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CE 1023

IVD For *in Vitro* Diagnostic Use

For Professional Use Only

HLA B*5701 Real-TM Handbook

Real Time PCR test for the detection of HLA-B (major histocompatibility complex, class I, B) Allele 5701

REF H53-100FRT

Σ 100

NAME
HLA B*5701 Real-TM

INTRODUCTION
Abacavir is a nucleoside reverse-transcriptase inhibitor with activity against the human immunodeficiency virus (HIV), available for once-daily use in combination with other antiretroviral agents, that has shown efficacy, few drug interactions, and a favorable long-term toxicity profile. The most important adverse effect of abacavir that limits its use in therapy and mandates a high degree of clinical vigilance is an immunologically mediated hypersensitivity reaction affecting 5 to 8% of patients during the first 6 weeks of treatment. Symptoms of a hypersensitivity reaction to abacavir are nonspecific and include combinations of fever, rash, constitutional symptoms, gastrointestinal tract symptoms, and respiratory symptoms that become more severe with continued dosing. Immediate and permanent discontinuation of abacavir is mandated, resulting in a rapid reversal of symptoms. Subsequent rechallenge with abacavir is contraindicated, since it can result in a more severe, rapid, and potentially life-threatening reaction. In 2002, an association between a diagnosis of hypersensitivity reaction to abacavir and carriage of the major histocompatibility complex class I allelee HLA-B*5701 was reported independently by several independent studies.
Studies of cohorts with HIV infection have also shown that avoiding abacavir in HLA-B*5701–positive patients significantly reduced the incidence of suspected hypersensitivity reaction up to 0,5%. Many clinical studies recommend for this reason, the pharmacogenetic molecular testing of the carriage of the major histocompatibility complex class I allelee HLA-B*5701 in all HIV positive patients treated with abacovir.
HLA-B*5701 Real-TM test can predict who will develop a severe allergic reaction to the anti-HIV drug abacavir as the presence of HLA-B*5701 is significantly associated with an abacavir hypersensitivity.

INTENDED USE
HLA B*5701 Real-TM is a Real-Time amplification test for the detection of HLA-B (major histocompatibility complex, class I, B) Allele 5701 in the biological materials.
The kit **HLA B*5701 Real-TM** can be used as screening test for the prevention of abacavir hypersensitivity reactions.

PRINCIPLE OF ASSAY
HLA B*5701 Real-TM Test is based on two major processes: isolation of genomic DNA from specimens and Real Time amplification with allele specific primers. The real-time PCR monitoring of fluorescence intensities allows the accumulating product detection without reopening of reaction tubes after the PCR run. **HLA B*5701 Real-TM** PCR kit is a qualitative test which contains the Internal Control IC (human beta-globine gene), which allows to control the presence of cellular material in the sample.

PRODUCT USE LIMITATIONS
All reagents may exclusively be used in in vitro diagnostics. Use of this product should be limited to personnel trained in the techniques of DNA amplification (EN375). Strict compliance with the user manual is required for optimal PCR results. Attention should be paid to expiration dates printed on the box and labels of all components. Do not use a kit after its expiration date.

QUALITY CONTROL
In accordance with Sacace's ISO 13485-Certified Quality Management System, each lot is tested against predetermined specifications to ensure consistent product quality.

MATERIALS PROVIDED

Reagent	*Description*	*Volume, ml*	*Amount*
PCR-mix-1-FRT HLA	colorless clear liquid	0.6	2 tubes
RT-PCR-mix-2-FL	colorless clear liquid	0.3	2 tubes
Polymerase (TaqF)	colorless clear liquid	0.03	2 tubes
TE-buffer	colorless clear liquid	0.07	2 tubes
Positive Control DNA HLA B*5701 and human DNA (C+)	colorless clear liquid	0.2	1 tube
Negative Control (C–)*	colorless clear liquid	0.5	4 tubes

* must be used in the isolation procedure as Negative Control of Extraction.

MATERIALS REQUIRED BUT NOT PROVIDED

- DNA extraction kit.
- Disposable powder-free gloves.
- Pipettes (adjustable).
- Sterile pipette tips with aerosol filters up to 200 µl.
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with a rotor for 2-ml reaction tubes.
- PCR box.
- Personal thermocycler.
- Disposable polypropylene microtubes for PCR or PCR-plate
- Refrigerator for 2–8 °C.
- Deep-freezer for ≤ –16 °C.
- Waste bin for used tips.

STORAGE INSTRUCTIONS

The kit **HLA B*5701 Real-TM** must be stored at or below minus 16 °C when not in use.
The kit can be shipped at 2-8°C for 3-4 days but should be stored at -20°C immediately on receipt.

PCR-mix-1-FRT HLA is to be kept away from light.

STABILITY

HLA B*5701 Real-TM is stable up to the expiration date indicated on the kit label. The product will maintain performance through the control date printed on the label. The shelf life of reagents before and after the first use is the same, unless otherwise stated.
Exposure to light, heat or humidity may affect the shelf life of some of the kit components and should be avoided. Repeated thawing and freezing of these reagents should be avoided, as this may reduce the sensitivity.

WARNINGS AND PRECAUTIONS

IVD ***In Vitro* Diagnostic Medical Device**

For *In Vitro* Diagnostic Use Only

1. Wear disposable gloves, laboratory coats and eye protection when handling specimens and reagents. Thoroughly wash hands afterward.
2. Do not pipette by mouth.
3. Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
4. Do not use a kit after its expiration date.
5. Dispose of all specimens and unused reagents in accordance with local regulations.
6. Biosafety Level 2 should be used for materials that contain or are suspected of containing infectious agents.
7. Clean and disinfect all spills of specimens or reagents using a disinfectant such as 0,5% sodium hypochlorite, or other suitable disinfectant.
8. Avoid contact of specimens and reagents with the skin, eyes and mucous membranes. If these solutions come into contact, rinse immediately with water and seek medical advice immediately.
9. Material Safety Data Sheets (MSDS) are available on request.
10. Use of this product should be limited to personnel trained in the techniques of DNA amplification.
11. PCR reactions are sensitive to contamination. Measures to reduce the risk of contamination in the laboratory include physically separating the activities involved in performing PCR in compliance with good laboratory practice.
12. Workflow in the laboratory must proceed in a uni-directional manner, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where you performed previous step.

Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

Sampling of biological materials for PCR-analysis, transportation, and storage are described in details in the handbook of the manufacturer. It is recommended that this handbook is read before beginning of the work.

SAMPLE COLLECTION, STORAGE AND TRANSPORT

HLA B*5701 Real-TM can analyze genomic DNA extracted from:

- *whole blood:* Collect 2 ml of blood to a tube with 0.2 ml of 3% EDTA solution. Invert a closed tube several times to ensure proper mixing. Blood samples should be stored at 2–8 °C for up to 48 h
- *Oropharyngeal swabs* are taken with a sterile probe with a cotton tip. After swabbing, the probe should be placed to a tube with 0.5 ml of “Transport Medium for Storage and Transportation Respiratory Swabs” (REF 958). The probe should be broken off at the score mark so that the tube is tightly closed. The sample should be stored at 2–8 °C for up to 3 days.

Transportation of clinical specimens must comply with country, federal, state and local regulations for the transport of etiologic agents.

DNA ISOLATION

Any commercial RNA/DNA isolation kit, if IVD-CE validated for the specimen types indicated herein at the "SAMPLE COLLECTION, STORAGE AND TRANSPORT" paragraph, could be used.

Sacace Biotechnologies recommends to use the following kits:

- **SaMag Blood DNA extraction kit** (Sacace, REF SM001);
- **DNA/RNA-Prep** (Sacace, REF K-2-9).

Extract DNA according to the manufacturer's instruction.

Whole blood samples should be treated with "Hemolytic" (**REF** 137-CE) before adding the lysis solution. To do this, add 1.0 ml of "Hemolytic" and 0.1 ml of whole blood to a 1.5-ml tube. Carefully vortex. Incubate the tubes at room temperature for 5 min, vortex, and incubate for 5 min once again. Centrifuge (8,000 rpm, 2 min). Remove and discard the supernatant. Leukocyte sediment should be immediately lysed; otherwise, it should be stored frozen at or below minus 16°C for up to 3 days or at or below minus 68°C for a long time.

Prior to DNA extraction from throat swabs placed in "Transport Medium for Storage and Transportation of Respiratory Swabs" (**REF** 957-CE), thoroughly mix, and then briefly vortex the samples.

REAGENT PREPARATION

1. Prepare the **reaction mixture**. Per **one** reaction:
   - **10 µl of PCR-mix-1-FRT HLA**
   - **5 µl of RT-PCR-mix-2-FL**
   - **0.5 µl of polymerase (TaqF)**

Add one extra reaction when calculating the reaction mixture volume.

Number of samples	Volume of the reagents for specified number of samples, µl (one extra reaction is included)		
	PCR-mix-1-FRT HLA	**RT-PCR-mix-2-FL**	**Polymerase (TaqF)**
6	**70**	**35**	**3.5**
11	**120**	**60**	**6.0**
18	**190**	**95**	**9.5**

2. Thoroughly vortex prepared mixture, make sure there are no drops on the wall of the tubes.
3. Take the required number of the PCR tubes for amplification of clinical and control samples. Transfer **15 µl** of prepared reaction mix to each tube.
4. **Add 10 µl** of **DNA samples** obtained from clinical or control samples at the stage of DNA extraction into prepared tubes.
5. Carry out control amplification reactions:

NCA - Add **10 µl** of **TE-buffer** to the tube labeled NCA (Negative Control of Amplification).

C+ - Add **10 µl** of **Positive Control DNA HLA B*5701 and human DNA** to the tube labeled C+ (Positive Control of Amplification).

Create a temperature profile on your Real-time instrument as follows:

	Rotor type instruments[1]				*Plate type or modular instruments*[2]			
Stage	*Temp, °C*	*Time*	*Fluorescence detection*	*Cycle repeats*	*Temp,°C*	*Time*	*Fluorescence detection*	*Cycle repeats*
Hold	95	15 min	–	1	95	15 min	–	1
Cycling	95	5 s	–	5	95	5 s	–	5
	60	20 s	–		60	20 s	–	
Cycling 2	95	5 s	–	40	95	5 s	–	40
	60	40 s	FAM(Green), JOE(Yellow)		60	50 s	FAM, JOE/HEX/Cy3	

[1] *For example Rotor-Gene™ 3000/6000/Q (Corbett Research, Qiagen)*

[2] *For example, SaCycler-96™ (Sacace), CFX/iQ5™ (BioRad); Mx3005P™ (Agilent), ABI® 7300/7500/StepOne Real Time PCR (Applied Biosystems), SmartCycler® (Cepheid), LineGeneK® (Bioer)*

RESULTS ANALYSIS

The results are interpreted by the device software through the presence of crossing of fluorescence curve with the threshold line.

*DNA HLA*B5701* is detected on the JOE (Yellow)/HEX/Cy3 channel and *IC* on the FAM (Green) channel.

Results are accepted as relevant if both positive and negative controls of amplification along with negative control of extraction are passed (see table 1).

Table 1. Results for controls

Control	Stage for control	Ct in channel		Interpretation
		FAM /Green	JOE/Yellow/HEX	
C–	DNA extraction	Neg	Neg	OK
NCA	Amplification	Neg	Neg	OK
C+	Amplification	Pos (< boundary value*)	Pos (< boundary value*)	OK

- The sample is considered to be positive if in the channel Joe (Yellow)/HEX/Cy3 the result is positive and the value of Ct on this channel is higher than Ct on the Fam (Green) channel but not more than 5 cycles (see table 2).
- The sample is considered to be negative if in the channel Joe (Yellow)/HEX/Cy3 value is negative or if the value of Ct on this channel is higher than Ct on the Fam (Green) of more than 5 cycles.
- Normal difference between Joe (Yellow) and Fam (Green) Ct values is 2-3 cycles.

Table 1. Results for samples

Sample	Ct value and result			
	RotorGene		iQ, iQ5, Mx3005, ABI	
	FAM, IC	JOE, HLA	FAM, IC	HEX, HLA
C+	< 25 (positive)	< 25 (positive)	< 29 (positive)	< 29 (positive)
Clinical sample	< 25 (positive)	< Ct (FAM)+5 (positive)	< 29 (positive)	< Ct (FAM)+5 (positive)

QUALITY CONTROL PROCEDURE

HLA B*5701 Real-TM PCR kit is a qualitative test which contains the Internal Control IC (human beta-globine gene), which allows to control the presence of cellular material in the sample. If the sample is not correctly prepared or it is an insufficient quantity of epithelial cells the Internal Control will not be detected.

A negative control of extraction (NCE), negative amplification control (NCA), positive amplification control (C+) are required for every run to verify that the specimen preparation, the amplification and the detection steps are performed correctly.

If the controls are out of their expected range (see table Results for Controls), all of the specimens and controls from that run must be processed beginning from the sample preparation step.

TROUBLESHOOTING

1. Absent signal of the IC (Fam (Green) channel): retesting of the sample is required.
    - The PCR was inhibited.
        ⇒ Make sure that you use a recommended DNA extraction method and follow the manufacturer's instructions.
    - The reagents storage conditions didn't comply with the instructions.
        ⇒ Check the storage conditions
    - The PCR conditions didn't comply with the instructions.
        ⇒ Check the PCR conditions and for the IC detection select the fluorescence channel reported in the protocol.
    - No correct sample collection or preparation.
2. No signal on the Joe (Yellow)/Cy3/HEX and Fam (Green) channels with Positive Control.
    - The reagents storage conditions didn't comply with the instructions.
        ⇒ Check the storage conditions
    - The PCR conditions didn't comply with the instructions.
        ⇒ Check the temperature profile and select the fluorescence channel reported in the protocol.
    - Incorrect configuration of the PCR reaction:
        ⇒ Check the reagents preparation step.
3. Any signal with Negative Control.
    - Contamination during PCR preparation procedure. All samples results are invalid.
        ⇒ Decontaminate all surfaces and instruments with sodium hypochlorite and ethanol or special DNA decontamination reagents.
        ⇒ Pipette the Positive controls at the end.
        ⇒ Repeat the PCR preparation with the new set of reagents.

PERFORMANCE CHARACTERISTICS

Sensitivity

Analytical Sensitivity of **HLA B*5701 Real-TM** PCR kit is not less than 1 x 10^3 cells per 1 ml of a sample (cells/ml).

The claimed analytical features of **HLA B*5701 Real-TM** PCR kit are guaranteed only when additional reagents kit, "**DNA/RNA-Prep** (Sacace, REF K-2-9), is used.

Specificity

Specificity of **HLA B*5701 Real-TM** PCR kit is assured by selection of specific primers and probes, as well as the selection of strict reaction conditions. The primers and probes were checked for possible homologies to all in gene banks published sequences by sequence comparison analysis. Specificity of **HLA B*5701 Real-TM** PCR kit was confirmed in laboratory clinical trials.

KEY TO SYMBOLS USED

REF	List Number		Caution!
LOT	Lot Number		Contains sufficient for <n> tests
IVD	For *in Vitro* Diagnostic Use	VER	Version
	Store at	**NCA**	Negative Control of Amplification
	Manufacturer	**C–**	Negative control of Extraction
	Consult instructions for use	**C+**	Positive Control of Amplification
	Expiration Date	**IC**	Internal Control

References

- *High sensitivity of human leukocyte antigen-B*5701 as a marker of immunologically confirmed abacavir hypersensitivity in white and black patients*. Saag M et al. Clin Infect Dis 46: 1111 – 1118, 2008.
- *Association between presence of HLA-B*5701, HLA-DR7, and HLA-DQ3 and hypersensitivity to HIV-1 reverse-transcriptase inhibitor abacavir. Mallal S, Nolan D et al. Lancet. 2002 Mar 2;359(9308):722-3.*
- *HLA-B*5701 screening for hypersensitivity to abacavir. Mallal et al N Engl J Med. 2008 Feb 7;358(6):568-79.*
- *Value of the HLA-B*5701 allele to predict abacavir hypersensitivity in Spaniards. Rodríguez-Nóvoa S, García-Gascó P, Blanco F, González-Pardo G, Castellares C, Moreno V, Jiménez-Nácher I, González-Lahoz J, Soriano V. AIDS Res Hum Retroviruses. 2007 Nov;23(11):1374-6.*
- *Prospective HLA-B*5701 screening and abacavir hypersensitivity: a single centre experience.Waters LJ, Mandalia S, Gazzard B, Nelson M. AIDS. 2007 Nov 30;21(18):2533-4.*
- *Abacavir hypersensitivity reaction in primary HIV infection. Stekler J, Maenza J, Stevens C, Holte S, Malhotra U, McElrath MJ, Corey L, Collier AC. AIDS. 2006 Jun 12;20(9):1269-74.*
- *The pharmacogenetics of antiretroviral therapy. Phillips EJ. Curr Opin HIV AIDS. 2006 May;1(3):249-56.*

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* SmartCycler® is a registered trademark of Cepheid

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Sacace BIOTECHNOLOGIES

IVD For *in Vitro* Diagnostic Use

CE

DNA/RNA Prep NA

Nucleic acid extraction kit for the extraction and purification of total RNA/DNA from clinical materials

USER MANUAL

REF **K-2-9/2**

Σ **100**

NAME

DNA/RNA Prep NA

INTENDED USE

The **DNA/RNA Prep NA** nucleic acid extraction kit is intended for the extraction and purification of total RNA/DNA from clinical materials (plasma, serum, swabs and washouts from nasal and oropharyngeal cavities, saliva, urine, sperm, prostate fluid, cerebrospinal fluid, epithelial scrapes from posterior pharyngeal wall, urethra, cervical canal, posterior vaginal vault, faeces).

PRINCIPLE OF ASSAY

DNA/RNA Prep NA nucleic acid extraction kit is a reagent kit for rapid and efficient manual extraction and purification of DNA/RNA from various biological materials. Lysis Sol contains chaotropic agent (guanidine thiocyanate) that lyses cells and denaturates cell proteins. The nucleic acids are then precipitated in 2-propanol solution. The nucleic acids are eluted in low salt buffer and are ready-for use in subsequent reactions. The prepared nucleic acids are suitable for applications like automated fluorescent DNA sequencing, RT-PCR, or any kind of enzymatic manipulation. We highly recommend the use of internal standards as well as positive and negative controls in order to monitor the purification, amplification and detection processes.

MATERIALS PROVIDED

- **Lysis Sol**, 30 ml;
- **Prec Sol**, 40 ml;
- **Washing Sol 3**, 50 ml;
- **Washing Sol 4**, 30 ml
- **RE-buffer**, 4 x 1,25 ml;

Contains reagents for 100 tests.

MATERIALS REQUIRED BUT NOT PROVIDED

- Biological cabinet
- Vortex
- Tube racks
- Desktop microcentrifuge for “eppendorf” type tubes
- 65°C ± 2°C dry heat block
- Microcentrifuge tubes, 1,5 - 2,0 ml
- Pipettes
- Sterile, RNase-free pipette tips with filters
- Biohazard waste container
- Disposable gloves, powderless
- Refrigerator
- Freezer

WARNINGS AND PRECAUTIONS

-  Component Lysis Sol contains guanidine thiocyanate. Guanidine thiocyanate is harmful if inhaled, or comes into contact with skin or if swallowed. Contact with acid releases toxic gas. (Xn; R: 20/21/22-36/38; S: 36/37/39).

  Risk Phrases

  R 20/21/22 Harmful by inhalation, in contact with the skin and if swallowed

  R 22 Harmful if swallowed

  R 36/38 Irritating to eyes and skin

  Safety Phrases

  S 13 Keep away from food, drink and animal feedstuffs

-  Component Prec Sol contains 2-propanol solution. Irritant. (R36-67, S7-16-24/25-26)

  Risk Phrases

  R36/37/38: Irritating to eyes, respiratory system and skin

  R67: Vapors may cause drowsiness and dizziness

  Safety Phrases

  S7: Keep container tightly closed

  S24/25: Avoid contact with skin and eyes

  S26: In case of contact with eyes, rinse immediately with plenty of water and seek medical advice;

- Wear disposable gloves, laboratory coats and eye protection when handling specimens and reagents. Thoroughly wash hands afterward.
- Do not pipette by mouth.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Specimens should be considered potentially infectious and handled in biological cabinet in accordance with Biosafety Level 2 or other appropriate biosafety practices.
- Clean and disinfect all spills of specimens or reagents using a disinfectant such as 0,5% sodium hypochlorite, or other suitable disinfectant.
- Avoid contact of specimens and reagents with the skin, eyes and mucous membranes. If these solutions come into contact, rinse immediately with water and seek medical advice immediately.
- Material Safety Data Sheets (MSDS) are available on request.
- Use of this product should be limited to personnel trained in the techniques of DNA amplification.
- Workflow in the laboratory must proceed in a uni-directional manner, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where you performed previous step.

SPECIMEN COLLECTION AND CONSERVATION

DNA/RNA Prep NA nucleic acid extraction kit is recommended for **RNA and DNA** extraction and purification from:

- *whole blood*;
- *serum*;
- *plasma*;
- *bone marrow aspirate*; *cerebrospinal fluid (liquor)*;
- *amniotic liquid*;
- *sinovial liquid*;
- *peritoneal and pleuric versament*;
- *tissue* homogenized with mechanical homogenizer and dissolved in PBS sterile;
- *urine (sediment)*;
- *prostatic liquid*; *seminal liquid*;
- *swabs;*
- *sputum:*
    - Collect sputum into 50 mL single-use PP tubes with a screw cap.
    - In a biological safety cabinet, homogenize samples after mixing with equal volume of 4% NaOH solution. (*N-acetyl-L-cysteine may be added if required in the amount of 50-70 mg per sample).* Mix intensely with a tube rotator for 5-20 minutes (depending on the density of a sample).
    - Centrifuge samples at 3000 rpm (2800-3000 g) for 15 min and carefully discard the supernatant leaving 500-1000 µl in the tube. Resuspend sediment and transfer it into a 1.5 ml tube.
    - Centrifuge samples at 12000 rpm for 5-10 min, discard the supernatant and use the same 1,5 ml sample tube for DNA isolation from sample sediment.
- *bronco aspirate:* transfer 1,0 ml to a polypropylene tube (1,5 ml) and centrifuge at 10000g/min for 10 min. Discard the supernatant and leave about 100 µl of solution for DNA extraction;
- *feces:*
    - Prepare 10-20% feces suspension, for instance adding 4ml of Saline Solution and 1,0 gr (approx. 1,0 ml) of feces in 5 ml tube (the same can be done in 2,0 ml tube). The DNA/RNA purification must be done immediately, if it is not possible add 20% Glycerol sterile solution (cryoprotective agent that provides intracellular and extracellular protection against freezing) and store at -20°C.
    - Vortex to get an homogeneous suspension and centrifuge for 5 min to 7000-12000g. Use the supernatant for the extraction of the viral DNA/RNA and the bacterial fraction (white-yellowish line between the sediment and the supernatant) for the extraction of bacterial DNA.

All kind of biological fluids or semi-fluid samples can be processed. For successful nucleic acid purification, it is important to obtain a homogeneous, clear and non-viscous sample before loading into the corresponding isolation tube. Therefore, check all samples (especially old or frozen ones) for the presence of precipitates.

Note: Handle all specimens as if they are potentially infectious agents.

It is recommended to process samples immediately after collection. Store samples at 2–8 °C for no longer than 24 hours, or freeze at –20/80°C. Transportation of clinical specimens must comply with country, federal, state and local regulations for the transport of etiologic agents.

STORAGE CONDITIONS AND PREPARATION OF WORKING SOLUTIONS

DNA/RNA Prep NA kit should be stored dry at +2-8°C; storage at higher temperatures should be avoided. If crystals are observed Sol Lys reagent bottle upon opening, allow the reagent to equilibrate at room temperature until the crystals disappear or prewarm at 60°C for a maximum of 5 min.

STABILITY

DNA/RNA Prep NA is stable up to the expiration date indicated on the kit label. The product will maintain performance through the control date printed on the label. All components of the DNA/RNA Prep NA nucleic acid extraction kit are stable until labeled expiration date. The shelf life of reagents before and after the first use is the same, unless otherwise stated**.** Components stored under conditions other than those stated on the labels may not perform properly and may adversely affect the assay results.

PROTOCOL

1. Prepare required number of 1.5 ml disposable polypropylene micro centrifuge tubes including one tube for Negative Control of Extraction (**Negative Control**, **C-**) and one tube for Positive Control of Extraction (**Positive Control** (RNA or DNA), if provided with the amplification kit).
2. Add to each tube **10 µl** of **Internal Control** (if provided with the amplification kit) and **300 µl** of **Lysis Sol**
3. Add **100 µl** of samples to the appropriate tubes using pipette tips with aerosol barriers.
4. Prepare Controls as follows:
    - add **100 µl** of **C–** (**Neg Control** provided with the amplification kit) to the tube labeled C*neg*
    - add **90 µl** of **Negative Control** (provided with the amplification kit) and **10 µl** of **Positive Control** to the tube labeled C*pos.*
5. Vortex the tubes and incubate for 15 min at 65°C. Centrifuge for 7-10 sec.
6. Add **400 µl** of **Prec Sol** and mix by vortex. Centrifuge all tubes at 13,000 r/min for 5 min and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between the tubes.
7. Add **500 µl of Wash Sol 3** into each tube. Vortex vigorously to ensure pellet washing. Centrifuge all tubes at 13,000 r/min for 2 min and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between the tubes.
8. Add **300 µl of Wash Sol 4** into each tube. Vortex vigorously to ensure pellet washing. Centrifuge all tubes at 13,000 r/min for 2 min and using a micropipette with a plugged aerosol barrier tip, carefully remove and discard supernatant from each tube without disturbing the pellet. Change tips between the tubes.
9. Incubate all tubes with open caps at **65 °C for 5 min**.
10. Resuspend the pellet in **50 µl of RE-buffer** (elution volume can be increased up to 90 µl). Incubate for 5 min at 65°C and vortex periodically.
11. Centrifuge the tubes at 13000g for 60 sec.

The supernatant contains RNA/DNA ready for amplification. If amplification is not performed the same day of extraction, the processed samples can be stored at 2-8°C for at maximum period of 5 days or frozen at - 20°/-80°C.

TROUBLESHOOTING

These troubleshooting rules may be helpful in explaining any questions that may arise.

False negatives with extraction product:

- Degradation of the nucleic acid contained in the sample. It's necessary to use a new sample. Store samples under appropriate conditions. Use plastic free from DNAses and RNAses
- Loss of pellet. Carefully draw off the washing solutions and try not to remove the nucleic acid residue.

False positives with extraction product:

- Contamination during sample extraction. Open one test tube at time. Avoid spilling the contents of the test tube, always change tips. Use only filter tips during the extraction procedure. Change tips between tubes.
- Contamination of the reagents prepared for the step. Repeat the test with the new set of reagents.
- Contamination of the extraction zone by amplicons. Decontaminate all surfaces and instruments with sodium hypochlorite and ethanol, wash lab coats, replace test tubes and tips in use. Use different laboratory coats in different Amplification areas.

KEY TO SYMBOLS USED

	List Number		Caution!
	Lot Number		Contains sufficient for <n> tests
	For *in Vitro* Diagnostic Use		Version
	Store at		Expiration Date
	Manufacturer		Warning
	Consult instructions for use		

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