

Immunoglobulin E (IgE) Test System Product Code: 2525-300

1.0 INTRODUCTION

Microplate Enzyme Immunoassay, Colorimetric Intended Use: tended Use: The Quantitative Determination of immunoglobulin E (IgE) Concentration in Human Serum by a

2.0 SUMMARY AND EXPLANATION OF THE TEST

mast cells and basophilic granulocytes. Subsequently the binding of altergen to cell-bound type causes these cells to release histamines and other vasoache substances. The release of histamines in the body results initiates what is commonly known measurement of immunoglobulin E (IgE) in serum is widely used in the diagnosis of allegic reactions and parasitic infections. Many allegies are caused by the immunoglobulins of subclass IgE acting as point of contact between the allergen and specialized cells. The IgE molecules (MMV 200,000) bind to the surface of the symptoms. In vitro and in vivo testing, however, play a key role in confirming clinical suspicions and tailoring treatment. The as an allergic reaction. Allergic reactions, which are becoming more widespread, diagnosed on the basis of medical history and clinical

along with other supporting diagnostic information, can help to make that determination. Measurement of total circulating light may also be of value in the early detection of allergy in infants and as a means of predicting future atopic manifestations. Before deciding on any therapy it is important to take into consideration by specific allergy testing. all the relevant clinical information as well as information supplied however, to know whether the aliergic reaction is IgE mediated or non-IgE mediated. Measurement of total IgE in serum sample, Before making any therapeutic determination it is important

igE ieveis show a slow increase during childhood, reaching adult eveis in the second decade of life. In general, the total lgE leveis increase with the allergies a person has and the number of thres of exposure to the relevant allergens. Significant elevations may be seen in the sensitized individuals, but also in cases of myeloma, purmonay aspergillosis, and during the active stages of parasitic intections

In this method, IgE calibrator, patient specimen or control is first added to a streptavidin coated well. Bioiniyated monoclonal antibody (specific for IgE) is added and the reactants mixed. Reaction between the IgE antibodies and native IgE forms complex that binds with the streptavidin coated to the well. The intensity of the color generation is directly proportional concentration of the IgE in the sample. monoclonal antibody. Excess enzyme is washed off via a wash step. A color is generated by the addition of a substrate. The the wells. The enzyme labeled antibody binds to the IgE already immobilized on the well through its binding with the biotinylated excess serum proteins are washed away via a wash step. Another enzyme labeled monocional antibody specific to IgE is added to

3.0 PRINCIPLE

takes place during the assay at the surface of a microplate well through the interaction of streptavidin coated on the well and exogenously added biolinylated monoclonal anti-IgE antibody. assay include high affinity and specificity antibodies (enzyme and immobilized), with different and distinct epitope recognition, in excess, and native antigen. In this procedure, the immobilization The essential reagents required for an immunoenzymometric Immunoenzymometric sequential assay (TYPE 4):

Upon mixing monoclonal biotinylated antibody, and a serum containing the native antigen, reaction results between the native antigen and the antibody, forming an antibody-antigen complex. The interaction is illustrated by the following equation:

^{Bin}Ab_(m) = Biotinylated Monoclonal Antibody (Excess Quantity)

k_a = Rate Constant of Disassociation Ag_{t ys;} = Native Antigen (Variable Quantity)
Ag_{t ys;} - ⁸ⁿAb_{rm} = Antigen-Antibody complex (Variable Quantity)
k_a = Rate Constant of Association

Simultaneously, the complex is deposited to the well through the high affinity reaction of streptawdin and biotinylated antibody. This interaction is illustrated below.

Ag_{(gB} - BthAb_(m) + <u>Streptavidinc, w</u> → <u>Immobilized complex</u> (IC)

<u>Streptavidinc, w</u> = Streptavidin immobilized on well immobilized complex (IC) = Ag-Ab bound to the well

fraction is separated from unbound antigen by decaration or aspiration. Another antibody (directed at a different epitope) labeled with an enzyme is added. Another interaction occurs to form an enzyme labeled antibody-antigen-biotinylated-antibody complex on the surface of the wells. Excess enzyme is washed off via a wash step. A suitable substrate is added to produce color measurable with the use of a micropiate spectrophotometer. The 9 references of known antigen concentration, a dose response curve can be generated from which the antigen concentration of enzyme activity on the well is directly proportional to the native antigen concentration. By utilizing several different serum After a suitable incubation period, the antibody-antigen bound fraction is separated from unbound antigen by decantation or can be ascertained

(IC) + ^{6rz}Ab_(x-gE) Enz Ab(x-IgE) - IC

EnzAtix-56 = Enzyme labeled Antibody (Excess Quantity)
EnzAb_[x-56] = IC = Antinen-Antibodica Caracteristics

***Ab_{K-kgE} – IC = Antigen-Antibodies Complex k_b = Rate Constant of Association k_b = Rate Constant of Dissociation

4.0

Materials Provided: A IgE Calibrators – 1.0 ml/vial - Icons A-F Six (6) vials of human serum based reference calibrators at

concentrations of 0 (A), 5 (B), 25 (C), 50 (D), 150 (E) and 400 (F) Iu/m. Store at 2-8°C. A preservative has been added. Note: The Calificators are standardized against WHO's 2ndRP 75/502 for IgE

IgE Biotin Reagent – 13 ml/vial – Icon V One (1) vial containing biotinylated anti) vial containing biotinylated anti-human IgE mlgG presented in a protein-stabilized matrix. A preservative

has been added. Store at 2-8°C

One (1) vial containing anti-human lgE-HRP incorporated complex in a protein-stabilized matrix. A preservative has been IgE Enzyme Reagent - 13 ml/vial - Icon (E)
One (1) vial containing anti-human IgE-H added. Store at 2-8°C

Streptavidin Plate - 96 wells - Icon U
One 96-well microplate coated with streptavidin and packaged in an aluminum bag with a drying agent. Store at 2-8°C.

Ö

Substrate A – 7.0ml/vial - Icon S^A Wash Solution Concentrate – 20ml/vial - Icon ♠ One (1) vial containing a surfactant in buffered saline.

One (1) vial containing tetramethylbenzidine (TMB) in acetate

G. Substrate B – 7.0ml/vial - loon S⁶
One (1) vial containing hydrogen peroxide (H₂O₂) in acetate buffer. Store at 2-8°C

H. Stop Solution - 8.0ml/vial - Icon

One (1) vial containing a strong acid (1N HCl). Store at 2-8°C

Note 2: Avoid extended exposure to heat and light. Opens reagents are stable for sixty (60) days when stored 2-8°C. Kit and component stability are identified on label. Note 3: Above reagents are for a single 96-well microplate. Note 1: Do not use reagents beyond the kit expiration date. Opened

4.1 Required But Not Provided:

Pipette capable of delivering 0.025 and 0.050ml (25 & 50µl) volumes with a precision of better than 1.5%

Dispenser(s) for repetitive deliveries of 0.100 and 0.350ml (100 & 350µl) volumes with a precision of better than 1.5%.

ω Microplate Reader with 450nm and 620nm wavelength Microplate washers or a squeeze bottle (optional)

absorbance capability

Absorbent Paper for blotting the microplate wells.

Plastic wrap or microplate cover for incubation steps.

Vacuum aspirator (optional) for wash steps.

Quality control materials

5.0 **PRECAUTIONS**

Not for Internal or External Use in Humans or Animals For In Vitro Diagnostic Use

hazardous and capable or uarisinuing uncountered in the procedures for handling blood products can be found in the procedures for handling blood products can be found in the Edition, 1988, HHS Publication No. (CDC) 88-8395. Center for Disease Control / National Institute of H "Biosafety in Microbiological and Biomedical Laboratories," human serum products snown to hazardous and capable of transmitting disease. Good laboratory hazardous and capable of transmitting disease. Good laboratory offer complete assurance that infectious agents are absent, all human serum products should be handled as potentially The number of the security of the security have been found to be non-reactive for Hepatitis B Surface Antigen, HIV 182 and HCV Antibodies by FDA licensed reagents. Since the nown test can offer complete assurance that infortions control to the security of the security o All products that contain human serum have been found to

Safe disposal of kit components must be according to local regulatory and statutory requirement.

SPECIMEN COLLECTION AND PREPARATION

should be collected in a plain redtop venipuncture tube without a fasting morning serum sample should be obtained. observed. For accurate comparison to established normal values precautions in the collection of venipuncture samples should be Centrifuge the specimen to separate the serum from the cells additives or anti-coagulants. Allow the blood to clot for samples. The specimens shall be blood serum in type and the usual . The blood

ensure fasting sample. >5mg/day), no sample should be taken until at least 8 hours after the last biotin administration, preferably overnight to in patients receiving therapy with high biotin doses (i.e.

of the specimen is required. Samples may be refrigerated at 2-8°C for a maximum period of five (5) days. If the specimen(s) cannot be assayed within this time, the sample(s) may be stored at temperatures of -20°C for up to 30 days. Avoid use of contaminated devices. Avoid repetitive reezing and thawing. When assayed in duplicate, 0.050ml (50µl)

7.0 QUALITY CONTROL

Each laboratory should assay controls at levels in the low, normal and elevated range for monitoring assay performance. These controls should be treated as unknowns and values determined in every lest procedure performed. Quality control charts should be trends. Significant deviation from established performance can maintained to follow the performance of the supplied reagents. Pertinent statistical methods should be employed to ascertain determine the reason for the variations degradation of kit reagents. Fresh reagents should be unnoticed change in experimental conditions used to or

8.0 REAGENT PREPARATION

deionized water in a suitable storage container. Storediluted Dilute contents of wash concentrate to 1000ml with distilled or buffer at 2-30°C for up to 60 days.

reagent for easy identification. Mix and label accordingly. Store Pour the contents of vial labeled Solution 'A' into the labeled Solution 'B'. Place the yellow cap on the mi Working Substrate Solution - Stable for one year mixed Via.

Note 2: Do not use reagents that are contaminated or have Note 1: Do not use the working substrate if it looks blue.

9.0 TEST PROCEDURE

Before proceeding with the assay, bring all reagents, serum reference calibrators and controls to room temperature (20-27°C).
Test procedure should be performed by a skilled individual. or trained professional

calibrator, control and patient specimen to be assayed in duplicate. Replace any unused microwell strips back into Format the micropiates' wells for each serum reference

Pipette 0.025 ml (25µl) of the appropriate serum reference calibrator, control or specimen into the assigned well. the aluminum bag, seal and store at 2-8°C

ω Add 0.100 ml (100µl) of the IgE Biotin Reagent to each well. It

bottom of the coated well. is very important to dispense all reagents close to the

0 0 4 Incubate 30 minutes at room temperature. Swirl the microplate gently for 20-30 seconds to mix and cover

Discard the contents of the microplate If decanting, tap and blot the plate dry decantation

container (avoiding air bubbles) to dispense the wash. Decart the wash and repeat two (2) additional times. Add 0.100 ml (100µl) of the IgE Enzyme Reagent labeled Add 0.350ml (350µl) of wash buffer (see Reagent Preparation Section), decant (tap and blot) or aspirate. Repeat two (2) additional times for a total of three (3) washes. An automatic manufacturer's instruction for proper usage. If a squeeze bottle is employed, fill each well by depressing the or manual plate washer can be used. Follow the

antibody to each well.

DO NOT SHAKE THE PLATE AFTER ENZYME ADDITION

10. Discard the contents of the microplate by decantation Cover and incubate 30 minutes at room temperature. aspiration. If decanting, blot the plate dry with absorbent

11. Add 0.350ml (350µl) of wash buffer (see Reagent Preparation additional times for a total of three (3) washes. An automatic or manual plate washer can be used. Follow the manufacturer's instruction for proper usage. If a squeeze bottle is employed, fill each well by depressing the container (avoiding air bubbles) to dispense the wash. Decant the wash and repeat two (2) additional times Section), decant (tap and blot) or aspirate. Repeat two (2)

12. Add 0.100 ml (100µl) of working substrate solution to all wells (see Reagent Preparation Section). Always add reagents in

14. Add 0.050ml (50µl) of stop solution to each well and gently mix DO NOT SHAKE THE PLATE AFTER SUBSTRATE ADDITION 13. Incubate at room temperature for fifteen (15) minutes.

15. Read the absorbance in each well at 450nm (using a reference wavelength of 620-630nm to minimize well imperfections) in a microplate reader. The results should be read within thirty (30) minutes of adding the stop solution.

CALCULATION OF RESULTS

A dose response curve is used to ascertain the concentration of in unknown specimens.

microplate reader as outlined in Example 1.

2. Plot the absorbance for each duplicate serum reference versus Record the absorbance obtained from the printout of the

the corresponding IgE concentration in IU/ml on linear

paper (do not average the duplicates of the serum references before plotting).

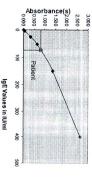
3. Draw the best-fit curve through the plotted points.

4. To determine the concentration of IgE for an unknown, locate the average absorbance of the duplicates for each unknown on the vertical axis of the graph, find the intersecting point on the curve, and read the concentration (in IUImi) from the horizontal axis of the graph (the duplicates of the unknown may be averaged as indicated). In the following example, the average absorbance (1.323) intersects the dose response curve at 142 IUIm IgE concentration (See Figure 1).

Note: Computer data reduction software designed for ELISA software is utilized, the validation of the software should be assays may also be used for the data reduction. If such

i angiit i	Patient 1	1	<u> </u>	-	<u>.</u>	-	<u></u>	Cair	<u> </u>	Ç.	ا ا	o o		Car	G B	Og 7	C3 A	I.D.	
B3	A3	H2	G2	F2	E2	D2	C2	B2	22	표	G1	FI	E1	D1	C1	B1	A1	Well	
1.324	1.322	0.807	0.818	2.549	2.575	2.682	2.601	1.388	1.340	0.614	0.663	0.326	0.364	0.074	0.072	0.016	0.014	Abs	EXAMPLE !
1.323	1 222	0.010	0.813	2.302	C33 C	2.041	1130	1.304	1361	0.000	0630	0.040	372 U	0.073	0 073	0.015	0.015	Mean Abs (B)	
142.0	0.077	11.2	74 2	3/3.3	275 2	400	100	130	150	٤	50	22	SE.	c		c	0	Conc	

Figure 1



*The data presented in Example 1 and Figure 1 is for illustration only and should not be used in lieu of a standard curve prepared with each assay.

11.0 Q.C. PARAMETERS

In order for the assay results to be considered valid the ollowing criteria should be met:

- The absorbance (OD) of calibrator 'A' should be \leq 0.05. The absorbance (OD) of calibrator 'F' should be \geq 1.8. Four out of six quality control pools should be with

12.0 RISK ANALYSIS established ranges.

on request from Monobind Inc. The MSDS and Risk Analysis Form for this product are available

- 12.1 As say Performance It is important that the time of reaction in each well is held
- constant to achieve reproducible results.

 Pipeting of samples should not extend beyond ten (10) minutes to avoid assay drift.
- 4 Highly hemolyzed 9
- 5 specimen(s) should not be used.
 If more than one (1) plate is used, it is recommended to repeat the dose response curve.
 The addition of substrate solution initiates a kinetic reaction, terminated by the addition of the stop solution. Therefore, the

substrate and stop solution should be added in the same sequence to eliminate any time-deviation during reaction. Pate readers measure vertically. Do not touch the bottom of

- aspiration or decantation wash step(s) may result in poor Failure to remove adhering the wells.
- replication and spurious results.

 Use components from the same lot. No intermixing of reagents from different batches
- Any deviation from Monobind IFU may yield inaccurate results.

 10. All applicable national standards, regulations and laws, including, but not limited to, good laboratory procedures, must be strictly followed to ensure compliance and proper devices Accurate and precise pipetting, as well as following the exact time and temperature requirements prescribed are essential.
- usage.

 11. It is important to calibrate all the equipment e.g. Pipettes Readers, with this Washers and/or the automated instruments s device, and to perform routine preven used
- Risk Analysis as required by CE Mark IVD Directive 98/79/EC for this and other devices, made by Monobind, can be requested via email from Monobind@monobind.com.

12.2 Interpretation

performed by a skilled individual or trained professional. and interpretation of results must 8

Laboratory results alone are only one aspect for determining patient care and should not be the sole basis for therapy,

ambodies: a problem for all immunoassays' Clin. Chem. 1983;347-33). For diagnostic purposes, the results from this assay should be in combination with clinical examination, patient history and all other clinical findings. interactions and have been known to be problems for all kinds of immunoassays (Boscato LM, Stuart MC. 'Heterophilic between rare serum specimens and test reagents can cause erroneous results. Heterophilic antibodies often cause these eliminate maximal interference; however, potential interaction particularly if the results conflict with other determinants.

The reagents for the test system have been formulated to

If test kits are aftered, such as by mixing parts of different kits, which could produce false test results, or if results are must be within the listed ranges and assay requirements.

incorrectly interpreted, <u>Monobind shall have no liability</u>. If computer controlled data reduction is used to interpret the results of the test, it is imperative that the predicted values for the calibrators fall within 10% of the assigned concentrations. Serum IgE concentration is dependent upon a multiplicity of factors: including if the patient is sensitized, how many times the patient has been exposed to a specific allergen etc. Total IgE concentration alone is not sufficient to assess the clinical clinical status of the patient. status. All the clinical findings especially specific allergy testing should be taken into consideration while determining the

Since all atopic reactions are not IgE mediated, all relevant making any determination for patients who may be in clinical information should be taken into consideration before the

13.0 EXPECTED RANGES OF VALUES

A study of population from different age groups was conducted to evaluate the IgE AccuBind® ELISA test system. The results are

Expected Values for the IgE (In IU/ml) Number (n) 31 43 145 Median 6.4 25.0 Absolute Range ND - 46 ND - 280 0 - 200

Age (Yrs) 0-3

3-16 Adult

Sample Pool 1

reasons each laboratory should depend upon the range of expected values established by the Manufacturer only until in-house range can be determined by the analysts using the method with a population indigenous to the area in which the It is important to keep in mind that establishment of a range of values which can be expected to be found by a given method for a population of "normal"-persons is dependent upon a multiplicity of factors: the specificity of the method, the population tested and the precision of the method in the hands of the analyst. For these

Pool 1/8 Pool 1/16 Pool 2 Pool 2/2 Pool 2/4 Pool 2/8 Pool 2/16

laboratory is located

14.0 PERFORMANCE CHARACTERISTICS

14.1 Precision The within and

The within and between assay precision of the IgE AccuBind® ELISA Test System were determined by analyses on three different levels of pool control sera. The number, mean value, standard deviation and coefficient of variation for each of these control sera are presented in Table 2 and Table 3.

=	tra-Assa	i ABLE 2 Intra-Assay Precision (in IU/ml)	on (in IU/ı	크
SAMPLE	z	×	q	C.V.%
Low	20	48.9	2.87	5.87
Medium	20	160.5	6.47	4.03
High	20	297.6	5.81	1.95
	Inter A	Inter Assay Precision (in IU/ml	ision (in I	U/ml)
SAMPLE	z	×	a	C.V.%
Low	10	46.3	3.9	8.42
Medium	10	157.0	7.3	4.64
Han	6	301.0	10.6	3.52

14.2 Sensitivity

The IgE AccuBind® ELISA test system has a sensitivity of 0.125 IU/m. The sensitivity was ascertained by determining the variability of the 0 IU/m serum calibrator and using the 2σ (95% certainty) statistics to calculate the minimum dose.

The IgE Accuracy

least square regression equation and the correlation coefficient were computed for this IgE AccuBind® ELISA method in reference method. Biological specimens with $\lg E$ levels in the low medium and high ranges were used. The values ranged from 0.8comparison with the predicate method (Table 4): to 3100 IIU/ml. The total number of such specimens was 219. The IgE AccuBind® ELISA test system was compared with a 5

		LABCE #	
Method	Mean	Least Square	Square Correlation
		Regression Analysis	Coefficient
Monobind (X)	179	x=-12.9+1.21(Y)	0.967
Predicate (Y)	157		

values. The least square regression equation and correlation coefficient indicates excellent method agreement. Only slight amounts of bias between this method and the reference method are indicated by the closeness of the mean

14.4 Specificity

The specificity of the IgE AccuBind® ELISA test system, to closely related immunogobulins was evaluated by adding those at twice the physiological concentrations to a serum matrix. No cross-reaction between the antibodies used and the related molecules was detected.

14.5. High Dose Effect

Since the assay is sequential in design, high concentrations of IgE do not show the hook effect. Myeloma IgE patient samples with concentrations over 8 million IU/ml demonstrated extremely high lands of characters. levels of absorbance.

4.6 Linearity

Two patient pools were as undiluted with the IgE Acc observed and expected vi

EC	104.2	24.7	25.8
	96.9	49.5	48.0
	107.2	98.9	106.1
(Experators De	95.8	197.9	189.5
Used By			395.9
	98.5	6.7	6.6
	100.6	13.3	13.4
Musican	94.8	26.7	25.3
Catalogu	95.1	53.4	50.8
777			106.8
	(O/E)	(E) (IU/ml)	(O) (IU/ml)
Sheeke	% Recovery	Expected	Observed
in Vigo. Diagnosts		ABLE 5	TAI
GV.	in Table 5:	pected values are listed below in Table 5:	ected values a
	stem. The	IgE AccuBind® ELISA test system. The	IgE AccuBind
	Calibrator) and	s were assayed diluted (in 'A' Calibrator) and	were assayed

14.7 Recovery
Two patient pools were spiked with known amounts of IgE assayed with the IgE AccuBind® ELISA test system observed and expected values are listed below in Table 6. and The

	7,4	ABLE 6	
Sample	Observed (O) (IU/ml)	Expected (E) (IU/ml)	% Recovery (O/E)
Pool 1	25.7		
Pool 1+ 25	50.7	50.7	100.0
Pool 1+ 50	74.8	75.7	101.2
Pool 1+ 100	122.7	125.7	97.6
Pool 1+ 200	232.0	225.7	102.7
Pool 2	12.3		
Pool 2 + 25	41.7	37.3	111.2
Pool 2+ 50	62.6	62.3	100.6
Pool 2+ 100	109.4	112.3	97.4
Pool 2+ 200	197.2	212.3	92.8

15.0 REFERENCES

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For Orders and inquires, please contact

** Monobind Inc.

100 North Pointe Drive
Lake Forest, CA 92830 USA

Tel: +1 949.951.2686 Fax: +1 949.951.3539 Mail: info@monobind.com Fax: www.monobind.com

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