</b></li> </ul>	If the <b>"In Use"</b> option is selected in the <b>"In Use"</b> pull down list of QC file, the QC results will be stored in the file.
<b>QC Sample ID</b>	If you are used to analyze control together with samples, you can set a unique ID for the control. The analyzer will recognize the sample as control when it reads the unique ID. After the analysis completes, the results will be saved into the QC file of the QC sample ID.	<ul style="list-style-type: none"> <li>You can enter letters, digits and all other characters on the keyboard (including special characters) for QC sample ID. Chinese and other languages are not supported (e.g. Japanese, Korean, etc.)</li> <li>If you are using external barcode scanner to scan sample IDs, make sure the QC sample IDs set on the QC file screen are the same as that of the Lot No. labels on the control vials.</li> <li>If you are manually entering sample IDs, make sure the QC sample IDs set on the QC file screen are the same as the sample IDs you entered on the "Mode" screen.</li> </ul>
<b>Communication ID</b>	If you are using 2-way LIS, LIS identifies QC results by the communication ID set here.	/
<b>Target</b>	The target values for the QC parameters Find the targets in the target sheet of the controls	/
<b>Limit</b>	The allowed deviation limit for each QC parameter Find the limits information in the target sheet of the controls	<ul style="list-style-type: none"> <li>The limits are represented by SD or by CV.</li> <li>Tap <b>"Set Limits"</b> and select to represent the deviations <b>"By SD"</b> or <b>"By CV"</b>.</li> </ul>

### 9.2.1.2 Setting up new L-J QC file

Before running a new lot of controls, you must set up a QC file for each lot of controls.

1. Tap **"Menu"-"QC"-"L-J QC"-"Setup"** in turn, to enter the L-J QC file setup screen.
2. Tap **"New"** to enter the new L-JQC file screen.
3. Enter the necessary QC file information.

You shall enter the required **"File Info."** and **"Target/Limit"** using one of the following ways:

- Reading the information provided by the manufacturer
  - a Insert the USB device saving the QC files to the USB port on the analyzer.
  - b On the new QC file screen, tap **"Import File"** and follow the software instruction to import the QC file.
- Manually enter the necessary QC file information.

#### NOTE

- For the introduction of the L-JQC file setup, refer to *Table 9-1L-J QC File*.
  - The user should ensure the data safety of the USB devices connecting to the analyzer.
4. Define **"QC Sample ID"** and **"Communication ID"**.
  5. (Optional) If necessary, set the QC file to **"In Use"** in the **"In Use"** pull-down list.

**NOTE**

- You can check or uncheck to "In Use" option on the QC file table screen to activate or deactivate the QC files.
- For files having the same "QC Sample ID" and presentation mode, only one of them can be "In Use".
- For files having the same QC type and level, only one of them can be "In Use".

6. Saving the QC file.
  - a Tap "Return" or other buttons on the screen.
- ✓ A confirm dialog box displays.
  - b Tap "Yes" to save the new QC file.

**9.2.1.3 Editing L-J QC files**

You can only edit empty L-JQC files.

**NOTE**

- You cannot edit the QC files that already have QC data.

1. Tap "Menu"-"QC"-"L-J QC"-"Setup" in turn, to enter the L-JQC file setup screen.
2. Tap to select the QC file to edit.
- ✓ The "\*" mark displays next to the "File No." of the selected QC file.
3. Tap "Edit" to enter the L-JQC file editing screen.
4. Edit the QC file as necessary.

**NOTE**

- For the introduction of the L-JQC file setup, refer to *Table 9-1L-J QC File*.

5. (Optional) If necessary, set the QC file to "In Use" in the "In Use" pull-down list.

**NOTE**

- You can check or uncheck to "In Use" option on the QC file table screen to activate or deactivate the QC files.
- For files having the same "QC Sample ID" and presentation mode, only one of them can be "In Use".
- For files having the same QC type and level, only one of them can be "In Use".

6. Saving the QC file.
  - a Tap "Return" or other buttons on the screen.
- ✓ A confirm dialog box displays.
  - b Tap "Yes" to save the new QC file.

**9.2.2 Running L-J QC Tests**

You can select one of the two ways below to run controls:

- Run controls under the "QC" screen.
- Put controls together with normal samples, and run the controls on the "Count" screen

**9.2.2.1 Running controls on the QC count screen**

Check the following before running QC analysis:

- Make sure you have set up a suitable and correct QC file for the control to be run, and the QC file is "In Use". (Refer to **9.2.1 Setting up L-J QC Files (Administrators)**.)

- Make sure you have prepared the controls in accordance with your laboratory protocols, and the requirements in the Instruction for Use of the controls.
- Make sure the analysis system is without error.

### AL-WB

Follow below instructions:

1. Select "**Menu**" > "**QC**" > "**L-J QC**" > "**Count**" or directly tap "**QC**" to enter the QC count screen.
2. Select "**File No.**" of the desired QC file corresponding to the control to be run from the "**File No.**" pull-down list.
3. Make sure the QC file information displayed on the screen is correct. Make sure the level of the control to be run is the same with the current QC file, and the control is not expired.

### NOTE

- 
- **The expiration date of expired controls is displayed in red.**
- 
4. Prepare the control as instructed by instructions for use of the controls, and paste barcode labels to the control vials (for details about pasting barcodes, see **7.5.4Placing Barcode Labels**).
  5. To run QC counts:
    - a Place the prepared controls into the tube rack.
    - b Place racks loading controls on the right tray of the autoloader, with the back of "MINDRAY" mark on the carrier facing the analyzer.
    - c Tap "**Count**" to start running.
  - √ The analyzer automatically analyzes the sample.
    - d When running finishes, you may remove the racks from the left of the autoloader.
  - √ When analysis finishes, the QC results will be displayed in the current screen and be saved in the QC file automatically.

### NOTE

- 
- **Up to 372 QC results can be saved in each QC file.**
- 
6. Perform the above procedures to continue running QC analysis if necessary.

### CT-WB/CT-BF

Follow below instructions:

1. Select "**Menu**" > "**QC**" > "**L-J QC**" > "**Count**" or directly tap "**QC**" to enter the QC count screen.
2. Select "**File No.**" of the desired QC file corresponding to the control to be run from the "**File No.**" pull-down list.
3. Make sure the QC file information displayed on the screen is correct. Make sure the level of the control to be run is the same with the current QC file, and the control is not expired.

### NOTE

- 
- **The expiration date of expired controls is displayed in red.**
- 
4. Prepare the control as instructed by instructions for use of the controls, and paste barcode labels to the control vials (for details about pasting barcodes, see **7.5.4Placing Barcode Labels**).
  5. To run QC counts:
    - a Place racks loading controls into the appropriate tube position in the sample compartment. Make sure the tube position switch is at micro-WB tube side, then place the controls tube into the regular tubes of compartment.
    - b Tap "**Count**" to start running.
  - √ The sample compartment closes, and the sampling probe automatically aspirates control.
    - c After the analyzer finishes aspirating sample, the sample compartment opens. You can remove the control safely.

- ✓ The analyzer automatically analyzes the control. During this process, the analyzer indicator is flickering in green.
- ✓ When analysis finishes, the QC results will be displayed in the current screen and be saved in the QC file automatically.

## NOTE

- Up to 372 QC results can be saved in each QC file.

6. Do the above procedures to continue running QC analysis if necessary.

### 9.2.2.2 Putting controls together with normal samples, and run the controls on the "Count" screen

After setting special "QC Sample ID" for a control under the QC setup screen, you can put the control together with normal samples, and run it on the "Count" screen.

Check the following before analysis:

- Make sure you have set up a suitable and correct QC file for the control to be run, and the QC file is "In Use". (Refer to **9.2.1 Setting up L-J QC Files (Administrators)**.)
- Make sure you have prepared the controls in accordance with your laboratory protocols, and the requirements in the Instruction for Use of the controls.
- The analyzer is ready to run samples (i.e. the analyzer indicator stays in green).
- Make sure the analysis system is without error.

Note when you set sample ID for the control:

- If you are using external barcode scanner to scan sample IDs, make sure the QC sample IDs set on the QC file screen are the same as that of the Lot No. labels on the control vials.
- If you are manually entering sample IDs, make sure the QC sample IDs set on the QC file screen are the same as the sample IDs you entered on the "Mode" screen.

Follow below instructions:

1. On the "Count" screen, tap the "Mode" button.
- ✓ The "Mode" dialog box displays.
2. Select the desired mode.

## NOTE

- Make sure the set mode is the same with that you set in the QC file for the control.

3. In the "Sample ID" column, manually enter the Sample ID, or use an external barcode scanner to scan the barcode label on the tube to enter Sample ID into the "Sample ID" field.

## NOTE

- Make sure the Sample ID you entered is the same with the QC sample ID you set in the QC file for the control.

4. Run the samples in accordance with the normal sample analysis procedure.
- ✓ After the analysis, the QC results will be automatically saved to the corresponding QC file.

## NOTE

- For the normal sample analysis procedures, refer to **7.6 Running Samples under Closed-tube Mode**.
- When the sample mode or sampling mode changes, the analyzer will switch the mode automatically and a message will be displayed.
- Up to 372 QC results can be saved in each QC file.



## 9.2.3 Reviewing L-J QC Results

After QC analysis, you can review the QC results in the "QC Table" review or "QC Graph" review.

### 9.2.3.1 QC Graph Review

#### Reviewing QC Graph

1. Tap "Menu"—"QC"—"L-J QC"—"Setup" in turn, to enter the L-JQC file setup screen.
2. Select the desired QC file to review.
- ✓ The "\*" mark displays next to the "File No." of the selected QC file.
3. Tap "QC Graph" to enter the QC graph review screen of the selected QC file.

#### Introduction to the L-J Blood QC Graph Screen

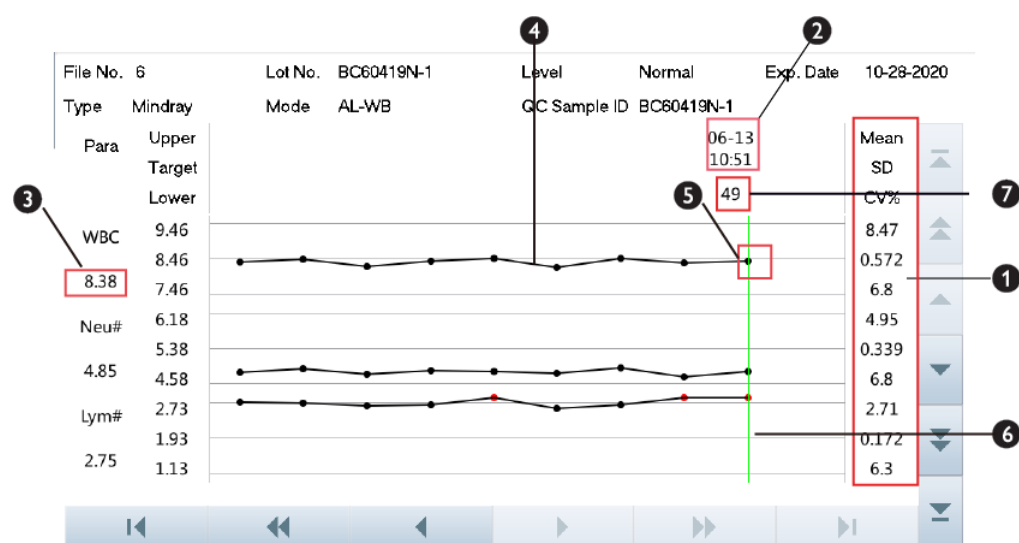
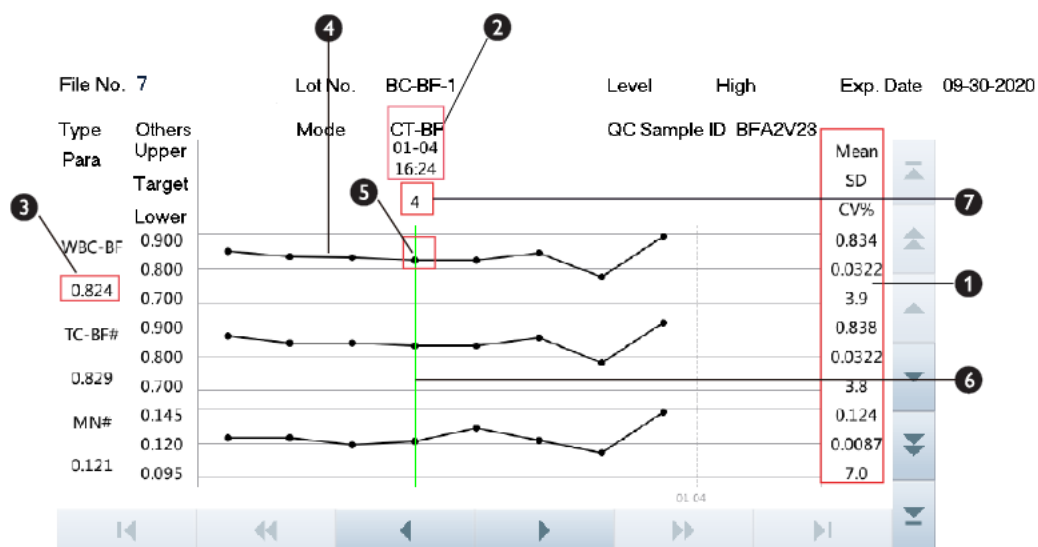


Figure 9-2 L-J Blood QC Graph

- 1—The Mean, SD and CV% of all the QC results of each parameter in the current graph.
- 2— The saving date and time of the QC point on the green line.
- 3— The QC result of the QC point on the green line.
- 4—The line connecting all QC points of the same parameter to show the trend. The QC points in each graph are displayed from left to right according to the sequence from the earliest to the latest.
- 5— Currently selected QC point. The analysis result of the selected QC point is displayed under the parameter. A black QC point indicates the value is within the limit; a red QC point indicates the value is out of the limit.
- 6— The green vertical line is used to identify a selected QC point and all parameter values of the QC point.
- 7— The sequence number of the QC point on the green line among all the QC points in the current QC file.

## Introduction to the L-J BF QC Graph Screen



**Figure 9-3 L-J BF QC Graph**

- 1—The Mean, SD and CV% of all the QC results of each parameter in the current graph.
- 2— The saving date and time of the QC point on the green line.
- 3— The QC result of the QC point on the green line.
- 4—The line connecting all QC points of the same parameter to show the trend. The QC points in each graph are displayed from left to right according to the sequence from the earliest to the latest.
- 5— Currently selected QC point. The analysis result of the selected QC point is displayed under the parameter. A black QC point indicates the value is within the limit; a red QC point indicates the value is out of the limit.
- 6— The green vertical line is used to identify a selected QC point and all parameter values of the QC point.
- 7— The sequence number of the QC point on the green line among all the QC points in the current QC file.

### Entering the Causes of Outliers (Administrators)

A red QC point indicates the value is out of the limit.

If necessary, follow below instruction to enter the reasons for the outliers.

1. Tap **"Menu">"QC">"L-J QC">"Setup"** and tap the **"Routine Blood Test QC"** tab to enter the L-J blood and BF QC file setting screen.
2. Select the desired QC file to review.
- ✓ The "\*" mark displays next to the "File No." of the selected QC file.
3. Tap **"QC Graph"** to enter the QC graph screen of the selected QC file.
4. Moving the green line to the desired QC point, and tap the "Outliers" button.
- ✓ A dialog box displays the QC results, targets and limits of all the parameters.

**Outliers**

	ESR	RBC-O	WBC-O	WBC-D	TNC-D
Target	2.00				
Limit	1.00				
Outliers	****				

Navigation: [Left Arrow] [Double Left Arrow] [Single Left Arrow] [Single Right Arrow] [Double Right Arrow] [Right Arrow]

**Cause of Outliers**

☐ Control not well mixed   
 ☐ Control deteriorated   
 ☐ Control expired  
☐ Reagent contaminated   
 ☐ Reagent expired  
☐ Others

OK Cancel

5. Enter the reasons for outliers.

Check the suitable causes for outliers;

or check **"Others"** and enter the causes of outliers in the edit box.

**Cause of Outliers**

☐ Control not well mixed   
 ☐ Control deteriorated   
 ☐ Control expired  
☐ Reagent contaminated   
 ☐ Reagent expired  
☒ Others

Navigation: [Left Arrow] [Double Left Arrow] [Single Left Arrow] [Single Right Arrow] [Double Right Arrow] [Right Arrow]

## NOTE

- You may enter up to 200 characters to the "Others" edit box.

6. Tap "OK" to save the reasons and exit the dialog box.

### 9.2.3.2 QC Table Review

#### Reviewing QC Table

1. Tap **"Menu"-"QC"-"L-J QC"-"Setup"** in turn, to enter the L-J QC file setup screen.
2. Select the desired QC file to review.
- ✓ The "\*" mark displays next to the **"File No."** of the selected QC file.
3. Tap **"QC Table"** to enter the QC Table screen of the specified QC file.

## Introduction to the L-J Blood QC Table Screen

File No. 6

Lot No. BC60419N-1

Level Normal

Exp. Date 10-28-2020

Type Mindray

Mode AL-WB

QC Sample ID BC60419N-1

	Date	Time	WBC	Neu#	Lym#	Mon#	
Target	/	/					
Limit	/	/					
6*	08-04-2020	10:30	10.01	1.00	1.00	1.00	
5	08-03-2020	20:23	10.00	1.00	1.00	1.00	
4	08-03-2020	20:23	10.07	1.00	1.00	1.00	
3	08-03-2020	20:23	10.02	1.00	1.00	1.00	
2	08-03-2020	20:23	10.08	1.00	1.00	1.00	
1	08-03-2020	20:23	10.06	1.00	1.00	1.00	

1

2

Figure 9-4 L-J Blood QC Table

- 1— The sequence number of the QC results saved in the QC file (earliest to the latest from top to down)  
 2— QC result  
 3— QC parameters (displayed in the same order as those on the QC graph screen)

## Introduction to the L-J BF QC Table Screen

File No.	7	Lot No.	BC-BF-1	Level	High	Exp. Date	09-30-2020
Type	Others	Mode	CT-BF	QC Sample ID BFA2V23			
	Date	Time	WBC	Neu#	Lym#	Mon#	
Target	/	/					
Limit	/	/					
8*	2020-01-04	16:30	0.891	0.894	0.142	15.9	
7	2020-01-04	16:28	0.775	0.779	0.113	14.6	
6	2020-01-04	16:27	0.844	0.850	0.122	14.5	
5	2020-01-04	16:25	0.824	0.827	0.131	15.9	
4	2020-01-04	16:24	0.824	0.829	0.121	14.7	
3	2020-01-04	16:22	0.831	0.835	0.119	14.4	
2	2020-01-04	16:20	0.832	0.836	0.124	14.9	
1	2020-01-04	16:19	0.849	0.855	0.124	14.6	

Figure 9-5 L-J BF QC Table

- 1—The sequence number of the QC results saved in the QC file (earliest to the latest from top to down)  
 2— QC result  
 3— QC parameters (displayed in the same order as those on the QC graph screen)

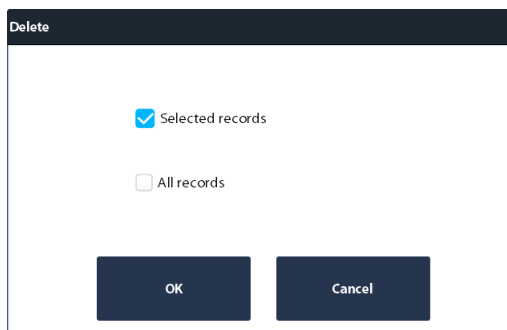
## Deleting QC Records (Administrators)

Administrators may delete selected or all QC record in the QC file.

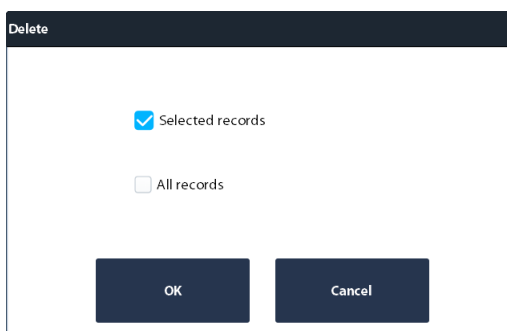
### ■ Deleting selected QC record(s)

1. Tap **"Menu"** - **"QC"** - **"L-J QC"** - **"Setup"** in turn, to enter the L-J QC file setup screen.
  2. Select the desired QC file.
- ✓ The "\*" mark displays next to the **"File No."** of the selected QC file.

3. Tap the **"QC Table"** button to enter the QC table screen of the corresponding QC file.
4. Select the QC record(s) you want to delete.
- √ The selected QC records are highlighted.
5. Tap **"Delete"**.
- √ The following dialog box displays.



6. Tap to select **"Selected records"** and then tap **"OK"** to delete selected records.
- Delete all QC records
1. Tap **"Menu"-"QC"-"L-J QC"-"Setup"** in turn, to enter the L-JQC file setup screen.
2. Tap the **"QC Table"** button to enter the QC table screen of the corresponding QC file.
3. Tap **"Delete"**.
- √ The following dialog box displays.



4. Tap to select **"All records"** and then tap **"OK"** to delete all records.

### Communication

You can transmit QC data to the external data management software or LIS/HIS.

Before transmitting QC data, make sure the network is properly connected.

1. Tap **"Menu"-"QC"-"L-J QC"-"Setup"** in turn, to enter the L-J QC file setup screen.
2. Select the desired QC file.
- √ The "\*" mark displays next to the **"File No."** of the selected QC file.
3. Tap the **"QC Table"** button to enter the QC table screen of the corresponding QC file.
4. Tap **"Comm."**.
- √ Communication starts. All QC data of the selected QC files will be transmitted by default.

### NOTE

- **The QC data saved in the process of transmission will not be transmitted.**
- **If auto-communication is enabled and a sample is run during the transmission of the QC data, then only when the QC data transmission finished will the auto-communication of the sample result start.**

## Exporting data (administrators)

Administrators may export selected or all QC records to an external USB device.

Before exporting data, make sure the USB device is firmly connected to the USB port on the side of the analyzer.

### NOTE

- **The user should ensure the data safety of the USB devices connecting to the analyzer.**

#### ■ Export selected records

Follow below instructions:

1. Tap **"Menu"**-"QC"-"L-J QC"-"Setup" in turn, to enter the L-J QC file setup screen.
2. Select the desired QC file.
- ✓ The "\*" mark displays next to the **"File No."** of the selected QC file.
3. Tap the **"QC Table"** button to enter the QC table screen of the corresponding QC file.
4. Select the QC record(s) you want to export.
- ✓ The selected QC records are highlighted.
5. Tap **"Export"**.
- ✓ The following dialog box displays.

The dialog box is titled "Export". It contains two sections: "Export Range" and "Export Content". In the "Export Range" section, there are two options: "All records" (unchecked) and "Selected records" (checked with a blue checkmark). In the "Export Content" section, there is one option: "Sample data" (checked with a green checkmark). At the bottom, there are two buttons: "OK" and "Cancel".

6. Tap to select **"Selected records"** and then tap **"OK"** to export the selected records.

#### ■ Exporting all QC records

1. Tap **"Menu"**-"QC"-"L-J QC"-"Setup" in turn, to enter the L-J QC file setup screen.
2. Tap the **"QC Table"** button to enter the QC table screen of the corresponding QC file.
3. Tap **"Export"**.
- ✓ The following dialog box displays.

The dialog box is titled "Export". It contains two sections: "Export Range" and "Export Content". In the "Export Range" section, there are two options: "All records" (unchecked) and "Selected records" (checked with a blue checkmark). In the "Export Content" section, there is one option: "Sample data" (checked with a green checkmark). At the bottom, there are two buttons: "OK" and "Cancel".

4. Tap to select **"All records"** and then tap **"OK"** to export all records from the QC Table.

### 9.2.3.3 Editing QC results (administrators)

When necessary, administrators can edit the QC results on the QC count screen.


Follow below instructions:

1. Select **"Menu"** > **"QC"** > **"L-J QC"** > **"Count"** or directly tap **"QC"** to enter the L-J QC count screen.
  2. In the **"File No."** pull-down list, select the QC file **"File No."**.
  3. (Optional) If necessary, tap **"Previous"** or **"Next"** to switch QC samples.
  4. Tap **"Edit Result"** to enter the **"Edit Result"** screen.
  5. Modify the results of the parameters as needed.
  6. Tap **"OK"** to return to the QC count screen.
- √ The modified results are saved on the count screen.
- √ "E" is marked behind the parameter result data directly modified manually. If modifying a parameter result directly and manually leads to the modification of its related parameter result, "e" is marked behind its result data.
7. (Optional) If necessary, tap **"Restore"**.
- √ The edited results will be changed back into the initial values.

## 9.2.4 Viewing Results of Single QC Count

If necessary, you can view the test results, RUO parameter results, histogram, scattergram, and other information of a QC analysis on the QC count screen.

Follow below instructions:

1. Select **"Menu"** > **"QC"** > **"L-J QC"** > **"Count"** or directly tap **"QC"** to enter the L-J QC count screen.
  2. In the **"File No."** pull-down list, select the QC file **"File No."**.
  3. (Optional) If necessary, tap **"Previous"** or **"Previous"** to switch QC samples.
- √ The screen displays the test results, scattergrams and histograms of corresponding QC samples.
4. Tap  button, and then tap **"RUO Para."** to go to the **"RUO Para."** screen.
- √ The screen displays the QC **"RUO Para."** results.

## 9.3 X-B QC

The X-B analysis is a weighted moving average analysis that uses values obtained from patient samples. It uses the 3 red cell indices, MCV, MCH and MCHC to indicate the hematology instrument performance.

It is recommended the X-B QC be activated when the sample volume of your laboratory is greater than 100 samples per day. Effective use of X-B requires randomization of samples and a normal cross section of patients to prevent skewing of indices. It observes the trend of QC results in the reference range formed by the specified target and limits.

The analyzer implements X-B QC on the three parameters: MCV, MCH and MCHC, each group of samples for X-B QC analysis consists of 20-200 sample results obtained from normal analysis of both WB and PD modes. The analyzer can save up to 3,000 X-B QC results. When the saved QC results have reached the maximum number, the newest result will overwrite the oldest result.

### 9.3.1 Validity Determination for X-B QC Samples

In X-B QC, sample results conforming to any of the following conditions will be considered as invalid and cannot be used in the QC calculation.

- Sample results exceeding the linearity range;
- Background results;
- Sample results not conforming to the "Sample Validity Setup";
- L-J QC results
- Calibration data.

### 9.3.2 Setting up X-B QC Rules

Administrators may set up X-B QC rules on the XB QC setup screen.

1. Tap "**Menu**" > "**QC**" > "**X-B QC**" > "**Setup**" in turn to enter the X-B QC file setup screen.
2. Set up X-B QC rules as needed.

See below for setting descriptions.

**Table 9-2 X-B QC count setup items**

<b>X-B QC On/Off</b>	Select " <b>On</b> " or " <b>Off</b> " to enable or disable the X-B QC.	When you select " <b>Off</b> ", the analyzer does not run X-B QC program.
<b>Samples/Batch</b>	The amount of samples to be included in calculating for an X-B QC point.	Setup range: 20 to 200; For example, when " <b>Samples/Batch</b> " is set to 20, and there are 20 valid samples, the analyzer starts to run X-B QC.
<b>Target/Limits Setup</b>	Enter the target/limit for each parameter	<ul style="list-style-type: none"> <li>• Do not leave any of the targets and limits for the QC parameters blank.</li> <li>• The units of target/limits of all parameters are the same as those on the "<b>Parameter Unit Setup</b>" screen of the hematology analyzer.</li> </ul>
<b>Sample Validity Setup</b>	In the "Sample Validity Setup" of the X-B QC setup screen, set the upper and lower limits of the 4 parameters in the sample validity setup area.	<ul style="list-style-type: none"> <li>• "<b>Sample Validity Setup</b>" is to set up the ranges of valid RBC, MCV, MCH and MCHC results. Only when the results of all these four parameters are within the specified ranges, the sample results can be used for X-B QC calculation.</li> <li>• In the sample validity setup, the upper limit shall be no smaller than the lower limit. Otherwise, there will be a message asking you to revise.</li> <li>• The valid ranges of the RBC parameters are their linearity ranges; the valid ranges of other parameters are their display ranges.</li> <li>• All the entries shall be numbers with only one decimal point. The length of the number entered cannot be longer than the length of the text box.</li> <li>• The units of lower and upper limits of all parameters are the same as those on the "<b>Parameter Unit Setup</b>" screen.</li> </ul>
<b>Restore Defaults</b>	Tap " <b>Restore Defaults</b> " to restore the targets and limits to the default values.	/
<b>Set Limits</b>	Tap " <b>Set Limits</b> " and select "By SD (#)" or "By CV (%)"	When " <b>By SD(#)</b> " is selected, the limits are displayed in the format of SD value; When " <b>By CV(%)</b> " is selected, the limits are displayed in the format of CV percent.



#### **BIOLOGICAL RISK**

- **All the samples, controls, calibrators, wastes and areas contacting them are potentially biohazardous. Wear proper personal protective equipment (e.g., gloves, lab coat, and glasses) and follow safe laboratory procedures when handling them and the contacted areas in the laboratory.**

After editing X-B setup, the system will start X-B QC automatically.

After every 20 to 200 valid sample results (determined by the "**Samples/Batch**" setup, the system will perform the X-B calculation once automatically. You can review the result in X-B QC graph or X-B QC table.

### 9.3.3 Reviewing X-B QC Results

After QC analysis, you can review the QC results in the "QC Table" review or "QC Graph" review.



### 9.3.3.1 QC Graph Review

Tap "Menu" > "QC" > "X-B QC" > "Graph" in turn to enter the X-B QC Graph screen.

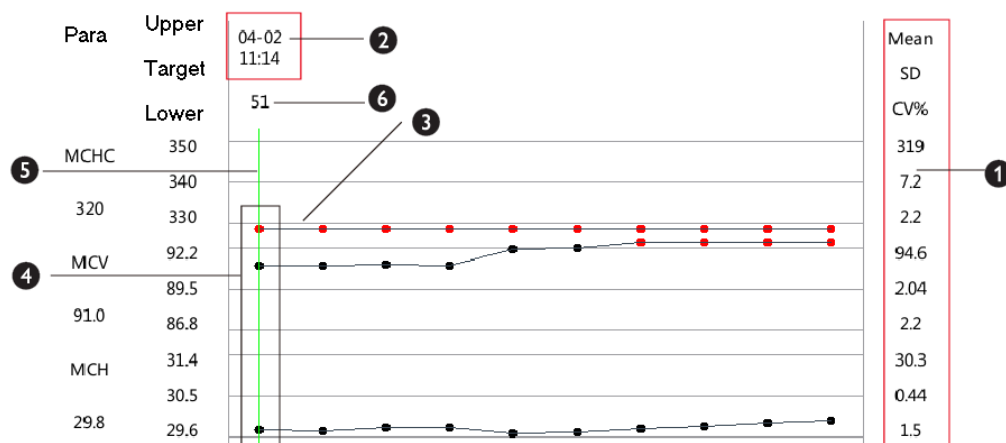


Figure 9-6 X-B QC Graph Screen

- 1—The Mean, SD and CV% of all the QC results of each parameter in the current graph.
- 2— The saving date and time of the QC points on the green line.
- 3—The line connecting all QC points of the same parameter to show the trend. The QC points in each graph are displayed from left to right according to the sequence from the earliest to the latest.
- 4— Currently selected QC point. The analysis result of the selected QC point is displayed under the parameter. A black QC point indicates the value is within the limit; a red QC point indicates the value is out of the limit.
- 5—The green vertical line is used to identify the QC points of the same analysis, all of which are displayed on the line when you select one of them.
- 6—The relative position of the QC point where the green vertical line is located in all QC points of the current QC file.

### 9.3.3.2 QC Table Review

1. Tap "Menu" > "QC" > "X-B QC" > "Graph" in turn to enter the X-B QC Graph screen.
2. Tap "QC Table" to go to the "QC Table" screen.

	Date	Time	MCV	MCH	MCHC
Target	/	/	89.5	30.5	340
Limit	/	/	92.2	31.4	350
389	2017-04-01	19:40	92.1	29.9	L 320
388	2017-04-01	19:20	92.2	30.0	L 325
387	2017-04-01	18:55	92.1	30.0	L 324
386	2017-04-01	18:36	92.1	30.0	L 325
385	2017-04-01	18:18	H 92.3	30.1	L 327
384	2017-04-01	17:59	92.1	30.1	L 327
383	2017-04-01	16:53	92.1	30.1	L 327
382	2017-04-01	16:20	92.2	30.1	L 327
381	2017-04-01	15:37	H 92.9	30.1	L 326
380	2017-04-01	15:00	H 93.2	29.9	L 325

Figure 9-7 X-B QC table screen

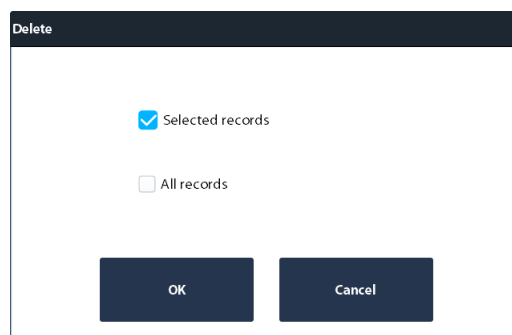
- 1— The sequence number of the QC results saved in the QC file (the earliest to the latest from top to down)
- 2— QC result
- 3— QC parameters (displayed in the same order as the Graph screen)
- 4— QC flags: The flag "H" or "L" (or other flags configured) will mark the results that are out of the limits

### Deleting QC Records (Administrators)

Administrators may delete selected or all QC record in the QC file.

#### ■ Deleting selected QC record(s)

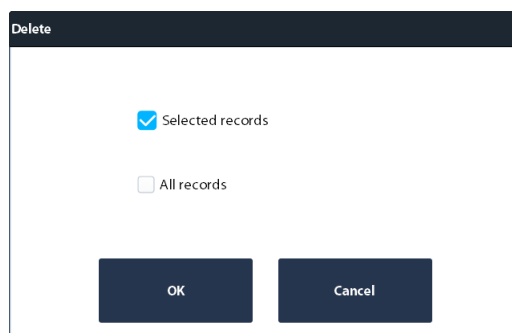
1. Tap "**Menu**" > "**QC**" > "**X-B QC**" > "**Graph**" in turn to enter the X-B QC Graph screen.
2. Tap "**QC Table**" to go to the "**QC Table**" screen.
3. Select the QC record(s) you want to delete.
- ✓ The selected QC records are highlighted.
4. Tap "**Delete**".
- ✓ The following dialog box displays.



5. Tap to select "**Selected records**" and then tap "**OK**" to delete selected records.

#### ■ Deleting all QC records

1. Tap "**Menu**" > "**QC**" > "**X-B QC**" > "**Graph**" in turn to enter the X-B QC Graph screen.
2. Tap "**QC Table**" to go to the "**QC Table**" screen.
3. Tap "**Delete**".
- ✓ The following dialog box displays.



4. Tap to select "**All records**" and then tap "**OK**" to delete selected records.

### Exporting data (administrators)

Administrators may export selected or all QC records to an external USB device.

Before exporting data, make sure the USB device is firmly connected to the USB port on the side of the analyzer.

## CAUTION

- **The user should ensure the data safety of the USB devices connecting to the analyzer.**

### ■ Exporting selected records

Follow below instructions:

1. Tap **"Menu"** > **"QC"** > **"X-B QC"** > **"Graph"** in turn to enter the X-B QC Graph screen.
2. Tap **"QC Table"** to go to the **"QC Table"** screen.
3. Select the QC record(s) you want to export.
- ✓ The selected QC records are highlighted.
4. Tap **"Export"**.
- ✓ The **"Export"** dialog box displays.
5. Tap to select **"Selected records"** and then tap **"OK"** to export the selected records.

### ■ Exporting all QC records

1. Tap **"Menu"** > **"QC"** > **"X-B QC"** > **"Graph"** in turn to enter the X-B QC Graph screen.
2. Tap **"QC Table"** to go to the **"QC Table"** screen.
3. Tap **"Export"**.
- ✓ The **"Export"** dialog box displays.
4. Tap to select **"All records"** and then tap **"OK"** to export all records.

## 9.4 When QC Results are Out of Range

If a QC result falls outside the control range, the **"QC"** button on the software screen lights in orange.

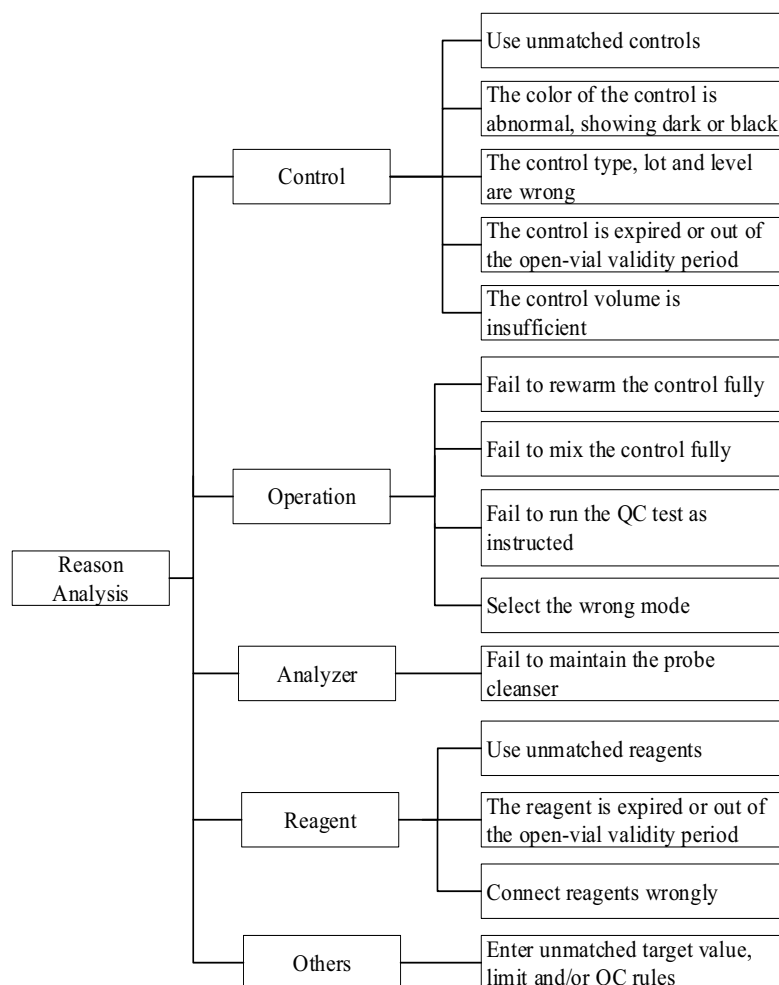
### Troubleshooting

When a QC result falls outside the control range, follow below steps to solve the problem.

- Analyze the cause of outliers, take corrective measures, and verify effectiveness of the corrective measures.
- If the corrective measures fail, report it according to the laboratory protocol.
- Contact Mindray Customer Service Department.

## Analyzing the Causes

Retest the samples with outliers. If the retest result still contains outliers and the trend is the same as that of the previous result with outliers, find out the cause of outliers by sequence referring to **Figure 9-8 Analyze the Cause of QC Outliers**.



**Figure 9-8 Analyze the Cause of QC Outliers**

## Taking Corrective Measures

Analyze the possible causes of QC outliers and make corrections in time according to **Figure 9-8 Analyze the Cause of QC Outliers**.

## Verifying Effectiveness of Corrective Measures

After taking corrective measures, retest the QC samples and verify whether the QC results are within the range.

If the cause of outliers is still not determined after all factors are analyzed, replace the controls with new one for verification, or directly contact Mindray Customer Service Department

# 10 Calibrating Your Analyzer

---

## 10.1 Overview

Calibration is a procedure to standardize the analyzer by determining its deviation under certain specified conditions. In order to get accurate sample analysis results, you should calibrate the analyzer per the procedure below when necessary.

There are four calibration programs available on this analyzer: auto calibration, manual calibration, ESR manual calibration, auto calibration using calibrators, auto calibration using fresh blood samples and protein gain calibration.

All the parameters or part of the parameters of WBC, RBC, HGB, MCV, ESR and, PLT can be calibrated by the calibration programs.

## 10.2 When to Calibrate

Your analyzer has been calibrated at the factory just before shipment. It is electronically stable and does not require frequent recalibration if you operate and maintain it as instructed by this manual. It is recommended that you run the calibration program every half year. You only need to recalibrate this analyzer if:

- you are going to use this analyzer for the first time (usually done by a Mindray-authorized representative when installing the analyzer).
- a major analytical component (including sampling probe, syringe, etc.) has been changed.
- you are going to re-use the analyzer after a long-term storage.
- the quality control results indicate there may be a problem.

---

### CAUTION

- **The instrument must be within the calibration validity period; otherwise, the measurement results may be inaccurate.**
- 

## 10.3 Checking before Calibration

Before calibration, follow the CLSI standards or your laboratory protocol to do tests, and make sure the analyzer's background (blank count) results, repeatability results and carryover results are all within the specified ranges.

If any of the above items is not in the range, check if the analyzer is in error. Remove the errors (if there are) and check again. If the problem persists, contact Mindray Customer Service Department.

### NOTE

- **For information of blank count, repeatability, and carryover of the analyzer, refer to Appendix B.6 Performance Specifications.**
- 

## 10.4 Running the Calibration Programs

The analyzer supports the following calibration programs:

- **Manual**
- **ESR**(administrators)
- **Calibrator**(administrators)
- **Fresh Blood**(administrators)

### 10.4.1 Notes before Calibration

Before calibration, check and make sure the analyzer works properly and enough reagents have been prepared for the calibration. You need to start over the calibration if the reagents run out during the process.

It is recommended that you create a log table for your analyzer. This log table should contain all necessary information that is pertinent to your analyzer. Suggested items that you may want to include in the log table are: calibration date, supplier of calibrator, lot number, expected results and limits, and result of background check.

### 10.4.2 Manual Calibration

#### NOTE

- **If you log in at the operator access level, you can only review calibration factors on the manual calibration screen. You cannot edit the calibration factors.**

#### 10.4.2.1 Sample requirements for manual calibration

You can use Mindray-specified calibrators or value-assigned fresh blood samples to perform manual calibration.

If you are using value-assigned fresh blood samples, make sure the samples meet the following requirements:

- has normal erythrocyte, leukocyte, and platelet morphology
- the test results are within corresponding ranges
- has sufficient sample volume for the whole calibration procedure

#### 10.4.2.2 Performing manual calibration

Do as follows to calibrate the analyzer.

1. Select a calibrator or a value-assigned fresh blood sample which meets the sample requirements for manual calibration, and run the sample consecutively for n times (not less 3 times), in whole blood or predilute mode.
2. Calculate the CV values and the Mean values for the n tests.

#### NOTE

- **You may review the CV and Mean values through the "Table Review" screen. See 8.2.7Calculating CV Values for more details.**

3. Check if the CV values are in the acceptable ranges.

#### NOTE

- **When the CV value for any parameter exceeds the acceptable range, check if the analyzer is in error. If there are errors, remove the errors and test again. If the problem cannot be solved, contact Mindray Customer Service Department.**

4. Tap **"Menu" > "Calibration" > "Manual"** in turn to enter the **"Manual"** screen.
5. Calculate the new calibration factors for the parameters according to the following equation. The calculated factors should show 2 decimal places.

$$\text{New calibration factor} = \frac{\text{Current calibration factor} \times \text{Reference value}}{\text{Mean}}$$

For example: Suppose the WBC reference value of a calibrator is 8.40, and the current calibration factor of the whole blood mode is 98.90%.

Run the sample under whole blood mode for 10 consecutive times (n=10) and take the WBC results of the 10 runs to calculate: 8.10, 8.00, 8.10, 8.30, 8.30, 8.20, 8.00, 8.10, 8.30. The obtained CV is 1.5% and Mean is 8.16, which meet the requirements.

Therefore:

$$\text{New Calibration Factor} = \frac{98.90\% \times 8.40}{8.16} = 101.81\%$$

6. The calculated calibration factors shall be between 75.00% - 125.00%.

If not, the calibration factor is invalid. In case of an invalid calibration factor, try to find out the reason (e.g. calibration material not thoroughly mixed, misoperation, etc.). Then, recalibrate the analyzer and recalculate the calibration factors.

7. Enter the new calibration factors into the factor cell of the parameter that require calibration.

√ The "Date" cells automatically display the date when the new calibration factors are entered.

If the entered calibration factors are invalid, the factors will be highlighted in red.

## NOTE

- **The calibration factor entered must be in the range of 75.00% to 125.00%, and only two decimal places can be reserved.**

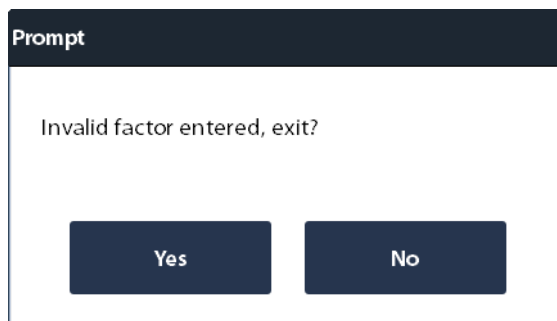
8. Save calibration factors.

a Tap another button on the software screen.

√ The following dialog box displays.

b Tap **"Yes"** to save new calibration factors.

9. If the entered calibration factors are invalid, a dialog box will display prompting "Invalid entry" when you are switching to another screen.



a If you want to re-do the calibration, tap **"No"**, and perform the calibration procedures again.

b Tap **"Yes"** to close the dialog box and switch to another screen without saving the changes; the original calibration factors and dates will remain unchanged.

### 10.4.3 Calibrating with ESR (administrators)

The ESR calibrators are used to calibrate the analyzer deviation so as to align the measurement results and the target value.

## NOTE

- **Perform ESR calibrators function under the guidance of engineers from Mindray Customer Service Department.**

## 10.4.4 Calibrating with Calibrators (administrators)

### 10.4.4.1 Sample requirements for calibrators

You must use Mindray-specified calibrators for calibration.

### 10.4.4.2 Performing calibration with calibrators

#### NOTE

- **The calibration with calibrators can only be performed in AL-WB mode.**

1. Make sure you have prepared the calibrators in accordance with your laboratory protocols, and the requirements in the use instruction of the calibrators.
2. Tap **"Menu"** > **"Calibration"** > **"Calibrator"** in turn to enter the **"Calibrator"** screen.
3. Set up calibrator information.

Enter the **"Lot No."**, **"Exp. Date"** and the target for each parameter.

See below for setting descriptions.

<b>Lot No.</b>	Find the lot No. on the label on the vial of the calibrator, or on the target sheet of the corresponding calibrator.
<b>Exp. Date</b>	The entered expiration date should be either the expiration date printed on the labeling or the open-vial expiration date, whichever is earlier. The open-vial expiration date is calculated as follows: the date that container is opened + the open-vial stability days.
Parameter Targets	Find the parameter targets on the target sheet of the corresponding calibrator.

4. Place racks loading calibrators on the right tray of the autoloader, with the back of "MINDRAY" mark on the carrier facing the analyzer.
  5. Perform the calibrator calibration.
    - a Tap **"Start Count"** on the calibrator calibration screen.
- ✓ The analyzer starts the calibration.

#### NOTE

- **To obtain valid calibration factors, we need 3 to 10 valid calibration results.**

- b When the first calibration finished, removed the tube rack with the calibrator from the unloading tray, and place it on the loading tray again.
  - c Press the **"Start Count"** key on the calibrator calibration screen to start the second analysis.
  - d Repeat steps a to c for the remaining tests.
- ✓ The analyzer automatically calculates the calibration factors for the parameters.
6. (Optional) You may select to use which calibration results to calculate the calibration factors.

Check the **"Select"** cells of the calibration results that are to be involved in the calculation of calibration factors. Select at least 3 groups of calibration results.

The calibration results are invalid under the following circumstances.

- if there is a parameter whose calibration data is out of its linearity range but still within the display range;
- if there is a parameter whose calibration data is out of the display range, then the non-numeric parameter values "\*\*\*\*" will be displayed in the list;

When there are invalid calibration results, a dialog box displays. Tap "OK" to close the message box, and the data will be deleted from the table without saving automatically.

7. Save calibration factors.
    - a Tap another button on the software screen.
- ✓ The analyzer gives different suggestions depending on the calculated calibration factors.



- b Read the software messages, and save the new calibration factors or exit the screen directly as prompted.

## 10.4.5 Calibrating with Fresh Blood Samples (administrators)

### 10.4.5.1 Requirements for fresh blood samples used for calibration

If you are using value-assigned fresh blood samples, make sure the samples meet the following requirements:

- has normal erythrocyte, leukocyte, and platelet morphology
- the test results are within corresponding ranges
- has sufficient sample volume for the whole calibration procedure

### 10.4.5.2 Calibrating with fresh blood samples

#### NOTE

- **The calibration with fresh blood samples can only be performed under AL-WB mode.**

Do as follows to calibrate the analyzer with fresh blood:

1. Follow the instruction in **7.5.1 Preparing Whole Blood Samples (For WB Mode)** to prepare sufficient fresh blood samples.
2. Tap **"Menu" > "Calibration" > "Fresh Blood"** in turn to enter the **"Fresh Blood"** screen.
3. Select a sample ID from the **"Current Sample ID:"** pull-down list.
4. Enter the targets corresponding to the parameters for calibration into the **"Target"** text boxes.
5. Place the prepared whole blood sample in a tube, and place the racks with tubes on the right tray of the autoloader, with the back of "MINDRAY" mark on the carrier facing the analyzer.
6. Perform the fresh blood sample calibration.

#### NOTE

- **To obtain valid calibration factors, we need 3 to 10 valid calibration results.**

- a Tap **"Start Count"** on the fresh blood sample calibration screen.
- √ The analyzer starts the calibration.
- b When the first calibration finished, removed the tube rack with the calibrator from the unloading tray, and place it on the loading tray again.
- c Press the **"Start Count"** key on the fresh blood sample calibrator screen to start the second analysis.
- d Repeat steps a to c for the remaining tests.
- √ The analyzer automatically calculates the calibration factors for the parameters.
7. (Optional) You may select to use which calibration results to calculate the calibration factors.

Check the **"Select"** cells of the calibration results that are to be involved in the calculation of calibration factors. Select at least 3 groups of calibration results.

- √ The analyzer automatically calculates the calibration factors for the parameters.

The calibration results are invalid under the following circumstances.

- if there is a parameter whose calibration data is out of its linearity range but still within the display range;
- If there is a parameter whose calibration data is out of the display range, then the non-numeric parameter values "\*\*\*" will be displayed in the list.

When there are invalid calibration results, a dialog box displays. Tap "OK" to close the message box, and the data will be deleted from the table without saving automatically.

Under the following circumstances, the calculated calibration factors are invalid:

- When the calculated calibration factor for a blood sample is not within the valid range (75% to 125%).

- The CV% value of any calibration parameter exceeds the precision index of the analyzer.

When there are invalid calibration factors, a dialog box displays. Tap "OK" to close the message box, and the data will be deleted from the table without saving automatically.

- Repeat steps 3 to 7 to complete the analysis of other blood samples.
- Calculate the mean calibration factor.

You must have at least 3 groups of valid calibration factors to calculate the mean calibration factors.

- Tap "**Calculate**" button to enter the screen for calculating the mean calibration factor of fresh blood samples.
  - Tap to select at least 3 groups of calibration factors that are to be involved in the calculation of a mean calibration factor.
- ✓ The analyzer automatically calculates the mean calibration factor.

Under the following circumstances, the calculated mean calibration factors are invalid.

- when the deviation of absolute value between the calibration factors included in calculating the mean and the original calibration factors reaches or exceeds 5%;
- when the calculated calibration factor is not within the specified range (75%-125%).

- Save calibration factors.
  - Tap another button on the software screen.

- ✓ The analyzer gives different suggestions depending on the calculated calibration factors.
- Read the software messages, and save the new calibration factors or exit the screen directly as prompted.
  - ✓ Read the software messages, and save the new calibration factors or exit the screen directly as prompted.

#### 10.4.6 Verifying Calibration Factors

Verify the calibration factors after calibration.

Verify the calibration factors using any of the following methods:

- Run the calibration at least 3 times, and check if the results are within the allowed range.
- Run the controls of high, normal, and low levees at least 3 times, and check if the results are within the allowed range.
- Run at least 3 fresh blood samples from normal patients, each sample for at least 3 times, and check if the results are within the allowed range.

### 10.5 Calibration History ("Menu" > "Calibrate" > "Calibration History")

Only administrators can view the calibration history.

Tap "**Menu**" > "**Calibration**" > "**Calibration History**" in turn to enter the "**Calibration History**" screen.

The relevant items are described below:

<b>Details</b>	Select a calibration record and tap " <b>Details</b> " to view the detailed calibration information.
<b>Go to</b>	Tap " <b>Go to</b> " to view the calibration history of the specified time period.
<b>Export</b>	Tap " <b>Export</b> " to export the specified or all calibration records to a USB device.

#### NOTE

- The user should ensure the data safety of the USB devices connecting to the analyzer.

# 11 Printing

You can set up the print templates for sample results report, graphs, QC results, QC graphs, manual calibration factors etc., and print them using the print templates.

## 11.1 Setting up Print Template

You can set up print settings on the "Print Setup" screen.

### 11.1.1 Operation Procedure

1. Tap "Menu" > "Setup" > "System Setup" > "Print Setup" to enter the "Print Setup" screen.
2. Tap the "Print Setup", "Printing Content", and "Auto print after sample analysis" tabs as needed to perform print settings as needed.

See below for setting descriptions

<b>Print device</b>	Select a print device on the network from the pull-down list:	For questions about the print device and print drive settings, contact the customer service personnel of your print device supplier. (administrators)
<b>Printer driver</b>	Select a proper print drive from the pull-down list: <ul style="list-style-type: none"><li>• <b>Auto Identification</b></li><li>• <b>PCL6</b></li><li>• <b>Raster print</b></li></ul>	
<b>Paper</b>	Select the desired paper type from the pull-down list: The analyzer supports two paper types: <ul style="list-style-type: none"><li>• <b>A4</b></li><li>• <b>A5</b></li></ul>	
<b>Blood Sample Report Title/ Body Fluid Sample Report Title</b>	<ol style="list-style-type: none"><li>1. Select "Blood Sample Report Title" or "Body Fluid Sample Report Title" from the pull-down list.</li><li>2. Enter the desired title in the edit box.</li></ol> <p>✓ The report titles set up here will display on the printed analysis reports.</p>	
<b>Blood Sample Report Template/ Body Fluid Sample Report Template</b>	<ol style="list-style-type: none"><li>1. Select "Blood Sample Report Template" or "Body Fluid Sample Report Template" from the pull-down list.</li><li>2. Select the desired template format from the pull-down list:<ul style="list-style-type: none"><li>• <b>One page with histogram</b></li><li>• <b>One page without histogram</b></li></ul></li></ol>	<ul style="list-style-type: none"><li>• When "One page with histogram" is selected, the print results include the parameter results and graphs.</li><li>• When "One page without histogram" is selected, the print results include only the parameter results.</li></ul>
<b>Para. Language</b>	Select a parameter language from the pull-down list: <ul style="list-style-type: none"><li>• <b>Chinese</b></li><li>• <b>Chinese/English</b></li></ul>	<ul style="list-style-type: none"><li>• When "Chinese" is selected, the printed report displays the Chinese parameter names.</li><li>• When "Chinese/English" is selected, the printed report displays Chinese and English parameter names in a pair.</li></ul>
<b>Copies</b>	Enter the number of report copies to be printed when you tap "Print". The default value is 1 copy, and the setting range is [1 to 20].	

<b>Printing Content</b>	Check the options you want to display on the printed report: <ul style="list-style-type: none"> <li>• <b>Print flags of edited result</b></li> <li>• <b>Print high/low result flags</b></li> <li>• <b>Print suspect flags</b></li> <li>• <b>Print result edit flag</b></li> <li>• <b>Print out of linearity range flag</b></li> <li>• <b>Print flags</b></li> <li>• <b>Print reference range</b></li> <li>• <b>Monochrome Print</b></li> <li>• <b>Print RUO Parameters</b></li> </ul>	<ul style="list-style-type: none"> <li>• For the description of setting up high/low results as well as suspect results flags, refer to section <b>6.3.3.2 Other Settings</b>.</li> <li>• For the description of setting up reference ranges, refer to section <b>6.3.4.2 Ref. Range Setup (Administrators)</b>.</li> </ul>
<b>Auto print after sample analysis</b>	Check to enable one or more desired auto print settings: <ul style="list-style-type: none"> <li>• <b>Auto print after analysis</b></li> <li>• <b>Auto print after validating</b></li> <li>• <b>Auto print after QC count</b></li> </ul>	<ul style="list-style-type: none"> <li>• When <b>"Auto print after analysis"</b> is selected, the analyzer automatically print the sample results after each analysis.</li> <li>• When <b>"Auto print after validating"</b> is selected, the analyzer automatically print the validated sample results.</li> <li>• When <b>"Auto print after QC count"</b> is selected, the analyzer automatically print the results of QC count results after each QC count ends.</li> </ul>

## 11.2 Printing Sample Result Report

### NOTE

- **The analyzer prints at most 700 records at one time.**

Before printing a sample test report, confirm the following items:

- The printer has been set up and connected correctly.
- There are enough paper in the printer.

### 11.2.1 Printing Current Sample Result Report

When a cycle of sample analysis finishes, tap the **"Print"** button on the **"Count"** screen.

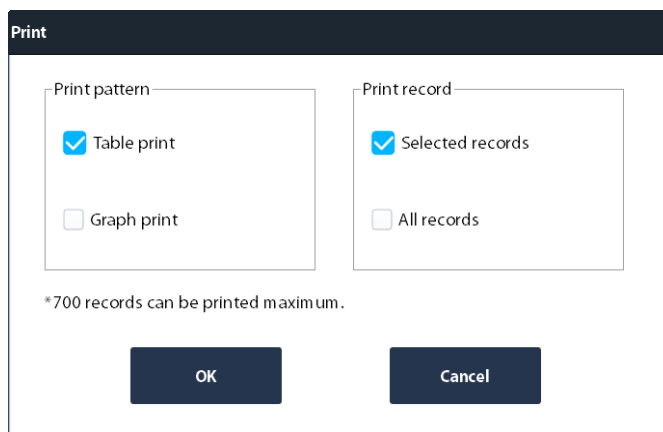
✓ The analyzer automatically prints the results of the current sample according to the print setup.

### 11.2.2 Printing from the Table Review Screen

Follow below instructions:

1. Select **"Menu- Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. (Optional) Tap to select one or more sample records to be printed on the test result report.  
If you are going to print all sample records, skip this step.
3. Tap **"Print"** in the utility button area.

- √ A dialog box for printing displays.



4. Set up the print methods.

See below for setting descriptions.

<b>Print pattern</b>	<b>Table print</b>	Select " <b>Table print</b> ", only to print sample results, but not to print graphs (histograms and scattergrams).
	<b>Graph print</b>	Select " <b>Graph print</b> ", print sample results as well as the graphs (histograms and scattergrams).
<b>Print record</b>	<b>Selected records</b>	Check " <b>Selected records</b> " to print the selected sample records.
	<b>All records</b>	Check " <b>All records</b> " to print all the sample records.

5. Tap "OK".

- √ The analyzer prints the sample result reports of the selected samples.

### 11.2.3 Printing from the Graph Review Screen

Follow below instructions:

1. Select "**Menu**"- "**Table Review**" or tap the "**Table Review**" utility button to enter the "**Table Review**" screen.
2. Select one or more sample records of which you want to review the graph data.
 

√ The selected sample record is highlighted.
3. Tap "**Graph**" to go to the "**Graph**" screen.
4. Tap "**Print**" in the utility button area.
 

√ The analyzer automatically prints the results of the current sample according to the print setup.

### 11.2.4 Printing RUO Parameter Results

Follow below instructions:

1. Tap "**Menu**"- "**Setup**"- "**System Setup**"- "**Print Setup**".
2. Tap "**Printing Content**" and check "**Print RUO Parameters**".

#### NOTE

- If you select to print "RUO parameters", the statement will be printed by default. The statement cannot be edited.

3. Tap another button on the screen to save the settings and exit the "**Printing Content**" setup screen.
 

√ The report will display the RUO parameter results .

## 11.2.5 Printing Microscopic Parameter Results

Follow below instructions:

1. Select **"Menu"**- **"Table Review"** or tap the **"Table Review"** utility button to enter the **"Table Review"** screen.
2. Select one or more sample records of which you want to review the microscopic parameters.
- ✓ The selected sample record is highlighted.
3. Tap **"Graph"** to go to the **"Graph"** screen.
4. Tap **"Other Para."** > **"Microscopic Para."** to enter the **"Microscopic Para."** screen.
5. Tap the **"Print"** button to print the microscopic parameter results.

## 11.3 Printing QC Result Report

Before printing a QC test result report, confirm the following items:

- The printer has been set up and connected correctly.
- There is enough paper in the printer.
- To print the L-J QC results, confirm that there are results in the L-J QC file.
- To print the X-B QC results, confirm that X-B QC is enabled and there are valid X-B results.

### 11.3.1 Printing L-J QC Result from the L-J QC Table Screen

Follow below instructions:

1. Tap **"Menu"**-**"QC"**- **"L-J QC"**- **"Setup"** to enter the L-J QC file setup screen.
2. Select the desired QC file.
3. Tap **"QC Table"** to enter the QC Table screen of the specified QC file.
4. Tap **"Print"** in the utility button area.
- ✓ The analyzer prints all the QC test results under the specified QC file.

#### NOTE

- **When the QC test results are printed from the QC table screen, the printed report does not include QC graph.**

### 11.3.2 Printing L-J QC Graph from the L-J QC Graph Screen

Follow below instructions:

1. Tap **"Menu"**-**"QC"**- **"L-J QC"**- **"Setup"** in turn to enter the L-J QC file setup screen.
2. Select the desired QC file.
3. Tap **"QC Graph"** to enter the QC graph review screen of the selected QC file.
4. Tap **"Print"** in the utility button area.
- ✓ The analyzer prints the QC graph in the specified QC file.

### 11.3.3 Printing X-B QC Graph from the X-B QC Graph Screen

Follow below instructions:

1. Tap **"Menu"** > **"QC"** > **"X-B QC Graph"** > **"Graph"** in turn to enter the X-B QC Graph screen.
2. Tap **"Print"** in the utility button area.
- ✓ The analyzer prints the X-B QC graph.

### 11.3.4 Print X-B QC Parameter Result from the X-B QC Table Screen

Follow below instructions:

1. Tap **"Menu"** > **"QC"** > **"X-B QC Graph"** > **"Graph"** in turn to enter the X-B QC Graph screen.
  2. Tap **"QC Table"** to go to the **"QC Table"** screen.
  3. Tap **"Print"** in the utility button area.
- √ The analyzer prints the X-B QC parameter result report.

## 11.4 Printing Manual Calibration Factors

Before printing manual calibration factors, confirm the following items:

- The printer has been set up and connected correctly.
- There is enough paper in the printer.

Follow below instructions:

1. Tap **"Menu"** > **"Calibration"** > **"Manual"** in turn to enter the **"Manual"** screen.
  2. Tap **"Print"** in the utility button area.
- √ The analyzer prints the manual calibration factors.

**This page intentionally left blank.**



# 12 Service

## 12.1 Overview

Preventive and corrective maintenance procedures are required to keep the analyzer in good operating conditions. This analyzer provides multiple maintenance functions for this purpose.

This chapter introduces how to use the provided functions to maintain and troubleshoot your analyzer.

## 12.2 When and Why to Perform the Maintenance

### 12.2.1 Maintenance of Parts and Components

Table 12-1 Maintenance of parts and components

Item/Software access			Purpose
Self-Test (Service-Debug & Self-Test-Self-Test)	Valve	Perform the self-test under the instruction of Mindray's service people	/
	Fluidics Debug	Perform the self-test under the instruction of Mindray's service people	/
Probe Cleanser Maint. (Service-Maintenance-Probe Cleanser Maint.)	Flow Cell	<ul style="list-style-type: none"><li>when there are a great amount of bad blood sample differential cases</li><li>when there are flow cell errors reported</li></ul>	Clean the flow cell
	Apertures	when clogging error is frequently reported	Clean the RBC channel
Auto-cleaning the Parts and Components (Service-Maintenance-Cleaning)	Clean fluidics	When the background background results are unqualified	Fluidics cleaning
Fluidics(Service-Maintenance-Fluidics)	Probe Cleanser Maint.	When the background results are unqualified or repeatability results continuously do not meet requirements	Thoroughly clean the fluidics, parts and components
	Pack-up	When the analyzer will not be used for a long time (over 10 days)	Remove the residue reagents in the pipelines
Sensor Maint.(Service-Maintenance-Sensor Maint.)	Unclog Aperture	when clogging error is frequently reported	clean the aperture
	Clean Flow Cell	<ul style="list-style-type: none"><li>when there are a great amount of bad blood sample differential cases</li><li>when there are flow cell errors reported</li></ul>	Clean the flow cell

Item/Software access			Purpose
Screen Cal.(Service-Screen Cal.)	Screen Cal.	when the touch screen response to touching is not sensitive or accurate	Calibrate touch screen

Table 12-2 Replacing and priming with reagents

Item/Software access		Timing	Purpose of maintenance
Replace Reagent (Service > Maintenance > Reagent)	DS Diluent	when the reagent runs out or insufficient	Replace the residue reagent in the pipelines
	Replace HGB lyse		
	Replace DIFF lyse		
	*Replace RET diluent		
	Replace DIFF dye		
	*Replace RET dye		
	ESR Solution Reagent		
Prime Reagent (Service > Maintenance > Reagent)	DS Diluent	when reagents are contaminated or expire.	Replace the residue reagents in the fluidic system
	Prime HGB lyse		
	Prime DIFF lyse		
	*Prime RET diluent		
	Prime DIFF dye		
	*Prime RET dye		
	ESR Solution Reagent		

**NOTE**

- The item with \* only applies to the BC-760[R]/BC-780[R] model.

**12.2.2 Manual Cleaning**

Table 12-3 Manual cleaning

Program	Timing	Purpose
Clean the probe wipe, floating blood barrier and the blood barrier bracket	Every month	Remove the blood sample and other residues in the probe wipe
Cover cleaning	As needed	Remove contaminants on the cover
Clean the filtering net	As needed/Recommend monthly cleaning	Clean the dust on the surface of air inlet filtering net

## 12.2.3 Replacing the Parts and Components

**Table 12-4 Replacing parts and components**

Program	Timing	
Replace Sampling probe	The sampling probe is damaged	Contact service engineer to replace the component.
Replace the filters	The analyzer reports a pressure fault, and the automatic drainage device fails to work normally	Contact service engineer to replace the component.

**Table 12-5 Replacing wearing parts**

Program	Timing	
Replace the sampling probe and probe wipe	120,000 times of work	When the sampling probe, the probe wipe filter or the HGB waste filter are near their service life, the software will give alarm. Contact service engineer to replace the component.
Replace the probe wipe filter	3 years or 60,000 times of work	
Replace the HGB waste filter	60,000 times of work	
Replace the DIFF waste filter	60,000 times of work	
Replace the RET waste filter	60,000 times of work	

### **WARNING**

- To avoid analyzer damage or personal injury, only Mindray-supplied parts can be used for maintenance.

## 12.3 Reagent Management

### 12.3.1 Viewing Reagent Information ("Menu" > "Setup" > "Reagent Setup")

On the "Reagent Setup" screen, you may review the expiration dates, use before dates, open dates, valid days and remaining volumes.

Tap "Menu" > "Setup" > "Reagent Setup" to go to the Reagent Setup screen.

You may review the expiration dates, use before dates, open dates, valid days and remaining volumes of the analyzing reagents on the "Reagent Setup" screen.

### 12.3.2 Replace the Reagents

Replace the reagent when the reagent runs out, is insufficient or expired.

The whole reagent replacing procedure includes 3 steps:

1. Install a new reagent;
2. Enter the new reagent information into the analyzer;
3. Replace the old reagent in the fluidic system.

### 12.3.2.1 Install a new reagent

#### Replacing the DS Diluent Container

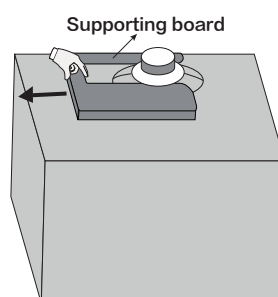
##### BIOLOGICAL RISK

- After replacing the reagent container/bag, check the tubing connected to the cap assembly and make sure it is not bent over.

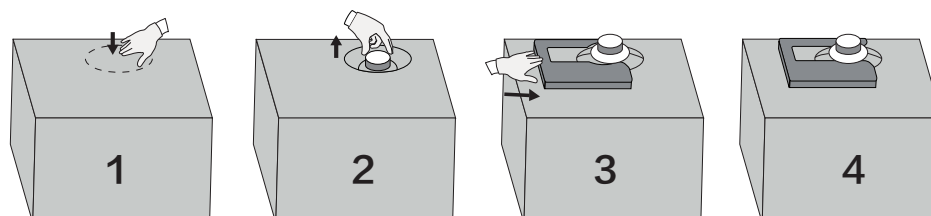
##### NOTE

- Keep the diluent container from severe shock or crashing against other object. Otherwise, the alarming would be unreliable.

1. Remove the cap of a new DS diluent container, and place the container next to the one to be replaced.
2. Remove the supporting board under the cap of the old container.



3. Turn the cap of the old container counterclockwise, and then take out the cap assembly with caution.
4. Insert the pickup tube of the cap assembly into the new reagent container, and then turn the cap clockwise until it is secured.
5. Install the supporting board under the new container's cap as shown below.



#### Replacing the Fluorescent Reagent Bag

##### BIOLOGICAL RISK

- After replacing the reagent container/bag, check the tubing connected to the cap assembly and make sure it is not bent over.

Before replacing the fluorescent reagent, confirm that the reagent name on the reagent bag label is consistent with that on the connecting cable label of the cap assembly.

1. Open the fluorescent dye compartment.
2. Get a new bag of fluorescent dye, open the cap and the aluminum film sealing the bag.

##### WARNING

- While installing or replacing the fluorescent dye bag, hold the upper corners of the bag or the part under the bag mouth (where the interior tube is located), in order not to extrude the reagent out.

3. Take out the bag to be replaced along the direction of the supporting rack.
4. Turn the cap of the old reagent container counterclockwise, and then take out the cap assembly with caution.

#### NOTE

- **If the pickup tube of the cap assembly is stuck when it is taken out of the fluorescent reagent bag, slightly adjust the position of the pickup tube and then take it out without pulling by force.**

5. Insert the pickup tube of the cap assembly vertically into the new container, and then turn the cap clockwise until it is secured.

#### NOTE

- **During replacement, make sure that the pickup tube of the cap assembly does not reach the bottom of the reagent bag, otherwise the reagent cannot be aspirated normally.**

6. Put the sealed new bag back on the support rack, making sure the bag is securely accommodated.
7. Cap the old bag using the cap of the new bag and dispose of the old bag properly.

#### Replacing Other Reagent Containers (Except DS Diluent and Fluorescent Reagent)

1. Remove the cap of a new reagent container, and place the container next to the one to be replaced.
2. Turn the cap of the old container counterclockwise, and then take out the cap assembly with caution.
3. Insert the pickup tube of the cap assembly into the new container, and then turn the cap clockwise until it is secured.
4. Cap the old container with the cap of the new container and dispose of the container properly.

### 12.3.2.2 Entering reagent information

Follow below instructions:

1. Enter the "Reagent Setup" dialog box.

You can enter the "Reagent Setup" dialog box in either of the following ways:

- Tap "Menu" > "Setup" > "Reagent Setup" or the "Reagent Setup" button directly to enter the "Reagent Setup" screen, and then tap "Setup".
- Tap the reagent error alarm on the right lower corner on the screen to enter the "Remove Error" screen, and then tap "Remove Error".

√ The "Reagent Setup" dialog box pops up, and the top part displays the information of the reagent to be replaced.

2. Enter the reagent information in one of the following ways:

If...	Then...
Barcode entry is selected	Tap "Enter Reagent Information" edit box and use a pop-up keyboard to enter the reagent barcode; or scan the reagent barcode labels with an external barcode scanner.
Information entry through RFID is selected	Swipe the reagent on RFID region of "Reagent Setup" screen.

#### NOTE

- **If you want to change the way of entering reagent information, please contact Mindray Customer Service Department.**
- **Only when replacing the dye or ESR cleanser, reagent information can be entered through RFID.**

√ If the barcode is valid, the reagent information will be refreshed.

## NOTE

- If the barcode is entered/scanned in the "Barcode Entry" edit box of the "Reagent Setup" dialog box, and the reagent corresponding to the barcode is not in the reagent list displayed at the top of the dialog box, the software will prompt an alarm. Check and make sure that you have selected the correct reagent for replacement.
- If necessary, you can manually modify the reagent validity period and reagent volume information, and the software will automatically save and refresh the information.

3. (Optional) If more than one reagent needs to be replaced, repeat step 3 to complete the setup of all reagents.
  4. Replace the old reagent in the pipelines.
- If "Auto replace reagent after setup" is checked on the "Setup" > "Auxiliary Setup" > "Other Settings" screen, the analyzer will automatically start the program to replace the old reagent in the pipelines at defined time.

As shown in the figure below, when "Auto replace reagent after setup" is enabled, the count-down timer displays on the "Replace" button after the reagent setup is completed. After the countdown ends, the analyzer automatically starts the program to replace the old reagent in the fluidic system.

The screenshot shows the "Reagent Setup" dialog box. At the top, there is a table with the following data:

Reagent Name	Expiration Date	Volume	Barcode
DS DILUENT	01-01-2036	20000.000(mL)	1009043601011111111

Below the table is a section titled "Barcode Entry" with the instruction: "Scan the reagent barcode, or manually enter the figures below the barcode." Below this instruction is a row of 12 empty input boxes. At the bottom of the dialog, there is a green status message: "All reagent setup is completed". At the very bottom is a button labeled "Replace(27)", where the number 27 is in a red box, indicating a countdown timer.

Figure 12-1 Countdown - replacing reagents through barcode entry

The screenshot shows the "Reagent Setup" dialog box. At the top, there is a table with the following data:

Reagent Name	Expiration Date	Volume(mL)	Barcode
ESR Solution Reagent	01-01-2036	1000.000	****

Below the table is a section titled "Enter Reagent Information" which contains an illustration of an RFID reader and an ESR reagent bottle. At the bottom of the dialog, there is a green status message: "All reagent setup is completed". At the very bottom is a button labeled "Replace(42)", where the number 42 is in a red box, indicating a countdown timer.

Figure 12-2 Countdown - replacing reagents through RFID

- If "Auto replace reagent after setup" is unchecked on the "Setup" > "Auxiliary Setup" > "Other Settings" screen, and you tap "Replace", the analyzer will start the program to replace the old reagent in the pipelines.

## NOTE

- For the instruction to enable "Auto replace reagent after setup", refer to section 6.3.3.2 *Other Settings*.

✓ The analyzer automatically eliminates the fault, loads new reagent information, and replaces the reagent in the remove the error. When you are replacing expired reagents, the analyzer will also perform the reagent priming process.

### 12.3.2.3 Replacing the reagent in the fluidic system and priming reagent

Under the following circumstances, you can replace the old reagent or prime reagent in the pipelines on the reagent maintenance screen.

Program	Timing	Purpose
Reagent Replacement	when the reagent runs out, is insufficient or expired	Replace the residue reagent in the pipelines
Reagent Priming	when reagents are contaminated or expire, or all reagents in the machine need to be replaced under special circumstances	Replace the residue reagent in the pipelines

Follow below instructions:

1. Tap "Menu" > "Service" > "Maintenance" > "Reagent" in turn to enter the reagent maintenance screen.
  2. Tap the reagent for replacing or priming in the pipelines.
- ✓ The analyzer automatically completes the operation.

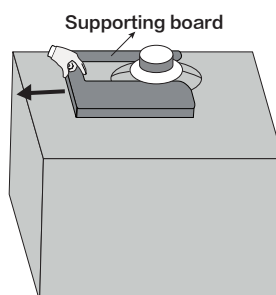
### 12.3.3 Replacing the Waste Container



#### BIOLOGICAL RISK

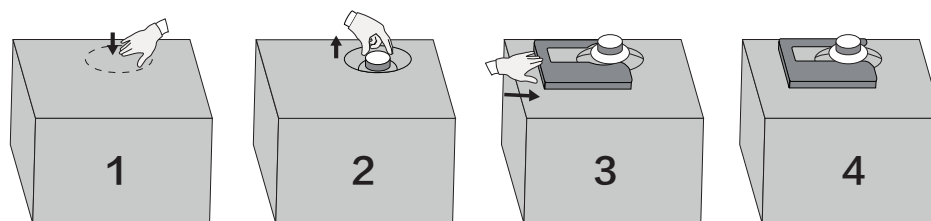
- Remove the waste container cap and replace the waste container only when the power indicator is not flickering, in order not to make the waste overflow from the container.
- If the waste is discharged using waste container, make sure the pickup tube of the waste container cap assembly is above, and the tube is smooth and not bent.

1. Get an empty waste container, remove the cap and place it next to the one to be replaced.
2. Remove the supporting board under the cap of the old container.



3. Turn the cap counterclockwise and remove the cap assembly from the old container with caution.
4. Insert the old cap assembly into the new container as vertically as possible, and secure the cap by turning it clockwise.

5. Install the supporting board under the new container's cap as shown below.



6. Cap the old container with the cap of the new one, and then dispose of the waste properly.

## 12.4 Probe Cleanser Maintenance

### 12.4.1 Daily Probe Cleanser Maintenance

When the Probe Cleanser maintenance conditions are met, the prompt for Probe Cleanser maintenance pops up:

#### NOTE

- For more details, see 6.3.5.2 *Probe cleanser maintenance*.
- For information on Probe Cleanser maintenance, contact Mindray Customer Service Department.

You can perform Probe Cleanser maintenance when the **"Time for maintenance. Perform Probe Cleanser maintenance now?"** dialog box displays or complete maintenance directly on the **"Fluidics"** screen.

Follow procedures below:

1. Start the Probe Cleanser maintenance.
  - When the **"Time for maintenance. Perform Probe Cleanser maintenance now?"** dialog box displays, tap **"Yes"**.
  - Tap **"Menu"**-"**Service**"-"**Maintenance**"-"**Fluidics**" in turn to enter the reagent maintenance screen, and tap **"Probe Cleanser Maint."** button.
  - ✓ The analyzer prepares for Probe Cleanser maintenance. After the preparation for Probe Cleanser maintenance completes, a dialog box displays.
  - ✓ The sample compartment opens.
2. Pick up a tube as instructed, and slowly pour at least 4mL Probe Cleanser into the tube. Place the tube with the Probe Cleanser uncapped into the regular tubes in the compartment.
3. Start the Probe Cleanser maintenance procedure.
  - ✓ The analyzer aspirates Probe Cleanser.
4. Remove the tube with Probe Cleanser when the sample compartment opens.
  - ✓ The analyzer automatically completes Probe Cleanser maintenance.

### 12.4.2 Probe Cleanser Maintenance to Parts and Components

You may perform Probe Cleanser maintenance to parts and components when necessary.

Follow below instructions:

1. Tap **"Menu"**-"**Service**"-"**Maintenance**"-"**Probe Cleanser Maint.**" in turn to enter the **"Probe Cleanser Maint."** screen.
2. Tap the buttons of the parts and components that need Probe Cleanser maintenance.
  - ✓ The analyzer prepares for Probe Cleanser maintenance. After the preparation for Probe Cleanser maintenance completes, a dialog box displays.
  - ✓ The sample compartment opens.



3. Pick up a clean anticoagulant collection tubes as instructed, and slowly pour at least 4ml Probe Cleanser into the tube. Place the tube with the Probe Cleanser uncapped into the regular tubes in the compartment.
4. Start the Probe Cleanser maintenance procedure.
  - ✓ The analyzer aspirates Probe Cleanser.
5. Remove the tube with Probe Cleanser when the sample compartment opens.
  - ✓ The analyzer automatically completes Probe Cleanser maintenance.

## 12.5 Auto-cleaning the Parts and Components

You should clean the following parts or components when:

Follow below instructions:

1. Tap "**Menu**"-"**Service**"-"**Maintenance**"-"**Cleaning**" to enter the cleaning screen.
2. Tap the corresponding cleaning program.
  - ✓ The analyzer automatically completes the operation.

## 12.6 Manual Cleaning of Parts and Components

You should clean the following parts or components when:

Program	Timing	Purpose	Tools needed
Clean the probe wipe, floating blood barrier and the blood barrier bracket	Every month	Remove the blood sample and other residues in the probe wipe	Alcohol, Probe Cleanser, clean water, and sterilized cotton swab
Cover cleaning	As needed	Remove contaminants on the cover	Disinfectant (see <b>12.6.2Cleaning the Analyzer Front Cover</b> for types of disinfectant)
Clean the filtering net	As needed/ Recommend monthly cleaning	Clean the dust on the surface of air inlet filtering net	Clean water

### 12.6.1 Cleaning the Probe Wipe, Floating Blood Barrier and the Blood Barrier Bracket



#### BIOLOGICAL RISK

- The probe wipe, floating blood barrier and blood barrier bracket are potentially infectious as there may be residue blood sample or reagent on them. Wear proper personal protective equipment (e.g. gloves, lab coat, glasses, etc.) and follow safe laboratory procedures when handling them and the contacted areas in the laboratory.



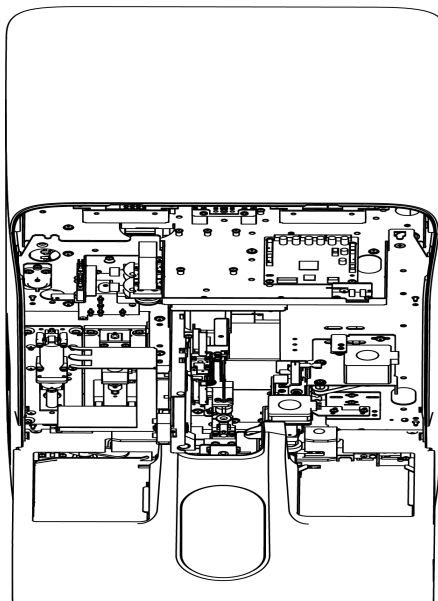
#### WARNING

- Before cleaning the probe wipe, blood barrier, and blood barrier bracket, make sure the analyzer is shut down and the power supply is turned off.

Follow the following instructions:

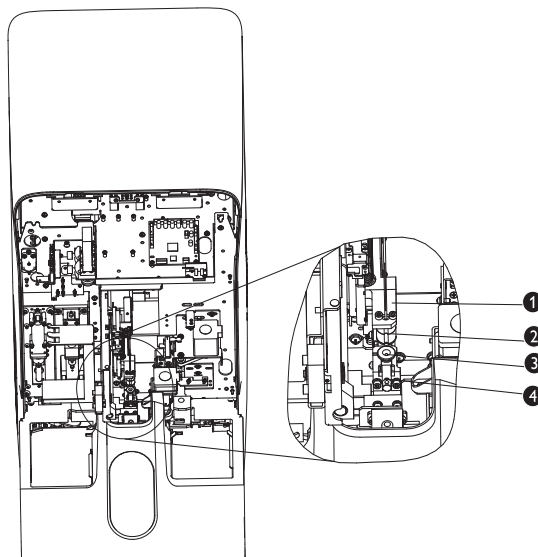
### Opening the front cover of the analyzer

Hold the protruding parts at the bottom of the front cover with both hands and open the front cover upward.



### Cleaning the probe wipe, floating blood barrier and the blood barrier bracket

1. Push the sampling probe assembly forward slightly to reserve sufficient operation and maintenance space. See the arrow direction shown in the figure below.



① Sampling probe assembly

② Probe wipe

③ Floating blood barrier

④ Blood barrier bracket

2. Use a sterilized cotton swab dipped with Probe Cleanser to wipe the surface of the probe wipe. Then, use a sterilized cotton swab dipped with clean water to wipe the surface until no blood residue or other residues are visible.

---

**⚠ CAUTION**


---

- After dipping a cotton swab in the Probe Cleanser or clean water, ensure that the cotton swab has no droplet. Otherwise, the droplet may drop into the autoloader, damaging components.
- 

3. Use a sterilized cotton swab dipped with alcohol to wipe the surface of the blood barrier and blood barrier bracket until no blood residue or other residues are visible.
- 

**⚠ CAUTION**


---

- After dipping a cotton swab in the alcohol or clean water, ensure that the cotton swab has no droplet. Otherwise, the droplet may drop into the autoloader, damaging components.
- 

**Closing the Front Cover of the Analyzer**

Gently lower the cover and close it.

---

**⚠ CAUTION**


---

- Closing the front cover gently to prevent the vibration from damaging internal parts of the analyzer.
- 

## 12.6.2 Cleaning the Analyzer Front Cover

---

**⚠ BIOLOGICAL RISK**


---

- Mindray does not claim the validity of the listed chemicals in infection control. For effective control of infection, please consult the Infection Prevention Department of the hospital or the epidemic professionals.
- 

**⚠ CAUTION**


---

- The user shall perform regular cleaning and sterilization to the cover of the instrument. Use the specified materials to sterilize the instrument only. For any damage to the instrument or other accidents caused by using materials other than specified, Mindray will not provide any warranty.
  - The cleaning and sterilization may damage the instrument to some extent. It is recommended to perform sterilization only when necessary according to your laboratory protocol. Remember to clean the instrument before sterilizing.
  - Do not use any decontamination or cleaning agents which could cause a HAZARD as a result of a reaction with parts of the instrument or with material contained in it.
  - If you accidentally spill hazardous material (for example, reagents or samples) on the instrument, clean the instrument with specified disinfectant. Wear proper personal protective equipment (e.g. gloves, lab coat, etc.) and follow safe laboratory procedures when handling them and the contacted areas in the laboratory.
- 

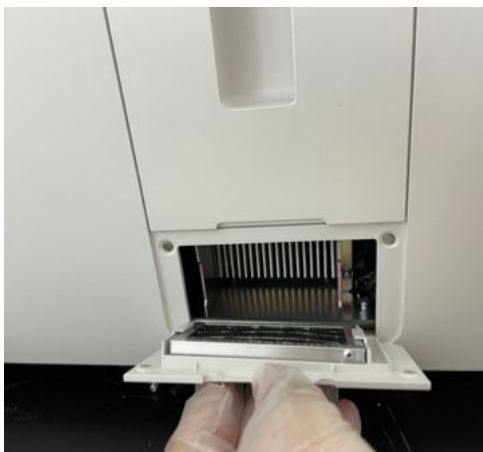
Perform regular cleaning on the covers of the analyzer.

Recommended disinfectant: water, 75% ethanol.

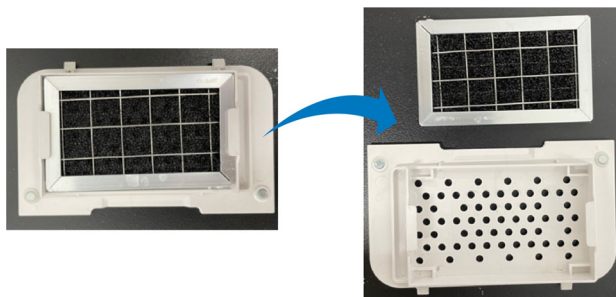
Prohibited disinfectant: 3% hydrogen peroxide.

### 12.6.3 Cleaning the Filter

1. Open the filtermesh compartment on the left side of the analyzer.



2. Pull out the air inlet filter.



3. Wash the filter with clean water, and use a piece of clean and dry cloth to wipe up the water in the filter.



4. Install the filter to its original position, and then close the filtermesh compartment.

## 12.7 Preparing to Ship

If the analyzer is not to be used for a long time (over 10 days), you should perform this procedure.

Follow below instructions:

1. Tap **"Menu"**-**"Service"**-**"Maintenance"**-**"Fluidics"** to enter the **"Fluidics"** screen.
2. Tap **"Prepare to Ship"**, and follow the software instruction to complete the pack-up procedure.

## 12.8 Screen Calibration

If the touch screen does not correctly respond to the positions you touched, perform the procedure to calibrate the touch screen.

### CAUTION

- Do not use anything sharp on the touch screen or strike on it.

### NOTE

- Do not click with the mouse to calibrate the touch screen.

Follow below instructions:

- Tap **"Menu"**-**"Service"**-**"Screen Cal."** to enter the **"Screen Cal."** screen.
  - Tap **"Screen Cal."** in the middle of the screen.
  - Tap the black plus sign at the upper left corner of the screen as instructed by the screen display to start the calibration.
- ✓ After the calibration is completed, the software displays **"Calibration succeeded."** on the screen.

## 12.9 Viewing and Exporting Logs

The "Log" screen records all activities of the analyzer. It contributes significantly to searching for operation history and troubleshooting the analyzer.

The analyzer can save logs of the recent two years. If number of logs exceeds the upper limit, the latest log will overwrite the oldest one. You can browse and print logs, but cannot delete them.

Administrators and common users have different authorities:

**Table 12-6 Log Types**

	Administrator's level	Operator's level
<b>All Logs</b>	Review all types of logs	View the logs for analyzer startup and shutdown, user logging in and logging out at the operator's level.
<b>General Logs</b>	View all operation-related logs at both administrator's and operator's levels.	View the logs for analyzer startup and shutdown, user logging in and logging out at the operator's level.
<b>Setup Adjustment</b>	View all setting adjustment logs at both administrator's and operator's levels.	Cannot review
<b>Error Information</b>	Review error information and troubleshooting information of the analyzer.	Cannot review

### 12.9.1 Viewing Logs

Follow below instructions:

- Tap **"Menu"**-**"Service"**-**"Log"** in turn to enter the **"Log"** screen.
- Tap a type of logs to be viewed.



- (Optional) Review the logs at specified date range.
  - Tap **"Go to"**.

- ✓ A confirm dialog box displays.

- b In the "**Date**" edit box, specify the date on which the logs need to be viewed.
  - c Tap "**OK**".
- ✓ The screen displays the logs at the specified date.

## 12.9.2 Exporting Logs

You may export the logs in specified time range to the USB device.

Before exporting sample records, make sure that you have inserted a safe USB flash drive into the USB port on the analyzer.

### CAUTION

- **The user should ensure the data safety of the USB devices connecting to the analyzer.**

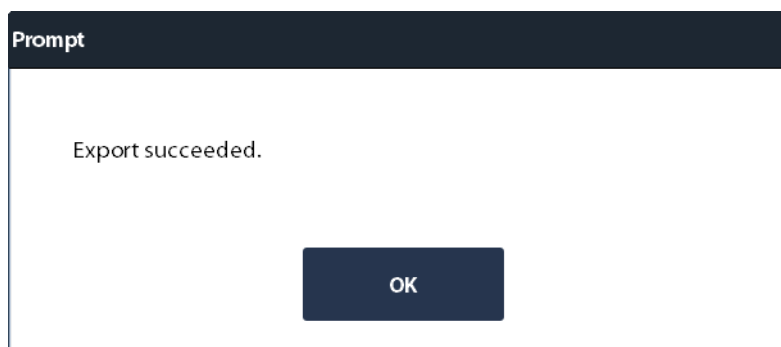
Follow below instructions:

1. Tap "**Menu**"-"**Service**"-"**Log**" in turn to enter the "**Log**" screen.
2. Tap "**Export**".

- ✓ A confirm dialog box displays.

3. In the "**Start date:**" and "**End date:**" edit boxes, specify the time range in which logs need to be exported.
  4. Tap "**OK**".
- ✓ The analyzer automatically exports the logs in specified time range to the USB device.

- √ After export ends, the screen displays the "**Export succeeded.**" dialog box.



## 12.10 Upgrading Analyzer

---

### CAUTION

---

- Once upgraded, the analyzer cannot be restored to the last version.
  - Analyzer upgrade may only be performed under the instruction from a Mindray-authorized service personnel. Do not perform the procedure by yourself.
-

**This page intentionally left blank.**



# 13 Troubleshooting

## 13.1 Overview

This chapter contains information that is helpful in locating and correcting problems that may occur during operation of your analyzer.

### NOTE

- This chapter is not a complete service manual and is limited to problems that are readily diagnosed and/or corrected by the user of the analyzer.

## 13.2 Checking Analyzer Status

You can check analyzer information from the “**Status**” menu, including statistics, temperature and pressure, floater status, sensor status, voltage and current, as well as version information. Checking the status information on the “**Status**” screen is significant for you to locate and remove errors of the analyzer.

Follow the path below to review analyzer status:

Item/Software access	Status	Path	Access Level Requirement
Statistics	Valid Runs	Menu-Status-Statistics	All
	Runs since latest initialization		
	Sample Runs		
	QC Runs		
	Calibration Runs		
	Valid Sample Runs		
	Valid Runs After Latest Startup		
	Runs after Probe Cleanser Maintain		
	Clogs in Impedance Channel		
	Background Runs		
	Wearing Parts		
Temp.&Pressure	Displays the current temperature and pressure as well as the acceptable range for various items. Out-of-range values are highlighted in red background.	Menu-Status-Temp.&Pressure	Administrator's level
Floater Status	Displays the full or empty status of the bathes and waste cistern.	Menu-Status-Floater Status	Administrator's level
Sensor	Displays the status of the sensors of the fluorescent dye detection, and the sensors on the drive board and the main control board.	Menu-Status-Sensor	Administrator's level

Item/Software access	Status	Path	Access Level Requirement
<b>Voltage &amp; Current</b>	Displays the voltage and current information Out-of-range values are highlighted in red background.	<b>Menu-Status-Voltage &amp; Current</b>	Administrator's level
<b>Version Info.</b>	Review the analyzer software version information	<b>Menu-Status-Version Info.</b>	All

### 13.3 Error Messages and Solutions

During the operation, if error(s) is detected, "**Error Information**" dialog box will pop up and the analyzer will beep.

The background colors of error messages turn red, orange, blue, and green according to error severity.

- Red: fatal error when this kind of error occurs, the analyzer will stop running immediately, and any further operation is prohibited.
- Orange: error that stops operation. When this kind of error occurs, the analyzer will stop running immediately.
- Blue: error that restricts certain operations. When this kind of error occurs, the analyzer can still continue with the current operation, but any other operations related to the error will be restricted.
- Green: prompting error when this kind of error occurs, the analyzer can still continue with the current operation, and other operations are not restricted.

The name and troubleshooting method of the errors are displayed in the "**Error Information**" dialog box. Names of the errors are displayed by the order of their occurrence.

You may tap to select the error, and view its troubleshooting information in the "**Error Description**". The troubleshooting information of the first error is displayed by default. Follow the instructions in the dialog box to remove error(s).

The following functions are provided:

- Remove error

Tap the "Remove Error" button to clear all the errors that can be removed automatically. For the errors that cannot be removed automatically, follow the troubleshooting method to solve them.

- To mute the alarm sound

Tap the touch screen to eliminate the alarm sound of the main unit.

- Close the "**Error Information**" dialog box

Tap "**Off**" to close the dialog box, but the errors will still be displayed in the error info. area on the screen. Tap the error info. area again, the dialog box will be displayed.

Error ID	Error Message	Description	Solution
0x10103	Waste Container Full	Waste Container Full	1. Replace the waste container with an empty one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10000	No DS Diluent. Replace the reagent	No DS Diluent. Replace the reagent	1. Replace the DS Diluent. 2. Click the "Remove Error" button to remove this error. 3. If the error persists, contact our Customer Service Department.

Error ID	Error Message	Description	Solution
0x10001	No LD Lyse. Replace the reagent	No LD Lyse. Replace the reagent	1. Replace the LD Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10003	No LH Lyse. Replace the reagent	No LH Lyse. Replace the reagent	1. Replace the LH Lyse with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10002	No DR Diluent. Replace the reagent	No DR Diluent. Replace the reagent	1. Replace the DR Diluent with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10009	No ESR Solution Reagent. Replace the reagent	No ESR Solution Reagent. Replace the reagent	1. Replace the ESR Solution Reagent; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10005	No FD Dye. Replace the reagent	No FD Dye. Replace the reagent	1. Replace the FD Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10006	No FR Dye. Replace the reagent	No FR Dye. Replace the reagent	1. Replace the FR Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10200	DS Diluent expires. Replace the reagent	DS Diluent expires. Replace the reagent	1. Replace the DS Diluent. 2. Click the "Remove Error" button to remove this error. 3. If the error persists, contact our Customer Service Department.
0x10201	LD Lyse expires. Replace the reagent	LD Lyse expires. Replace the reagent	1. Replace the LD Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10202	LH Lyse expires. Replace the reagent	LH Lyse expires. Replace the reagent	1. Replace the LH Lyse with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10203	DR Diluent expires. Replace the reagent	DR Diluent expires. Replace the reagent	1. Replace the DR Diluent with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x10209	ESR Solution Reagent expires. Replace the reagent	ESR Solution Reagent expires. Replace the reagent	1. Replace the ESR Solution Reagent; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10204	FD Dye expires. Replace the reagent	FD Dye expires. Replace the reagent	1. Replace the FD Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10205	FR Dye expires. Replace the reagent	FR Dye expires. Replace the reagent	1. Replace the FR Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10400	DS Diluent low volume. Replace the reagent	DS Diluent low volume. Replace the reagent	1. Replace the DS Diluent. 2. Click the "Remove Error" button to remove this error. 3. If the error persists, contact our Customer Service Department.
0x10401	LD Lyse low volume. Replace the reagent	LD Lyse low volume. Replace the reagent	1. Replace the LD Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10402	LH Lyse low volume. Replace the reagent	LH Lyse low volume. Replace the reagent	1. Replace the LH Lyse with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10403	DR Diluent low volume. Replace the reagent	DR Diluent low volume. Replace the reagent	1. Replace the DR Diluent with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10409	ESR reagent low volume. Replace the reagent	ESR reagent low volume. Replace the reagent	1. Replace the ESR Solution Reagent; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10404	FD Dye low volume. Replace the reagent	FD Dye low volume. Replace the reagent	1. Replace the FD Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x10405	FR Dye low volume. Replace the reagent	FR Dye low volume. Replace the reagent	1. Replace the FR Dye with a new one; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x40003	Import "Key." file	Import "Key." file	Tap Menu - "Service" - "Advanced Toolbox" - "Debug Setup" - "Import Password"; otherwise, reagents cannot be replaced when built-in authorization run out.
0x10100	DIL preheating bath sensor abnormal	DIL preheating bath sensor abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, tap "Precise Fault Diagnosis" to enter the "Precise Fault Diagnosis" screen to locate the error. 3. If no error is found, re-start the analyzer; otherwise, contact our Customer Service department.
0x10102	Waste cistern floater status abnormal	WC2 waste cistern floater status abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x10101	Cistern floater status abnormal	SCI cistern floater status abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, tap "Precise Fault Diagnosis" to enter the "Precise Fault Diagnosis" screen to locate the error. 3. If no error is found, re-start the analyzer; otherwise, contact our Customer Service department.
0x30105	FS baseline abnormal	FS baseline abnormal	1. Tap the "Remove Error" button to remove the error; 2. Switch off and then switch on the instrument power; 3. If the error still exists after the restart, contact our customer service department.
0x30106	FS baseline abnormal	SS baseline abnormal	1. Tap the "Remove Error" button to remove the error; 2. Switch off and then switch on the instrument power; 3. If the error still exists after the restart, contact our customer service department.
0x30107	FS baseline abnormal	FL baseline abnormal	1. Tap the "Remove Error" button to remove the error; 2. Switch off and then switch on the instrument power; 3. If the error still exists after the restart, contact our customer service department.
0x30200	HGB baseline abnormal	HGB baseline abnormal	1. Tap the "Remove Error" button to remove the error; 2. Switch off and then switch on the instrument power; 3. If the error still exists after the restart, contact our customer service department.

Error ID	Error Message	Description	Solution
0x30401	ESR signal abnormal	ESR signal abnormal	1.Tap the "Remove Error" button to remove the error; 2.Switch off and then switch on the instrument power; 3.If the error still exists after the restart, contact our customer service department.
0x00300	DIL syringe action abnormal	Invalid command to DIL syringe	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00301	DIL syringe action abnormal	Conflicting DIL syringe actions	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00303	DIL syringe action abnormal	Error occurs when DIL syringe leaves sensor area	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00302	DIL syringe action abnormal	Error occurs when DIL syringe returns to home position	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00305	DIL syringe action abnormal	DIL syringe aspiration/ dispensation action failure 1	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00304	DIL syringe action abnormal	DIL syringe aspiration/ dispensation action failure 2	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00307	DIL syringe action abnormal	DIL syringe aspiration/ dispensation action not allowed 1	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00306	DIL syringe action abnormal	DIL syringe aspiration/ dispensation action not allowed 2	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00308	DIL syringe action abnormal	DIL syringe aspirated volume too high	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00310	DIL syringe action abnormal	DIL syringe action time out	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00309	DIL syringe action abnormal	DIL syringe dispensed volume too high	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x00200	SP syringe action abnormal	Invalid command to SP syringe	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00201	SP syringe action abnormal	Conflicting SP syringe actions	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00203	SP syringe action abnormal	Error occurs when SP syringe leaves sensor area	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00202	SP syringe action abnormal	Error occurs when SP syringe returns to home position	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00205	SP syringe action abnormal	SP syringe aspiration/ dispensation action failure 1	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00204	SP syringe action abnormal	SP syringe aspiration/ dispensation action failure 2	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00206	SP syringe action abnormal	SP syringe aspiration/ dispensation action not allowed 1	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00207	SP syringe action abnormal	SP syringe aspiration/ dispensation action not allowed 2	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00208	SP syringe action abnormal	SP syringe aspirated volume too high	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00210	SP syringe action abnormal	SP syringe action time out	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x00209	SP syringe action abnormal	SP syringe dispensed volume too high	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07000	Sampling assembly action abnormal	Invalid command to sample probe	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x07100	Sampling assembly action abnormal	Sampling assembly failed to return to home position in vertical direction	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07101	Sampling assembly action abnormal	Sampling assembly failed to leave home position in vertical direction	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07200	Sampling assembly action abnormal	Sampling assembly failed to return to home position in horizontal direction	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07201	Sampling assembly action abnormal	Sampling assembly failed to leave home position in horizontal direction	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07202	Sampling assembly action abnormal	Sampling assembly confirmation sensors in horizontal direction abnormally blocked	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07203	Sampling assembly action abnormal	Sampling assembly notch count jumps in horizontal direction abnormal	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07102	Sampling assembly action abnormal	Sample probe pierce action loses steps	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07103	Sampling assembly action abnormal	Sample probe lifting action loses steps	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07001	Sampling assembly action abnormal	Conflicting command to sampling assembly	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07204	Sampling assembly action abnormal	Sampling assembly actions in horizontal direction time out	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07104	Sampling assembly action abnormal	Sampling assembly actions in vertical direction time out	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07105	Sampling assembly action abnormal	Vertical home position sensors are not blocked before sampling assembly actions in horizontal direction	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.



Error ID	Error Message	Description	Solution
0x01000	Mixing gripper action error	Mixing gripper action error	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01001	Mixing gripper action error	Mixing gripper action error	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x04100	Loading mechanism action error	Loading mechanism fails to leave sensor area in initialization	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x04101	Loading mechanism action error	Loading mechanism fails to return to sensor area in initiation	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x04102	Loading mechanism action error	Fails to load tube racks	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x04103	Loading mechanism action error	Loading mechanism home position sensor status abnormal	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x04104	Loading mechanism action error	Loading mechanism end position sensor status abnormal	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x04105	Loading mechanism action error	Sensor mistakenly triggered during loading mechanism returning to home position	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>

Error ID	Error Message	Description	Solution
0x04106	Loading mechanism action error	Loading end position sensor is mistakenly triggered	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04107	Auto loading sensor abnormal	Auto loading sensor abnormal	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04200	Loading mechanism action error	Unloading mechanism fails to leave sensor area in initialization	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04201	Loading mechanism action error	Unloading mechanism fails to return to sensor area in initiation	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04202	Remove all the tube racks on the unload tray	Unloading tray is full	1.Remove all the tube racks on the unload tray, and the instrument will automatically remove the error; 2.If the error still exists, contact our customer service department.
0x04203	Loading mechanism action error	Sensor mistakenly triggered during unloading mechanism returning to home position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04204	Loading mechanism action error	Unloading mechanism protruding failed	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04300	Loading mechanism action error	Feeding mechanism fails to leave sensor area in initialization	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04301	Loading mechanism action error	Feeding mechanism fails to return to sensor area in initiation	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04302	Loading mechanism action error	Sensor failed during feeding mechanism returning to right position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x04303	Loading mechanism action error	Feeding mechanism failed to leave home position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04304	Loading mechanism action error	Sensor mistakenly triggered during feeding mechanism returning to home position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04305	Loading mechanism action error	Sensor mistakenly triggered during feeding mechanism returning to right position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04306	Loading mechanism action error	Feeding mechanism loses step at the left side	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04307	Loading mechanism action error	Feeding mechanism loses step at the right side	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04308	Loading mechanism action error	Left sensor abnormal during feeding mechanism initialization	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04309	Loading mechanism action error	Sensor failed during feeding mechanism reaching left position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04310	Loading mechanism action error	Sensor mistakenly triggered during feeding mechanism reaching left position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04002	Loading mechanism action error	Manually move of tube rack	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04311	Adjacent tubes with the same barcode in the tube rack	Adjacent tubes with the same barcode in the tube rack	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x04312	Loading mechanism action error	Feeding mechanism loses step when returning	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x02000	Scanning mechanism error	Scanning mechanism error	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x02002	Rotary scanning mechanism action error	Tube clamping mechanism stretch failed	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x02003	Rotary scanning mechanism action error	Tube clamping mechanism retraction failed	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x02004	Rotary scanning mechanism action error	Tube fixing mechanism failed to leave home position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x02005	Rotary scanning mechanism action error	Tube fixing mechanism failed to return to home position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x02006	Rotary scanning mechanism action error	Sensor mistakenly triggered during tube fixing mechanism returning to home position	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x02001	Scanner communication abnormal	Scanner communication abnormal	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x02007	Rotary scanning mechanism action error	Tube fixing mechanism slide rail is stuck	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x04500	Sample compartment action abnormal	Sample compartment failed to leave home position	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x04501	Sample compartment action abnormal	Sample compartment failed to return to home position	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x04502	Sample compartment action abnormal	Sample compartment failed to open	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x04503	Sample compartment action abnormal	Sensor mistakenly triggered during sample compartment returning to home position	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x04504	Sample compartment action abnormal	Sample compartment home position sensor status abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x01100	Mixing gripper action error	Sensor mistakenly triggered during tube gripper returning to home position in Y direction	1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes. 2. Remove all tube racks from the loading platform. 3. Click the "Remove Error" button to remove this error. 4. If the error persists, contact our Customer Service Department.
0x01101	Mixing gripper action error	Tube gripper failed to leave home position in Y direction	1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes. 2. Remove all tube racks from the loading platform. 3. Click the "Remove Error" button to remove this error. 4. If the error persists, contact our Customer Service Department.
0x01102	Mixing gripper action error	Tube gripper failed to return to home position in Y direction	1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes. 2. Remove all tube racks from the loading platform. 3. Click the "Remove Error" button to remove this error. 4. If the error persists, contact our Customer Service Department.
0x01104	Mixing gripper action error	Tube gripper Y direction home position sensor status abnormal	1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes. 2. Remove all tube racks from the loading platform. 3. Click the "Remove Error" button to remove this error. 4. If the error persists, contact our Customer Service Department.

Error ID	Error Message	Description	Solution
0x01300	Mixing gripper action error	Tube gripper Y direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01305	Mixing gripper action error	Tube gripper Y direction end position sensor mistakenly triggered	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01302	Mixing gripper action error	Mixing gripper failed to stretch to front position in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01109	Mixing gripper action error	Mixing gripper failed to retract to rear position in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01200	Mixing gripper action error	Mixing gripper failed to stretch to midposition in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01201	Mixing gripper action error	Sensor mistakenly triggered during mixing gripper stretching to midposition in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01202	Mixing gripper action error	Mixing gripper failed to retract to midposition in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01203	Mixing gripper action error	Sensor mistakenly triggered during mixing gripper retracting to midposition in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01204	Mixing gripper action error	Mixing gripper Y direction midposition sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01205	Mixing gripper action error	Mixing gripper Y direction midposition sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01103	Mixing gripper action error	Tube gripper Y direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01105	Mixing gripper action error	Mixing gripper failed to return to home position in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01106	Mixing gripper action error	Tube gripper Y direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01107	Mixing gripper action error	Mixing gripper Y direction home position sensor mistakenly triggered	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01303	Mixing gripper action error	Mixing gripper failed to reach end position in Y direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>



Error ID	Error Message	Description	Solution
0x01304	Mixing gripper action error	Tube gripper Y direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01301	Mixing gripper action error	Tube gripper Y direction end position sensor mistakenly triggered	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01306	Mixing gripper action error	Tube gripper Y direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01307	Mixing gripper action error	Tube gripper Y direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01108	Mixing gripper action error	Tube gripper Y direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01406	Mixing gripper action error	Sensor mistakenly triggered during tube gripper returning to home position in Z direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01402	Mixing gripper action error	Tube gripper failed to leave home position in Z direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01403	Mixing gripper action error	Tube gripper failed to return to home position in Z direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01411	Mixing gripper action error	Tube gripper Z direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01500	Mixing gripper action error	Tube gripper Z direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01510	Mixing gripper action error	Tube gripper Z direction end position sensor mistakenly triggered	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01502	Mixing gripper action error	Tube gripper failed to lift up in Z direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01405	Mixing gripper action error	Tube gripper failed to lower down in Z direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01401	Mixing gripper action error	Sensor mistakenly triggered during tube gripper returning to home position in Z direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01511	Mixing gripper action error	Tube gripper Z direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01504	Mixing gripper action error	Tube gripper Z direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01412	Mixing gripper action error	Tube gripper Z direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01512	Mixing gripper action error	Tube gripper Z direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01404	Mixing gripper action error	Tube gripper Z direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01409	Mixing gripper action error	Mixing gripper failed to descend in Z direction at capillary blood position (micro-WB position)	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01410	Mixing gripper action error	Mixing gripper Z direction home position sensor at micro-WB position mistakenly triggered	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01407	Mixing gripper action error	Tube gripper Z direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01508	Mixing gripper action error	Sensor mistakenly triggered at end position in Z direction of the micro-WB position of the mixing gripper	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01509	Mixing gripper action error	Mixing gripper failed to ascend in Z direction at micro-WB position	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01408	Mixing gripper action error	Tube gripper Z direction home position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01600	Mixing gripper action error	Sensor mistakenly triggered during tube gripper returning to home position in R direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01601	Mixing gripper action error	Tube gripper failed to leave home position in R direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01602	Mixing gripper action error	Tube gripper failed to return to home position in R direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01603	Mixing gripper action error	Tube gripper R direction end position sensor status abnormal	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x01604	Mixing gripper action error	Tube gripper Z direction end position sensor status abnormal during mixing	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>

Error ID	Error Message	Description	Solution
0x01605	Mixing gripper action error	Tube gripper failed to move to home position in R direction	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x04600	Micro-WB mixer action abnormal	Sensor mistakenly triggered during micro-WB mixer returning to home position	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04601	Micro-WB mixer action abnormal	Micro-WB mixer failed to leave home position	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04602	Micro-WB mixer action abnormal	Micro-WB mixer failed to return to home position	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04603	Micro-WB mixer action abnormal	Micro-WB mixer sensor status abnormal	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04604	Micro-WB mixer action abnormal	Micro-WB mixer sensor jump	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04605	Micro-WB mixer action abnormal	Sensor mistakenly triggered during micro-WB mixer returning to home position	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04606	Micro-WB mixer action abnormal	Micro-WB mixer failed to leave home position	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04607	Micro-WB mixer action abnormal	Micro-WB mixer failed to return to home position	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x04400	Counter error	Counter sensor status error	<ol style="list-style-type: none"> <li>1. Remove all the tube racks on the load tray;</li> <li>2. Tap the "Remove Error" button to remove this error;</li> <li>3. If the error still exists, contact our customer service department.</li> </ol>
0x04003	Counter error	Wrong counter count jump times	<ol style="list-style-type: none"> <li>1. Remove all the tube racks on the load tray;</li> <li>2. Tap the "Remove Error" button to remove this error;</li> <li>3. If the error still exists, contact our customer service department.</li> </ol>

Error ID	Error Message	Description	Solution
0x04004	Counter error	Manually move of tube rack	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x04000	Autoloading status error	Autoloading status error	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x04505	Sample compartment status error	Sample compartment status error	<ol style="list-style-type: none"> <li>1.Tap the "Remove Error" button to remove this error;</li> <li>2.If the error still exists, contact our customer service department.</li> </ol>
0x01002	Mix assembly status error	Mix assembly status error	<ol style="list-style-type: none"> <li>1. Open the front cover and check whether there are test tubes in the mixing gripper and micro-WB mixer. If yes, remove the test tubes.</li> <li>2. Remove all tube racks from the loading platform.</li> <li>3. Click the "Remove Error" button to remove this error.</li> <li>4. If the error persists, contact our Customer Service Department.</li> </ol>
0x07002	Motor debug error	Motor debug error	<ol style="list-style-type: none"> <li>1.Tap the "Remove Error" button to remove this error;</li> <li>2.If the error still exists, contact our customer service department.</li> </ol>
0x04001	Clear all the tube racks on the feeding channel	There are tube racks on the feeding channel	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>
0x10308	Auto pressure building out of time	Auto pressure building out of time	<ol style="list-style-type: none"> <li>1.Tap the "Remove Error" button to remove this error;</li> <li>2.If the error still exists, contact our customer service department.</li> </ol>
0x99800	Startup initiation not performed	Startup initiation not performed	<ol style="list-style-type: none"> <li>1.Tap the "Remove Error" button to remove this error;</li> <li>2.If the error still exists, contact our customer service department.</li> </ol>
0x04005	Autoloader board communication out of time	Autoloader board communication out of time	<ol style="list-style-type: none"> <li>1.Remove all the tube racks on the load tray;</li> <li>2.Tap the "Remove Error" button to remove this error;</li> <li>3.If the error still exists, contact our customer service department.</li> </ol>



Error ID	Error Message	Description	Solution
0x20012	Drive board communication out of time	Drive board communication out of time	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10107	SCI priming out of time	SCI priming out of time	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x99805	Background abnormal	Background abnormal	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10306	Fluidic system status abnormal	Sampling probe clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10307	Fluidic system status abnormal	Flow cell clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x99801	Exiting standby status failed	Exiting standby status failed	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x99802	Exiting standby status failed	Exiting standby status failed	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x99803	Auto startup failed	Auto startup failed	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10111	SCI priming failed	SCI syringe is busy	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x20008	Closed-reagent RFID board communication timeout	Closed-reagent RFID board communication timeout	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x01003	Mix motor board communication timeout	Mix motor board communication timeout	1.Remove all the tube racks on the load tray; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x30109	Optical signal board communication timeout	Optical signal board communication timeout	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x30400	ESR board communication timeout	ESR board communication timeout	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x20011	Power supply board communication timeout	Power supply board communication timeout	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x10300	50kPa pressure out of range	50kPa pressure out of range	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, tap "Precise Fault Diagnosis" to enter the "Precise Fault Diagnosis" screen to locate the error. 3. If no error is found, re-start the analyzer; otherwise, contact our Customer Service department.
0x10304	-40kPa pressure out of range	-40kPa pressure out of range	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, tap "Precise Fault Diagnosis" to enter the "Precise Fault Diagnosis" screen to locate the error. 3. If no error is found, re-start the analyzer; otherwise, contact our Customer Service department.
0x10302	40 kPa pressure out of range	40 kPa pressure out of range	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, tap "Precise Fault Diagnosis" to enter the "Precise Fault Diagnosis" screen to locate the error. 3. If no error is found, re-start the analyzer; otherwise, contact our Customer Service department.
0x10104	Waste channel abnormal	Waste channel abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x10301	Pressure cell pressure release abnormal	Pressure cell pressure release abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x10303	SCI bath pressure release abnormal	SCI bath pressure release abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x10305	WC2 bath pressure release abnormal	WC2 bath pressure release abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x30000	Reaction bath temperature high	Reaction bath temperature high	1. Click "Remove Error" button and recheck the temperature ; 2. If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x30001	Preheating bath temperature control abnormal	Preheating bath temperature out of the upper limit for counting	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30005	Ambient temperature is high	Ambient temperature is high	1. Make sure the ambient temperature is within acceptable range. 2.Tap the"Remove Error "button to re-test the temperature. 3.If the error still exists, contact our customer service department.
0x30002	Temperature inside analyzer out of range	Temperature inside analyzer out of range	1. Make sure the analyzer is placed in a place with good ventilation, heat dispersion and with no direct sunlight. 2.Tap the"Remove Error"button to re-test the temperature. 3.If the error still exists, contact our customer service department.
0x30404	ESR assembly temperature too high	ESR assembly temperature too high	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30003	Reaction bath temperature low	Reaction bath temperature low	1.Click"Remove Error"button and recheck the temperature ; 2.If the error still exists, contact our customer service department.
0x30004	Preheating bath temperature control abnormal	Preheating bath temperature out of the lower limit for counting	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30006	Ambient temperature low	Ambient temperature low	1. Make sure the ambient temperature is within acceptable range. 2.Tap the"Remove Error "button to re-test the temperature. 3.If the error still exists, contact our customer service department.
0x30403	ESR assembly temp. too low	ESR assembly temp. too low	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30007	Ambient temperature is high	Ambient temperature is high	1. Make sure the ambient temperature is within acceptable range. 2.Tap the"Remove Error "button to re-test the temperature. 3.If the error still exists, contact our customer service department.
0x30008	Ambient temperature low	Ambient temperature low	1. Make sure the ambient temperature is within acceptable range. 2.Tap the"Remove Error "button to re-test the temperature. 3.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x30009	Reaction bath temperature control assembly is damaged	Reaction bath temperature control assembly is damaged	1.Click"Remove Error"button and recheck the temperature ; 2.If the error still exists, contact our customer service department.
0x30010	Preheating bath temperature control abnormal	Preheating bath does not achieve target temperature after startup procedure	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30100	Optical system working voltage abnormal	PMT voltage abnormal	1.Tap the "Remove Error" button to remove the error; 2.Switch off and then switch on the instrument power; 3.If the error still exists after the restart, contact our customer service department.
0x30201	HGB blank voltage abnormal	HGB blank voltage abnormal	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, tap "Precise Fault Diagnosis" to enter the "Precise Fault Diagnosis" screen to locate the error. 3. If no error is found, re-start the analyzer; otherwise, contact our Customer Service department.
0x30402	ESR background intensity abnormal	ESR background intensity abnormal	1.Click"Remove Error"button and recheck the temperature ; 2.If the error still exists, contact our customer service department.
0x30103	Flow cell contaminated	DIFF channel FS blank voltage abnormal	1.On the instrument main unit software, tap Menu - "Service" - "Maintenance" - "Fluidics" to enter the "Fluidics" maintenance screen, and perform the daily probe cleanser maintenance procedure; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x30104	Flow cell contaminated	RET channel FS blank voltage abnormal	1.On the instrument main unit software, tap Menu - "Service" - "Maintenance" - "Fluidics" to enter the "Fluidics" maintenance screen, and perform the daily probe cleanser maintenance procedure; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.
0x20203	Fan in the analyzer faulty	Radiator fan in the analyzer is blocked	1.Check if the fan located on the back of the analyzer main unit is stuck by any foreign objects; 2.Tap the "Remove Error" button to remove the error; 3.If the error still exists after restarting the instrument, contact our customer service department.

Error ID	Error Message	Description	Solution
0x99804	Front cover is open	Front cover is open	1. Tap the "Remove Error" button to remove this error; 2. If the error still exists, contact our customer service department.
0x30108	Optical system shielding box is open	Optical system shielding box is open	1. Close the Optical system shielding box; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x40001	System time error	System time error	1. On the instrument main unit screen, tap Menu"- "Setup" - "Date/Time Setup" to enter the "Date/Time Setup" screen and set up the correct system time; 2. Tap the "Remove Error" button to remove this error; 3. If the error still exists, contact our customer service department.
0x30300	Clog	Aperture voltage abnormal	1. Aperture voltage abnormal. 2. Tap the "Remove Error" button to remove this error. 3. If the error still exists, contact our customer service department.
0x30301	Clog	Aperture voltage abnormal	1. Aperture voltage abnormal. 2. Tap the "Remove Error" button to remove this error. 3. If the error still exists, contact our customer service department.
0x30302	Clog	RBC sample preparation abnormal	1. RBC sample preparation abnormal. 2. Tap the "Remove Error" button to remove this error. 3. If the error still exists, contact our customer service department.
0x30303	Clog	RBC sample preparation abnormal	1. RBC sample preparation abnormal. 2. Tap the "Remove Error" button to remove this error. 3. If the error still exists, contact our customer service department.
0x20200	Power fan error	Power fan blocked	1. Check whether the power fan is stuck. 2. If the error persists, contact our Customer Service Department.
0x20201	Board fan faulty	Board radiator fan is blocked	1. Check if the fan located on the back of the analyzer main unit is stuck by any foreign objects; 2. Tap the "Remove Error" button to remove the error; 3. If the error still exists after restarting the instrument, contact our customer service department.

Error ID	Error Message	Description	Solution
0x20202	Board fan faulty	Board radiator fan is blocked	<ol style="list-style-type: none"> <li>1. Check if the fan located on the back of the analyzer main unit is stuck by any foreign objects;</li> <li>2. Tap the "Remove Error" button to remove the error;</li> <li>3. If the error still exists after restarting the instrument, contact our customer service department.</li> </ol>
0x20002	Air pressure detection board error	Air pressure detection board communication error	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove the error;</li> <li>2. Switch off and then switch on the instrument power;</li> <li>3. If the error still exists after the restart, contact our customer service department.</li> </ol>
0x20003	Air pressure detection board error	Air pressure detection board calibration parameter error	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove the error;</li> <li>2. Switch off and then switch on the instrument power;</li> <li>3. If the error still exists after the restart, contact our customer service department.</li> </ol>
0x03001	Tube in wrong tube rack	Tube in wrong tube rack	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. Place the tube in the correct type of tube rack.</li> <li>3. If the error persists, contact our Customer Service Department.</li> </ol>
0x03000	Sample type detection sensor abnormal	Sample type detection sensor abnormal	<ol style="list-style-type: none"> <li>1. Click the "Remove Error" button to remove this error.</li> <li>2. If the error persists, contact our Customer Service Department.</li> </ol>
0x30202	Waste channel abnormal	HGB waste channel clogged	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>
0x30203	Waste channel abnormal	HGB waste channel clogged	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>
0x30204	Waste channel abnormal	HGB waste channel clogged	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>
0x30600	Waste channel abnormal	DIFF waste channel clogged	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>
0x30601	Waste channel abnormal	DIFF waste channel clogged	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>

Error ID	Error Message	Description	Solution
0x30602	Waste channel abnormal	DIFF waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30603	Waste channel abnormal	RET waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30604	Waste channel abnormal	RET waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30605	Waste channel abnormal	RET waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10108	Waste channel abnormal	Probe wipe waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10109	Waste channel abnormal	Probe wipe waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x10110	Waste channel abnormal	Probe wipe waste channel clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30304	Waste channel abnormal	Cleaning channel of RBC sample preparation clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30305	Waste channel abnormal	Cleaning channel of RBC sample preparation clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x30306	Waste channel abnormal	Cleaning channel of RBC sample preparation clogged	1.Tap the "Remove Error" button to remove this error; 2.If the error still exists, contact our customer service department.
0x07005	Probe wipe filter near service life	Probe wipe filter near service life	1. The probe wipe filter will reach the end of their service life after 1500 more tests. Customer Service to replace the component; 2.Tap the "Remove Error" button to remove this error; 3.If the error still exists, contact our customer service department.

Error ID	Error Message	Description	Solution
0x07006	Probe wipe filter reached service life	Probe wipe filter reached service life	<ol style="list-style-type: none"> <li>1. The probe wipe filter have reached the end of their service life. Customer Service to replace the component;</li> <li>2. Tap the "Remove Error" button to remove this error;</li> <li>3. If the error still exists, contact our customer service department.</li> </ol>
0x99806	Maintenance failed	Maintenance failed	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>
0x07003	Sample probe near service life	Sample probe near service life	<ol style="list-style-type: none"> <li>1. The sample probe will reach the end of its service life after 3000 more tests. Contact Mindray's Customer Service to replace the component;</li> <li>2. Tap the "Remove Error" button to remove this error;</li> <li>3. If the error still exists, contact our customer service department.</li> </ol>
0x07004	Sample probe reached service life	Sample probe reached service life	<ol style="list-style-type: none"> <li>1. The sample probe has reached the end of its service life. Contact Mindray's Customer Service to replace the component;</li> <li>2. Tap the "Remove Error" button to remove this error;</li> <li>3. If the error still exists, contact our customer service department.</li> </ol>
0x30405	ESR assembly version not match	ESR assembly version not match	Please contact Mindray Customer Service Department.
0x99807	Maintenance failed	Cleaning failed	<ol style="list-style-type: none"> <li>1. Tap the "Remove Error" button to remove this error;</li> <li>2. If the error still exists, contact our customer service department.</li> </ol>
0x10309	Probe cleanser aspirated volume not enough	Probe cleanser aspirated volume not enough	The Probe Cleanser volume is not enough for maintenance, which may cause frequent auto-cleaning when the instrument is in measurement. For the next time maintenance, make sure the Probe Cleanser volume is no less than 4mL.



# A Index

---

## A

Analyzer shutdown 7-21

## B

Blood Group 8-5  
Blood Sample Test Scattergrams  
    DIFF Scattergram 4-3, 8-9

## C

Calibrate  
    Calibrate with Calibrators 10-4  
    Calibrate with fresh blood samples 10-5  
    Calibration History 10-6  
    Calibration Programs 10-1  
    Manual calibration 10-2  
    Notes before Calibration 10-2  
    Verify calibration factors 10-6  
    When to Calibrate 10-1  
Customizing the Analyzer Software 6-1

## D

Daily Operation  
    Sample Preparation  
        Body Fluid Samples 7-8  
        Prediluted Samples 7-7  
        Whole Blood Samples 7-6  
Daily Operations  
    Preparations 7-4  
DIFF Scattergram 4-3  
DIFF scattergram 8-9

## E

ESR 8-5

## F

Flags  
    Abnormal Blood Cell Differential Or Morphology 8-9  
    Parameter Flags 8-9

## J

Installation Requirements 5-1  
    Environment Requirements 5-2  
    Power Requirements 5-2  
    Space Requirements 5-1

## M

Maintenance and Service  
    Probe Cleanser Maintenance to Parts and Components  
        12-8  
Maintenance and service  
    Maintenance of parts and components 12-1  
maintenance and service  
    daily probe cleanser maintenance 12-8  
    manual cleaning 12-2, 12-9  
    replace and prime reagent 12-7  
    replacing parts and components 12-3  
    timing and purpose 12-1

## P

Parameters 3-1  
    Blood Sample Test Histograms 3-6  
    Blood Sample Test Report Parameters 3-2  
    Blood Sample Test RUO Parameters 3-3  
    Blood Sample Test Scattergrams 3-6  
    Body Fluid Parameters 4-8  
    Body Fluid Sample Test Histograms 3-7  
    Body Fluid Sample Test Report Parameters 3-7  
    Body Fluid Sample Test RUO Parameters 3-7  
    Body Fluid Sample Test Scattergram 3-8  
    HGB 4-5  
    Microscopic Parameters  
        Define Microscopic Parameters 6-11  
        Print Microscopic Parameter Results 11-4  
    PLT-related Parameters 4-6  
    RBC-related Parameters 4-6  
    Reticulocyte Parameters 4-8  
    WBC-Related Parameters 4-3  
Prepare to Ship 12-12  
Print  
    Auto Print 11-2  
    Print Manual Calibration Factors 11-5  
    Print QC Result Report 11-4  
    Print Sample Test Result Report 11-2  
    Print Setup 11-1  
print 3-24  
Probe Cleanser Maintenance 12-8

## Q

QC  
    L-J CBC QC  
        entering causes of outliers 9-8  
    L-J QC  
        Delete QC Records 9-10  
        Exporting QC Data 9-12  
        Print QC Results 11-4  
        Review QC Results  
            Communicating QC Data 9-11  
            QC Graph 9-7  
            QC Table 9-9  
        Run Controls on the QC Count Screen 9-4  
        Run Controls on the Sample Count Screen 9-6  
        Set QC Files 9-2  
    X-B QC  
        Print QC Results 11-4, 11-5  
        Delete QC Records 9-16  
        Review QC Results  
            QC Graph 9-15, 9-16, 9-17  
            QC Table 9-15  
        Sample Validity 9-13  
        X-B QC Rules 9-14

## R

Reagent 3-24  
    Controls and Calibrators 3-25  
    LH lyse for hematology analysis 3-25

- Probe Cleanser 3-25
- Replace Reagent 12-3
- Reagent Connection
  - Connecting Lyses and Diluent 5-5

## S

- Sample Analysis
  - OV-BF Sample Analysis Procedure 7-15
  - OV-PD Sample Analysis Procedure 7-14
  - OV-WB Sample Analysis Procedure 7-12
- Sample Result Review
  - Calculate CV 8-7
  - Communicate Sample Records 8-6
  - Delete Sample Records 8-8
  - Edit Analysis Results 8-5
  - Export Sample Results 8-7
  - Graph Review 8-4
  - Review Special Information 8-6
  - Review Traceability Information 8-6
  - RUO Parameter Results 8-4
  - Validate Sample Results 8-8
- Screen Cal. 12-13
- Setting
  - Lab Info. 6-4
- Setup
  - Auto Standby 7-21
  - Communication
    - Transmission Mode 6-3
  - Date/Time 6-4
  - Flag Alarm Sensitivity 6-4
  - HGB Gains 6-13
  - Permissions 6-1
  - Save Changes 6-2
- System Principles
  - SF CUBE Cell Analysis Technology 4-5
  - Sheath Flow Impedance Method 4-5

## T

- Troubleshooting
  - Check Analyzer Status 13-1
  - Error List 13-2

## W

- WBC Measurement 4-1
- Working Principles
  - Hemoglobin Concentration Measurement 4-4
  - RBC/PLT Measurement 4-5

# B Specification

## B.1 Classification

According to the CE classification, the device belongs to Class B in vitro diagnostic medical device according to rule 6, annex VIII of REGULATION (EU) 2017/746.

## B.2 Reagent

The analyzer can be used with the following reagents, controls, and calibrators.

### NOTE

- For any questions related to reagents, controls, and calibrators, please consult your local distributor.

Table B-1 Reagents

Applicable Channel	BC-760[B]	BC-760[R]/BC-780[R]
HGB channel	LH Lyse	LH Lyse
DIFF channel	LD Lyse	LD Lyse
	FD Dye	FD Dye
RET channel	/	DR Diluent
	/	FR Dye
/	DS Diluent	DS Diluent
	Probe Cleanser	Probe Cleanser
ESR channel	ESR Solution Reagent	ESR Solution Reagent

Table B-2 Controls/calibrators for complete blood tests

Name	Model	Applicable Model
Hematology control	BR60	BC-760[B]/BC-760[R]/BC-780[R]
Hematology control	BC-6D	BC-760[B]/BC-760[R]/BC-780[R]
Hematology control	BC-RET	BC-760[R]/BC-780[R]
Hematology calibrator	SC-CAL PLUS	BC-760[B]/BC-760[R]/BC-780[R]
Hematology calibrator	SC-CAL RET	BC-760[R]/BC-780[R]

Table B-3 Controls for body fluid tests

Name	Model	Applicable Model
Hematology control	BC-BF	BC-760[B]/BC-760[R]/BC-780[R]

**Table B-4 Controls/calibrators for ESR tests**

Name	Model	Applicable Model
Hematology control	BC-6D	BC-760[B]/BC-760[R]/BC-780[R]
Hematology calibrator	SC-CAL PLUS	BC-760[B]/BC-760[R]/BC-780[R]

**Table B-5 Part number for reagent controls/calibrators**

Name	Model	Part Number
LH Lyse	M-6	105-012291-00 (4L*1)
		105-012292-00 (1L*4)
LD Lyse	M-6	105-012287-00 (4L*1)
		105-012288-00 (1L*4)
FD Dye	M-6	105-012297-00 (48mL*1)
		105-012298-00 (12mL*4)
*DR Diluent	M-6	105-012285-00 (4L*1)
		105-012286-00 (1L*4)
*FR Dye	M-6	105-012295-00 (48mL*1)
		105-012296-00 (12mL*4)
DS Diluent	/	105-012283-00 (20L)
		105-012284-00 (10L)
Probe Cleanser	/	105-002225-00 (50 mL*1)
		105-009432-00 (25mL*6)
Hematology control	BC-6D	105-002421-00 (High/4.5mL*6)
		105-002422-00 (Normal/4.5mL*6)
		105-002423-00 (Low/4.5mL*6)
		105-002424-00 (High/Normal/Low)*2/4.5mL*6)
		105-003219-00 (High/Normal/Low)*1/4.5mL*3)
Hematology control	BR60	105-009074-00 (4ml*6/H/M/L)
		105-009073-00 (4ml*3/H/M/L)
		105-009072-00 (4ml*6/L)
		105-009068-00 (4ml*6/M)
		105-009064-00 (4ml*6/H)

Name	Model	Part Number
*Hematology control	BC-RET	105-002429-00 (High 4.5mL*6)
		105-002430-00 (Normal 4.5mL*6)
		105-002431-00 (Low 4.5mL*6)
		105-002432-00 ((High/Normal/Low)*2/ 4.5mL*6)
		105-003221-00 ((High/Normal/Low)*1/4.5mL*3)
		105-004088-00 (High/4.5mL*1)
		105-004089-00 (Normal/4.5mL*1)
		105-004090-00 (Low/4.5mL*1)
C-reaction protein control	/	105-018801-00 (I/II level/1.5ml*2)
		105-018803-00 (I/II level/1.5ml*6)
+Serum amyloid A control	/	105-021028-00 (Level I/II 1.5ml*2)
		105-021029-00 (Level I/II 1.5ml*6)
*Hematology control	BC-BF	105-018564-00 (H/N/L level/3ml*6)
		105-018567-00 ((N level/3ml*6)
		105-018572-00 (H/N/L level/3ml*3)
		105-018573-00 (N level/3ml*1)
		105-018582-00 (L level/3ml*6)
		105-018586-00 (H level/3ml*1)
		105-018587-00 (L level/3ml*1)
		105-018588-00 (H level/3ml*6)
Hematology calibrator	SC-CAL PLUS	105-003223-00 (3mL*2)
		105-004091-00 (3mL*1)
ESR Solution Reagent	ESR	105-026688-00 (1L*1)
		105-026689-00 (1L*4)

## NOTE

- The items with \* only apply to the BC-760[R]/BC-780[R] model.

## B.3 Models

### NOTE

- For details of tube specifications, refer to **3.4 Supported Tubes, Tube Racks and Adapters**.
- For the details of tube information not specified in the manual, contact Mindray Customer Service Department or the local distributor.

## B.4 Parameters

**Table B-6 Blood sample test report parameters**

Group	Parameter name	Abbreviation	Applicable Model
WBC series	White blood cell count	WBC	General
	Basophil count	Bas#	General
	Basophil percentage	Bas%	General
	Neutrophil count	Neu#	General
	Neutrophil percentage	Neu%	General
	Eosinophil count	Eos#	General
	Eosinophil percentage	Eos%	General
	Lymphocyte count	Lym#	General
	Lymphocyte percentage	Lym%	General
	Monocyte count	Mon#	General
	Monocyte percentage	Mon%	General
	Immature granulocyte count	IMG#	General
	Immature granulocyte percentage	IMG%	General
RET group	Reticulocyte percentage	RET%	BC-760[R]/BC-780[R]
	Reticulocyte count	RET%	
	Reticulocyte hemoglobin expression	RHE	
	Immature reticulocyte fraction	IRF	
	Low fluorescent ratio	LFR	
	Middle fluorescent ratio	MFR	
	High fluorescent ratio	HFR	
RBC group	Red blood cell count	RBC	General
	Hemoglobin concentration	HGB	General
	Mean corpuscular volume	MCV	General
	Mean corpuscular hemoglobin	MCH	General
	Mean corpuscular hemoglobin concentration	MCHC	General
	Red blood cell distribution width coefficient of variation	RDW-CV	General
	Red blood cell distribution width standard deviation	RDW-SD	General
	Hematocrit	HCT	General
	Nucleated red blood cell count	NRBC#	General
	Nucleated red blood cell percentage	NRBC%	General

Group	Parameter name	Abbreviation	Applicable Model
Platelet group	Platelet count	PLT	General
	Mean platelet volume	MPV	General
	Platelet distribution width	PDW	General
	Plateletcrit	PCT	General
	Platelet-large cell ratio	P-LCR	General
	Platelet-large cell count	P-LCC	General
	Immature platelet fraction	IPF	BC-760[R]/BC-780[R] For BC-760[B], this parameter is optional. For more information, consult Mindray Customer Service Department
	Platelet count- impedance	PLT-I	General (find the parameter result on the "Other Para."- "Analysis Para.")
	Platelet count hybrid	PLT-H	BC-760[R]/BC-780[R] For BC-760[B], this parameter is optional. For more information, consult Mindray Customer Service Department
	Optical platelet count	PLT-O	BC-760[R]/BC-780[R]
ESR	Erythrocyte Sedimentation Rate	ESR	General

Table B-7 Body fluid sample test report parameters

Group	Parameter name	Abbreviation	Applicable Model
WBC group	White blood cell count-body fluid	WBC-BF	General
	Total nucleated cell count-body fluid	TC-BF#	General
	Mononuclear cell count	MN#	General
	Mononuclear cell percentage	MN%	General
	Polymorphonuclear cell count	PMN#	General
	Polymorphonuclear cell percentage	PMN%	General
RBC group	Red blood cell count-body fluid	RBC-BF	General

## B.5 Sampling Features

### B.5.1 Sample modes, test panel, and applicable model

**Table B-8 List of sample modes, test panel, and applicable models**

Sample Mode		Test Panel	Applicable Model
Whole blood sample	CT-WB	CBC,CD,Ret,CR,CDR; ESR,CBC+ESR,CD+ESR,CDR+ESR	BC-760[R] BC-780[R]
		CBC,CD; ESR,CBC+ESR,CD+ESR	BC-760[B]
	CT-Low WBC	CD/WBC-3X	BC-760[B]
		CD/WBC-3X, CDR/WBC-3X	BC-760[R] BC-780[R]
	CT-Low PLT	CR/PLT-5X,CDR/PLT-5X	BC-760[R] BC-780[R]
CT-PD		CBC,CD,Ret,CR,CDR	BC-760[R] BC-780[R]
		CBC,CD	BC-760[B]
CT-BF		CD	General
Whole blood samples	AL-WB	CBC,CD,Ret,CR,CDR; ESR,CBC+ESR,CD+ESR,CDR+ESR	BC-760[R] BC-780[R]
		CBC,CD; ESR,CBC+ESR,CD+ESR	BC-760[B]

## B.5.2 Sample Volumes Required for Each Analysis

Sample Mode	Test Panel	Sample Volume (μl) Required for Each Analysis	Applicable Model
Whole blood	CBC, CD	25±2	BC-760[B]/BC-760[R]/BC-780[R]
	Ret, CR, CDR	33	BC-760[R]/BC-780[R]
	ESR	140	BC-760[B]/BC-760[R]/BC-780[R]
	CBC+ESR, CD+ESR	160	BC-760[B]/BC-760[R]/BC-780[R]
	CDR+ESR	160	BC-760[R]/BC-780[R]
Low WBC	CD/WBC-3X	37	BC-760[B]/BC-760[R]/BC-780[R]
	CDR/WBC-3X	42	BC-760[R]/BC-780[R]
Low PLT	CR/PLT-5X,CDR/PLT-5X	33	BC-760[R]/BC-780[R]
Predilute	CBC,CD	20	BC-760[B]/BC-760[R]/BC-780[R]
	Ret, CR, CDR	20	BC-760[R]/BC-780[R]
Body fluid	CD	85	BC-760[B]/BC-760[R]/BC-780[R]

## B.5.3 Throughput



Table B-9 Throughput

Sample Mode	Test Panel	Throughput (test/hour)	Applicable Model
Whole blood	CBC, CD	At least 80	BC-760[B]/BC-760[R]/BC-780[R]
	ESR	At least 50	BC-760[B]/BC-760[R]/BC-780[R]
	Ret, CR, CDR	At least 45	BC-760[R]/BC-780[R]
	CBC+ESR, CD+ESR	At least 40	BC-760[B]/BC-760[R]/BC-780[R]
	CDR+ESR	At least 30	BC-760[R]/BC-780[R]
Low WBC	CD/WBC-3X	At least 60	BC-760[B]/BC-760[R]/BC-780[R]
	CDR/WBC-3X	At least 40	BC-760[R]/BC-780[R]
Low PLT	CR/PLT-5X, CDR/PLT-5X	At least 35	BC-760[R]/BC-780[R]
Predilute	CBC, CD	At least 50	BC-760[B]/BC-760[R]/BC-780[R]
	Ret, CR, CDR	At least 33	BC-760[R]/BC-780[R]
Body fluid	CD	At least 50	BC-760[B]/BC-760[R]/BC-780[R]

## B.6 Performance Specifications

### B.6.1 Background/Blank Count Requirements

Table B-10 Background/blank count requirements for blood samples

Parameters	Acceptable Range
WBC	$\leq 0.10 \times 10^9 / L$
RBC	$\leq 0.02 \times 10^{12} / L$
RBC-O	$\leq 0.02 \times 10^{12} / L$
HGB	$\leq 1 \text{ g/L}$
PLT	$\leq 3 \times 10^9 / L$

Table B-11 Background/blank count requirements for body fluid samples

Parameters	Acceptable Range
WBC-BF/ TC-BF#	$\leq 0.001 \times 10^9 / L$
RBC-BF	$\leq 0.003 \times 10^{12} / L$

## B.6.2 Linearity Ranges

**Table B-12 Linearity requirements for blood samples**

Parameters	Linearity Range	Acceptable Deviation Range (WB)	Acceptable Deviation Range (PD)	Correlation Coefficient
WBC	(0 ~ 100.00)×10 <sup>9</sup> /L	±0.20×10 <sup>9</sup> /L±2%	±0.50×10 <sup>9</sup> /L±5%	≥ 0.990
	(100.01 ~ 350.00)×10 <sup>9</sup> /L	±6%	±6%	≥ 0.990
	(350.01 ~ 500.00)×10 <sup>9</sup> /L	±11%	±11%	≥ 0.990
RBC	(0 ~ 8.60)×10 <sup>12</sup> /L	±0.03×10 <sup>12</sup> /L±2%	±0.05×10 <sup>12</sup> /L ±5%	≥ 0.990
HGB	(0 ~ 260) g/L	±2g/L±2%	±2g/L or ±3%	≥ 0.990
HCT	(0.0 ~ 75.0)%	±1.0% (HCT) or ±2% (percentage error)	±2.0% (HCT) or ±4% (percentage error)	/
PLT	(0 ~ 1000)×10 <sup>9</sup> /L	±10×10 <sup>9</sup> /L±5%	±10×10 <sup>9</sup> /L±10%	≥ 0.990
	(1001 ~ 5000)×10 <sup>9</sup> /L	±6%	±10%	≥ 0.990
RET%	(0.00 ~ 30.00)%	±0.30% (RET% value) or ±20% (percentage error)	/	/
RET%	(0.0000~0.8000)×10 <sup>12</sup> /L	±0.0150×10 <sup>12</sup> /L±20%	/	/

**Table B-13 Linearity requirements for body fluid samples**

Parameters	Linearity Range	Deviation range
WBC-BF/TC-BF#	(0~0.050)×10 <sup>9</sup> /L	±0.010×10 <sup>9</sup> /L
	(0.051~1.000)×10 <sup>9</sup> /L	±20%
	(1.001~10.000)×10 <sup>9</sup> /L	±20%
RBC-BF	(0.000~0.100)×10 <sup>12</sup> /L	±0.010×10 <sup>12</sup> /L or ±5%
	(0.101~5.000)×10 <sup>12</sup> /L	±0.030×10 <sup>12</sup> /L or ±2%

## B.6.3 Accuracy

### B.6.3.1 Blood parameter accuracy

**Table B-14 Accuracy requirements**

Parameters	Measurement Range	Acceptable Relative Deviation Range/%
WBC	3.50×10 <sup>9</sup> /L~9.50×10 <sup>9</sup> /L	Within ±10.0
RBC	3.80×10 <sup>12</sup> /L~5.80×10 <sup>12</sup> /L	Within ±6.0
HGB	115g/L~175g/L	Within ±6.0
PLT	125×10 <sup>9</sup> /L~350×10 <sup>9</sup> /L	Within ±20.0
HCT/MCV	35%~50% (HCT) or 82.0f~100.0fL (MCV)	Within ±9.0 (HCT) or ±7.0 (MCV)

## B.6.4 Repeatability

**Table B-15 Repeatability requirements for blood samples**

Parameters	Range	Whole Blood (CV/Absolute Deviation d*/SD)	Predilute (CV/Absolute Deviation d*)
WBC	$(3.50 \sim 4.50) \times 10^9/L$	$\leq 3.0\%$	$\leq 4.0\%$
	$\geq 4.51 \times 10^9/L$	$\leq 2.5\%$	$\leq 3.5\%$
RBC	$\geq 3.50 \times 10^{12}/L$	$\leq 1.5\%$	$\leq 2.0\%$
HGB	$(110 \sim 180)g/L$	$\leq 1.0\%$	$\leq 2.0\%$
MCV	$(80.0 \sim 100.0) fL$	$\leq 1.0\%$	$\leq 3.0\%$
HCT	$(30.0 \sim 50.0)\%$	$\leq 1.5\%$	$\leq 3.0\%$
MCH	/	$\leq 1.5\%$	/
MCHC	/	$\leq 1.5\%$	/
RDW-SD	/	$\leq 2.0\%$	/
RDW-CV	/	$\leq 2.0\%$	/
PLT	$\geq 100 \times 10^9/L$	$\leq 4.0\%$	$\leq 8.0\%$
		$\leq 2.5\%$ Applicable to the CDR/PLT-5X and CR/PLT-5X test panels	/
	$(20 \sim 100) \times 10^9/L$	$\leq 5.0\%$ Applicable to the CDR/PLT-5X and CR/PLT-5X test panels	/
	$\leq 20 \times 10^9/L$	$\leq 1.5(SD)$ Applicable to the CDR/PLT-5X and CR/PLT-5X test panels	/
PDW	$PLT \geq 100 \times 10^9/L$	$\leq 10.0\%$	/
MPV	$PLT \geq 100 \times 10^9/L$	$\leq 3.0\%$	/
P-LCR	$PLT \geq 100 \times 10^9/L$	$\leq 15.0\%$	/
P-LCC	$PLT \geq 100 \times 10^9/L$	$\leq 15.0\%$	/
PCT	$PLT \geq 100 \times 10^9/L$	$\leq 5.0\%$	/
Neu%	Neu% $\geq 30.0\%$ $WBC \geq 4.00 \times 10^9/L$	$\leq 6.0\%$	$\leq 12.0\%$
Lym%	Lym % $\geq 15.0\%$ $WBC \geq 4.00 \times 10^9/L$	$\leq 6.0\%$	$\leq 12.0\%$
Mon%	Mon % $\geq 5.0\%$ $WBC \geq 4.00 \times 10^9/L$	$\leq 16.0\%$	$\leq 32.0\%$
Eos%	$WBC \geq 4.00 \times 10^9/L$	$\leq 20.0\%$ or $\pm 1.5\%(d)$	$\leq 40.0\%$ or $\pm 3.0\%(d)$
Bas%	$WBC \geq 4.00 \times 10^9/L$	$\leq 30.0\%$ or $\pm 1.0\%(d)$	$\leq 60.0\%$ or $\pm 2.0\%(d)$
Neu#	$\geq 1.20 \times 10^9/L$	$\leq 6.0\%$	$\leq 12.0\%$
Lym#	$\geq 0.60 \times 10^9/L$	$\leq 6.0\%$	$\leq 12.0\%$
Mon#	$\geq 0.20 \times 10^9/L$	$\leq 16.0\%$	$\leq 32.0\%$
Eos#	$WBC \geq 4.00 \times 10^9/L$	$\leq 20.0\%$ or $\pm 0.12 \times 10^9/L(d)$	$\leq 40.0\%$ or $\pm 0.24 \times 10^9/L(d)$
Bas#	$WBC \geq 4.00 \times 10^9/L$	$\leq 30.0\%$ or $\pm 0.06 \times 10^9/L(d)$	$\leq 60.0\%$ or $\pm 0.12 \times 10^9/L(d)$

Parameters	Range	Whole Blood (CV/Absolute Deviation d*/SD)	Predilute (CV/Absolute Deviation d*)
IMG%	WBC $\geq 4.00 \times 10^9/L$ IMG% $\geq 2.0\%$	$\leq 25.0\%$ or $\pm 1.5\%(d)$	/
IMG#	$\geq 0.10 \times 10^9/L$	$\leq 25.0\%$ or $\pm 0.12 \times 10^9/L(d)$	/
RET%	RBC $\geq 3.00 \times 10^{12}/L$ RET%: 1.00% ~ 4.00%	$\leq 15\%$	$\leq 30\%$
RET%	RBC $\geq 3.00 \times 10^{12}/L$ RET%: 1.00% ~ 4.00%	$\leq 15\%$	$\leq 30\%$
RHE	RET# $\geq 0.0200 \times 10^{12}/L$	$\leq 5\%$	/
LFR	RBC $\geq 3.00 \times 10^{12}/L$ RET%: 1.00% ~ 4.00% LFR $\geq 20\%$	$\leq 30\%$	/
MFR	RBC $\geq 3.00 \times 10^{12}/L$ RET%: 1.00% ~ 4.00% MFR $\geq 20\%$	$\leq 50\%$	/
HFR	RBC $\geq 3.00 \times 10^{12}/L$ RET%: 1.00% ~ 4.00%	$\leq 100\%$ or $\pm 2.0\%(d)$	/
IRF	RBC $\geq 3.00 \times 10^{12}/L$ RET%: 1.00% ~ 4.00% IRF $\geq 20\%$	$\leq 30\%$	/
IPF	PLT $\geq 50 \times 10^9/L$ IPF $\geq 3.0\%$	$\leq 25\%$	/
ESR	0 ~ 20mm/h	$\leq 1.8(SD)$	/
	> 20mm/h	$\leq 9\%$	/

Note:

1. \*Absolute deviation d = MAX|Measured value - measured mean|
2. \*\*Range = Maximum measurement value – Minimum measurement value

**Table B-16 Repeatability Requirements for Body Fluid Samples**

Parameters	Measurement Range	CV or Range
WBC-BF/TC-BF#	$(0.015-0.100) \times 10^9/L$	$\leq 30\%$
RBC-BF	$(0.003-0.050) \times 10^{12}/L$	$\leq 40\%$ or range $\leq 7000/\mu L$

## B.6.5 Carryover

**Table B-17 Carryover requirements for blood samples**

Parameters	Carryover
WBC	$\leq 1.0\%$
RBC	$\leq 1.0\%$
HGB	$\leq 1.0\%$
HCT	$\leq 1.0\%$
PLT	$\leq 1.0\%$

Parameters	Carryover
ESR	≤1.0%

**Table B-18 Carryover requirements for body fluid samples**

Parameters	Carryover
WBC-BF / TC-BF#	≤0.3% or $\leq 0.001 \times 10^9/L$
RBC-BF	≤0.3% or $\leq 0.003 \times 10^{12}/L$

### B.6.6 Correlation

Correlation with reference instrument must meet the requirements of *Table B-19 Correlation requirements for blood sample test*.

**Table B-19 Correlation requirements for blood sample test**

Parameters	Correlation with Reference Instrument
WBC	≥ 0.99
Neu%	≥ 0.90
Lym%	≥ 0.90
Mon%	≥ 0.80
Eos%	≥ 0.90
Bas%	≥ 0.60
RBC	≥ 0.99
HGB	≥ 0.98
MCV	≥ 0.98
HCT	≥ 0.95
RDW-CV	≥ 0.85
RDW-SD	≥ 0.85
PLT	≥ 0.95
MPV	≥ 0.80
RET%	≥ 0.90
RET#	≥ 0.90
RHE	≥ 0.90
IPF	≥ 0.80
IMG%	≥ 0.80
NRBC%	≥ 0.80
ESR	≥ 0.90

## B.7 Requirements for Input/Output Devices

The analyzer can be connected to a USB flash drive, keyboard, mouse, and barcode scanner as required.

## WARNING

- Be sure to use the specified devices only.
- External equipment connected to the analyzer and digital interfaces must be authorized and complied with relevant safety and EMC standards (e.g. IEC 60950 Safety of Information Technology Equipment Standard and CISPR 22 EMC of Information Technology Equipment Standard (Class B)). Any persons who connects additional equipment to the signal input or output ports and configures an IVD system, is responsible for ensuring that the system works normally and complies within the safety and EMC requirements. If you have any questions, consult the technical service department of your local representative. .

### B.7.1 Keyboard

USB port (supporting the protocol of USB2.0 and above) keyboard.

### B.7.2 Mouse

USB port (supporting the protocol of USB2.0 and above) mouse.

### B.7.3 External barcode scanner

USB port (supporting the protocol of USB2.0 and above) hand-held barcode scanner.

### B.7.4 Printer

USB port (supporting the protocol of USB2.0 and above) printer.

### B.7.5 USB Drive

Supporting the protocol of USB2.0 and above.

### B.7.6 Electronic Interface Specifications

#### NOTE

- Connection of the electronic interface to an IT network that includes other equipment could result in previously unidentified risks to patients, users or third parties; the responsible organization should identify, analyse, evaluate and control these risks.

	Specifications	
Communication format (protocol) and relevant standards	USB ports	Type A interface, complied with USB 2.0/3.0 standard.
	Network ports	TCP/IP protocol bottom layer. DICOM/HL7 protocol application layer. RJ45 interface, supporting wired network 10 M/100 M/1000 M, and complied with technical standard IEEE802.3. NTP/SNTP Calibration protocol of TCP/IP.
Time setting	The time on the analyzer should be set to the correct local time.	

## B.8 Interfaces

### NOTE

- The USB interfaces on the back of the analyzer shall only be used to connect the peripheral devices specified in this manual. See Appendix B.7 *Requirements for Input/Output Devices* for details about supported devices and models.
- 
- One network port (compatible with 10/100/1000M Ethernet and complying with the 802.3u/802.3ab standard)
  - Four USB ports including three supporting USB2.0 and one supporting USB3.0 (specification: DC 5V; 500 mA)

## B.9 Power supply

	Voltage	Input power	Frequency
Main unit	100V-240V~ (±10%)	600 VA	50 Hz/60 Hz (±1 Hz)

## B.10 Fuse

### WARNING

- The fuse used in the instrument is not a replaceable one. If there is any problem with the fuse, contact Mindray Customer Service Department or your local distributor.

## B.11 Electromagnetic compatibility (EMC)

The IVD device complies with the EMC standard IEC 61326-1 and IEC 61326-2-6.

For EMISSIONS and IMMUNITY specific requirements, see *Table B-20 GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC EMISSIONS* and *Table B-21 GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC IMMUNITY*.

### WARNING

- The IVD MEDICAL EQUIPMENT complies with the emission and immunity requirements described in this part of IEC 61326.
- This equipment is not intended for use in residential environments and may not provide adequate protection to radio reception in such environments.
- This equipment is designed for use in a PROFESSIONAL HEALTHCARE FACILITY ENVIRONMENT. It is likely to perform incorrectly if used in a HOME HEALTHCARE ENVIRONMENT. If it is suspected that performance is affected by electromagnetic interference, correct operation may be restored by increasing the distance between the equipment and the source of the interference.
- The electromagnetic environment should be evaluated prior to operation of the device.
- Do not use this device in proximity to sources of strong electromagnetic radiation (e.g. unshielded intentional RF sources), as these can interfere with proper operation.

### NOTE

- It is the manufacturer's responsibility to provide equipment electromagnetic compatibility information to the customer or user.
- It is the user's responsibility to ensure that a compatible electromagnetic environment for the equipment can be maintained in order that the equipment can be maintained in order that the device will perform as intended.

- The calculation formula to determine the separation distance between an IVD MEDICAL EQUIPMENT and a mobile phone is given by  $d = 6/E \cdot \sqrt{P}$ , where  $d$  is the minimum separation distance in meters,  $P$  is the maximum power in watts, and  $E$  is the immunity test level in V/m.

**Table B-20 GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC EMISSIONS**

GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC EMISSIONS	
The system is intended for use in the electromagnetic environment specified below. The customer or the user of system should assure that it is used in such an environment.	
EMISSIONS TEST	COMPLIANCE
RF emissions CISPR 11	Group 1
RF emissions CISPR 11	Class A
Harmonic Emissions IEC 61000-3-2	N/A
Voltage Fluctuations/ Flicker Emissions IEC 61000-3-3	

**Table B-21 GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC IMMUNITY**

GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC IMMUNITY			
The system is intended for use in the electromagnetic environment specified below. The customer or the user of system should assure that it is used in such an environment.			
IMMUNITY TEST	BASIC STANDARD	TEST VALUE	PERFORMANCE CRITERION
Electrostatic Discharge (ESD)	IEC 61000-4-2	± 4 kV contact ± 2 kV, ± 4 kV, ± 8 kV air	B B
Electromagnetic field	IEC 61000-4-3	3 V/m (80 MHz to 6 GHz)	A
Electrical fast Transient / burst	IEC 61000-4-4	AC Power: ± 1 kV I/O single/Control <sup>1</sup> : ± 0.5 kV	B
Surge	IEC 61000-4-5	line-to-line: ± 0.5kV, 1kV, ± 2kV line-to-ground: ± 0.5kV, 1 kV	B B
Conducted RF	IEC 61000-4-6	3 V (150 kHz to 80 MHz)	A
Voltage dips, Short interruptions and voltage variation on power supply input voltage	IEC 61000-4-11	0 % during 0,5 cycles 0 % during 1 cycle 70 % during 25/30 cycles <sup>b</sup> 0 % during 250/300 cycles	B B C C
Power frequency magnetic field	IEC 61000-4-8	3 A/m (50 Hz, 60 Hz)	A



**GUIDANCE AND MINDRAY DECLARATION—ELECTROMAGNETIC IMMUNITY****NOTE:**

<sup>a</sup> Only in case of line >3m

<sup>b</sup> "25/30 cycles" means "25 cycles for 50 Hz test" or "30 cycles for 60 Hz test".

**Performance criterion:**

A: The equipment shall continue to operate as intended during and after the test.

B: The equipment shall continue to operate as intended after the test.

C: LOSS OF FUNCTION is allowed, provided the function is self-recoverable or can be restored by the operation of the controls.

**B.12 Noise Level**

Maximal sound pressure: 80 dBA

**NOTE**

- Be sure to use and store the analyzer under the specified environment conditions.

**B.13 Normal Operating Environment**

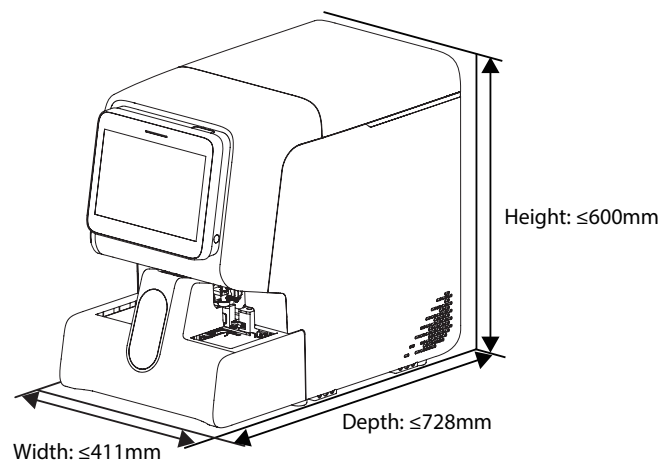
- Normal operating temperature range: 10°C to 35°C
- Normal operating humidity range: 30% to 85%
- Normal operating atmospheric pressure range: 70.0 kPa to 106.0 kPa

**B.14 Storage Environment**

- Ambient temperature range: -10°C to 40°C
- Relative humidity range: 10% to 90%
- Atmospheric pressure range: 50.0 kPa to 106.0 kPa

**B.15 Operating Environment**

- Ambient temperature range: 5°C to 40°C
- Relative humidity range: 10% to 90%
- Atmospheric pressure range: 70.0 kPa to 106.0 kPa

**B.16 Dimensions and Weight**

**Figure B-1 Main unit dimensions**

Main Unit Dimensions and Weight	Value
Width (autoloader included)	≤411mm
Height (autoloader included)	≤600mm
Depth (autoloader included)	≤728mm
Weight (autoloader included)	≤73kg

## B.17 Contraindication

None

## B.18 Barcode Specifications

The analyzer can read barcodes that are stuck to the test tubes. The sample ID read from the barcode will be stored and used as the only identification of the sample.

The barcodes used shall meet the specifications stated in this section.

### 1. Supported barcode types

All code types and check digit supported by the analyzer are listed as follows.

**Table B-22 Supported barcode types**

Code Type	Check Digit	Number of Digits
CODE128	Self-checking (check digit is always included)	No more than 20 digits (sample ID)
CODE93	Self-checking (check digit is always included)	No more than 20 digits (sample ID)
UPC/EAN/JAN	Self-checking (check digit is always included)	Fixed length: 8 or 13 digits
ITF	Not use check digit	No more than 20 digits (sample ID)
	Use check digit	No more than 19 digits (sample ID)+1 digit (check digit) = no more than 20 digits
CODE39	Not use check digit	No more than 20 digits (sample ID)
	Use check digit	No more than 19 digits (sample ID)+1 digit (check digit) = no more than 20 digits
CODABAR	Not use check digit	No more than 20 digits (sample ID)
	Use check digit	No more than 19 digits (sample ID)+1 digit (check digit) = no more than 20 digits

### NOTE

- Use check digit whenever possible, so as to reduce the possibility of misreading.

## 2. Barcode label dimensions

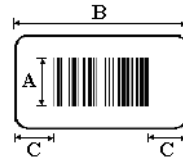
Barcode height:  $A \geq 10\text{mm}$

Label width:  $B \leq 50\text{mm}$

Clear area width:  $C \geq 5\text{mm}$

Width of the narrowest bar: above 0.152 mm

Code quality: According to ANSI MH10.8M standard, the code quality shall be Level C or above.



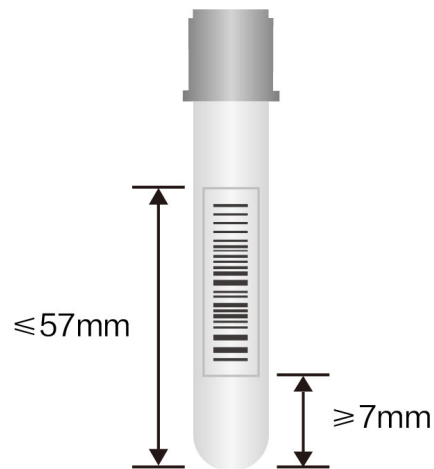
## 3. Samples of valid and invalid barcode labels:

Use of invalid barcode labels shown in the following figures will increase the possibility of misreading. To ensure good readability, use valid labels.

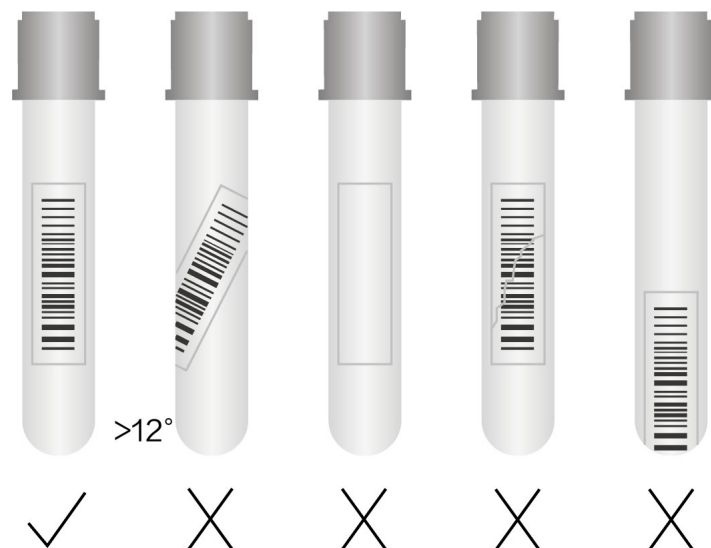


#### 4. Where to place the barcode label

You must place the label on the region as shown in *Figure B-2 Where to place the barcode label* in accordance with instructions as shown in *Figure B-3 How to place the barcode label*.



**Figure B-2 Where to place the barcode label**



**Figure B-3 How to place the barcode label**

#### 5. How to select the barcode label

It is recommended to use the matte label paper instead of the glossy label paper to print labels.

## B.19 Safety Classification

Level of transient overvoltage: Category II.

Rated pollution degree: 2.

## B.20 Limitations

### B.20.1 For Blood Routine Tests

**Table B-23 Blood Routine Tests**

Parameters	Interfering substances
WBC	<p>Where the following is present, the WBC may be reported falsely low:</p> <ul style="list-style-type: none"> <li>• WBC agglutination</li> </ul> <p>Where the following is present, the WBC may be reported falsely high:</p> <ul style="list-style-type: none"> <li>• Cryoglobulin</li> <li>• Fibrin</li> <li>• Lyse resistant RBC</li> <li>• Giant platelet</li> <li>• Platelet clumps</li> </ul>
RBC	<p>Where the following is present, the RBC may be reported falsely low:</p> <ul style="list-style-type: none"> <li>• RBC cold agglutination</li> <li>• Microcytosis</li> <li>• RBC fragment</li> </ul> <p>Where the following is present, the RBC may be reported falsely high:</p> <ul style="list-style-type: none"> <li>• Giant platelet</li> </ul>
HGB	<p>Where the following is present, the HGB result may be reported falsely low:</p> <ul style="list-style-type: none"> <li>• Lyse resistant RBC</li> <li>• RBC cold agglutination</li> </ul> <p>Where the following is present, the HGB result may be reported falsely high:</p> <p>Leukocytosis (&gt; 100000/<math>\mu</math>L)</p> <ul style="list-style-type: none"> <li>• Hyperlipidemia</li> </ul>
MCV	<p>Where the following is present, the MCV result may be reported falsely low:</p> <ul style="list-style-type: none"> <li>• Giant platelet</li> </ul> <p>Where the following is present, the MCV result may be reported falsely high:</p> <ul style="list-style-type: none"> <li>• RBC fragments</li> <li>• Microcytosis</li> <li>• RBC agglutination</li> <li>• Spherocyte</li> </ul>
PLT	<p>Where the following is present, the PLT count may be reported falsely low:</p> <ul style="list-style-type: none"> <li>• Platelet clump</li> <li>• Giant platelet</li> </ul> <p>Where the following is present, the PLT count may be reported falsely high:</p> <ul style="list-style-type: none"> <li>• Microcytosis</li> <li>• RBC fragments</li> <li>• WBC fragment</li> <li>• Cryoglobulin</li> </ul>
RET# (only available on BC-760 [R] and BC-780 [R])	<p>Where the following is present, the RET count may be reported falsely high:</p> <ul style="list-style-type: none"> <li>• Howell-Jolly body</li> <li>• Giant Platelet</li> <li>• RBC agglutination</li> <li>• Malaria</li> </ul>
NRBC#	<p>Where the following is present, the NRBC count may be reported falsely high:</p> <ul style="list-style-type: none"> <li>• Lyse resistant RBC</li> <li>• Malaria</li> </ul>

## B.20.2 ESR Tests

Erythrocyte sedimentation rate (ESR) is not only a non-specific inflammation index, but also one of indicators of erythrocyte aggregation. The analyzer measures the erythrocyte sedimentation rate by detecting the signals from erythrocyte aggregation process. However, the analyzer has limitations when analyzing the following samples:

1. Samples from patients with plasma cell diseases (e.g., multiple myeloma). In these samples, the erythrocytes may have already accumulated to rouleaux formation; therefore the analyzer may not be able to detect the process of erythrocyte aggregation, and may produce wrong results. The analyzer will give an alarm and may shield the results. To ensure accurate ESR results of these samples, it is recommended to use the traditional Westphal method to measure such samples.
2. Samples in which red blood cell agglutination has occurred (e.g., patient's blood containing cold agglutinin, and red blood cells aggregates in vitro cold environment). For such samples, the analyzer may not be able to detect the process of erythrocyte aggregation, and may produce wrong results. The analyzer will give an alarm and may shield the results. To ensure accurate ESR results of these samples, it is recommended to use the traditional Westphal method to measure such samples.
3. Other samples that may have rouleaux formation, blood coagulation, and abnormal erythrocyte morphology. The analyzer may give wrong results. Doctors should report based on the comprehensive judgment taking in consideration of analyzer-provided flags and prompts, as well as other clinical information of the patients.

# C Accessories and Packing List

---

## C.1 Accessories of the Analyzer

Name	Part Number
Handheld Barcode Scanner	023-000866-00
HP LaserJet Printer	023-001523-00
Tube Rack Assembly (CAL 8000)	115-070616-00
Capillary blood tube rack assembly	115-072433-00

### NOTE

- **The accessories actually attached to the product depend on your product configuration. For details about the configured/optional accessories, consult your sales representative.**

## C.2 Optional Accessories of the Analyzer

- Handheld barcode scanner
- Lyse cap assembly (black, die sinking connector)
- Waste container cap assembly
- Capillary blood tube rack assembly
- Tube rack assembly

## C.3 Packing List

- Main power cords
- Network cable
- Display
- Handheld barcode scanner (optional)
- Computer
- Diluent container supporting board
- Lyse cap assembly (green, die sinking connector)
- Lyse cap assembly (red, die sinking connector)
- Lyse cap assembly (black, die sinking connector) (optional)
- Lyse cap assembly (purple, die sinking connector)
- DS diluent cap assembly
- Waste container cap assembly (Optional)
- Operator's Manual
- Main Unit (Analyzer)
- Autoloader that may accommodate 6 rows of 5-position tube racks
- Capillary blood tube rack (optional)
- Tube rack assembly (optional)
- Handle bar
- Laboratory Data Management Software

**This page intentionally left blank.**



# D

## Communication

---

The LIS/HIS function of this analyzer enables the communication between the analyzer and the PC in laboratory through Ethernet, including sending analysis results to and receiving worklist from PC.

In the LIS/HIS communication process of the analyzer involves the HL7 communication protocol. For details about the connection control, and the introduction, message definition and examples, please contact Mindray Customer Service Department or your local distributor.

**This page intentionally left blank.**

# E

## Radio Regulatory Compliance

---

### RF Parameters

Radio Devices	RF ID
Operating Frequency	13.56MHz
Modulation mode	ASK
RF output power/Transmitter H-field	≤60 dBuA/m@10m



The radio device used in this product as well as our product is in compliance with the essential requirements and other relevant provisions of Directive 2014/53/EU.

## Declaration of Conformity V 2.0



## Declaration of Conformity

**Manufacturer:** Shenzhen Mindray Bio-Medical Electronics Co., Ltd.  
Mindray Building, Keji 12th Road South, High-tech Industrial  
Park, Nanshan, Shenzhen, 518057, P. R. China

**EC-Representative:** Shanghai International Holding Corp. GmbH (Europe)  
Eiffestraße 80  
20537 Hamburg, Germany

**Product:** Auto Hematology Analyzer

**Model:** BC-760[B], BC-760[R], BC-780[R], BC-700[B] 、  
BC-700[R]、BC-720[R]

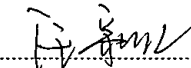
We herewith declare under our sole responsibility that the above mentioned products meet the provisions of the Council Directive 2014/53/EU concerning radio equipment. All supporting documentation is retained under the premises of the manufacturer.

## Standards Applied:

<input checked="" type="checkbox"/> EN 61326-1: 2020	<input checked="" type="checkbox"/> EN 50364: 2018
<input checked="" type="checkbox"/> ETSI EN 301 489-1 V2.2.3	<input checked="" type="checkbox"/> ETSI EN 301 489-3 V2.1.1
<input checked="" type="checkbox"/> ETSI EN 300 330 V2.1.1	

**Start of CE-Marking:** 2022-4-19

**Place, Date of Issue:** Shenzhen, 2022-12-20

**Signature:** 

**Name of Authorized Signatory:** Mr. Wang Xinbing

**Position Held in Company:** Deputy Director, Technical Regulation Department

# F

## References

---

1. CLSI. Interference Testing in Clinical Chemistry; Approved Guideline; Second Edition. CLSI document EP7-A2. Clinical and Laboratory Standards Institute; 2005.
2. Levey S, Jennings ER. The use of control charts in the clinical laboratory. *Am J Clin Pathol*. 1950;20: 1059-1066
3. Westgard, J.O., P.L. Barry, and M.R. Hunt (1981). "A Multi-rule Shewhart Chart for Quality Control in Clinical Chemistry," *Clinical Chemistry*, vol. 27, pp. 493-501.
4. Westgard, J.O., P.L. Barry (1986). "Cost-Effective Quality Control: Managing the Quality and Productivity of Analytical Processes" AACC Press.
5. Bull BS. A statistical approach to quality control. *Quality Control in Hematology*, Symposium of the International Committee for Standardization in Haematology. Lewis SM and Coster JF, eds, Academic Press, London, England, 1975.
6. International Committee for Standardization in Haematology. Lewis SM and Coster JF, eds, Academic Press, London, England, 1975.
7. Bull BS. A study of various estimations for the derivation of quality control procedures from patient erythrocyte indexes [J]. *Am J Clin Pathol* 1974.61(4):473-481
8. M.W. Rampling, G. Martin. A comparison of the Myrenne erythrocyte aggregometer with older techniques for estimating erythrocyte aggregation, *Biorheology* 9 (1989), 41–46.
9. M. Plebani, S. De Toni, M.C. Sanzari , et al., The TEST 1 automated system – A new method for measuring the erythrocyte sedimentation rate, *Am J Clin Pathol* 110 (1998), 334–340.
10. Bull BS. A study of various estimations for the derivation of quality control procedures from patient erythrocyte indexes [J]. *Am J Clin Pathol* 1974.61(4):473-481.

**This page intentionally left blank.**

# G Maintenance Logs

---

## NOTE

- You are advised to prepare a maintenance checklist suitable for the operating environment of the analyzer.
- For more information about the maintenance procedure, refer to *12Service*.

**This page intentionally left blank.**



**Routine maintenance items**

Date	Probe Cleanser Maintenance	Cleaning the Analyzer Front Cover	Probe wipe cleaning	Date	Probe Cleanser Maintenance	Cleaning the Analyzer Front Cover	Probe wipe cleaning
1				17			
2				18			
3				19			
4				20			
5				21			
6				22			
7				23			
8				24			
9				25			
10				26			
11				27			
12				28			
13				29			
14				30			
15				31			
16							





