

Technical Data

Moeller Decarboxylase Broth Base

M393

(Decarboxylase Broth Base, Moeller)

Moeller Decarboxylase Broth Base with the addition of appropriate L-amino acid is used to differentiate bacteria on the basis of their ability to decarboxylate the amino acids.

Composition**

Ingredients	Gms / Litre
Peptic digest of animal tissue	5.000
Beef extract	5.000
Dextrose	0.500
Bromocresol purple	0.010
Cresol red	0.005
Pyridoxal	0.005
Final pH (at 25°C)	6.0 ± 0.2

^{**}Formula adjusted, standardized to suit performance parameters

Directions

Suspend 10.52 grams in 1000 ml distilled water. Add 10 gm. of L-Lysine, L-Arginine, L-Ornithine or other L-amino acids. When using DL-amino acids, use 2% concentration. Heat if necessary to dissolve the medium completely. When L-Ornithine is added, readjustment of the pH is required. Dispense in 5 ml amount in screw-capped tubes and sterilize by autoclaving at 15 lbs pressure (121°C) for 10 minutes.

Principle And Interpretation

Moeller Decarboxylase Broth Base is used for differentiating gram-negative enteric bacilli on the basis of their ability to decarboxylate amino acids. Moeller introduced the Decarboxylase Broth for detecting the production of lysine and ornithine decarboxylase and arginine dihydrolase (1). Prior to Moellers work, bacterial amino acid decarboxylases were studied by Gale (2) and Gale and Epps (3). Production of ornithine decarboxylase is a helpful criterion in differentiating *Klebsiella* and *Enterobacter* species. *Klebsiella* are nonmotile and do not produce ornithine decarboxylase while *Enterobacter* are motile and produce ornithine decarboxylase except *Enterobacter agglomerans* (4).

This medium contains beef extract and peptic digest of animal tissue, which provide nitrogenous nutrients for the growth of bacteria. Dextrose is the fermentable carbohydrate and pyridoxal is the co-factor for the decarboxylase enzyme. Bromo cresol purple and cresol red are the pH indicators in this medium. When the medium is inoculated with the dextrose fermenting bacteria, the pH is lowered due to acid production, which changes the colour of the indicator from purple to yellow. Acid produced stimulates decarboxylase enzyme. Decarboxylation of lysine yields cadaverine while putrescine is produced due to ornithine decarboxylation. Arginine is first hydrolyzed to ornithine which is then decarboxylated to form putrescine. Formation of these amines increases the pH of the medium, changing the colour of the indicator from yellow to purple. If the organisms do not produce the appropriate enzyme, the medium remains acidic, yellow in colour. Each isolate to be tested should also be inoculated into Moeller Decarboxylase Broth Base medium tube lacking the amino acid.

Inoculated tubes must be protected from air with a layer of sterile mineral oil. Exposure to air may cause alkalinization at the surface of the medium which makes the test invalid.

Quality Control

Appearance

Light yellow to greenish yellow homogeneous free flowing powder

Colour and Clarity of prepared medium

Purple coloured, clear solution without any precipitate in tubes

Reaction

Reaction of 1.05% w/v aqueous solution at 25°C. pH: 6.0±0.2

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5.80-6.20

Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for upto 4 days with addition of appropriate amino acids and overlaying with sterile mineral oil.

Cultural Response

Cultural Response				
Organism	Inoculum (CFU)	Arginine decarboxylation	Ornithine decarboxylation	Lysine decarboxylation
Cultural Response		·	•	•
Citrobacter freundii ATCC	50-100	variable	variable	negative
8090		reaction	reaction	reaction, yellow
_				colour
Enterobacter aerogenes	50-100	negative	positive	positive
ATCC 13048		reaction, yellow	reaction, purple	reaction, purple
Englishing all ATCC	50 100	colour variable	colour variable	colour
Escherichia coli ATCC 25922	50-100	reaction	reaction	positive reaction, purple
23922		reaction	reaction	colour
Klebsiella pneumoniae	50-100	negative	negative	positive
ATCC 13883		reaction, yellow	reaction, yellow	reaction, purple
		colour	colour	colour
Proteus mirabilis ATCC	50-100	negative	positive	negative
25933		reaction, yellow	reaction, purple	reaction, yellow
		colour	colour	colour
Proteus vulgaris ATCC	50-100	negative	negative	negative
13315		reaction, yellow	reaction, yellow	reaction, yellow
		colour	colour	colour
Salmonella Paratyphi A	50-100	delayed	positive	negative
ATCC 9150		positive	reaction, purple	reaction, yellow
		reaction/	colour	colour
		positive		
		reaction,purple		
Salva are all a Troubi ATCC	50-100	colour	ma antivo	
Salmonella Typhi ATCC 6539	30-100	delayed positive	negative reaction, yellow	positive reaction, purple
0339		reaction /	colour	colour
		negative	Coloui	coloui
		reaction		
Serratia marcescens ATCC	50-100	negative	positive	positive
8100		reaction, yellow	reaction, purple	reaction, purple
		colour	colour	colour
Shigella dysenteriae ATCC	50-100	negative	negative	negative
13313		reaction/	reaction, yellow	reaction, yellow
		delayed	colour	colour
		positive		
		reaction		
Shigella flexneri ATCC	50-100	negative	negative	negative
12022		reaction/	reaction, yellow	reaction, yellow
		delayed	colour	colour
		positive reaction		
Shigella sonnei ATCC 2593.	<i>1.</i> 50 ₋ 100	variable	positive	negative
Singena somet ATCC 2393.	1 50-100	reaction	reaction, purple	reaction, yellow
		reaction	colour	colour
			201041	201041

Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

Reference

1. Moeller V., 1955, Acta Pathol. Microbiol. Scand. 36:158.

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- 2. Gale G. F., 1940, Biochem. J., 34:392.
- 3. Gale and Epps, 1943, Nature, 152:327.
- 4. MacFaddin J., 1980, Biochemical Tests for Identification of Medical Bacteria, 2nd ed., Williams and Wilkins, Baltimore.

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