

Certificate of Approval

This is to certify that the Management System of:

ELITechGroup B.V.

Van Rensselaerweg 4, 6956 AV Spankeren, The Netherlands

has been approved by Lloyd's Register to the following standards:

ISO 13485:2016

Approval number(s): ISO 13485 – 00020722

This certificate is valid only in association with the certificate schedule bearing the same number on which the locations applicable to this approval are listed.

The scope of this approval is applicable to:

Design, development and manufacturing of clinical chemistry analyzers, contract manufacturing of erythrocyte sedimentation rate analyzers and warehousing of erythrocyte sedimentation rate tubes for the in vitro diagnostic investigation of samples of human origin.



Paul Graaf

Chief Operating Officer, Management Systems, MSIS

Issued by: Lloyd's Register Nederland B.V.

for and on behalf of: Lloyd's Register Quality Assurance Limited



001

Certificate Schedule

Location	Activities
ELITechGroup B.V. Van Rensselaerweg 4, 6956 AV Spankeren, The Netherlands	ISO 13485:2016 Design, development and manufacturing of clinical chemistry analyzers, contract manufacturing of erythrocyte sedimentation rate analyzers and warehousing of erythrocyte sedimentation rate tubes for the in vitro diagnostic investigation of samples of human origin.
ELITechGroup B.V. Kanaaldijk 90, 6956 AX Spankeren, The Netherlands	ISO 13485:2016 Design, development and manufacturing of clinical chemistry analyzers, contract manufacturing of erythrocyte sedimentation rate analyzers and warehousing of erythrocyte sedimentation rate tubes for the in vitro diagnostic investigation of samples of human origin.



001

DECLARATION DE CONFORMITE CE

Nous, ELITech Clinical Systems SAS, zone industrielle 61500 SEES France, déclarons sous notre seule responsabilité que les réactifs référencés dans la liste ci-jointe (2 pages), sont conformes aux exigences essentielles des annexes I et III de la Directive Européenne 98/79/CE relative aux dispositifs médicaux de diagnostic *in vitro* et au code de la santé publique.

Ces dispositifs sont classés dans la catégorie « autre dispositif » puisqu'ils n'appartiennent ni à la liste A et liste B de l'annexe II et ni à la classe des autotests.

Cette déclaration est basée sur le contenu de chaque dossier technique et s'appuie sur la certification de notre système qualité selon la norme NF EN ISO 13485 : 2016 (Certification valable jusqu'au 27 juillet 2023).

DECLARATION OF EC CONFORMITY

We, ELITech Clinical Systems SAS, Zone Industrielle 61500 SEES France, hereby certify, under our own responsibility, that the reagents such as listed attached (2 pages), conform to the essential requirements of appendices I and III of European Directive 98/79/EC, relating to in vitro diagnostic medical devices and to the public health code.

These devices are classified in the "other device" category since they do not belong neither to list A or list B of annex II nor to self-testing class.

This declaration is based on the contents of each technical file and is supported by the certification of our quality system according to the standard NF EN ISO 13485 : 2016 (Certification valid until July 27th, 2023).

DECLARACIÓN CE DE CONFORMIDAD

Nosotros, ELITech Clinical Systems SAS, Zone Industrielle 61500 SEES France, declaramos bajo nuestra única responsabilidad que los reactivos referenciados en la lista adjunta (2 páginas), son conformes con los requisitos esenciales de los anexos I y III de la Directiva Europea 98/79/CE sobre dispositivos médicos para diagnóstico in vitro y el código de salud pública.

Estos dispositivos se clasifican en la categoría "otro dispositivo", ya que no pertenecen a la lista A ni a la lista B del anexo II, tampoco a la clase de autodiagnóstico.

Esta declaración se basa en el contenido de cada expediente técnico y está respaldado por la certificación de nuestro sistema de calidad según la norma NF EN ISO 13485 : 2016 (Certificación válida hasta el 27 de Julio 2023).

Sées, le 12 Mai 2021

Valérie LAMBERT,

Responsable des Affaires Réglementaires

Regulatory Affairs Manager

Responsable de los Asuntos Reglementarios



ELITech Clinical Systems SAS

Zone Industrielle

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Cécile GOUBAULT,

Directeur Général Délégué

Managing Director

Directora General



Société par actions simplifiée au capital de 1.688.392,33 € – SIREN : 318 365 228 – RCS ALENCON

REACTIFS - REAGENTS - REACTIVOS	RÉFÉRENCES - REFERENCES - REFERENCIAS	Code GMDN
Metabolites divers / Miscellaneous metabolites		
ALBUMIN	ALBU-0600/0700/0250/M830	
ALBUMIN ENVOY	ALBU-0850	53597
BILIRUBIN DIRECT 4+1	BIDI-0600/0250	53233
BILIRUBIN TOTAL 4+1	BITO-0600/0250	53229
BILIRUBIN TOTAL & DIRECT 4+1	BITD-0600	53229/53233
CREATININE ENVOY	CRSL-0850	53250
CREATININE JAFFE	CRCO-0600/0700	53251
CREATININE PAP	CRSL-M490	
CREATININE PAP SL	CRSL-0630/0250	53250
DIRECT BILIRUBIN	BIDI-M430	53233
DIRECT BILIRUBIN ENVOY	BIDV-0850	53233
GLUCOSE ENVOY	GPST-0850	
GLUCOSE HK	GHSL-M490	
GLUCOSE HK SL	GHSL-0600/0250	53301
GLUCOSE PAP	GPST-M690	
GLUCOSE PAP SL	GPST-0507/0500/0707/0700/0250/0455/0497	
LACTATE	LACT-0100	53342
MICROPROTEIN PLUS	PRTU-0600/0250	53481
PHOSPHORUS	PHOS-0600/0230/M430	
PHOSPHORUS ENVOY	PHOS-0850	59123
TOTAL BILIRUBIN	BITO-M430	53229
TOTAL BILIRUBIN ENVOY	BITV-0850	53229
TOTAL PROTEIN	PROB-M830	
TOTAL PROTEIN ENVOY	PROB-0850	53985
TOTAL PROTEIN PLUS	PROB-0600/0700/0250	
UREA	URSL-M830	
UREA ENVOY	URSL-0850	53587
UREA UV SL	URSL-0427/0420/0500/0507/0250/0455	
URIC ACID	AUML-M830	
URIC ACID ENVOY	AUVD-0850	
URIC ACID MONO SL	AUML-0497/0427/0420/0500/0507/0707/0250	53583
URIC ACID SL	AUSL-0250	
URINE PROTEIN	PRTU-M230	53481
Enzymes / Enzymes		
ALP (DEA) SL	PASL-0400/0420/0230	
ALP ENVOY	PIVD-0850	52928
ALP IFCC	ALPI-0230	
ALT ENVOY	ALSL-0850	
ALT/GPT	ALSL-M490	
ALT/GPT 4+1 SL	ALSL-0410/0430/0510/0250/0455	52923
AMYLASE	AMSL-M430	
AMYLASE ENVOY	AMSL-0850	52940
AMYLASE SL	AMSL-0390/0400/0230	
AST/GOT	ASSL-M490	
AST ENVOY	ASVD-0850	52954
AST/GOT 4+1 SL	ASSL-0410/0430/0510/0250/0455	
CHOLINESTERASE	CHES-0053	52971
CK ENVOY	CKSL-0850	53003
CK-MB ENVOY	CMSL-0850	52994
CK-MB SL / CKMB	CMSL-0410/0430/0230	
CK NAC	CKSL-M230	
CK NAC SL	CKSL-0410/0430/0230	53003
GAMMA-GT	GISL-M230	
GAMMA-GT PLUS SL	GISL-0400/0420/0250	53027
GGT ENVOY	GISL-0850	
LDH ENVOY	LLSL-0850	
LDH IFCC	LLSL-M230	53072
LDH-L SL	LLSL-0400/0420/0230	
LIPASE	LPSL-0250	
LIPASE ENVOY	LPSL-0850	53108
LIPASE SL	LPSL-0230	
Electrolytes / Oligo-éléments / Electrolytes / Trace-elements		
CALCIUM ARSENAZO	CALA-0600/0250/M430	
CALCIUM ENVOY	CALA-0850	45789
CHLORIDE	CHLO-0600/0250	60037
IRON ENVOY	FEFE-0850	
IRON FERENE	FEFE-0230/0600/M230	54758
MAGNESIUM ENVOY	MAGX-0850	
MAGNESIUM XB	MGXB-0250/0600/M430	46795
MAGNESIUM XYLIDYL	MAGX-0230/0600	
Lipides / Lipids		
CHOLESTEROL	CHSL-M690	
CHOLESTEROL ENVOY	CHSL-0850	53359
CHOLESTEROL HDL SL 2G	HDLL-0230/0380/0390	53391
CHOLESTEROL LDL SL 2G	LDLL-0230/0380/0390	53395
CHOLESTEROL SL	CHSL-0507/0500/0700/0707/0250/0455/0497	53359
HDL CHOLESTEROL	CHDL-0250/0600/M330	
HDL CHOLESTEROL ENVOY	HDLL-0850	53391
LDL CHOLESTEROL	CLDL-0250/M330	
LDL CHOLESTEROL ENVOY	LDLL-0850	53395
TRIGLYCERIDES	TGML-M690	
TRIGLYCERIDES ENVOY	TGML-0850	
TRIGLYCERIDES MONO SL NEW	TGML-0427/0425/0515/0700/0517/0707/0497	53460
TRIGLYCERIDES SL	TGML-0250/0455	

Vla


REACTIFS - REAGENTS - REACTIVOS	RÉFÉRENCES - REFERENCES - REFERENCIAS	Code GMDN
Contrôles-Calibrants-Standards / Controls-Calibrators-Standards		
CHOLESTEROL HDL 2G CALIBRATOR	HDLL-0011/0041	44696
CHOLESTEROL LDL 2G CALIBRATOR	LDLL-0011/0041	41728
CHOLESTEROL Standard 200 mg/dL	CHOL-0055	44698
CK-MB CONTROL	CKMB-0900	44693
ELICAL 2	CALI-0550	47868
ELITROL I	CONT-0060	47869
ELITROL II	CONT-0160	
GLUCOSE Standard 100 mg/dL	GLUP-0055	41818
HDL LDL CALIBRATOR	HLCA-0041	47868
ISE CONTROL I	ISCT-0046	47869
ISE CONTROL II	ISCT-0047	
MICROPROTEIN PLUS Standard 100 mg/dL	PRTU-0022	53482
TRIGLYCERIDES Standard 200 mg/dL	TRIG-0055	44702
UREA Standard 50 mg/dL	URUV-0055	53588
URIC ACID Standard 6 mg/dL	ACUR-0055	44704
Protéines spécifiques / Specific proteins		
ANTI-STREPTOLYSIN O	ASLO-0250	59055
CRP IP	ICRP-0400/M230	53705
CRP IP CALIBRATOR SET	ICRP-0043	41838
CRP IP CONTROL I	ICRP-0046	41839
CRP IP CONTROL II	ICRP-0047	
CRP WR	CRPW-0230	53705
CRP WR CALIBRATOR SET	CRPW-0043	41838
CRP WR CONTROL	CRPW-0045	41839
CRP WR ENVOY	CRPW-0850	53705
FERRITIN	IFRT-0230	53718
FERRITIN CALIBRATOR	IFRT-0042	41927
HAPTOGLOBIN IP	IHAP-0400	53737
HbA1c	HBAC-0240	59090
HbA1c CALIBRATOR SET	HBAC-0043	53315
HbA1c CONTROL L + H	HBAC-0049	44435
IgA IP	IIGA-0400	53760
IgG IP	IIGG-0400	53787
IgM IP	IIGM-0400	53795
µALBUMIN IP	IMAL-0400	53475
µALBUMIN IP CALIBRATOR SET	IMAL-0043	53477
µALBUMIN IP CONTROL I	IMAL-0046	53478
µALBUMIN IP CONTROL II	IMAL-0047	
OROSOMUCOID IP	IORO-0400	53606
PREALBUMIN IP	IPAL-0400	53957
PROTEIN IP CALIBRATOR SET	IPRO-0043	53593
RF CALIBRATOR	IRFA-0042	42230
RHEUMATOID FACTOR	IRFA-0230	55111
RHEUMATOLOGY CONTROL I	IRCT-0046	47869
RHEUMATOLOGY CONTROL II	IRCT-0047	
TRANSFERRIN IP	ITRF-0400	59041
Vitamines/Vitamins		
VITAMIN D	VITD-0250	54476
VITAMIN D CALIBRATOR SET	VITD-0043	54474
VITAMIN D CONTROL SET	VITD-0049	54475
ISE Solutions pour électrodes selectives d'ions / ISE Solutions for ion-selective electrodes		
ISE BASELINE SOLUTION ENVOY	ISBA-0850	59238
ISE CALIBRATORS	ISCA-0250	52867
ISE CALIBRATOR ENVOY	ISCV-0850	
ISE CLEANER/CONDITIONER	ISCC-0280	59058
ISE DILUENT	ISDI-0250	58237
ISE DILUENT ENVOY	ISDV-0850	
ISE REFERENCE SOLUTION	ISRS-0800	59238
ISE REFERENCE SOLUTION ENVOY	ISRS-0850	
Solutions de lavage pour les équipements ELITech Clinical Systems / Cleaning solutions for ELITech Clinical Systems Equipments		
ACID SOLUTION for ELITech Clinical Systems Analyzers	SLHC-5900	59058
SYSTEM CLEANING SOLUTION for ELITech Clinical Systems Analyzers	SLNA-5900	59058
SYSTEM SOLUTION	SLSY-5905	58236
SYSTEM SOLUTION for ELITech Clinical Systems Analyzers	SLSY-5900	
WASH SOLUTION A	SOLA-M163	59058
WASH SOLUTION B	WASH SOLUTION B	59058
Tests d'agglutination / Agglutination tests		
CRP LATEX	LXCR-0112	53707

Vla
CG

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To: Whom it May Concern

Regulatory status of parts & accessories

As mentioned on the current Declarations of Conformity of our Clinical Chemistry Analyzers also the accessories conform to the provisions of the EU Directive on In Vitro Diagnostic Medical Devices (98/79/EC). This applies to the parts and accessories as mentioned in the attached list.

'IVD accessory' means an article which, whilst not being an IVD medical device, is intended specifically by its manufacturer to be used together with an IVD device to enable that IVD device to be used in accordance with its intended purpose.

ELITechGroup B.V.



Adriaan P. Intveld
Manager Quality Assurance & Regulatory Affairs

Part number	Description	IVD medical device	IVD accessory	general laboratory use	spare part	supporting part
1540-001	Anti-Slip sheet					✓
2206-007	Cooling Liquid (1 L)					✓
3062-021	Sample cup (1000 pcs)		✓			
3062-033	Sample tube 6 ml (500 pcs)					✓
3062-040	Water container 10 L					✓
3062-041	Water container 5 L					✓
3066-155	Syringe 100 µl		✓			
3066-156	Syringe 1 ml		✓			
3069-040	Keyboard Dust cover					✓
3069-047	Keyboard Dust cover					✓
3070-518	Cap holder					✓
3070-538	Cap rotor Left					✓
3070-539	Cap rotor right					✓
3201-002	Dichromate 8 Abs (25ml)		✓			
3365-192	USB Stick					✓
3374-003	Mains cable (USA)					✓
3374-059	Pumpunit cable		✓			
3374-066	Mains cable					✓
3374-097	Serial Null-modem cable					✓
3374-286	USB Extension cable					✓
4804-038	Reagent identification Disc					✓
6001-826	Diluted Waste container		✓			
6001-827	Concentrated Waste container		✓			
6001-860	Water container		✓			
6001-861	Tube assy (analyser)		✓			
6001-872	Tube assy (cooling unit)		✓			
6002-102	Assorter unit				✓	
6002-386	System software on CD		✓			
6002-706	Reaction Rotor set (3 pcs)		✓			
6002-726	System Disc		✓			
6002-817	Bottle 30 ml (20 pcs)		✓			
6002-818	Bottle 15 ml (20 pcs)		✓			
6002-904	Water container 5 L		✓			
6002-910	Assorter unit				✓	
6002-913	External tubing		✓			
6003-074	System software on USB stick		✓			
6003-444	Diluted Waste Container 5 L		✓			
6003-466	Keyboard Support option					✓
6003-797	CW Waste Container 2 L		✓			
6003-808	Assorter unit				✓	

Instrument Training

Vital Scientific BV hereby declares that the participant has attended a four days seminar for service engineers and the participant is now a certified engineer for the declared instruments.

Participant: Mr. S. Sorocovici

Company: Global Biomarketing Group-Moldova SRL
Moldova

Instrument: Vitalab: XL Series
E Series
Junior Series
Dry ISE
Micro Series
ProXS

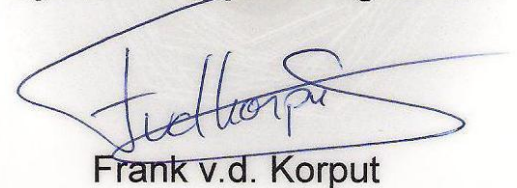
Date of training: April 20th – April 23rd, 2010

System Support Manager:



Jan Oostendorp

System Support Engineer:



Frank v.d. Korput

☞Referințe:
ALBU-0250
ALBU-0600
ALBU-0700

Compoziția trusei:
R 12 x 20 mL
R 2 x 125 mL + Std 1 x 2 mL
R 4 x 250 mL + Std 1 x 2 mL



FTRO-ALBU-v18 (09/2020)_PIT-ALBU-4-v18

☞SCOPUL UTILIZĂRII

ELITech Clinical Systems ALBUMIN este un reactiv de diagnostic *in vitro* destinat determinării cantitative a albuminei din probele serul uman și plasmă.

☞SEMNIIFICAȚIE CLINICĂ⁽¹⁻³⁾

Albumina este sintetizată în special de ficat și reprezintă aproximativ 50% din proteinele plasmatică. Funcția principală a albuminei este menținerea presiunii oncotice și transportul unui număr mare de compuși. Măsurarea albuminei serice sau plasmatică este indicată în special pentru a ajuta la diagnosticarea și monitorizarea bolilor cu pierdere de proteine sau sinteză scăzută (sindromul nefrotic, pierderea gastrointestinală, insuficiența hepatică), inflamația acută și cronică, și malnutriția severă.

☞METODĂ⁽⁴⁾

Verde de bromocresol (BCG). Punct final

PRINCIPIU⁽⁴⁾

Determinarea colorimetrică a albuminei utilizând verdele de bromocresol la pH 4,20.

Albumină + BCG $\xrightarrow{pH=4,20}$ Complex albumină - BCG

☞COMPOZIȚIA REACTIVULUI

Reactiv: R

Tampon succinat, pH 4,20

Verde de bromocresol 0,2 mmol/L

Surfactant

Standard: Std. (Ref.: ALBU-0600/0700)

Albumină bovină 3,5 g/dL

35 g/L

Azidă de sodiu < 0,1 %

☞MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

☞AVERTISMENTE ȘI PRECAUȚII

- Acest dispozitiv de diagnostic *in vitro* (Reactiv și Standardul) este destinat numai pentru uz profesional.
- Standardul conține azidă de sodiu care poate reacționa cu plumbul sau instalațiile din cupru pentru a forma potențiale azide metalice explozive. În momentul eliminării acestui standard, spălați întotdeauna cu apă din abundență pentru a preveni acumularea de azide.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Standardul trebuie să fie imediat închis cu capacul pentru a preveni contaminarea și evaporarea.
- Pentru mai multe informații, Fișa de date privind siguranța (SDS) este disponibilă la cerere pentru utilizatorul profesional.

☞STABILITATEA

A se depozita la 2-25°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii și Standardul sunt gata pentru utilizare.

☞DETERIORAREA PRODUSELOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

☞PROBE

Specimen⁽²⁾

- Ser.
- Plasmă (heparină de litiu)
- A nu se utiliza alte specimene.

Avertismente și precauții

Conform bunei practici de laborator, puncția venoasă trebuie efectuată înainte de administrarea de medicamente.

Depozitare⁽¹⁾

- Analizați serul proaspăt sau stocați-le la 2-8°C mai puțin de 72 de ore.
- Stocate la -20°C, probele sunt stabile 6 luni. Pentru o depozitare mai îndelungată, probele sunt înghețate la -70°C.

☞VALORI DE REFERINȚĂ⁽¹⁾

Ser, plasmă:

Pacienții în repaus

< 60 ani: 3.5 – 5.2 g/dL (35 – 52 g/L)

60-90 ani: 3.2 – 4.6 g/dL (32 – 46 g/L)

>90 ani: 2.9 – 4.5 g/dL 29 – 45 g/L)

În cazul adulților din ambulatoriu, valorile pot fi mai mari cu 0,3 - 0,5 g/dL (3 - 5 g/L).

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.



ALBUMIN

☞ Referințe:
ALBU-0250
ALBU-0600
ALBU-0700

Compoziția trusei:
R 12 x 20 mL
R 2 x 125 mL + Std 1 x 2 mL
R 4 x 250 mL + Std 1 x 2 mL



FTRO-ALBU-v18 (09/2020)_PIT-ALBU-4-v18

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems, aplicațiile sunt disponibile la cerere.
Lungime de undă 620 nm
Temperatură: 37°C
Citiți pe reactivul martor.

	MARTOR	CALIBRARE	TEST
Reactiv R	360 µL	360 µL	360 µL
Apă distilată	3 µL	-	-
Calibrator/ Standard	-	3 µL	-
Probă	-	-	3 µL

Amestecați și citiți absorbanțele (A) după o incubare de 4 minute și 30 de secunde.

- Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

☞ CALCUL

A Sample
_____ x n n = calibrator/standard concentration
A Standard/
Calibrator

Factor de conversie: g/dL x 10 = g/L

☞ CALIBRARE

Pentru referința ALBU-0600/0700: Pentru calibrare, trebuie utilizat fie calibratorul multiparametric ELICAL 2 fie Standardul Albumin 3.5 g/dL.
Pentru referința ALBU-0250: Pentru calibrare, utilizați calibratorul multiparametric ELICAL 2.

Valorile concentrației Standardului Albumin 3.5 g/dL și calibratorului multiparametric ELICAL 2 sunt trasabile conform materialului de referință ERM-DA 470k.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

☞ CONTROLUL CALITĂȚII

Pentru a verifica precizia testelor, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

☞ MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

☞ DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems

- **Interval de măsurare**
Determinat conform protocolului CLSI EP6-A⁽⁵⁾, intervalul de măsurare este între 1.6 și 6.0 g/dL (de la 16 la 60 g/L).

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁶⁾.
LoD= 0.003 g/dL (0,03 g/L).
LoQ= 0.50 g/dL (5.0 g/L).

- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁷⁾.

	n	Medie		În interiorul ciclului	Total
		g/dL	g/L		
Nivelul 1	80	2.54	25.4	0.9	2.3
Nivelul 2	80	3.53	35.3	0.5	2.1
Nivelul 2	80	4.98	49.8	0.8	2.1

- Corelație

A fost efectuat un studiu comparativ între un analizor ELITech Clinical Systems ProM și un sistem similar disponibil în comerț (metoda BCG) pe eşantioane de ser uman 100 determinată conform protocolului CLSI EP9-A2⁽⁸⁾.

Concentrațiile eşantioanelor au fost între 1.43 și 5.89 g/dL (14.3 – 58.9 g/L).

Rezultatele sunt după cum urmează:

Coeficient de corelație: (r)=0.997

Regresie liniară: y = 0.961x + 0.12 g/dL (1.2 g/L)

- Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽⁹⁾. Recuperarea este în intervalul ±10% din valoarea inițială a activității Albumina de 3.5 și 5.00 g/dL.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 µmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 µmol/L).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL.

Trigliceride: Nicio interferență semnificativă până la 3000 mg/dL (33.90 mmol/L).

Acid ascorbic: Nicio interferență semnificativă până la 20.0 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

Gamma globulină: Nicio interferență semnificativă până la 1500 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglubulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹⁰⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹¹⁻¹²⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

ALBUMIN

☞ Referințe:
 ALBU-0250
 ALBU-0600
 ALBU-0700

Compoziția trusei:
 R 12 x 20 mL
 R 2 x 125 mL + Std 1 x 2 mL
 R 4 x 250 mL + Std 1 x 2 mL



FTRO-ALBU-v18 (09/2020)_PIT-ALBU-4-v18

- **Stabilitatea la bord/Frecvența calibrării**

Stabilitatea la bord: 28 zile

Frecvența calibrării: 28 zile





Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

☞ **BIBLIOGRAFIE**

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5. Evaluation of the Linearity of the Measurement of Quantitative Procedures: a Statistical Approach; Approved Guideline. CLSI (NCCLS) document EP6-A (2003), **23** (16).
6. Protocols for Determination of Limits of Detection and Limits of Quantification; Approved Guideline. CLSI (NCCLS) document EP17-A (2004), **24** (34).
7. Evaluation of Precision Performance of Quantitative Measurement Methods; Approved Guideline—Second Edition. CLSI (NCCLS) document EP5-A2 (2004), **24** (25).
8. Method Comparison and Bias estimation Using Patient Samples; Approved Guideline—Second Edition. CLSI (NCCLS) document EP9-A2 (2002), **22** (19).
9. Interference Testing in Clinical Chemistry ; Approved Guideline—Second Edition. CLSI (NCCLS) document EP7-A2 (2005), **25**(27).
10. Berth, M. & Delanghe, J. Protein precipitation as a possible important pitfall in the clinical chemistry analysis of blood samples containing monoclonal immunoglobulins: 2 case reports and a review of literature, Acta Clin Belg., (2004), **59**, 263.
11. Young, D.S., Effects of preanalytical variables on clinical laboratory tests, 2nd edition, AACC Press (1997).
12. Young D.S., Effects of drugs on clinical laboratory tests, 4th edition, AACC Press (1995).

☞ **SIMBOLURI**

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

	Conținut
	Reactiv
	Standard
	Conformitate europeană
☞	Modificare față de versiunea precedentă

Notă

Doar pentru ref. **ALBU-0250**, utilizate cu software-ul Selectra TouchPro.



Albumin. 0
 101 PIT-ALBU

Referințe:

ALSL-0250	8 x 25 mL
ALSL-0455	4 x 55 mL
ALSL-0410	2 x 62,5 mL
ALSL-0430	4 x 62,5 mL
ALSL-0510	5 x 125 mL

Compoziția trusei:

R1	8 x 20 mL + R2 8 x 5 mL
R1	4 x 44 mL + R2 4 x 11 mL
R1	2 x 50 mL + R2 1 x 26 mL
R1	4 x 50 mL + R2 2 x 26 mL
R1	5 x 100 mL + R2 1 x 127 mL



FTRO-ALSL4+1-v18(12/2018)_PIT-ALSL4+1-4-v18

SCOPUL UTILIZĂRII

ALT/GPT 4+1 SL ELITech Clinical Systems este conceput pentru determinarea cantitativă de diagnosticare *in vitro* a alaninaminotransferazei (ALT) în serul uman și plasmă.

SEMNIFICAȚIE CLINICĂ ^(1-3,6)

Alaninaminotransferaza (ALT), cunoscută și ca glutamat piruvat transferaza (GPT), este o transaminază. ALT catalizează transferul grupului amino al L-alaninei la α -ketoglutarat pentru a rezulta L-glutamatul. Cele mai mari nivele se găsesc în ficat și rinichi, și cele mai mici cantități în inimă și mușchii scheletici. Concentrația ALT este crescută când celulele hepatice sunt deteriorate (necroză celulară hepatică sau leziune de orice cauză). Într-adevăr, hepatita virală și toxică induce o creștere marcată a activității ALT în ser. Aportul de alcool, delirium tremens, și administrarea diverselor medicamente induc creșterea ușoară sau moderată a ALT. Concentrația ALT în ser este, de asemenea, crescută ușor în diverse afecțiuni precum: distrofia musculară, boala hemolitică, infarctul miocardic...

ALT este mai specifică ficatului decât AST (aspartataminotransferaza). Măsurarea atât a AST, cât și a ALT are o anumită valoare în distingerea hepatitei de alte leziuni parenchimale.

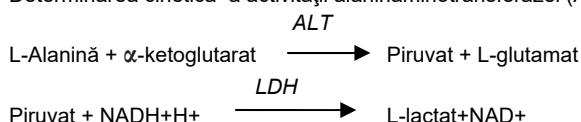
Nivelul ALT din ser poate fi scăzut în cazul deficienței de vitamina B6.

METODĂ ⁽⁴⁾

Metoda IFCC fără piridoxal fosfat (P-5'-P). cinetică. UV.

PRINCIPIU ⁽⁴⁾

Determinarea cinetică a activității alaninaminotransferazei (ALT).



LDH=lactat dehidrogenază

COMPOZIȚIA REACTIVULUI

Reactiv 1: R1

Tampon Tris, pH 7,5 (30°C)	125 mmol/L
L-alanină	680 mmol/L
LDH	≥ 2000 U/L
Azidă de sodiu	< 0,1 %

Reactiv 2: R2

□ Ketoglutarat	97 mmol/L
NADH	1,1 mmol/L
Azidă de sodiu	< 0,1 %

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550, ELICAL 2
- CONT-0060, ELITROL I
- CONT-0160, ELITROL II
- Echipamente generale de laborator.
- Soluție salină obișnuită (NaCl 9 g/L).
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest reactiv este conceput doar pentru utilizarea în scopul diagnosticării *in vitro*.
- Acești reactivi conțin azidă de sodiu care poate reacționa cu plumbul sau instalațiile sanitare din cupru și poate forma azide metalice explozibile. În cazul aruncării acestor reactivi, spălați întotdeauna cu cantități mari de apă pentru a preveni formarea de azide.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați fiolele de reactiv din truse diferite.
- Pentru mai multe informații, consultați Fișa de date privind siguranța (SDS).

STABILITATEA REACTIVILOR

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA REACTIVILOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.

AMBALAJ DETERIORAT

Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, recipient perforat).

PROBE ^(1,5)

Specimen

- Ser și plasmă heparinizată de litiu, libere de hemoliză.
- A nu se utiliza alte specimene.

Avertisment și precauții

Conform buneii practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.



Referințe:

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ALSL-0430	4 x 62,5 mL
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Compoziția trusei:

R1	8 x 20 mL + R2 8 x 5 mL
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R1	4 x 50 mL + R2 2 x 26 mL
R1	5 x 100 mL + R2 1 x 127 mL



FTRO-ALSL4+1-v18(12/2018)_PIT-ALSL4+1-4-v18

Depozitare

Eșantioanele sunt stabile timp de 3 zile la temperatura camerei și 7 zile la 2-8°C. Stabilitatea ALT este menținută mai bine la -70°C.

VALORI DE REFERINȚĂ ^(1,4)

Bărbați: ≤ 45 U/L

Femei: ≤ 34 U/L

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems.

Aplicațiile sunt disponibile la cerere

Lungime de undă 340 nm

Temperatură 37°C

Citiți pe reactivul maror.

Reactiv R1	240 μL
Proba	15 μL

Amestecați și așteptați 4 minute și 43 de secunde incubația.

Reactiv R2	60 μL
-------------------	-------

Amestecați și așteptați o incubație de 50 de secunde, măsurați modificarea absorbției pe minut (ΔA/min.) timp de 159 de secunde.

- Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

- Valorile mari ale ALT pot induce rezultate scăzute în mod fals datorită golirii substratului (consumul total de NADH înainte de citirea rezultatului). Pentru Analizoarele Selectra pro Series ELITech Clinical Systems, aplicația conține o alarmă specifică pentru a avertiza utilizatorii.

CALCUL

(ΔA) Proba x n n=concentrație calibrator

(ΔA) Calibrator

Factor de conversie: U/L x 0,0167 = μkat/L

CALIBRARE

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este trasabilă conform metodei de referință IFCC ⁽⁶⁾.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați și DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a asigura calitatea adecvată, vor fi utilizate serurile de control precum ELITROL I (control normal) și ELITROL II (control patologic). Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie

să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele locale și legale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems
- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁷⁾, intervalul de măsurare este între 10,0 și 450,0 U/L (de la 0,17 la 7,50 μkat/L). Probele care depășesc 450,0 U/L trebuie să fie diluate 1:10 cu soluție de NaCl 9 g/L (salină normală) și re-analizate. Utilizarea acestei proceduri extinde intervalul de măsurare de la 250,0 la 4500,0 U/L (de la 7,50 la 75,00 μkat/L).

Pentru utilizatorii Selectra TouchPro, funcția „reluare diluare” efectuează diluția automată a eșantionului. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁸⁾.

LoD= 2,9 U/L (0,05 μkat/L).

LoQ= 5,0 U/L (0,08 μkat/L).

Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁹⁾.

	n	Medie		În interiorul ciclului	Total
		U/L	μkat/L	CV (%)	
Nivel scăzut	80	34,2	0,57	1,1	4,4
Nivel mediu	80	71,2	1,19	1,2	2,9
Nivel înalt	80	367,0	6,12	0,5	1,8

- Corelație

A fost efectuat un studiu comparativ între Analizorul Selectra ProM ELITech Clinical Systems și un alt echipament al unui sistem aprobat de FDA (metoda IFCC) pe 100 de eșantioane de ser uman conform protocolului CLSI EP9-A2⁽¹⁰⁾.

Valorile acoperă domeniul de măsurare.

Parametrii regresiei liniare sunt după cum urmează:

Coeficient de corelație: (r)= 0,996

Regresie liniară: y= 1,017x + 0,6 U/L (0,01 μkat/L)

Referințe:

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ALSL-0410	2 x 62,5 mL
ALSL-0430	4 x 62,5 mL
ALSL-0510	5 x 125 mL

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
R1 4 x 44 mL + R2 4 x 11 mL
R1 2 x 50 mL + R2 1 x 26 mL
R1 4 x 50 mL + R2 2 x 26 mL
R1 5 x 100 mL + R2 1 x 127 mL



FTRO-ALSL4+1-v18(12/2018)_PIT-ALSL4+1-4-v18

- Limitări și interferențe

- Eșantioanele hemolizate nu trebuie să fie utilizate deoarece hemoliza semnificativă poate crește concentrația ALT din cauza nivelului ridicat de ALT în eritrocite.

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2 ⁽¹¹⁾ al CLSL. Recuperarea este în intervalul ±10% din valoarea inițială a lipazei de 35,0 și 350,0 U/L.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 μmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 μmol/L).

Trigliceride: Nicio interferență semnificativă până la 2300 mg/dL (25,99 mmol/L).

Piruvat: Nicio interferență semnificativă până la 3,0 mg/dL.

Acid ascorbic: Nicio interferență semnificativă până la 20 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200,0 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglubulinemia Waldenstrom) poate duce la rezultate nefiababile. ⁽¹²⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young și Glick. ⁽¹³⁻¹⁴⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 28 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit, și după o operație de întreținere.

BIBLIOGRAFIE

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4. Schumann, G., et al. *Clin Chem Lab Med*, (2002), **40**, 718.
5. Guder, W.G., et al., *Use of anticoagulants in diagnostic laboratory investigations*, WHO/DIL/LAB/99.1 Rev.2, (2002), 21.
6. Ward, M.K., Cockayne, S., *Enzymology. Clinical Chemistry : Concepts and Application*, Anderson, S.C., Cockayne, S. (W.B.Saunders eds. Philadelphia USA), (1993), 238.
7. Evaluation of the Linearity of the Measurement of Quantitative Procedures: a Statistical Approach; Approved Guideline. CLSI (NCCLS) document EP6-A (2003), **23** (16).

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9. Evaluation of Precision Performance of Quantitative Measurement Methods; Approved Guideline—Second Edition. CLSI (NCCLS) document EP5-A2 (2004), **24** (25).

10. Method Comparison and Bias estimation Using Patient Samples; Approved Guideline—Second Edition. CLSI (NCCLS) document EP9-A2 (2002), **22** (19).












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13. Young, D. S., *Effects of preanalytical variables on clinical laboratory tests*, 2nd Ed., AACC Press, (1997).

14. Young, D. S., *Effects of drugs on clinical laboratory tests*, 4th Ed., AACC Press, (1995)

SIMBOLURI

	Dispozitiv medical de diagnosticare in vitro.
	Consultați instrucțiunea de utilizare.
	Producător
	Limită de temperatură
	Număr de lot
	Data expirării
	Număr catalog
	Conținut
	Reactiv 1
	Reactiv 2
	Conformitate europeană

ALT/GPT 4+1 SL

Referințe:

ALSL-0250	8 x 25 mL
ALSL-0455	4 x 55 mL
ALSL-0410	2 x 62,5 mL
ALSL-0430	4 x 62,5 mL
ALSL-0510	5 x 125 mL

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
R1 4 x 44 mL + R2 4 x 11 mL
R1 2 x 50 mL + R2 1 x 26 mL
R1 4 x 50 mL + R2 2 x 26 mL
R1 5 x 100 mL + R2 1 x 127 mL



FTRO-ALSL4+1-v18(12/2018)_PIT-ALSL4+1-4-v18


Notă

Doar pentru ref. **ALSL-0250/ALSL-0455**, utilizată cu software-ul Selectra TouchPro.



ALT (GPT)
140

1
PIT-ALSL

:Modificare față de versiunea precedentă.

AMYLASE SL

Referințe:

AMSL-0230	6 x 20 mL
AMSL-0390	1 x 50 mL
AMSL-0400	6 x 50 mL

Compoziția trusei:

R 6 x 20 mL
R 1 x 50 mL
R 6 x 50 mL



FTRO-AMSL-v18 (12/2018)_PIT-AMSL-4-v18

SCOPUL UTILIZĂRII

AMYLASE SL ELITech Clinical Systems este conceput pentru determinarea cantitativă a amilazei în serul uman și plasmă pentru diagnosticare *in vitro*.

SEMNIFICAȚIE CLINICĂ⁽¹⁻²⁾

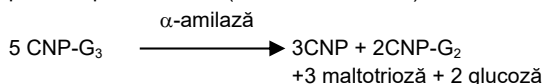
α -amilaza este o enzimă de origine pancreatică sau salivară care hidrolizează legăturile 1,4- α -glucozidice, ajutând astfel la digestia amidonului. Analiza amilazei serice este utilizată în special în diagnosticarea bolilor pancreatice (pancreatită acută sau cronică și complicațiile acestora, carcinoame). În timpul pancreatitei acute, se observă o creștere tranzitorie a amilazei serice, un vârf corespunzând unei creșteri de 4-6 ori fiind obținut în 12-72 de ore de la începere, activitatea revenind la normal după 3-5 zile. Cu toate acestea, o creștere a amilazei serice este de asemenea observată în alte patologii intra-abdominale, insuficiență renală, cancere ovariene, leziuni ale glandelor salivare, alcoolism acut, insuficiență renală sau macroamilazemie (prezența unui complex amilază-IgG nefiltrat de glomerul).

METODĂ⁽³⁾

Substrat: CNP-G₃ (2-cloro-4-nitrofenil- α -maltotriozidă)
 Enzimatică, cinetică

PRINCIPIU⁽³⁾

Substratul CNP-G₃ este hidrolizat prin acțiunea catalitică a α -amilazei pentru a produce CNP (2-cloro-4-nitrofenol).



CNP-G₂ = 2-cloro-4-nitrofenil- α -maltozidă

Rata de creștere a absorbanței este măsurată la 405 nm și este direct proporțională cu activitatea α -amilazei în probă,

COMPOZIȚIA REACTIVULUI

Reactiv: R

Tampon MES, pH 6,15	50	mmol/L
Clorură de sodiu	70	mmol/L
Clorură de calciu	6	mmol/L
Tiocianat de potasiu	900	mmol/L
CNP-G ₃	2,27	mmol/L
Azidă de sodiu	< 0,1	%

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2 4 x 3 mL
- CONT-0060 ELITROL I 10 x 5 mL
- CONT-0160 ELITROL II 10 x 5 mL
- Echipamente generale de laborator.
- Soluție salină obișnuită (NaCl 9 g/L).
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Această trusă de reactiv este concepută doar pentru utilizarea profesională în scopul diagnosticării *in vitro*.
- În contact cu acizi, degajă un gaz toxic.
- Reactivul conține azidă de sodiu, care poate reacționa cu plumbul sau instalațiile sanitare din cupru și poate forma azide metalice explozibile. În cazul aruncării acestor reactivi, spălați întotdeauna cu cantități mari de apă pentru a preveni formarea de azide.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Saliva și transpirația conține amilază. Astfel, se recomandă purtarea mănușilor și a unei măști pentru a evita contaminarea reactivului.
- Fișa cu date de securitate disponibilă la cerere.

STABILITATEA REACTIVILOR

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.
 (Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA REACTIVILOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.

AMBALAJ DETERIORAT

Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiole perforate).

PROBE⁽⁴⁾

Specimen

- Ser
- Plasmă heparinizată cu litiu
- A nu se utiliza alte specimene.

Avertisment și precauții

Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.

Depozitare și stabilitate

Probele sunt stabile timp de 1 săptămână la temperatura camerei, 1 săptămână la 2-8°C, și 1 an la -20°C.

VALORI DE REFERINȚĂ⁽⁵⁾

Ser, plasmă (37°C): 31-107 U/L

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.



AMYLASE SL

Referințe:

AMSL-0230	6 x 20 mL
AMSL-0390	1 x 50 mL
AMSL-0400	6 x 50 mL

Compoziția trusei:

R 6 x 20 mL
R 1 x 50 mL
R 6 x 50 mL

FTRO-AMSL-v18 (12/2018)_PIT-AMSL-4-v18



PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems.

Aplicațiile sunt disponibile la cerere
Lungime de undă 405 nm
Temperatură 37°C
Citiți pe reactivul martor.

Reactiv R	300 µL
Probă	3 µL

Amestecați și după o incubare de 50 de secunde, măsurați modificarea absorbantei pe minut ($\Delta A/\text{min.}$) timp de 159 de secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

CALCUL

$(\Delta A) \text{ Probă} \times n$ n=concentrație calibrator
 $(\Delta A) \text{ Calibrator}$

Factor de conversie: U/L x 0,0167 = $\mu\text{kat/L}$

CALIBRARE ⁽⁵⁾

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este trasabilă conform metodei IFCC.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a asigura precizia analizelor, vor fi utilizate serurile de control precum ELITROL I (control normal) și ELITROL II (control patologic). Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare și trebuie adaptată procedurilor de Controlul Calității fiecărui laborator și cerințelor reglementarilor în vigoare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele locale și legale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems

- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁶⁾, intervalul de măsurare este între 20 și 1500 U/L (de la 0,33 la 25,00 $\mu\text{kat/L}$). Probele care depășesc 1500 U/L trebuie să fie diluate 1:10 cu soluție de NaCl 9 g/L (salină normală) și re-analizate. Utilizarea acestei proceduri extinde intervalul de măsurare de la 1500 la 15000 U/L (de la 25,00 la 250,00 $\mu\text{kat/L}$).

Pentru utilizatorii Selectra TouchPro, funcția „diluare” efectuează diluția automată a probei. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁷⁾.
LoD = 6 U/L (0,10 $\mu\text{kat/L}$).
LoQ = 13 U/L (0,22 $\mu\text{kat/L}$).

- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁸⁾.

	n	Medie		În interiorul ciclului	Total
		U/L	$\mu\text{kat/L}$	CV (%)	
Nivel scăzut	80	82	1,37	1,3	2,7
Nivel mediu	80	204	3,40	0,9	2,2
Nivel înalt	80	992	16,53	1,5	2,6

- Corelație

A fost efectuat un studiu comparativ între Analizorul Selectra ProM ELITech Clinical Systems și un alt echipament al unui sistem aprobat de FDA (metoda IFCC) pe 100 de probe de ser uman conform protocolului CLSI EP9-A2⁽⁹⁾.

Valorile au fost între 21 și 1439 U/L (între 0,35 și 23,98 $\mu\text{kat/L}$).

Parametrii regresiei liniare sunt după cum urmează:

Coeficient de corelație: $(r)=0,999$

Regresie liniară: $y=0,976x - 1 \text{ U/L (0,02 } \mu\text{kat/L)}$

- Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽¹⁰⁾ al CLSL. Recuperarea este în intervalul $\pm 10\%$ din valoarea inițială a amilasa de 80 U/L și 1000 U/L.

Trigliceride: Nicio interferență semnificativă până la 3000 mg/dL (33,9 mmol/L).

Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 $\mu\text{mol/L}$).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 $\mu\text{mol/L}$).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL.

Acid ascorbic: Nicio interferență semnificativă până la 20,0 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglubulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹¹⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹²⁻¹³⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.



AMYLASE SL

Referințe:

AMSL-0230	6 x 20 mL
AMSL-0390	1 x 50 mL
AMSL-0400	6 x 50 mL

Compoziția trusei:

R 6 x 20 mL
R 1 x 50 mL
R 6 x 50 mL

FTRO-AMSL-v18 (12/2018)_PIT-AMSL-4-v18


Stabilitatea la bord/Frecvența calibrării
Stabilitatea la bord: 28 de zile











Frecvența calibrării: 28 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

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SIMBOLURI

	Dispozitiv medical de diagnosticare <i>in vitro</i> .
	Consultați instrucțiunea de utilizare.
	Producător
	Limită de temperatură
	Număr de lot
	Data expirării
	Număr catalog
	Conținut
	Reactiv
	Conformitate europeană

Notă

- Doar pentru ref. AMSL-0230, utilizată cu software-ul Selectra TouchPro.


 Amylase IFCC
 165

 0
 PIT-AMSL

 Modificare față de versiunea precedentă.


AST/GOT 4+1 SL

Referințe:

ASSL-0250	8 x 25 mL
ASSL-0455	4 x 55 mL
ASSL-0410	2 x 62,5 mL
ASSL-0430	4 x 62,5 mL
ASSL-0510	5 x 125 mL

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
R1 4 x 44 mL + R2 4 x 11 mL
R1 2 x 50 mL + R2 1 x 26 mL
R1 4 x 50 mL + R2 2 x 26 mL
R1 5 x 100 mL + R2 1 x 127 mL



FTRO-ASSL4+1-v19(12/2018)_PIT-ASSL4+1-4-v19

SCOPUL UTILIZĂRII

AST/GOT 4+1 SL ELITech Clinical Systems este conceput pentru determinarea cantitativă a aspartat aminotransferazei (AST) în serul uman și plasmă pentru diagnosticare *in vitro*.

SEMNIFICAȚIE CLINICĂ⁽¹⁻⁴⁾

Aspartat aminotransferaza (AST), cunoscută și ca glutamat oxalat transaminază (GOT), este o transaminază. AST catalizează transferul grupului amino al L-aspartatului la α-ketoglutarat pentru a rezulta L-glutamatul. AST este distribuită în mare măsură în organism, însă cele mai mari nivele se găsesc în inimă, ficat, mușchii scheletici și rinichi.

Deteriorarea celulelor acestor țesuturi induce creșterea AST în ser. În cazul formelor fulminante de hepatită, în special hepatita virală, nivelul enzimei este marcat ridicat. În cazul infarctului miocardic, activitatea AST crește și atinge un vârf după 18-24 ore. Activitatea scade din nou după 4-5 zile, cu condiția să nu aibă loc un infarct.

Următoarele stări patologice sunt exemple ale afecțiunilor care duc, de asemenea, la o creștere a activității enzimatice: necroza celulelor ficatului sau leziunea de orice cauză (de exemplu aportul de alcool, delirium tremens, și administrarea diverselor medicamente induc creșterea moderată a AST), hepatita alcoolică, distrofia musculară și gangrena, mononucleoza infecțioasă, pancreatita acută, afecțiunile cardiace precum miocardita sau pericardita, emboliile pulmonare...

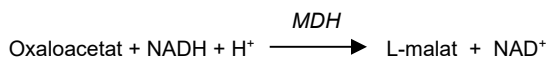
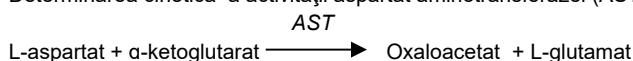
Nivelul AST în ser poate fi redus în cazul deficienței de vitamina B6.

METODĂ⁽⁵⁾

Metoda IFCC fără piridoxal fosfat (P-5'-P).
Cinetică. UV.

PRINCIPIU⁽⁵⁾

Determinarea cinetică a activității aspartat aminotransferazei (AST).



MDH=malat dehidrogenază

COMPOZIȚIA

Reactiv 1: R1

Tampon Tris, pH 7,80 (30°C)	100 mmol/L
L-aspartat	330 mmol/L
LDH	≥ 2000 U/L
MDH	≥ 1000 U/L
Azidă de sodiu	< 0,1%

Reactiv 2: R2

α-Ketoglutarat	78 mmol/L
NADH	1,1 mmol/L
Azidă de sodiu	< 0,1%

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550, ELICAL 2
- CONT-0060, ELITROL I
- CONT-0160, ELITROL II
- Soluție salină obișnuită (NaCl 9 g/L).
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest reactiv este conceput doar pentru utilizarea în scopul diagnosticării *in vitro*.
- Acești reactivi conțin azidă de sodiu care poate reacționa cu plumbul sau instalațiile sanitare din cupru și poate forma azide metalice explozibile. În cazul aruncării acestor reactivi, spălați întotdeauna cu cantități mari de apă pentru a preveni formarea de azide.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați fiolele de reactiv din truse diferite.
- Pentru mai multe informații, Fișa de date privind siguranța (SDS) este disponibilă la cerere pentru utilizatorul profesional.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele recipientelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, recipient perforat).

PROBE^(2,6)

Specimen

- Ser și plasmă heparinizată de litu, libere de hemoliză.
- A nu se utiliza aste specimene.

Avertisment și precauții

Conform bunei practici de laborator, puncția venoasă trebuie efectuată înainte de administrarea de medicamente.

AST/GOT 4+1 SL

Referințe:

ASSL-0250	8 x 25 mL
ASSL-0455	4 x 55 mL
ASSL-0410	2 x 62,5 mL
ASSL-0430	4 x 62,5 mL
ASSL-0510	5 x 125 mL

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
R1 4 x 44 mL + R2 4 x 11 mL
R1 2 x 50 mL + R2 1 x 26 mL
R1 4 x 50 mL + R2 2 x 26 mL
R1 5 x 100 mL + R2 1 x 127 mL



FTRO-ASSL4+1-v19(12/2018)_PIT-ASSL4+1-4-v19

Depozitare și stabilitate

Eșantioanele sunt stabile timp de 24 de ore la temperatura camerei, 7 zile la 2-8°C, și 3 luni la -20°C.

VALORI DE REFERINȚĂ (2,3)

Ser. plasmă (37°C): <40 U/L

Valorile de referință pentru infanți sunt mai mari decât pentru adulți.

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems.

Aplicațiile sunt disponibile la cerere

Lungime de undă 340 nm

Temperatură 37°C

Citiți pe reactivul martor.

Reactiv R1	240 µL
Proba	15 µL

Amestecați și așteptați o incubație de 4 minute și 43 de secunde, apoi adăugați:

Reactiv R2	60 µL
-------------------	-------

Amestecați și așteptați o incubație de 50 de secunde, măsurarea modificarea absorbantei per minut ($\Delta A/\text{min}$) timp de 159 de secunde.

- Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.
- Valorile mari ale AST pot induce rezultate scăzute în mod fals datorită golirii substratului (consumul total de NADH înainte de citirea rezultatului). Pentru Analizoarele Selectra pro Series ELITech Clinical Systems, aplicația conține o alarmă specifică pentru a avertiza utilizatorii.

CALCUL

$(\Delta A) \text{ Proba} \times n$ $n = \text{concentrație calibrator}$

$(\Delta A) \text{ Calibrator}$

Factor de conversie: U/L x 0.0167 = µkat/L

CALIBRARE

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este trasabilă conform metodei de referință IFCC⁽⁶⁾.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a asigura calitatea adecvată, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite.

Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems
- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁷⁾, intervalul de măsurare este între 10,0 și 450,0 U/L (de la 0,17 la 7,50 µkat/L). Probele care depășesc 450,0 U/L trebuie să fie diluate 1:10 cu soluție de NaCl 9 g/L (salină normală) și re-analizate. Utilizarea acestei proceduri extinde intervalul de măsurare de la 450,0 la 4500,0 U/L (de la 7,50 la 75,00 µkat/L).

Pentru utilizatorii Selectra TouchPro, funcția „dilute” efectuează diluția automată a eșantionului. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁸⁾.

LoD= 2,5 U/L (0,04 µkat/L).

LoQ= 5,0 U/L (0,08 µkat/L).

- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁹⁾.

	n	Medie		În interiorul ciclului	Total
		U/L	µkat/L	CV (%)	
Nivel scăzut	80	34,1	0,57	1,7	3,4
Nivel mediu	80	67,9	1,13	0,8	1,9
Nivel înalt	80	353,6	5,89	0,4	2,0

- Corelație

A fost efectuat un studiu comparativ între Analizorul Selectra ProM ELITech Clinical Systems și un alt echipament al unui sistem aprobat de FDA (metoda IFCC fără piridoxal fosfat) pe 114 de eșantioane de ser uman conform protocolului CLSI EP9-A2⁽¹⁰⁾.

Valorile acoperă domeniul de măsurare.

Parametrii regresiei liniare sunt după cum urmează:

Coefficient de corelație: (r)=0,999

Regresie liniară: $y = 0,927 x - 0,3 \text{ U/L (0,01 } \mu\text{kat/L)}$.

- Limitări și interferențe

- Eșantioanele hemolizate nu trebuie să fie utilizate deoarece hemoliza semnificativă poate crește concentrația AST din cauza nivelurilor ridicate de AST în eritrocite.

- Nu raportați rezultatele în afara intervalului utilizabil.

Referințe:

ASSL-0250	8 x 25 mL
ASSL-0455	4 x 55 mL
ASSL-0410	2 x 62,5 mL
ASSL-0430	4 x 62,5 mL
ASSL-0510	5 x 125 mL

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
R1 4 x 44 mL + R2 4 x 11 mL
R1 2 x 50 mL + R2 1 x 26 mL
R1 4 x 50 mL + R2 2 x 26 mL
R1 5 x 100 mL + R2 1 x 127 mL



FTRO-ASSL4+1-v19(12/2018)_PIT-ASSL4+1-4-v19

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2 ⁽¹¹⁾ al CLSL și recomandările SFBC⁽¹²⁾. Recuperarea este în intervalul ±10% din valoarea inițială a activității AST de 35,0 U/L și 350,0 U/L.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30 mg/dL (513 μmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 μmol/L).

Trigliceride: Nicio interferență semnificativă până la 2400 mg/dL (27,12 mmol/L) echivalent trigliceride.

Piruvat: Nicio interferență semnificativă până la 3 mg/dl.

Acid ascorbic: Nicio interferență semnificativă până la 20 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹³⁾
- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹⁴⁻¹⁵⁾
- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu aste rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 28 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit, și după o operație de întreținere.

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SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

CONT	Conținut
R1	Reactiv 1
R2	Reactiv 2
CE	Conformitate europeană

Notă:

Doar pentru ref. **ASSL-0250/ASSL-0455**, utilizată cu software-ul Selectra TouchPro.


 ASAT (GOT)
180

 1
PIT-ASSL

☞: Modificare față de versiunea precedentă.



BILIRUBIN TOTAL & DIRECT 4+1

Referințe:

BIDI-0250 Directă 4+1	8 x 25 mL
BITO-0250 Totală 4+1	8 x 25 mL
BIDI-0600 Directă 4+1	2 x 125 mL
BITO-0600 Totală 4+1	2 x 125 mL
BITD-0600 T&D 4+1	2 x 125 mL

Compoziția trusei:

R1 Directă	8 x 20 mL	+ R2	8 x 5 mL
R1 Totală	8 x 20 mL	+ R2	8 x 5 mL
R1 Directă	2 x 100 mL	+ R2	1 x 50 mL
R1 Totală	2 x 100 mL	+ R2	1 x 50 mL
R1 Totală	1 x 100 mL	+ R1 Directă	1 x 100 mL
		+ R2	1 x 50 mL

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SCOPUL UTILIZĂRII

Pentru Bilirubină Totală: BILIRUBIN TOTAL 4+1 ELITech Clinical Systems este conceput pentru determinarea cantitativă a bilirubinei totale în serul uman și plasmă în cazul adulților și copiilor cu vârsta de peste 10 zile pentru diagnosticare *in vitro*.

Pentru Bilirubină Directă: BILIRUBIN DIRECT 4+1 ELITech Clinical Systems este conceput pentru determinarea cantitativă a bilirubinei directe în serul uman și plasmă pentru diagnosticare *in vitro*.

SEMNIFICAȚIE CLINICĂ⁽¹⁻²⁾

Aproximativ 80-85% din bilirubina este produsă din fracțiunea hem a hemoglobinei eliberate de eritrocitele care îmbătrânesc în celulele reticuloendoteliale. Bilirubina, legată de albumină, este transportată în ficat, unde este conjugată rapid cu glucuronida pentru a-i mări solubilitatea. Apoi, aceasta este excretată în canaliculii biliari și hidrolizată în tractul gastrointestinal.

Concentrația serului de bilirubină neconjugată crește în cazul supra-producerii de bilirubină (anemie hemolitică acută și cronică) și în cazul afecțiunilor metabolismului bilirubinei și defectelor de transport (aport afectat de celulele hepatice: sindromul Gilbert; defectele în reacția de conjugare: sindromul Crigler-Najjar). Excreția redusă (deteriorare hepatocelulară: hepatită, ciroză...; sindromul Dubin-Johnson și Rotor) și obstrucția fluxului biliar (cel mai adesea produsă de calculii biliari sau de tumori) induc o creștere importantă a bilirubinei conjugate și într-o măsură minoră o creștere a bilirubinei neconjugate (hiperbilirubinemia conjugată).

METODĂ⁽²⁾

Malloy-Evelyn modificată. Punct final.

PRINCIPIU⁽¹⁻²⁾

Acidul sulfanilic reacționează cu nitritul de sodiu pentru a forma acidul sulfanilic diazotat. În prezența acceleratorului (cetrimidă), bilirubina conjugată și neconjugată reacționează cu acidul sulfanilic diazotat pentru a forma azobilirubina (Bilirubina totală 4+1). În absența acceleratorului, doar bilirubina conjugată reacționează (Bilirubină directă 4+1). Creșterea absorbanței la 546 nm este proporțională cu concentrația bilirubinei.

Acidul sulfanilic + NaNO₂ → Acid sulfanilic diazotat

Bilirubină + Acid sulfanilic diazotat → Azobilirubină

COMPOZIȚIA REACTIVULUI

BILIRUBINĂ TOTALĂ 4+1

Reactiv 1: R1

Acid sulfanilic	29	mmol/L
Cetrimidă	29	mmol/L

BILIRUBINĂ DIRECTĂ 4+1

Reactiv 1: R1

Acid sulfanilic	29	mmol/L
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BILIRUBINĂ TOTALĂ & DIRECTĂ 4+1

Reactiv 2: R2

Nitrit de sodiu	11	mmol/L
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MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Soluție salină obișnuită (NaCl 9 g/L)
- Echipamente generale de laborator.
- Analizor de biochimie echipat cu filtrele necesare. (Consultați § PROCEDURA).
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acești reactivi sunt doar pentru utilizarea profesională, în scopul diagnosticării *in vitro*.
- Reactivii R1 conține acid sulfanilic. Poate provoca o reacție alergică.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați fiolele de reactiv din truse diferite.
- Fișa cu date de securitate disponibilă la cerere.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Reactivul R1 din Bilirubină totală 4+1 poate fi ușor tulbure. Acesta conține un detergent care poate duce la formarea de spumă în unitățile de spălare ale unor echipamente. Aceste două caracteristici nu au consecințe asupra performanțelor produsului.
- Reactivii R1 din Bilirubină directă 4+1 și reactivul R2 din Bilirubină totală și directă 4+1 trebuie să fie limpezi. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

PROBE⁽³⁾

Specimen

- Ser sau plasmă heparinizată cu litiu.
- A nu se utiliza alte specimene.



BILIRUBIN TOTAL & DIRECT 4+1

Referințe:

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BITO-0250 Totală 4+1	8 x 25 mL
BIDI-0600 Directă 4+1	2 x 125 mL
BITO-0600 Totală 4+1	2 x 125 mL
BITD-0600 T&D 4+1	2 x 125 mL

Compoziția trusei:

R1 Directă 8 x 20 mL + R2 8 x 5 mL
R1 Totală 8 x 20 mL + R2 8 x 5 mL
R1 Directă 2 x 100 mL + R2 1 x 50 mL
R1 Totală 2 x 100 mL + R2 1 x 50 mL
R1 Totală 1 x 100 mL + R1 Directă 1 x 100 mL + R2 1 x 50 mL

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Avertisment și precauții

- Pentru bilirubina directă: Nu utilizați probe hemolizate.
- Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.
- Trebuie acordată o atenție specială umplerii tuburilor heparinizate conform instrucțiunilor producătorului. O umplere insuficientă poate duce la rezultate eronate.
- Protejați probele împotriva luminii înainte și în timpul analizei.

Depozitare și stabilitate

Dacă plasma și serul sunt protejate împotriva luminii, probele sunt stabile 1 zi (Bilirubină totală) sau 2 zile (Bilirubina directă) la temperatura camerei, 7 zile la 2-8°C și 6 luni la -20°C.

VALORI DE REFERINȚĂ ⁽⁴⁾

Ser, plasmă:

Bilirubină totală:

Adulți și copii peste 10 zile:

0,2-1,2 mg/dL (3,4-21 μmol/L)

Bilirubină directă:

<0,2 mg/dL (3,4 μmol/L)

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere.

A) Bilirubină totală

Lungime de undă 546-700 nm

Temperatură: 37°C

Citiți pe reactivul martor.

	CALIBRARE	TEST
Reactiv R1	240 μL	240 μL
Calibrator	15 μL	-
Probă	-	15 μL

Amestecați și citiți absorbanta (ΔA1) după o incubație de 4 minute 40 (proba martor), apoi adăugați:

Reactiv R2	60 μL
------------	-------

Amestecați și citiți absorbanta (ΔA2) după o incubație de 6 minute 30.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestei inserții.

Pentru utilizatorii Selectra ProXS, este nevoie de un filtru suplimentar de 700nm.

B) Bilirubină directă

Lungime de undă 546 nm

Temperatură: 37°C

Citiți pe reactivul martor.

	CALIBRARE	TEST	
Reactiv R1	240 μL	240 μL	
Calibrator	30 μL	-	
Probă	-	30 μL	

Amestecați și citiți absorbanta (A1) după o incubație de 4 minute 40 (proba martor), apoi adăugați:

Reactiv R2	60 μL
------------	-------

Amestecați și citiți absorbanta (A2) după o incubație de 50 de secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

În aplicație, compensarea trebuie setată la: -0,05 mg/dL (-0,9 μmol/L).

CALCUL

A) Bilirubină totală

$(\Delta A2 - \Delta A1) \text{ Probă} \times \text{concentrație calibrator}$

$(\Delta A2 - \Delta A1) \text{ Calibrator}$

B) Bilirubină directă

$(A2 - A1) \text{ Probă} \times \text{concentrație calibrator}$

$(A2 - A1) \text{ Calibrator}$

Factor de conversie: mg/dL x 17,1 = μmol/L

CALIBRARE

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este definită în raport cu materialul de referință SRM 916a (al Institutului Național de Standarde și Tehnologie).

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a verifica precizia testelor, vor fi utilizate serurile de control precum ELITROL I (control normal) și ELITROL II (control patologic). Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems

A) Bilirubină totală

- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁵⁾, intervalul de măsurare este între 0,25 mg/dL și 25,00 mg/dL (de la 4,3 la 427,6 μmol/L). Probele care depășesc 25,00 mg/dL trebuie diluate 1:5 cu soluție de NaCl 9 g/l (soluție salină normală) și re-testate. Utilizarea acestei



BILIRUBIN TOTAL & DIRECT 4+1

Referințe:

BIDI-0250 Directă 4+1 8 x 25 mL
 BITO-0250 Totală 4+1 8 x 25 mL
 BIDI-0600 Directă 4+1 2 x 125 mL
 BITO-0600 Totală 4+1 2 x 125 mL
 BITD-0600 T&D 4+1 2 x 125 mL

Compoziția trusei:

R1 Directă 8 x 20 mL + R2 8 x 5 mL
 R1 Totală 8 x 20 mL + R2 8 x 5 mL
 R1 Directă 2 x 100 mL + R2 1 x 50 mL
 R1 Totală 2 x 100 mL + R2 1 x 50 mL
 R1 Totală 1 x 100 mL + R1 Directă 1 x 100 mL + R2 1 x 50 mL



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proceduri extinde intervalul de măsurare între 25,00 și 60,00 mg/dL (de la 427,6 la 1026,3 μmol/L).

Pentru utilizatorii Selectra TouchPro, funcția „diluare” efectuează diluarea probelor automat. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁶⁾.

LoD=0,04 mg/dL (0,7 μmol/L)

LoQ=0,15 mg/dL (2,6 μmol/L)

- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁷⁾.

	n	Medie		În interiorul ciclului	Total
		mg/dL	μmol/L		
				CV (%)	
Nivelul 1	80	1,15	19,7	1,8	5,0
Nivelul 2	80	4,08	69,8	0,4	3,1
Nivelul 3	80	14,61	249,9	0,5	2,9

- Corelație

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda DCA) pe 100 de probe de ser uman conform protocolului CLSI EP9-A2⁽⁸⁾.

Concentrațiile probelor au fost între 0,32 și 23,02 mg/dL (5,5 și 393,7 μmol/L).

Parametrii regresiiilor liniare sunt după cum urmează:

Coeficient de corelație: (r)=0,999

Regresie liniară: y= 0,948 x -0,11 mg/dL (1,9 μmol/L)

- Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽⁹⁾. Recuperarea este în intervalul ±15% din valoarea inițială a concentrației bilirubinei totale de 1,00 mg/dL și 15,00 mg/dL.

Trigliceride: Nicio interferență semnificativă până la 2100 mg/dL (23,73 mmol/L).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

Acid ascorbic: Nicio interferență semnificativă până la 4 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹⁰⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹¹⁻¹²⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 28 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

B) Bilirubină directă

- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁵⁾, intervalul de măsurare este între 0,08 mg/dL și 10,55 mg/dL (de la 1,4 la 180,4 μmol/L). Probele care depășesc 10,55 mg/dL trebuie diluate cu soluție de NaCl 9 1:5 (soluție salină normală) și re-testate. Utilizarea acestei proceduri extinde intervalul de măsurare între 10,55 și 50,00 mg/dl (de la 180,4 la 855,2 μmol/L).

Pentru utilizatorii Selectra TouchPro, funcția „diluare” efectuează diluarea probelor automat. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁶⁾.

LoD=0,01 mg/dL (0,2 μmol/L)

LoQ=0,08 mg/dL (1,4 μmol/L)

- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁷⁾.

	n	Medie		În interiorul ciclului	Total
		mg/dL	μmol/L		
				CV (%)	
Nivelul 1	80	0,36	6,2	3,8	5,2
Nivelul 2	80	1,51	25,8	1,9	5,3
Nivelul 3	80	3,99	68,2	0,9	4,7

- Corelație

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda DCA) pe 100 de probe de ser uman conform protocolului CLSI EP9-A2⁽⁸⁾.

Concentrațiile probelor au fost între 0,09 și 10,52 mg/dL (1,5 și 179,9 μmol/L).

Parametrii regresiiilor liniare sunt după cum urmează:

Coeficient de corelație: (r)=0,998

Regresie liniară: y=0,926 x -0,03 mg/dL (0,5 μmol/L)

- Limitări și interferențe

- Concentrația acidului ascorbic mai mare de 0,5 mg/dL poate duce la rezultate fals pozitive ale bilirubinei directe.

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽⁹⁾. Recuperarea este în intervalul ±15% din valoarea inițială a concentrației bilirubinei directe de 0,40 mg/dL și 4,00 mg/dL.

Trigliceride: Nicio interferență semnificativă până la 2000 mg/dL (22,60 mmol/L).

Hemoglobină: Nicio interferență semnificativă până la 125 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.



BILIRUBIN TOTAL & DIRECT 4+1

Referințe:

BIDI-0250 Directă 4+1 8 x 25 mL
 BITO-0250 Totală 4+1 8 x 25 mL
 BIDI-0600 Directă 4+1 2 x 125 mL
 BITO-0600 Totală 4+1 2 x 125 mL
 BITD-0600 T&D 4+1 2 x 125 mL

Compoziția trusei:

R1 Directă 8 x 20 mL + R2 8 x 5 mL
 R1 Totală 8 x 20 mL + R2 8 x 5 mL
 R1 Directă 2 x 100 mL + R2 1 x 50 mL
 R1 Totală 2 x 100 mL + R2 1 x 50 mL
 R1 Totală 1 x 100 mL + R1 Directă 1 x 100 mL + R2 1 x 50 mL



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Acid ascorbic: Nicio interferență semnificativă până la 0,5 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹⁰⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹¹⁻¹²⁾

- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 28 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

CONT	Conținut
R1	Reactiv 1
R2	Reactiv 2
CE	Conformitate europeană

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- Young D.S., Effects of drugs on clinical laboratory tests, 4th edition, AACC Press (1995).

 :Modificare față de versiunea precedentă.



BILIRUBIN TOTAL & DIRECT 4+1

Referințe:

BIDI-0250 Directă 4+1 8 x 25 mL
BITO-0250 Totală 4+1 8 x 25 mL
BIDI-0600 Directă 4+1 2 x 125 mL
BITO-0600 Totală 4+1 2 x 125 mL
BITD-0600 T&D 4+1 2 x 125 mL

Compoziția trusei:

R1 Directă 8 x 20 mL + **R2** 8 x 5 mL
R1 Totală 8 x 20 mL + **R2** 8 x 5 mL
R1 Directă 2 x 100 mL + **R2** 1 x 50 mL
R1 Totală 2 x 100 mL + **R2** 1 x 50 mL
R1 Totală 1 x 100 mL + **R1 Directă** 1 x 100 mL + **R2** 1 x 50 mL

FTRO-BITD-v12 (12/2018)_PIT_BITD-4-v12



BILIRUBINĂ DIRECTĂ 4+1:

NOTĂ IMPORTANTĂ

- Doar pentru ref. **BIDI-0250**, utilizată cu software-ul Selectra TouchPro.

- **Vezi ȘPROCEDURĂ:** Este necesară introducerea manuală



Bilirubin Direct New 205 0
PIT-BITD

BILIRUBINĂ TOTALĂ 4+1:

- Doar pentru ref. **BITO-0250**, utilizată cu software-ul Selectra TouchPro.



Bilirubin Total New 225 0
PIT-BITD



☞ Referințe:
CALA-0600
CALA-0250

Compoziția trusei:
R 2 x 125 mL + Std 1 x 5 mL
R 12 x 20 mL



FTRO-CALA-v18 (09/2020)_PIT-CALA-4-v18

☞ SCOPUL UTILIZĂRII

ELITech Clinical Systems CALCIUM ARSENAZO este un reactiv de diagnostic *in vitro* destinat determinării cantitative a calciului total din probele serul uman, plasmă și urină.

SEMNIFICAȚIE CLINICĂ⁽¹⁻³⁾

În sânge, aproximativ 45% din calciul plasmatic este liber, 45% este legat de proteine, asociat în principal cu albumina și 10% formează complexe. Calcemia măsoară calciul total, însă doar calciul liber este activ din punct de vedere biologic. Calciul are un rol fiziologic activ în mineralizarea oaselor, excitabilitatea neuromusculară, contracția musculară și coagularea sângelui. Nivelurile de proteine serice sau albumina trebuie avute în vedere pentru interpretarea corespunzătoare a nivelurilor de calciu seric total. Hipocalcemia poate rezulta din insuficiența renală cronică cu hipoproteinemia și hiperfosfatemia, sau hipoparatiroidismul, sau deficiența de vitamina D (osteomalachia, rahitismul...). Cele mai frecvente cazuri de hipercalcemie sunt asociate cu hiperparatiroidismul sau supradoza de vitamina D...

Calciuria are o valoare practică mică în diverse diagnostice, cu excepția tubulopatiilor renale.

METODĂ

Testul direct colorimetric complexometric (Arsenazo III).
Punct final.

PRINCIPIU⁽⁴⁾

La un pH ușor acid, ionii Ca²⁺ formează cu Arsenazo III (acid 2,7-bis(2-aronofenilazo))-1,8-dihidroxi-naftalen-3,6-disulfonic) un complex a cărui absorbanta este direct proporțională cu concentrația totală de calciu.

COMPOZIȚIA REACTIVULUI

Reactiv: R

Tampon MES, pH 6,50 100 mmol/L
Arsenazo III 200 μmol/L

Standard: Std. (Ref.: CALA-0600)

Calciu 10 mg/dL
2.5 mmol/L

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Echipamente generale de laborator.
- Analizor de biochimie echipat cu filtrele necesare. (Consultați ȘI PROCEDURA).
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest dispozitiv de diagnostic *in vitro* (Reactiv și Standardul) este destinat numai pentru uz profesional.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.

- Standardul trebuie să fie imediat închis cu capacul pentru a preveni contaminarea și evaporarea.
- Pentru mai multe informații, Fișa de date privind siguranța (SDS) este disponibilă la cerere pentru utilizatorul profesional.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați ȘI DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii și Standardul sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Reactivul și soluția standard trebuie să fie limpezi. Turbiditatea ar indica deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, folie perforată).

PROBE^(1,2,5)

Specimen

- Ser
- Plasmă heparinizată cu litiu.
- Urină colectată peste 24 de ore.
- A nu se utiliza alte specimene.

Avertismente și precauții

- Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.
- Serul trebuie separat de celule cât mai repede posibil.
- După colectare, specișenele de urină trebuie să fie acidificate cu acid clorhidric 6N la un pH < 2, pentru a preveni precipitarea sării de calciu.

Depozitare

- Calciul total este stabil în ser și plasmă la temperatura camerei până la 7 zile, la 2-8°C timp de 3 săptămâni și în stare înghețată (-20°C) până la 8 luni.
- Urina poate fi conservată la temperatura camerei timp de 2 zile, la 2-8°C timp de 4 zile și în stare înghețată (-20°C) până la 3 săptămâni.

VALORI DE REFERINȚĂ^(1,6)

Ser, plasmă: 8,6-10,3 mg/dL
2,15-2,57 mmol/L
Urină: 100-300 mg/24 ore
2,50-7,50 mmol/24 ore
6,7-20,0 mg/dL*
1,67-5,00 mmol/L*

Calcemia este întotdeauna interpretată în funcție de ratele proteinei plasmatice.

*pentru un volum urinar de 1,5 L pe 24 de ore.

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

☞ Referințe:
CALA-0600
CALA-0250

Compoziția trusei:
R 2 x 125 mL + Std 1 x 5 mL
R 12 x 20 mL



FTRO-CALA-v18 (09/2020)_PIT-CALA-4-v18

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere.

Lungime de undă 660-700 nm

Temperatură: 37°C

Citiți pe reactivul maror.

	CALIBRARE	CALIBRARE	TEST
Reactiv R	300 µL	300 µL	300 µL
Apă distilată	6 µL	-	-
Standard/ Calibrator	-	6 µL	-
Proba	-	-	6 µL

Amestecați, așteptați 4 minute și 30 de secunde și apoi citiți absorbanta (A).

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

Pentru utilizatorii Selectra ProXS, este nevoie de un filtru suplimentar de 700 nm.

CALCUL

ΔA Proba

_____ x n n = concentrație calibrator/standard

ΔA Standard/
Calibrator

Factor de conversie: mg/dL x 0,25= mmol/L

☞ CALIBRARE

Pentru referința CALA-0600: Pentru calibrare, trebuie utilizat fie calibratorul multiparametric ELICAL 2 fie Standardul de calciu de 10 mg/dL.

Pentru referința CALA-0250: Pentru calibrare, utilizați calibratorul multiparametric ELICAL 2.

Valorile concentrației Standardului de calciu de 10 mg/dL și calibratorului multiparametric ELICAL 2 sunt trasabile conform Materialului Standard de Referință 956d (al Institutului Național de Standarde și Tehnologie).

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați și DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a verifica precizia testelor, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems

- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁷⁾.

a) Ser/Plasmă

Intervalul de măsurare este între 5,00 și 15,00 mg/dL (de la 1,25 la 3,74 mmol/L).

b) Urină

Intervalul de măsurare este între 1,50 și 18,00 mg/dL (0,37 - 4,49 mmol/L). care depășesc 18,00 mg/dL trebuie să fie diluate 1:5 cu soluție NaCl 9g/L (salină obișnuită) și re-evaluate. Utilizarea acestei proceduri extinde intervalul de măsurare până la 90,00 mg/dL (22,46 mmol/L.)

Pentru utilizatorii Selectra TouchPro, funcția „diluare” efectuează diluarea probelor automat. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁸⁾.

LoD=0,04 mg/dL (0,01 mmol/L)

LoQ=5,00 mg/dL (1,25 mmol/L)

a) Urină

LoD=0,15 mg/dL (0,04 mmol/L)

LoQ=1,50 mg/dL (0,37 mmol/L)

- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁹⁾.

a) Ser/Plasmă

	n	Medie		În interiorul ciclului	Total
		mg/dL	mmol/L		
				CV (%)	
Nivelul 1	80	8,28	2,07	1,1	1,7
Nivelul 2	80	10,32	2,57	0,5	1,4
Nivelul 3	80	12,96	3,23	0,5	1,0

b) Urină

	n	Medie		În interiorul ciclului	Total
		mg/dL	mmol/L		
				CV (%)	
Nivelul 1	80	4,53	1,13	1,3	1,8
Nivelul 2	80	10,89	2,72	0,5	1,2
Nivelul 3	80	17,51	4,37	0,3	0,8

☞ - Corelație

a) Ser/Plasmă

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda colorimetrică) pe 106 probe de ser uman conform protocolului CLSI EP9-A2⁽¹⁰⁾.

Valorile au fost între 5,33 și 15,53 mg/dL (1,33 și 3,87 mmol/L).

Parametrii regresiiilor liniare sunt după cum urmează:

Coefficient de corelație: (r)=0,993

Regresie liniară: y=0,996x +0,43 mg/dL (0,11 mmol/L)

☞ Referințe:
CALA-0600
CALA-0250

Compoziția trusei:
R 2 x 125 mL + Std 1 x 5 mL
R 12 x 20 mL



FTRO-CALA-v18 (09/2020)_PIT-CALA-4-v18

b) Urină

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda colorimetrică) pe 52 probe de urină conform protocolului CLSI EP9-A2⁽¹⁰⁾.

Valorile au fost între 1,57 și 17,99 mg/dL (0,39 și 4,49 mmol/L).

Parametrii regresii liniare sunt după cum urmează:

Coefficient de corelație: (r)=0,995

Regresie liniară: $y=0,983x + 0,21$ mg/dL (0,05 mmol/L)

- Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽¹¹⁾.

a) Ser/Plasmă

Recuperarea este în intervalul $\pm 10\%$ din valoarea inițială a concentrației calciului de 8,00 mg/dL și 12,00 mg/dL.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 μ mol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 μ mol/L).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL.

Trigliceride: Nicio interferență semnificativă până la 1726 mg/dL (19,50 mmol/L).

Magneziu: Nicio interferență semnificativă până la 12,0 mg/dL.

Acid ascorbic: Nicio interferență semnificativă până la 20,00 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabale.⁽¹²⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young.⁽¹³⁻¹⁴⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

b) Urină

Recuperarea este în intervalul $\pm 10\%$ din valoarea inițială a concentrației calciului de 4,00 mg/dL și 16,00 mg/dL.

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 μ mol/L).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL.

Acid ascorbic: Nicio interferență semnificativă până la 20,00 mg/dL.

Uree: Nicio interferență semnificativă până la 5000 mg/dL.

(832 mmol/L).

Acid uric: Nicio interferență semnificativă până la 100 mg/dL.

(5,9 mmol/L).

Magneziu: Nicio interferență semnificativă până la 1,0 mg/dL.

(4,1 mmol/L).

pH: Nicio interferență semnificativă pentru pH în intervalul 2,5 – 6,0.

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young.⁽¹³⁻¹⁴⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatări clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 28 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

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9. Evaluation of Precision Performance of Quantitative Measurement Methods; Approved Guideline - Second Edition. CLSI (NCCLS) document EP5-A2 (2004), **24** (25).
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CALCIUM ARSENAZO

☞ Referințe:
CALA-0600
CALA-0250






Compoziția trusei:
R 2 x 125 mL + Std 1 x 5 mL
R 12 x 20 mL



FTRO-CALA-v18 (09/2020)_PIT-CALA-4-v18

SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

	Conținut
	Reactiv
	Standard
	Conformitate europeană
	Modificare față de versiunea precedentă

Notă:

Doar pentru ref. **CALA-0250**, utilizată cu software-ul Selectra TouchPro.



Calcium New
245

0
PIT-CALA



Referințe:

CKSL-0230 4 x 25 mL

CKSL-0410 2 x 62,5 mL

CKSL-0430 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL + R2 4 x 5 mL

R1 2 x 50 mL + R2 1 x 26 mL

R1 4 x 50 mL + R2 2 x 26 mL



FTRO-CKSL-v21 (12/2018)_PIT-CKSL-4-v21

SCOPUL UTILIZĂRII

CK NAC SL ELITech Clinical Systems este conceput pentru determinarea cantitativă a creatinkinazei în serul uman și plasmă pentru diagnosticare *in vitro*.

SEMNIFICAȚIE CLINICĂ⁽¹⁻⁴⁾

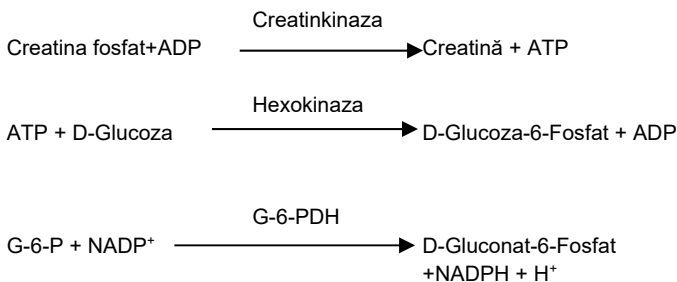
Creatinkinaza (CK) există în 3 forme citoplasmice: CK-MM (în mușchiul striat și cardiac), CK-MB (doar în mușchiul cardiac), și CK-BB (în special în creier). Determinarea CK este utilizată pentru diagnosticarea și urmărirea bolilor musculare (în special distrofiile musculare) și ale leziunilor mușchiului cardiac. În infarctul miocardic, ratele CK totale cresc rapid până este atins un vârf la 10-24 de ore de la debutul infarctului. Nivelurile revin la normal în 3-4 zile.

Anumite medicamente antipsihotice (olanzapina și quetiapina) și în unele cazuri hipotiroidismul, declanșează de asemenea creșterea activității CK.

METODĂ⁽⁵⁾

Metoda IFCC - cinetică. UV.

PRINCIPIU⁽⁵⁾



G-6-P: D-Glucoza-6-Fosfat

G-6-PDH: Glucoza-6-Fosfat Dehidrogenaza.

Creșterea concentrației NADPH este direct proporțională cu activitatea CK enzimatică.

COMPOZIȚIA REACTIVULUI

Reactiv 1: R1

Tampon imidazol, pH 6,10	125 mmol/L
D-Glucoză	25 mmol/L
N-Acetil-L-Cisteină	25 mmol/L
Acetat de magneziu	12,5 mmol/L
NADP	2,4 mmol/L
EDTA	2,0 mmol/L
Hexokinază	≥ 6800 U/L
Azidă de sodiu	< 0,1 %

Reactiv 2: R2

Tampon imidazol, pH 8,9	125 mmol/L
Creatină fosfat	250 mmol/L
ADP	15,2 mmol/L
AMP	23 mmol/L
Deadenozină pentafoșfat	103 μmol/L
G-6-PDH	≥ 8800 U/L
Azidă de sodiu	< 0,1 %

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Echipamente generale de laborator.
- Soluție salină obișnuită (NaCl 9 g/L).
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Această trusă de reactivi este doar pentru utilizarea profesională, în scopul diagnosticării *in vitro*.
- Reactivii R1 și R2 sunt clasificați ca fiind periculoși (Imidazol).



PERICOL: Poate dăuna fătului. Procurați instrucțiuni speciale înainte de utilizare. Purtați mănuși de protecție/îmbrăcăminte de protecție/echipament de protecție a ochilor/echipament de protecție a feței.

ÎN CAZ DE expunere sau de posibilă expunere: consultați medicul

- Reactivii conțin azidă de sodiu care poate reacționa cu plumbul sau instalațiile din cupru pentru a forma potențiale azide metalice explozive. În momentul eliminării acestor reactivi, spălați întotdeauna cu apă din abundență pentru a preveni acumularea de azide.
- Pentru mai multe informații, consultați Fișa de date privind siguranța (SDS).
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați reactivii din truse diferite.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactiv trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

Referințe:

CKSL-0230 4 x 25 mL

CKSL-0410 2 x 62,5 mL

CKSL-0430 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL + R2 4 x 5 mL

R1 2 x 50 mL + R2 1 x 26 mL

R1 4 x 50 mL + R2 2 x 26 mL



FTRO-CKSL-v21 (12/2018)_PIT-CKSL-4-v21

PROBE (1,5)

Specimen

- Ser liber din hemoliză (specimen recomandat de IFCC).
- Plasmă heparinizată liberă din hemoliză.
- A nu se utiliza alte specimene.

Avertismente și precauții

Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.

Depozitare și stabilitate

Probele trebuie analizate imediat sau depozitate protejate împotriva aerului și luminii timp de 8 ore la temperatura camerei, 2 zile la 2-8°C, sau 1 lună la -20°C.

VALORI DE REFERINȚĂ (5)

Bărbați < 171 U/L

Femei < 145 U/L

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere.

Lungime de undă 340 nm

Temperatură: 37°C

Citiți pe reactivul martor.

Reactiv 1	240 µL
Proba	12 µL

Amestecați și așteptați 4 minute și 43 de secunde, apoi adăugați:

Reactiv 2	60 µL
------------------	-------

Amestecați și după o incubație de 130 de secunde, măsurați modificarea absorbției pe minut ($\Delta A/\text{min.}$) timp de 106 secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

CALCUL
 $\Delta A \text{ Proba} \times n \quad n = \text{concentrație calibrator}$
 $\Delta A \text{ Calibrator}$

Factor de conversie: U/L x 0,0167 = µkat/L

CALIBRARE

Pentru calibrare, trebuie utilizat calibratorul multiparametric Elical 2. Valoarea sa este definită conform metodei IFCC⁽⁵⁾.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a verifica precizia testelor, vor fi utilizate serurile de control precum ELITROL I (control normal) și ELITROL II (control patologic). Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia

măsurii corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems
- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁶⁾, intervalul de măsurare este între 10 și 1714 U/L (de la 0,17 la 28,57 µkat/L). Probele care depășesc 1714 U/L trebuie diluate 1:10 cu soluție de NaCl 9 g/L (soluție salină normală) și re-testate. Utilizarea acestei proceduri extinde intervalul de măsurare între 1714 și 17140 U/L (de la 28,57 la 285,67 µkat/L).

Pentru utilizatorii Selectra TouchPro, funcția „diluare” efectuează diluarea probelor automat. Rezultatele iau în considerare diluția.

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁷⁾.

 $LoD = 1 \text{ U/L (0,02 } \mu\text{kat/L)}$
 $LoQ = 5 \text{ U/L (0,08 } \mu\text{kat/L)}$
- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁸⁾.

	n	Medie		În interiorul ciclului	Total
		mg/dL	µkat/L	CV (%)	
Nivel redus	80	147	2,45	0,7	1,7
Nivel mediu	80	406	6,77	1,1	2,4
Nivel înalt	80	1154	19,23	1,1	3,9

- Corelație

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda IFCC) pe 100 de probe de ser uman conform protocolului CLSI EP9-A2⁽⁹⁾.

Concentrațiile probelor au fost între 11 și 1712 U/L (0,18 și 28,53 µkat/L).

Parametrii regresiiilor liniare sunt după cum urmează:

Coeficient de corelație: (r)=0,998

Regresie liniară: $y = 1,012 \times x + 2 \text{ U/L (0,03 } \mu\text{kat/L)}$

- Limitări și interferențe

- Probele hemolizate nu trebuie utilizate deoarece hemoliza semnificativă poate duce la concentrație fals crescută a CK din cauza eliberării adenilat kinazei.

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽¹⁰⁾. Recuperarea este în intervalul $\pm 10\%$ din valoarea inițială a activității CK de 150 și 1200 U/L.

Trigliceride: Nicio interferență semnificativă până la 3000 mg/dL (33,9 mmol/L).



Referințe:

CKSL-0230 4 x 25 mL
 CKSL-0410 2 x 62,5 mL
 CKSL-0430 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL+R2 4 x 5 mL
 R1 2 x 50 mL+R2 1 x 26 mL
 R1 4 x 50 mL+R2 2 x 26 mL

FTRO-CKSL-v21 (12/2018)_PIT-CKSL-4-v21



Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 μmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 μmol/L).

Acid ascorbic: Nicio interferență semnificativă până la 20,0 mg/dL.

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabale. ⁽¹¹⁾

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹²⁻¹³⁾

- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 28 zile





Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

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SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.


	Conținut
	Reactiv 1
	Reactiv 2
	Conformitate europeană

Doar pentru ref. **CKSL-0230**, utilizată cu software-ul Selectra TouchPro.



CK Total
390

0
PIT-CKSL

 Modificare față de versiunea precedentă.



Referințe:

CMSL-0230	4 x 25 mL
CMSL-0410	2 x 62,5 mL
CMSL-0430	4 x 62,5 mL

Compoziția trusei:

R1	4 x 20 mL+	R2	1 x 20 mL
R1	2 x 50 mL+	R2	1 x 26 mL
R1	4 x 50 mL+	R2	2 x 26 mL

FTRO-CMSL-v23(12/2018)_PIT-CMSL-4-v23

SCOPUL UTILIZĂRII

CK-MB SL ELITech Clinical Systems este conceput pentru determinarea cantitativă de diagnosticare *in vitro* a formei CK-MB a creatinkinazei (CK) în serul uman.

SEMNIFICAȚIE CLINICĂ⁽¹⁻²⁾

Creatinkinaza există în 3 forme citoplasmice: CK-MB (doar în mușchiul cardiac), CK-MM (în mușchiul striat și cardiac) și CK-BB (în special în creier).

Determinarea CK în ser este utilizată pentru diagnosticarea și urmărirea leziunilor musculare cardiace. În infarctul miocardic, ratele CK totale și CK-MB cresc rapid până este atins un vârf la 10-24 de ore de la debutul infarctului. Nivelele revin la normal în 3-4 zile. Nivelele CK-MB mai mari decât cele normale pot fi, de asemenea, observate după deteriorările musculare.

METODĂ⁽³⁻⁴⁾

Imuno-inhibarea, Metoda IFCC.
Cinetică. UV.

PRINCIPIU⁽³⁻⁴⁾

Reactivul CK-MB SL conține un anticorp care inhibă în mod specific subunitățile CK-M (și anume 100% din CK-MM și 50% din izoenzimele CK-MB). Activitatea rămasă, corespunzând activității fracției CK-B, este măsurată conform metodei de referință IFCC pentru măsurarea activității CK. Activitatea CK-MB este apoi obținută prin înmulțirea cu 2 a activității rămase.

COMPOZIȚIA REACTIVULUI

Reactiv 1: R1

Tampon imidazol, pH 6,10	125 mmol/L
D-Glucoză	25 mmol/L
N-Acetil-L-Cisteină	25 mmol/L
Acetat de magneziu	12,5 mmol/L
NADP	2,4 mmol/L
EDTA	2,0 mmol/L
Hexokinază	≥ 6800 U/L
Azidă de sodiu	< 0,1 %

Concentrația anticorpului anti-CK-M conținut de reactivul 1 este suficient pentru a inhiba 2000 U/L din CK-M la 37°C.

Reactiv 2: R2

Tampon imidazol, pH 8,9	125 mmol/L
Creatină fosfat	250 mmol/L
ADP	15,2 mmol/L
AMP	23 mmol/L
Deadenozină pentafosfat	103 μmol/L
G-6-PDH	≥ 8800 U/L
Azidă de sodiu	< 0,1 %

MATERIALE NECESARE DAR NEFURNIZATE

- CKMB-0900, CK-MB CONTROL 4 x 3 mL
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Această trusă de reactivi este doar pentru utilizarea profesională, în scopul diagnosticării *in vitro*.
- Reactivii R1 și R2 sunt clasificați ca fiind periculoși (Imidazol).



PERICOL: Poate dăuna fătului. Procurați instrucțiuni speciale înainte de utilizare. Purtați mănuși de protecție/îmbrăcăminte de protecție/echipament de protecție a ochilor/echipament de protecție a feței. ÎN CAZ DE expunere sau de posibilă expunere: consultați medicul.

- Reactivii conțin azidă de sodiu care poate reacționa cu plumbul sau instalațiile din cupru pentru a forma potențiale azide metalice explozive. În momentul eliminării acestor reactivi, spălați întotdeauna cu apă din abundență pentru a preveni acumularea de azide.
- Pentru mai multe informații, consultați Fișa de date privind siguranța (SDS).
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați reactivii din truse diferite.

STABILITATEA REACTIVILOR

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

PREGĂTIREA ȘI STABILITATEA REACTIVULUI DE LUCRU

- Pregătire

Amestecați într-o fiolă goală (fiola goală furnizată cu ref. CMSL-0230) 4 volume de reactiv R1 cu 1 volum de reactiv R2.

- Stabilitate

1 zi la 20-25°C
2 săptămâni la 2-8°C

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor. (Consultați §DATE PRIVIND PERFORMANȚA).

DETERIORAREA REACTIVILOR

- Soluția de reactiv trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.

AMBALAJ DETERIORAT

Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

Referințe:

 CMSL-0230 4 x 25 mL
 CMSL-0410 2 x 62,5 mL
 CMSL-0430 4 x 62,5 mL

Compoziția trusei:
R1 4 x 20 mL+ **R2** 1 x 20 mL
R1 2 x 50 mL+ **R2** 1 x 26 mL
R1 4 x 50 mL+ **R2** 2 x 26 mL

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PROBE ⁽⁶⁾
Specimen

- Ser liber din hemoliză.
- A nu se utiliza alte specimene.

Avertismente și precauții

Conform buneii practici de laborator, puncția venoasă trebuie să fie efectuată înainte de administrarea de medicamente.

Depozitare și stabilitate

Eșantioanele trebuie analizate imediat sau depozitate protejate împotriva aerului și luminii 2 zile la 2-8°C, sau 1 lună la -20°C.

VALORI DE REFERINȚĂ ⁽³⁻⁵⁾

Ser (37°C): 0-25 U/L

Activitatea CK-MB trebuie să fie comparată cu activitatea CK totale

$$(CK-MB/CK \text{ totală}) \times 100 < 6\%$$

Următorii 3 factori sunt indicatori ai deteriorării mușchiului cardiac:

CK totală

Bărbați > 171 U/L

Femei > 145 U/L

CK-MB: > 25 U/L

Raport: (CK-MB/CK totală) x 100: 6 – 25%

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Acest reactiv poate fi utilizat pentru o procedură cu un singur reactiv.

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere.

ungime de undă 340 nm

Temperatură: 37°C

Citiți pe reactivul martor.

Reactiv de lucru	250 μL
Proba	10 μL

Amestecați și după o incubare de 130 de secunde, măsurați modificarea absorbției pe minut (ΔA/min.) timp de 159 secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

CALCUL
a) Activitatea CK totale:

Determinarea cu reactivul CK NAC SL.

b) Activitatea CK-MB:

La 340 nm, cu o cuveta cu calea luminii de 1 cm:

$$\text{Activitate (U/L)} = \Delta A / \text{min.} \times 8 \ 254$$

c) Procent activitate CK-MB în eșantion:

$$\% \text{ CK-MB} = \frac{\text{CK-MB}}{\text{CK totală}} \times 100$$

$$\text{Factor de conversie: U/L} \times 0,0167 = \mu\text{kat/L}$$

CONTROLUL CALITĂȚII

Pentru a verifica precizia testelor, vor fi utilizate serurile de control precum CK-MB CONTROL. Acest ser de control este pregătit din enzimă umană nemodificată.

Controlul calității trebuie efectuat și validat înainte ca eșantioanele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra E ale ELITech Clinical Systems

 - **Interval de măsurare**

Reactivul este liniar între 15 și 600 U/L.

 - **Limita de detecție** ⁽⁷⁾

Determinată conform protocolului SFBC, limita de detecție este egală cu 6 U/L.

 - **Precizie**

Reproductibilitate în interiorul ciclului	n	Medie	CV (%)
		U/L	
Nivel normal	20	11	16,3
Nivel patologic	20	152	1,0

Reproductibilitate între cicluri	n	Medie	CV (%)
		U/L	
Nivel normal	20	14	14,1
Nivel patologic	20	145	1,6

 - **Corelație**

A fost efectuat un studiu comparativ privind acest reactiv între Analizorul Selectra E ELITech Clinical Systems și Cobas Mira pe 60 de eșantioane de ser uman. Valorile acoperă intervalul linearității.

Concentrațiile eșantioanelor au fost între 11 și 1712 U/L (0,18 și 28,53 μkat/L).

Parametrii regresii liniare sunt după cum urmează:

Coeficient de corelație: (r)=0,9980

Regresie liniară: y=1,0007 x +1,3 U/L

 - **Limitări și interferențe** ^(2,7,8)

- Serurile hemolizate nu trebuie utilizate deoarece hemoliza semnificativă poate crește concentrația CK din cauza eliberării adenilat kinazei.

- metoda va măsura și orice izoenzimă CK-BB prezentă doar în ser sau complexată cu imunoglobuline (macro-CK).

Activitatea izoenzimei este de obicei neglijabilă, însă dacă este prezentă o cantitate semnificativă a activității CK-BB, activitatea CK-MB va fi supraestimată.

Referințe:

CMSL-0230 4 x 25 mL
 CMSL-0410 2 x 62,5 mL
 CMSL-0430 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL+ **R2** 1 x 20 mL
R1 2 x 50 mL+ **R2** 1 x 26 mL
R1 4 x 50 mL+ **R2** 2 x 26 mL



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- Nu raportați rezultatele în afara intervalului utilizabil.
- Conform recomandărilor SFBC, au fost efectuate unele studii pentru a stabili nivelul interferenței din diferiți compuși:
Bilirubină neconjugată: Polarizare negativă începând cu 9 mg/dL (153,9 μmol/L) pe serurile umane normale.
 Polarizare negativă începând cu 15 mg/dL (256,6 μmol/L) pe serurile umane patologice.
Bilirubină conjugată: Polarizare negativă începând cu 2 mg/dL (34,2 μmol/L) pe serurile umane normale.
 Polarizare negativă începând cu 6 mg/dL (102,6 μmol/L) pe serurile umane patologice.
Turbiditate: Nicio interferență semnificativă până la 600 mg/dL (6,78 mmol/L) echivalent trigliceride.
- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabale. ⁽⁹⁾
- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹⁰⁻¹¹⁾
- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.












- Stabilitatea la bord

Stabilitatea la bord: 7 zile

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- Young, D.S., *Effects of preanalytical variables on clinical laboratory tests*, 2nd Ed., AACC Press, (1997).
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SIMBOLURI

	Dispozitiv medical de diagnosticare in vitro.
	Consultați instrucțiunea de utilizare.
	Producător
	Limită de temperatură
	Număr de lot
	Data expirării
	Număr catalog
	Conținut
	Reactiv 1
	Reactiv 2
	Conformitate europeană

Notă :

 Doar pentru ref. **CMSL-0230**, utilizată cu software-ul Selectra TouchPro.

 CK-MB
400

 0
PIT-CMSL

 Modificare față de versiunea precedentă


Referințe:
FEFE-0230
FEFE-0600

Compoziția trusei:
R1 4 x 14,6 mL + R2 4 x 3,9 mL
R1 2 x 100 mL + R2 1 x 50 mL + Std 1 x 5 mL



FTRO-FEFE-v6 (12/2018)_FTCE-FEFE-4-v6

SCOPUL UTILIZĂRII

IRON FERENE ELITech Clinical Systems este conceput pentru deteriorarea cantitativ a fierului total în serul uman pentru diagnosticare *in vitro*.

SEMNIFICAȚIE CLINICĂ⁽¹⁻²⁾

În organism, 65-70 % din fier intră în compoziția hemoglobinei, 25% este stocat în celule sub forma unui complex fier-feritină și 3% este transportat prin transferină. Nivelele de fier seric cresc în hemocromatoză sau leziunile hepatice. Nivelele scăzute de fier seric pot fi asociate cu necesitățile crescute, o deficiență dietară sau afecțiuni gastrointestinale (diaree cronică, sângerare intestinală sau malabsorbție). Nivelele de fier seric sunt întotdeauna interpretate împreună cu datele privind saturația transferinei.

METODĂ

Colorimetrică – Ferene
Punct final.

PRINCIPIU⁽¹⁻²⁾

Fierul este eliberat din transferină în pH-ul acid ca ion feric Fe³⁺. Acesta este apoi redus de acidul ascorbic în ion feros Fe²⁺ și formează eventual un complex colorat cu Ferene. Absorbanța la 578 nm de complex Fier - Ferene este proporțională cu concentrația fierului din eșantion.

pH acid, Acid ascorbic

Transferină – (Fe³⁺)₂ → 2 Fe²⁺ + Transferină

Fe²⁺ + 3 Ferene → Albastru de Ferene (complex de fier)

COMPOZIȚIA REACTIVULUI

Reactiv: R1

Tiouree	120	mmol/L
Tampon de acetat (pH 4.5)	1	mol/L

Reactiv: R2

Ferene	3	mmol/L
Acid ascorbic	240	mmol/L
Tiouree	120	mmol/L

Standard: Std

Fier	100	µg/dL
	17,9	µmmol/L

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Echipamente generale de laborator.
- Analizor de biochimie echipat cu filtrele necesare. (Consultati § PROCEDURA).
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest reactiv este conceput doar pentru utilizarea profesională, în scopul diagnosticării *in vitro*.
- Reactivul R1 este clasificat ca periculos (C9-11-izozalcooli, C10-rich etoxilat; dodecan-1-ol etoxilat).



PERICOL: Provoacă iritarea pielii. Provoacă leziuni oculare grave. Purtați mănuși de protecție/îmbrăcăminte de protecție/echipament de protecție a ochilor/echipament de protecție a feței. ÎN CAZ DE CONTACT CU OCHII:

Clătiți cu atenție cu apă timp de mai multe minute. Scoateți lentilele de contact, dacă este cazul și dacă acest lucru se poate face cu ușurință. Continuați să clătiți. Sunați imediat la un CENTRU DE INFORMARE TOXICOLOGICĂ/un medic. ÎN CAZ DE CONTACT CU PIELEA: spălați cu multă apă și săpun. În caz de iritare a pielii: consultați medicul. Scoateți îmbrăcămintea contaminată și spălați-o înainte de reutilizare.

- Pentru mai multe informații, consultați Fișa de date privind siguranța (SDS).
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Standardul trebuie să fie imediat închis cu capacul pentru a preveni contaminarea și evaporarea.
- Nu interschimbați fiolele de reactiv din truse diferite.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiole perforate).

PROBE^(1,3)

Specimen

- Fără ser din hemoliză
- A nu se utiliza alte specimene.

Avvertisment și precauții

Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.



Referințe:
 FEFE-0230
 FEFE-0600

Compoziția trusei:
R1 4 x 14,6 mL + **R2** 4 x 3,9 mL
R1 2 x 100 mL + **R2** 1 x 50 mL + **Std** 1 x 5 mL



FTRO-FEFE-v6 (12/2018)_FTCE-FEFE-4-v6

Depozitare

Serurile sunt stabile timp de 7 zile la temperatura camerei, sau la 2-8°C sau 1 an la -20°C.

VALORI DE REFERINȚĂ (4)

Ser		
Nou-născuți	100-250 µg/dL	17,9-44,8 µmol/L
Infanți	40-100 µg/dL	7,2-17,9 µmol/L
Copii	50-120 µg/dL	9,0-21,5 µmol/L
Femei	50-170 µg/dL	9,0-30,4 µmol/L
Bărbați	65-175 µg/dL	11,6-31,3 µmol/L

Intervalul nivelelor de fier seric în cazul persoanelor sănătoase clinic poate fi influențat de un număr de factori bine-cunoscuți precum dieta, sexul, vârsta, ciclul menstrual, sarcina sau fluctuațiile circadiene.

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems.

Aplicațiile sunt disponibile la cerere

Lungime de undă 578-700 nm

Temperatură: 37°C

Citiți pe reactivul martor.

	CALIBRARE	TEST
Reactiv R1	240 µL	240 µL
Calibrator/Standard	30 µL	-
Proba	-	30 µL

Amestecați și citiți absorbanta (A1) după o incubare de 4 minute 40.

Reactiv R2	60 µL	60 µL
-------------------	-------	-------

Amestecați și citiți absorbanta (A2) după o incubare de 6 minute 30.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

Pentru utilizatorii Selectra ProXS, este nevoie de un filtru suplimentar de 700nm.

CALCUL

(A2-A1) Proba x n n=concentrație calibrator/standard

(A2-A1) Calibrator /Standard

Factor de conversie: µg/dL x 0,179 = µmol/L

CALIBRARE

Pentru referința FEFE-0600: Pentru calibrare, trebuie utilizat fie calibratorul multiparametric ELICAL 2 fie Standardul de fier de 100 µg/dL.

Pentru referința FEFE-0230: Pentru calibrare, utilizați calibratorul multiparametric ELICAL 2.

Valorile concentrației Standardului de fier de 10 µg/dL și calibratorului multiparametric ELICAL 2 sunt trasabile conform Materialului Standard de Referință SRM937 (al Institutului Național de Standarde și Tehnologie).

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este definită în raport cu materialul de referință NIST SRM937 (al Institutului Național de Standarde și Tehnologie).

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a asigura calitatea adecvată, vor fi utilizate serurile de control precum ELITROL I (control normal) și ELITROL II (control patologic). Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele locale și legale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems

- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁵⁾, intervalul de măsurare este între 20 și 1000 µg/dL (de la 3,6 la 179,1 µmol/L).

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁶⁾.

LoD=6 µg/dL (1,1 µmol/L)

LoQ=20 µg/dL (3,6 µmol/L)



Referințe:
FEFE-0230
FEFE-0600

Compoziția trusei:
R1 4 x 14,6 mL + R2 4 x 3,9 mL
R1 2 x 100 mL + R2 1 x 50 mL + Std 1 x 5 mL



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- Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁷⁾.

	n	Medie		În interiorul ciclului	Total
		µg/dL	µmol/L	CV (%)	
Nivel scăzut	80	43	7,7	2,0	5,5
Nivel mediu	80	137	24,5	0,4	3,2
Nivel înalt	80	248	44,4	0,7	3,1

- Corelație

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda colorimetrică Ferene) pe 99 de eşantioane de ser uman conform protocolului CLSI EP9-A2⁽⁸⁾.

Concentrațiile eşantioanelor au fost între 22 și 1048 µg/dL (3.9 și 187.7 µmol/L).

Parametrii regresiiilor liniare sunt după cum urmează:

Coefficient de corelație: (r)=1.000

Regresie liniară: $y=1,041 x - 2 \mu\text{g/dL}$
(0,4 µmol/L)

- Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽⁹⁾. Recuperarea este în intervalul ±10% din valoarea inițială a concentrației fierului de 40 µg/dL și 250 µg/dL.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30 mg/dL (513 µmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29.5 mg/dL (504 µmol/L).

Trigliceride: Nicio interferență semnificativă până la 3000 mg/dL (33.90 mmol/L).

Acid ascorbic: Nicio interferență semnificativă până la 20 mg/dL .

Cupru: Nicio interferență semnificativă până la 500 µg/dL (78,7 µmol/L).

Acid acetilsalicilic: Nicio interferență semnificativă până la 200 mg/dL.

Acetaminofen: Nicio interferență semnificativă până la 30.0 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabile.⁽¹⁰⁾
- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young.⁽¹¹⁻¹²⁾
- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Aceste date sunt definite pentru un volum de umplere de 14 ml într-un flacon de 25 ml pentru reactivul R1 și respectiv de 4 ml într-un flacon de 10 ml pentru reactivul R2 (Ref: FEFE-0230).

În cazul unui format diferit al flaconului și / sau a unui volum diferit de umplere, este responsabilitatea fiecărui laborator de a revalida stabilitatea la bord și frecvența de calibrare.

Stabilitatea la bord: 28 de zile

Frecvența calibrării: 14 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit, și după o operație de întreținere.

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IRON FERENE

Referințe:
FEFE-0230
FEFE-0600






Compoziția trusei:
R1 4 x 14,6 mL + **R2** 4 x 3,9 mL
R1 2 x 100 mL + **R2** 1 x 50 mL + **Std** 1 x 5 mL

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SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

	Conținut
	Reactiv 1
	Reactiv 2
	Standard
	Conformitate europeană


Notă:

Doar pentru ref. **FEFE-0230** , utilizată cu software-ul Selectra TouchPro.



Iron Ferene
510

0
FTCE-FEFE

 Modificare față de versiunea precedentă.

Referințe:

PASL-0230 4 x 25 mL
 PASL-0400 2 x 62,5 mL
 PASL-0420 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL + R2 4 x 5 mL
 R1 2 x 50 mL + R2 1 x 26 mL
 R1 4 x 50 mL + R2 2 x 26 mL



FTRO-PASL-v21(12/2018)_PIT-PASL-4-v21

SCOPUL UTILIZĂRII

ALP (DEA) SL ELITech Clinical Systems este conceput pentru determinarea cantitativă a fosfatazei alcaline în serul uman pentru de diagnosticare *in vitro*.

SEMNIFICAȚIE CLINICĂ⁽¹⁻²⁾

Fosfataza alcalină (ALP) corespunde unui grup de fosfataze care prezintă activitatea maximă la pH alcalin. ALP este larg distribuită în ficat, osteoblaste, epiteliul intestinal, rinichi și placentă.

Rata ALP crește fiziologic pentru copii și adolescenți în perioadele creșterii active, precum și pentru femeile în al treilea trimestru de sarcină.

Creșterile marcate ale ratei ALP sunt observate în cazul obstrucției extra-hepatice (calculi biliari, tumori...) și bolile osoase precum boala Paget și cancerul osteogenic osos.

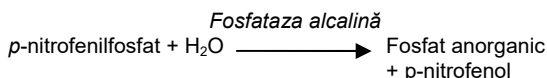
Activitatea PAL poate crește, de asemenea, moderat în cazul obstrucției intra-hepatice, hepatitei, cirozei, sau în cazul rahitismului, osteomalaciei, hiperparatiroidismului, vindecarea fracturilor osoase.

METODĂ⁽³⁻⁴⁾

Bazată pe metoda DGKC și SCE.
 Enzimatică. CINETICĂ.

PRINCIPIU⁽³⁻⁴⁾

În prezența Mg²⁺ și dietanolaminei ca acceptor al fosfatului, *p*-nitrofenilfosfatul este transformat de fosfatazele alcaline în fosfat și *p*-nitrofenol (compus galben).



COMPOZIȚIA REACTIVULUI

Reactiv1: R1

Dietanolamină, pH 10,2 1,4 mol/L
 Clorură de magneziu 0,625 mmol/L
 Azidă de sodiu < 0,1%

Reactiv 2: R2

p-nitrofenilfosfat 50 mmol/L
 Azidă de sodiu < 0,1 %

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550, ELICAL 2
- CONT-0060, ELITROL I
- CONT-0160, ELITROL II
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest reactiv este conceput doar pentru utilizarea profesională, în scopul diagnosticării *in vitro*.
- Reactivul R1 este clasificat ca periculos (2,2'-iminodietanol).



PERICOL: Poate provoca leziuni ale organelor în caz de expunere prelungită sau repetată. Provoacă leziuni oculare grave. Provoacă iritarea pielii. Nu inspirați ceața/vaporii/ spray-ul. Purtați mănuși de



protecție/îmbrăcăminte de protecție/echipament de protecție a ochilor/echipament de protecție a feței.

ÎN CAZ DE CONTACT CU OCHII: Clătiți cu atenție cu apă timp de mai multe minute. Scoateți lentilele de contact, dacă este cazul și dacă acest lucru se poate face cu ușurință. Continuați să clătiți. Sunați imediat la un CENTRU DE INFORMARE TOXICOLOGICĂ/un medic. ÎN CAZ DE CONTACT CU PIELEA: spălați cu multă apă. În caz de iritare a pielii: consultați medicul.

- Reactivii conțin azidă de sodiu care poate reacționa cu plumbul sau instalațiile din cupru pentru a forma potențiale azide metalice explozive. În momentul eliminării acestor reactivi, spălați întotdeauna cu apă din abundență pentru a preveni acumularea de azide.
- Pentru mai multe informații, consultați Fișa de date privind siguranța (SDS).
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați fiolele de reactiv din truse diferite.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

PROBE^(1,2,5)

Specimen

- Fără ser de hemoliză
- A nu se utiliza alte specimene.

Avertismente și precauții

- Conform bunei practici de laborator, puncția venoasă trebuie efectuată înainte de administrarea de medicamente.
- Este mai bine să analizați specimenul proaspăt (nu mai târziu de 4 ore după prelevare) și să le păstrați la temperatura camerei.

Depozitare și stabilitate

- Activitatea ALP poate crește în cazul în care specimenul refrigerat (2-8°C) sau înghețat (-20°C) este pus la temperatura camerei.
- Dacă speciimenele sunt înghețate în scopul depozitării prelungite, mutați-le la temperatura camerei cu 18 - 24 de ore înainte de analiză, pentru a activa complet enzima.
- Eșantioanele sunt stabile 1 săptămână la temperatura camerei, 1 săptămână la 2-8°C și 2 luni la -20°C.



Referințe:

PASL-0230 4 x 25 mL
 PASL-0400 2 x 62,5 mL
 PASL-0420 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL + R2 4 x 5 mL
 R1 2 x 50 mL + R2 1 x 26 mL
 R1 4 x 50 mL + R2 2 x 26 mL



FTRO-PASL-v21(12/2018)_PIT-PASL-4-v21

VALORI DE REFERINȚĂ ⁽⁶⁾

Ser (37°C):

Bărbați <270 U/L
 Femei <240 U/L

Valorile de referință pentru copii și adolescenți în timpul creșterii oaselor sunt mai mari decât pentru adulți.

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere.

Lungime de undă 405 nm

Temperatură: 37°C

Citiți pe reactivul martor.

Reactiv R1	200 µL
Proba	5 µL

Amestecați, așteptați 4 minute și 43 de secunde și adăugați:

Reactiv R2	50 µL
------------	-------

Amestecați și așteptați o incubație de 50 de secunde, măsurată variația absorbției pe minut ($\Delta A/\text{min.}$) timp de 133 secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert. .

CALCUL

$\Delta A \text{ Proba} \times n$ n=concentrație calibrator

$\Delta A \text{ Calibrator}$

Factor de conversie: U/L x 0,0167 = µkat/L

CALIBRARE

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este trasabilă conform măsurătorii manuale.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați Ş DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a asigura calitatea adecvată, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca eşantioanele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform liniilor directoare locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra XL ale ELITech Clinical Systems
- Interval de măsurare

Reactivul este liniar de la 20 la 900 U/L.

- Limita de detecție ⁽⁷⁾

Determinată conform protocolului SFBC, limita de detecție este egală cu 6 U/L.

- Precizie

Reproductibilitate în interiorul ciclului	n	Medie	CV (%)
		U/L	
Nivel 1	20	42	2,9
Nivel 2	20	146	0,8
Nivel 3	20	739	0,6

Reproductibilitate între cicluri	n	Medie	CV (%)
		U/L	
Nivel 1	20	38	5,5
Nivel 2	20	147	1,1
Nivel 3	20	760	1,3

- Corelație

A fost efectuat un studiu comparativ pe analizorul Selectra XL ELITech Clinical Systems între procedura cu un reactiv și procedura cu doi reactivi pe 30 de eşantioane de ser.

Valorile au fost între 17 și 886 U/L.

Parametrii regresiei liniare sunt după cum urmează:

Coefficient de corelație: (r)=0,9999

Regresie liniară: $y=1,9800x + 3,19$ U/L

- Limitări și interferențe ⁽⁷⁻⁸⁾

- Nu raportați rezultatele în afara intervalului utilizabil.

- Conform recomandărilor SFBC, au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși:

Bilirubină neconjugată: Nicio interferență semnificativă până la 36 mg/dL (615,8 µmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 25 mg/dL (427,6 µmol/L).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL (5 g/L).

Turbiditate: Nicio interferență semnificativă până la 600 mg/dL (6,78 mmol/L) echivalent trigliceride.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽⁹⁾
- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹⁰⁻¹¹⁾
- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 14 zile

Frecvența calibrării: 7 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit, și după o operație de întreținere.



Referințe:

PASL-0230 4 x 25 mL
 PASL-0400 2 x 62,5 mL
 PASL-0420 4 x 62,5 mL

Compoziția trusei:

R1 4 x 20 mL + R2 4 x 5 mL
 R1 2 x 50 mL + R2 1 x 26 mL
 R1 4 x 50 mL + R2 2 x 26 mL







FTRO-PASL-v21(12/2018)_PIT-PASL-4-v21

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- Young, D.S., *Effects of preanalytical variables on clinical laboratory tests*, 2nd Ed., AACC Press, (1997).
- Young, D.S., *Effects of drugs on clinical laboratory tests*, 4th Ed., AACC Press, (1995).

SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu exceptia celor prezentate mai jos.


	Conținut
	Reactiv 1
	Reactiv 2
	Conformitate europeană

Notă:

Doar pentru ref. **PASL-0230**, utilizată cu software-ul Selectra TouchPro.



Alkaline Phosphatase 0
 120 PIT-PASL

 :Modificare față de versiunea precedentă.



Referințe:

TGML-0425
TGML-0515
TGML-0700
TGML-0427
TGML-0497
TGML-0517
TGML-0707

Compoziția trusei:

R 6 x 50 mL
R 6 x 100 mL
R 4 x 250 mL
R 6 x 50 mL + Std 1 x 5 mL
R 1 x 100 mL + Std 1 x 5 mL
R 6 x 100 mL + Std 1 x 5 mL
R 4 x 250 mL + Std 1 x 5 mL

Referințe:

TGML-0250
TGML-0455

Compoziția trusei:

R 12 x 20 mL
R 6 x 45 mL

FTRO-TGML-v24 (10/2020)_PIT-TGML-4-v24



SCOPUL UTILIZĂRII

ELITech Clinical Systems TRIGLYCERIDES SL și TRIGLYCERIDES MONO SL NEW sunt reactiv de diagnostic *in vitro* destinat determinării cantitative a trigliceridelor din probele serul uman și plasmă.

SEMNIFICAȚIE CLINICĂ ⁽¹⁻²⁾

Trigliceridele constituie 95% din grăsimile depozitate în țesuturi și rolul lor principal este de a furniza energia pentru celulă. Acestea sunt sintetizate atât în intestin, din grăsimile dietetice și în ficat din carbohidrații dietetici, și sunt apoi transportate în sânge prin chilomicroni și VLDL.

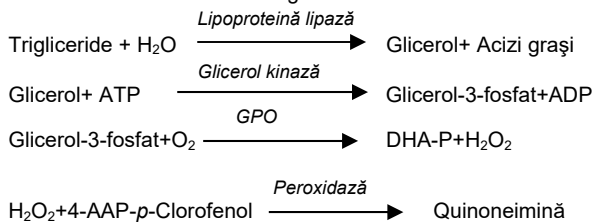
Nivelele înalte de trigliceride serice sunt asociate cu riscurile importante de ateroscleroză. Acestea pot fi cauzate de bolile precum diferitele afecțiuni ale metabolismului lipidic (hiperlipoproteinemia, deficiența activității lipazei, deficiența apolipoproteinei C-II), însă și de diabet, afecțiuni renale sau endocrine.

METODĂ ⁽³⁾

Enzimatică - colorimetrică. Punct final.

PRINCIPIU ⁽³⁾

Determinarea enzimatică a trigliceridelor conform următoarelor reacții:



GPO=Gliceori-3-fosfat oxidază
DHA-P= Dihidroxiaceton-fosfat
4-AAP=Amino-4-antipirină

COMPOZIȚIA

Reactiv: R

Tampon Good, pH 7,00

p-Clorofenol	2,7	mmol/L
ATP	3,15	mmol/L
4-Aminoantipirină	0,31	mmol/L
Lipoproteină lipază	≥ 2000	U/L
Glicerol kinază	≥ 500	U/L
Glicerol-3-fosfat oxidază	≥ 4000	U/L
Peroxidază	≥ 500	U/L
Azidă de sodiu	< 0,1	%

De asemenea, conține săruri de magneziu, FAD și surfactanți pentru performanță optimă.

Standard: Std. (Ref.: TGML-0427/0497/0517/0707)

Glicerol (echivalent trigliceride)	200	mg/dL
	2,26	mmol/L
Azidă de sodiu	< 0,1	%

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Soluție salină obișnuită (NaCl 9 g/L)
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest dispozitiv de diagnostic *in vitro* (Reactiv și Standardul) este destinat numai pentru uz profesional.
- Reactivul R și Standardul conține azidă de sodiu care poate reacționa cu plumbul sau instalațiile din cupru pentru a forma potențiale azide metalice explozive. În momentul eliminării acestor reactivi, spălați întotdeauna cu apă din abundență pentru a preveni acumularea de azide.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Standardul trebuie să fie imediat închis cu capacul pentru a preveni contaminarea și evaporarea.
- Pentru mai multe informații, Fișa de date privind siguranța (SDS) este disponibilă la cerere pentru utilizatorul profesional.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii și Standardul sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactiv trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

PROBE ⁽⁴⁾

Specimen

- Ser sau plasmă heparinizată de litiu de la pacienții care țin post (≥12 ore).
- Nu utilizați eșantioane icterice sau hemolizate.
- A nu se utiliza alte specimene.

Referințe:	Compoziția trusei:
TGML-0425	R 6 x 50 mL
TGML-0515	R 6 x 100 mL
TGML-0700	R 4 x 250 mL
TGML-0427	R 6 x 50 mL + Std 1 x 5 mL
TGML-0497	R 1 x 100 mL + Std 1 x 5 mL
TGML-0517	R 6 x 100 mL + Std 1 x 5 mL
TGML-0707	R 4 x 250 mL + Std 1 x 5 mL

Referințe:	Compoziția trusei:
TGML-0250	R 12 x 20 mL
TGML-0455	R 6 x 45 mL

FTRO-TGML-v24 (10/2020)_PIT-TGML-4-v24



Avertismente și precauții

- Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente. Prelevarea poate duce la rezultate false dacă este efectuată în timpul sau imediat după administrarea anumitor medicamente.
 - Colectați mostrele în tuburi și tampoane fără glicerol.
 - Separați de celule în termen de 2 ore.
- Depozitare și stabilitate**
- Eșantioanele sunt stabile între 5 și 7 zile, dacă sunt depozitate la 2-8°C, 3 luni de la -15°C până la -20°C și mai mulți ani la -70°C. Evitați înghețarea și dezghețarea repetate.

VALORI DE REFERINȚĂ (2)

NCEP (Programul Național American de Educație privind Colesterolul) a stabilit următoarea clasificare pentru nivelele de colesterol total, în funcție de riscul dezvoltării cardiopatiei coronariene:

Clasificarea riscurilor:

	Nivel (mg/dl)	Nivel (mmol/L)
Normal	< 150	1,69
Limită superioară	150-199	1,69-2,25
Mare	200-499	2,26-5,64
Foarte mare	≥ 500	5,65

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere.

Lungime de undă 505 nm

Temperatură: 37°C

Citiți pe reactivul maror.

	CALIBRARE	CALIBRARE	TEST
Reactiv R	300 µL	300 µL	300 µL
Apă distilată	3 µL	-	-
Calibrator	-	3 µL	-
Proba	-	-	3 µL

Amestecați și citiți absorbantele (A) după o incubare de 11 minute și 30 secunde.

- Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestei inserții.
- **Reactivii Triglycerides SL și Triglyceride Mono SL New pot fi contaminați de reactivul Cholesterol HDL SL 2G.**

Pentru a evita contaminarea pe Selectra ProM și ProXL, **programați incompatibilitățile după cum urmează:**

Software	Meniu	Parametru
TouchPro	Probe incompatibilities	Link / Cholecterol HDL SL 2G – Acide Solution
Altele	Needle incompatibility	Cholesterol HDL SL 2G <<HCl

Pentru alte instrumente Selectra Pro, repetați orice rezultate absurde după programarea unei spăiri a acelor.

- **Reactivul LIPASE SL este puternic contaminat cu reactivul Triglycerides SL.**

Pentru a evita contaminarea cuvetei pe instrumentele Selectra Pro, **programați următoarele incompatibilități:**

Software	Meniu	Parametru
TouchPro	Test Incompatibilities	Link / Triglycerides SL – Acid Solution
Altele	Cuvette Incompatibility	Triglycerides SL <<HCl

Pentru a evita contaminarea acelor pe instrumentele Selectra Pro, **nu programați Lipase SL și Triglycerides SL în același ciclu. Asigurați-vă că instrumentul revine la statusul „stand-by” înainte de începerea unui ciclu care conține Lipase SL.**

CALCUL

A Proba

_____ x n n = concentrație calibrator/standard

A Calibrator/
Standard

Factor de conversie: mg/dL x 0,0113= mmol/L
mg/dL x 0,01= g/L

CALIBRARE

Pentru referința TGML-0427/0497/0517/0707: Pentru calibrare, trebuie utilizat fie calibratorul multiparametric ELICAL 2 fie Standardul Triglycerides 200 mg/dL.

Pentru referința TGML-0250/0455/0425/0515/0700: Pentru calibrare, utilizați calibratorul multiparametric ELICAL 2.

Valorile concentrației Standardului Triglycerides 200 mg/dL și calibratorului multiparametric ELICAL 2 sunt trasabile în raport metoda de referință ID-GC-MS (Diluția izotopică – Spectrometria de masă cromatografie de gaz - spectrometrie de masă).

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a verifica precizia testelor, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca eșantioanele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform liniilor directe locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

Referințe:	Compoziția trusei:
TGML-0425	R 6 x 50 mL
TGML-0515	R 6 x 100 mL
TGML-0700	R 4 x 250 mL
TGML-0427	R 6 x 50 mL + Std 1 x 5 mL
TGML-0497	R 1 x 100 mL + Std 1 x 5 mL
TGML-0517	R 6 x 100 mL + Std 1 x 5 mL
TGML-0707	R 4 x 250 mL + Std 1 x 5 mL

Referințe:	Compoziția trusei:
TGML-0250	R 12 x 20 mL
TGML-0455	R 6 x 45 mL

FTRO-TGML-v24 (10/2020)_PIT-TGML-4-v24



DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems

Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁶⁾, Intervalul de măsurare este între 30 și 1000 mg/dL (0,34 – 11,30 mmol/L). Probele cu concentrații mai mari trebuie să fie diluate 1:5 cu soluție NaCl 9g/L (salină obișnuită) și re-evaluate. Utilizarea acestei proceduri extinde intervalul de măsurare până la 5 000 mg/dL (56,50 mmol/L.)

Pentru utilizatorii Selectra TouchPro, funcția „dilute” efectuează diluarea eșantioanelor automat. Rezultatele iau în considerare diluția.

Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁶⁾.

	Medie			În interiorul ciclului	Total
	n	mg/dL	mmol/L		
Nivel redus	80	44	0,50	2,0	3,8
Nivel mediu	80	131	1,48	0,9	2,3
Nivel înalt	80	267	3,02	1,2	2,4

Corelație

A fost efectuat un studiu comparativ între un Analizor Selectra ProM ELITech Clinical Systems și un alt echipament al sistemului aprobat de FDA (metoda enzimatică și colorimetrică) pe 99 eșantioane de ser uman conform protocolului CLSI EP9-A2⁽⁷⁾.

Valorile au fost între 30 și 957 mg/dL (0,34 și 10,81 mmol/L).

Parametrii regresiiilor liniare sunt după cum urmează:

Coeficient de corelație: (r)=0,999

Regresie liniară: $y=1,019 \cdot x + 1 \text{ mg/dL}$ (0,10 mmol/L)

Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽⁸⁾. Recuperarea este în intervalul ±10% din valoarea inițială a concentrației trigliceridelor de 133 mg/dL și 266 mg/dL.

Bilirubină neconjugată: Nicio interferență semnificativă până la 15 mg/dL (257 μmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 5,9 mg/dL (101 μmol/L)

Hemoglobină: Nicio interferență semnificativă până la 125 mg/dL.

Acid uric: Nicio interferență semnificativă până la 24,2 mg/dL (1440 μmol/L).

Acid ascorbic: Nicio interferență semnificativă până la 2,0 mg/dL. Concentrațiile peste nivelele terapeutice vor interfera și cauza rezultate eronate.

Metil-dopa: Nicio interferență semnificativă până la 1,0 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽⁹⁾

- Rezultatele pot fi fals reduse de nivele semnificative ale eșantionului de NAC (N-Acetil-Cisteină), NAPQI (metabolit de acetaminofen (paracetamol) sau metamizol).

- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹⁰⁻¹¹⁾

- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 28 zile

Frecvența calibrării: 14 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit, și după o operație de întreținere.

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- Young, D.S., *Effects of preanalytical variables on clinical laboratory tests*, 2nd edition, AACC Press (1997).
- Young D.S., *Effects of drugs on clinical laboratory tests*, 4th edition, AACC Press (1995).

Referințe:	Compoziția trusei:
TGML-0425	R 6 x 50 mL
TGML-0515	R 6 x 100 mL
TGML-0700	R 4 x 250 mL
TGML-0427	R 6 x 50 mL + Std 1 x 5 mL
TGML-0497	R 1 x 100 mL + Std 1 x 5 mL
TGML-0517	R 6 x 100 mL + Std 1 x 5 mL
TGML-0707	R 4 x 250 mL + Std 1 x 5 mL

Referințe:	Compoziția trusei:
TGML-0250	R 12 x 20 mL
TGML-0455	R 6 x 45 mL

FTRO-TGML-v24 (10/2020)_PIT-TGML-4-v24



☞ SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

CONT	Conținut
R	Reactiv
std	Standard
CE	Conformitate europeană
☞	Modificare față de versiunea precedentă

Notă

- Doar pentru ref. **TGML-0250/TGML-0455**, utilizată cu software-ul Selectra TouchPro.
- **Vezi ȘPROCEDURĂ:**
Risc de contaminare



Triglycerides 0
620 PIT-TGML

Os resultados devem estar dentro dos intervalos definidos. Se os valores ficarem fora dos intervalos definidos, cada laboratório deve tomar as medidas corretivas necessárias.

TRATAMENTO DOS RESÍDUOS

O descarte de todo material residual deve estar de acordo com os requisitos regulamentares locais, estaduais e federais (consulte a Ficha de dados de segurança (SDS)).

DESEMPENHO

Os desempenhos foram obtidos no Selectra ProM, seguindo as recomendações técnicas do CLSI, sob condições ambientais controladas.

- Precisão de medição

20.0 - 400.0 mg/dL (1.11 - 22.20 mmol/L).
As amostras com maiores concentrações devem ser diluídas 1:5 com solução de NaCl 9 g/L e ensaiado novamente. Este procedimento estabelece a faixa de medição até 2 000.0 mg/dL (111.01 mmol/L).

Não relatar resultados fora do intervalo de medição.

Para utilizadores do Selectra TouchPro, a função de «diluir» realiza a diluição do amostras automaticamente. Os resultados são tomados em consideração na diluição.

- Limite de detecção (LoD) e limite de quantificação (LoQ)

LoD = 0,2 mg/dL (0.01 mmol/L)
LoQ = 10,0 mg/dL (0.56 mmol/L)

- Precisão

Dados de imprecisão foram obtidos em 2 analisadores Selectra ProM ao longo de 20 dias (2 corridas por dia, testes realizados em duplicata). Os resultados representativos são apresentados abaixo :

	n	Média		Intra-série	Total
		mg/dL	mmol/L	CV (%)	
Nível 1	80	37.4	2.08	0.7	1.6
Nível 2	80	113.1	6.28	0.5	0.9
Nível 3	80	284.0	15.76	0.7	1.3

- Correlação

Foi realizado um estudo comparativo entre o reagente GLUCOSE PAP SL em um analisador Selectra ProM e um sistema similar disponível comercialmente em 100 amostras de soro humano.

As concentrações da amostra variaram de 22.2 para 384.9 mg/dL (1.23 - 21.36 mmol/L).

Os resultados são os seguintes:
Coeficiente de correlação: (r) = 1.000
Regressão linear: y = 0.989x + 1.1 mg/dL (0.06 mmol/L)

Limitações/Interferências

Estudos foram realizados para determinar o nível de interferência de diferentes compostos.

Os seguintes níveis de glicose foram testados : 36.0 mg/dL, 108.1 mg/dL e 400.0 mg/dL.

Uma interferência não significativa é definida por uma recuperação $\pm 10\%$ do valor inicial.

Bilirrubina não conjugada: Nenhuma interferência significativa até 6.0 mg/dL (103 $\mu\text{mol/L}$).

Bilirrubina conjugada: Nenhuma interferência significativa até 5.9 mg/dL (101 $\mu\text{mol/L}$).

Hemoglobina: Nenhuma interferência significativa até 300 mg/dL.

Triglicéridos : Nenhuma interferência significativa até 920 mg/dL (10.40 mmol/L).

Ácido ascórbico: Nenhuma interferência significativa até 2.0 mg/dL.

Ácido úrico : Nenhuma interferência significativa até 23.0 mg/dL (1 368 $\mu\text{mol/L}$).

Metil dopa: Nenhuma interferência significativa até 0.8 mg/dL.

L-Dopa: Induz falsamente resultados baixos em concentrações terapêuticas.

Tolazamida: Nenhuma interferência significativa até 40.0 mg/dL.

Acetaminofeno : Nenhuma interferência significativa até 30.0 mg/dL.

Não use amostras ictericas ou hemolisadas.

- Em casos muito raros, as gamopatas monoclonais (mieloma múltiplo), em particular, tipo IgM (macroglobulinemia de Waldenstrom) podem causar resultados não confiáveis.⁽⁷⁾

- Os resultados podem ser falsamente reduzidos em níveis significativos na amostra de NAC (*N*-acetilcisteína), NAPQI (metabólito do acetaminofeno (paracetamol)) ou metamazolo.

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young.⁽⁸⁻⁹⁾

- Estabilidade a bordo / frequência de calibração

Estabilidade a bordo: 28 dias

Frequência de calibração: 28 dias

Recalibre quando os lotes de reagentes mudarem, quando os resultados do controle de qualidade estiverem fora da faixa estabelecida e após uma operação de manutenção.

Estes desempenhos foram obtidos utilizando o analisador ELITech Selectra ProM. Os resultados podem variar se um instrumento diferente ou um procedimento manual for usado.

Os desempenhos de aplicações não validados pela ELITech não são garantidos e devem ser definidos pelo usuário.

DECLARAÇÃO DE INCIDENTE GRAVE

Notifique o fabricante (através do seu distribuidor) e a autoridade competente do Estado-Membro da união europeia em que o usuário e / ou o paciente está estabelecido, de qualquer incidente grave que tenha ocorrido em relação ao dispositivo.
Para outras jurisdições, a declaração de incidente grave deve estar de acordo com os requisitos regulamentares locais, estaduais e federais.
Ao relatar um incidente grave, você fornece informações que podem contribuir para a segurança de dispositivos médicos *in vitro*.

ASSISTÊNCIA TÉCNICA

Entre em contato com o seu distribuidor local ou com a ELITech Clinical Systems SAS. (CCsupport@elitechgroup.com).

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SYMBOLS/SYMBOLS/ SÍMBOLOS/SÍMBOLOS

- Les symboles utilisés sont décrits dans la norme ISO 15223-1 hormis ceux présentés ci-dessous.

- Symbols used are defined on ISO 15223-1 standard, except those presented below.

- Los símbolos utilizados son descritos en la norma ISO 15223-1 a la excepción de los presentados a continuación.

- Os símbolos utilizados são definidos na norma ISO 15223-1, exceto os apresentados abaixo.

CONT	Contient Contient Contiene Conteúdo
R	Réactif Reagent Reactivo Reagente
Std	Standard Standard Estándar Padrão
	Modification par rapport à la version précédente Modification from previous version Modificación con respecto a la versión anterior Modificação relativamente à versão anterior
CE	Conformité Européenne European Conformity Conformidad Europea Conformidade Europeia

GLUCOSE PAP SL

GPSL-0497
GPSL-0507
GPSL-0707
GPSL-0250
GPSL-0455
GPSL-0500
GPSL-0700

GPSL

R	1 x 100 mL	+	Std	1 x 5 mL
R	6 x 100 mL	+	Std	1 x 5 mL
R	4 x 250 mL	+	Std	1 x 5 mL
R	12 x 20 mL			
R	6 x 45 mL			
R	6 x 100 mL			
R	4 x 250 mL			



Acétaminophène: Aucune interférence significative jusqu'à 30.0 mg/dL.

Ne pas utiliser d'échantillons icteriques ou hémolysés.

- Dans des cas très rares, les gammopathies monoclonales (myélome multiple), en particulier de type IgM (Macroglobulinémie de Waldenström) peuvent être à l'origine de résultats peu fiables.⁽⁷⁾

- Les résultats peuvent être faussement abaissés dans les échantillons contenant des niveaux significatifs de NAC (*N*-Acétyl-Cystéine), de NAPQI (métabolite de l'acétaminophène (paracétamol)) ou de metamazolo.

- D'autres substances et médicaments peuvent interférer. Certains d'entre eux sont répertoriés dans les revues publiées par Young.⁽⁸⁻⁹⁾

Stabilité à bord / fréquence de calibration

Stabilité à bord : 28 jours

Frequéncia de calibração : 28 jours

La nouvelle calibration doit être effectuée après chaque changement de lot de réactif, lorsque les résultats du ou des contrôles de qualité sont hors de l'intervalle établi, et après une opération de maintenance.

Ces performances ont été définies sur un automate ELITech Selectra ProM. Les résultats peuvent varier si le réactif est utilisé sur un automate différent ou en méthode manuelle.

Les performances obtenues à partir d'applications non validées par ELITech ne peuvent être garanties et doivent être définies par l'utilisateur.

DECLARATION DES INCIDENTS GRAVES

Veillez notifier au fabricant (par l'intermédiaire de votre distributeur) et à l'autorité compétente de l'Etat membre de l'union européenne dans lequel l'utilisateur et/ou le patient est établi, les cas d'incident grave survenu en lien avec le dispositif.

Pour les autres juridictions, la déclaration d'incident grave doit être effectuée conformément aux exigences réglementaires locales, d'état et fédérales.

En signalant les incidents graves, vous contribuez à fournir davantage d'informations sur la sécurité des dispositifs médicaux de diagnostic *in vitro*.

ASSISTANCE TECHNIQUE

Contactez votre distributeur local ou ELITech Clinical Systems SAS (CCsupport@elitechgroup.com).

English - EN

INTENDED USE

ELITech Clinical Systems GLUCOSE PAP SL is an *in vitro* diagnostic reagent intended for the quantitative determination of glucose in human serum and plasma samples on analyzers or semi-automatic analyzers. The standard is intended for the calibration of the reagent.

These *in vitro* diagnostic devices are for professional use only.

CLINICAL SIGNIFICANCE (1-3)

Glucose is the main source of energy for the human body. Glucose is converted either into glycogen or into triglycerides to be stored. Glucose blood level is mainly regulated by two antagonist hormones: insulin and glucagon.

Glycemia disorders appear mostly in type I or type II diabetes as well as in gestational diabetes. They can be also associated to various endocrinal, pancreatic or hepatic disorders, or linked to drugs.

In normal health condition, glucose is filtered then reabsorbed by kidneys and is therefore not present in urine. Elevated concentrations in urine are observed when the blood concentration is high or in case of impaired tubular reabsorption. Glucose measurement in blood is indicated for diabetes, in screening, diagnosis or follow-up of patients. It is also indicated to monitor patients with symptoms of hyperglycemia or hypoglycemia.

LIMITATION OF USE

For diabetes assessment, the collection conditions and interpretation of serum glucose concentrations should follow local recommendations such as those published by WHO.⁽⁴⁾

The quantitative assay of glucose alone can not be used to diagnose a disease or a specific pathology. The results must be interpreted in conjunction with other diagnostic test results, clinical findings and the patient's medical history.

PRÉPARATION

Le réactif et le standard sont prêts à l'emploi.

DÉTÉRIORATION DU PRODUIT

- Le produit doit être limpide. Tout trouble serait le signe d'une détérioration du produit.
- Ne pas utiliser le produit s'il y a des signes évidents de contamination ou de détérioration (ex : particules).
- Un flacon endommagé peut avoir un impact sur les performances du produit. Ne pas utiliser le produit si les flacons présentent des signes physiques de détérioration (par exemple, fuite, flacon percé).

ÉCHANTILLONS

Echantillons requis (1)

- Sérum.
- Plasma (héparine de lithium).
- Plasma (fluorure de sodium / oxalate de potassium (inhibiteurs de la glycolyse)).
- L'utilisation de toute autre type d'échantillon doit être validée par le laboratoire.

Avvertissements et précautions

- Les échantillons prélevés sans inhibiteur de la glycolyse doivent être séparés des cellules rapidement après le prélèvement pour limiter la perte de glucose (diminution de 5-7% par heure dans le sang total à température ambiante).⁽¹⁾
- La méthode PAP n'est pas appropriée pour la mesure du glucose dans l'urine en raison des quantités importantes d'interfèrents endogènes présents dans cette matrice.^(1,2)
- Les échantillons doivent être prélevés selon les Bonnes Pratiques de Laboratoire et les guides appropriés qui sont mis en place.
Stockage et stabilité (1,6)
Sérum / Plasma (héparine de lithium)
- 8 heures à température ambiante
- 3 jours à 2-8°C
Plasma (fluorure de sodium / oxalate de potassium)
- 2 jours à température ambiante
- 7 jours à 2-8°C

VALEURS DE RÉFÉRENCE (3)

Sérum/plasma	mg/dL	mmol/L
Nouveau-nés	30 – 60	1.7 – 3.3
Enfants	60 – 100	3.3 – 5.6
Adultes 18-60 ans	74 – 106	4.1 – 5.9
Adultes 60-90 ans	82 – 115	4.6 – 6.4

Remarque : Les valeurs ci-dessus ne sont données qu'à titre indicatif. Il est recommandé à chaque laboratoire d'établir et de maintenir ses propres valeurs de référence par rapport à la population visée.

Remarque : Les valeurs ci-dessus ne sont données qu'à titre indicatif. Il est recommandé à chaque laboratoire d'établir et de maintenir ses propres valeurs de référence par rapport à la population visée.

PROCÉDURE

Procédure manuelle

Longueur d'onde : 505 nm
Trajet optique : 1 cm
Ratio échantillon/réactif : 1:100
Température : 37 °C
Lire contre le blanc réactif.

	CALIBRATION	DOSAGE
Réactif R	1 000 μL	1 000 μL
Calibrant/Standard	10 μL	-
Echantillon	-	10 μL

Mélanger et lire les absorbances (A) après 10 minutes d'incubation.

Procédure sur automate

Ces réactifs peuvent être utilisés sur différents automates. Pour les automates ELITech Selectra, les applications validées sont disponibles sur demande. Avec le logiciel Selectra TouchPro, utilisez l'application incluse dans le code barre disponible à la fin de cette notice.

CALCUL

$$\frac{A_{\text{Echantillon}}}{A_{\text{Standard/Calibrant}}} \times n \quad n = \text{concentration du standard/Calibrant}$$

Facteur de conversion: mg/dL x 0.055= mmol/L

CALIBRATION

Pour les références GPSL-0497/0507/0707 : ELICAL 2 et Glucose Standard 100 mg/dL sont traçables par rapport à la méthode de référence ID-MS (Dilution Isotopique - Spectrométrie de Masse).

Pour les références 0250/0455/0500/0700 : ELICAL 2 est traçable par rapport à la méthode de référence ID-MS (Dilution Isotopique - Spectrométrie de Masse).

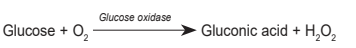
Frequéncia de calibração : La fréquence de calibration est spécifique à chaque automate (se référer au § PERFORMANCES).

Note/Nota	
- Uniquement pour les réf. GPSL-0455/GPSL-0250, utilisées avec le logiciel Selectra TouchPro.	
- Only for ref. GPSL-0455/GPSL-0250, used with Selectra TouchPro software.	
- Únicamente para las ref. GPSL-0455/GPSL-0250, utilizadas con el software Selectra TouchPro.	
- Somente para ref. GPSL-0455/GPSL-0250, usados com o Selectra TouchPro.	
	
Glucose	0
480	PIT-GPSL



► METHOD & PRINCIPLE ⁽⁶⁾

Enzymatic / PAP - End Point.



► COMPOSITION

Reagent: R
Phosphate buffer, pH 7.4
Phenol 10 mmol/L
4-Aminoantipyrine 0.3 mmol/L
Glucose Oxidase ≥ 10 000 U/L
Peroxidase ≥ 700 U/L
Sodium azide < 0.1 % (w/w)
Standard: Std (ref : G^{PSL-0497/0507/0707})
D-Glucose 100 mg/dL 5.55 mmol/L

MATERIALS REQUIRED BUT NOT PROVIDED

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II

- Normal saline solution (NaCl 9 g/L).
- Analyzers or semi-automatic analyzers.
- General Laboratory equipment (e.g. pipette).
- Do not use materials that are not required as indicated above.

PRECAUTIONS FOR USE AND WARNINGS - Consult Safety Data Sheet (SDS) for a proper handling.

- Reagent R contains sodium azide which may react with lead or copper plumbing to form potentially explosive metal azides. When disposing of these reagents always flush with copious amounts of water to prevent azide buildup.

- Take normal precautions and adhere to good laboratory practice.

- Use clean or single use laboratory equipment only to avoid contamination.

►STABILITY

Store at 2-8 °C and protect from light. Do not freeze. Do not use after expiration dates indicated on the vial labels.

The standard should be immediately and tightly capped to prevent contamination and evaporation.

On board stability :

The on-board stability is specific for each analyzer. (Refer to § PERFORMANCE DATA).

PREPARATION

The reagent and standard are ready to use.

PRODUCT DETERIORATION

- The product should be clear. Cloudiness would indicate deterioration.

- Do not use the product if there is visible evidence of contamination or damage (e.g. particle matter).

- Damage to the product container may impact on product performance. Do not use the product if there is physical evidence of deterioration (e.g. leakages or punctured container).

►SAMPLES

Specimen ⁽¹⁾
– Serum.
- Plasma (lithium heparin).
- Plasma (Sodium fluoride / potassium oxalate (glycolysis inhibitors)).
- Using any other specimen type should be validated by the laboratory.

Warnings and precautions

- In samples collected without glycolysis inhibitors, blood cells must be removed rapidly to prevent the glucose loss (5% to 7% per hour in whole blood at room temperature).⁽¹⁾

- PAP method is not suitable for glucose measurement in urine due to the large amount of interfering endogenous substances present in this matrix. ^(1,2)

- Samples should be collected in accordance with Good Laboratory Practice and appropriate guidelines that may be in place.

Storage and stability ^(1,6)
Serum / Plasma (lithium heparin)
- 8 hours at room temperature
- 3 days at 2-8°C
Plasma (sodium fluoride / potassium oxalate)
- 2 days at room temperature
- 7 days at 2-8°C

►REFERENCE VALUES ⁽⁶⁾

<i>Serum/plasma</i>	mg/dL	mmol/L			
Neonates	30 – 60	1.7 – 3.3			
Children	60 – 100	3.3 – 5.6			
Adults 18-60 y/o	74 – 106	4.1 – 5.9			
Adults 60-90 y/o	82 – 115	4.6 – 6.4			

Note : The quoted range should serve as a guide only. It is recommended that each laboratory verifies this range or establishes a reference interval for the intended population.

PROCEDURE

Manual Procedure

Wavelength : 505 nm
Optical path : 1 cm
Sample/ Reagent ratio : 1:100
Temperature: 37 °C
Read against reagent blank.

	CALIBRATION	TEST
Reagent R	1 000 µL	1 000 µL
Standard/ Calibrator	10 µL	-
Sample	-	10 µL

Mix and read the absorbances (A) after an incubation of 10 minutes.

Automatic Procedure

These reagents may be used on several automatic analyzers. For ELITech Selectra Analyzers, validated applications are available on request. For Selectra TouchPro software, use the application included in the barcode available at the end of this insert.

CALCULATION

A Sample _____ x n n = Calibrator/ standard
A Calibrator/ _____ concentration
Standard

Conversion factor : mg/dL x 0.0555 = mmol/L

CALIBRATION

For the references G^{PSL-0497/0507/0707}: ELICAL 2 and *Glucose Standard 100 mg/dL* are traceable to ID-MS (Isotope Dilution - Mass Spectrometry) reference method.

For the references G^{PSL-0250/0455/0500/0700} : ELICAL 2 is traceable to ID-MS (Isotopie Dilution - Mass Spectrometry) reference method.

Calibration frequency : The calibration is specific for each analyzer. (Refer to § PERFORMANCE DATA).

QUALITY CONTROL

It is recommended that quality control sera such as ELITROL I and ELITROL II be used to monitor the performance of the assay.

Controls have to be performed :
- prior to assaying patient samples,
- at least once per day,
- after every calibration,
- and/or in accordance with laboratory and regulatory requirements.

Results should be within the defined ranges. If values fall outside of the defined ranges, each laboratory should take necessary corrective measures.

WASTE MANAGEMENT

Disposal of all waste material should be in accordance with local, state and federal regulatory requirements (please refer to the Safety Data Sheet (SDS)).

PERFORMANCES

Performances were obtained on Selectra ProM, following CLSI technical recommendations, under controlled environmental conditions.

- Measuring range

20.0 – 400.0 mg/dL (1.11 - 22.20 mmol/L)
Samples having greater concentrations should be diluted 1:5 with NaCl 9 g/L solution and re-assayed. This procedure extends the measuring range up to 2 000.0 mg/dL (111.01 mmol/L).

Do not report results outside this extended range.

For users with Selectra TouchPro software, the «dilute» function performs the sample dilution automatically. Results take the dilution into account.

Limit of Detection (LoD) and Limit of Quantification (LoQ)

LoD = 0.2 mg/dL (0.01 mmol/L)
LoQ = 10.0 mg/dL (0.56 mmol/L)

- Precision

Imprecision data has been obtained on 2 Selectra ProM analyzers over 20 days (2 runs per day, tests performed in duplicate).

Representative results are presented below :

		Mean	Within-run	Total	
	n	mg/dL	mmol/L	CV (%)	
Level 1	80	37.4	2.08	0.7	1.6
Level 2	80	113.1	6.28	0.5	0.9
Level 3	80	284.0	15.76	0.7	1.3

- Correlation

A comparative study has been performed between GLUCOSE PAP SL reagent on a Selectra ProM analyzer and a similar commercially available system on 100 human serum samples.

The sample concentrations ranged from 22.2 to 384.9 mg/dL (1.23 - 21.36 mmol/L).

The results are as follows :
Correlation coefficient : (r) = 1.000
Linear regression: y = 0.989x + 1.1 mg/dL (0.06 mmol/L)

►. Limitations/Interferences

- Studies have been performed to determine the level of interference from different compounds.

The following glucose levels were tested: 36.0 mg/dL, 108.1 mg/dL and 400.0 mg/dL.

No significant interference is defined by a recovery ±10% of the initial value.

Unconjugated bilirubin: No significant interference up to 6.0 mg/dL (103 µmol/L).

Conjugated bilirubin: No significant interference up to 5.9 mg/dL (101 µmol/L).

Hemoglobin: No significant interference up to 300 mg/dL.

Triglycerides: No significant interference up to 920 mg/dL (10.40 mmol/L).

Ascorbic acid: No significant interference up to 2.0 mg/dL.

Uric acid: No significant interference up to 23.0 mg/dL (1 368 µmol/L).

Methyl dopa: No significant interference up to 0.8 mg/dL.

L-Dopa: Induces falsely low results at therapeutic concentrations.

Tolazamide: No significant interference up to 40.0 mg/dL.

Acetaminophen: No significant interference up to 30.0 mg/dL.

Do not use icteric or hemolyzed samples.

- In very rare cases, monoclonal gammopathies (multiple myeloma), in particular IgM type (Waldenström’s macroglobulinemia) can cause unreliable results.⁽⁷⁾

- Results can be falsely lowered by significant levels in the sample of NAC (*N*-Acetyl-Cysteine), NAPQI (metabolite of acetaminophene (paracetamol)) or metanzolol.

- Many other substances and drugs may interfere. Some of them are listed in reviews published by Young.⁽⁸⁻⁹⁾

- On board stability/Calibration frequency

On Board Stability: 28 days

Calibration frequency: 28 days

Recalibrate when reagent lots change, when quality control results fall outside the established range and after a maintenance operation.

These performances have been obtained using ELITech Selectra ProM analyzer. Results may vary if a different instrument or a manual procedure is used. The performances of applications not validated by ELITech are not warranted and must be defined by the user.

►DECLARATION OF SERIOUS INCIDENT

Please notify the manufacturer (through your distributor) and competent authority of the Member State of the european union in which the user and/or the patient is established, of any serious incident that has occurred in relation to the device. For other jurisdictions, the declaration of serious incident should be in accordance with local, state and federal regulatory requirements. By reporting a serious incident, you provide information that can contribute to the safety of *in vitro* medical devices.

►TECHNICAL ASSISTANCE

Contact your local distributor or ELITech Clinical Systems SAS (CCsupport@elitechgroup.com).

Español - ES

►USO PREVISTO

ELITech Clinical Systems GLUCOSE PAP SL es un reactivo de diagnóstico *in vitro* diseñado para la determinación cuantitativa de glucosa en muestras de suero y plasma humanos en equipos automatizados o equipos semiautomáticos. El estándar está diseñado para la calibración del reactivo.

Estos dispositivos de diagnóstico *in vitro* están destinados únicamente para los profesionales.

►SIGNIFICADO CLÍNICO ⁽¹⁻³⁾

La glucosa es la principal fuente de energía para el cuerpo humano. La glucosa se convierte en glucógeno o en triglicéridos para ser almacenados. El nivel de glucosa en sangre está regulado principalmente por dos hormonas antagonistas: la insulina y glucagon. Los trastornos de la glucemia aparecen principalmente en la diabetes tipo I o tipo II, así como en la diabetes gestacional. Pueden ser también asociado a diversos trastornos endocrinos, pancreáticos o hepáticos, o vinculado a fármacos.

En condición de salud normal, la glucosa se filtra y luego es absorbida por los riñones y, por lo tanto, no está presente en la orina. Se observan concentraciones elevadas en la orina cuando la concentración en sangre es alta o en caso de reabsorción tubular alterada.

La medición de glucosa en sangre es indicada para la diabetes, en la detección, el diagnóstico o el seguimiento de los pacientes. También es adecuada para monitorear pacientes con síntomas de hiperglucemia o hipoglucemia.

►LÍMITE DE UTILIZACIÓN

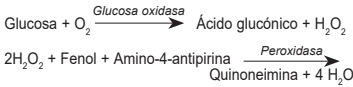
Para la evaluación de la diabetes, las condiciones de muestreo y la interpretación de las concentraciones de glucosa en suero deben seguir las recomendaciones locales publicadas por la OMS (WHO).⁽⁴⁾

La cuantificación de glucosa no puede ser utilizado solo para diagnosticar una enfermedad o patología específica.

Los resultados siempre deben compararse con los resultados de otras pruebas de diagnóstico, exámenes clínicos y el historial médico del paciente.

►MÉTODO & PRINCIPIO ⁽⁴⁾

Enzimático / PAP - Point final.



►COMPOSICIÓN

Reactivo : R

Tampón Fosfato pH 7.4

Fenol	10 mmol/L
Amino-4-antipirina	0.3 mmol/L
Glucosa oxidasa	≥ 10 000 U/L
Peroxidasa	≥ 700 U/L
Azida sódica	< 0.1 % (p/p)
E estándar : Std (ref : G^{PSL-0497/0507/0707})	
D-Glucosa	100 mg/dL 5.5 mmol/L

MATERIALES REQUERIDOS PERO NO INCLUIDOS

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II

- Solución salina normal (NaCl 9 g/L).
- Equipos automatizados o equipos semiautomáticos.
- Equipamiento general de laboratorio (p. ej. pipeta).
- No utilice materiales que no se requieren, tal como se indica anteriormente.

PRECAUCIONES DE USO Y ADVERTENCIAS

- Consulte la Hoja de Datos de Seguridad (SDS) para un manejo adecuado.

- El reactivo R contiene azida sódica que puede reaccionar con el plomo o el cobre de la tubería y formar potencialmente azidas metálicas explosivas. Cuando se elimine el reactivo enjuague con agua abundantemente para prevenir la acumulación de azidas.
- Tome las precauciones normales y respete las buenas prácticas de laboratorio.
- Para evitar contaminaciones utilizar equipo nuevo o completamente limpio.

►ESTABILIDAD

Conservar a 2-8 °C y protegidos de la luz. No congelar.

No utilice después de la fecha de caducidad indicada en la etiqueta de los frascos.

El estándar debe cerrarse inmediatamente y correctamente para evitar contaminación y evaporación.

Estabilidad en el equipo:

La estabilidad es específica para cada equipo. (Referirse al § DATOS DE RENDIMIENTO).

PREPARACIÓN

El reactivo y el estándar están listos para su uso.

DETERIORACIÓN DEL PRODUCTO

- El producto debe ser claro. Turbidez indicaría deterioro.

- No utilice el producto si este presenta signos evidentes de contaminación o deterioro (p. ej. partículas).

- Un frasco dañado puede tener un impacto en el rendimiento del producto. No utilice el producto si este tiene signos físicos de deterioro (p. ej, fugas, frasco perforado).

►MUESTRAS

Muestras requeridas ⁽¹⁾

- Suero
- Plasma (heparina de litio).
- Plasma (fluoruro de sodio / oxalato de potasio (Inhibidor de la glicólisis)).
- El uso de cualquier otro tipo de muestra debe ser validado por el laboratorio.

Advertencias y precauciones

- Las muestras tomadas sin un inhibidor de la glicólisis deben separarse de las células inmediatamente después de la recolección para limitar la pérdida de glucosa (5-7% de disminución por hora en sangre total a temperatura ambiente). ⁽¹⁾

- El método PAP no es adecuado para la medición de glucosa en la orina debido a la gran cantidad de sustancias endógenas interferentes presentes en esta matriz.^(1,2)

- Las muestras deben de tomarse de acuerdo con las Buenas Prácticas de Laboratorio y las guías apropiadas establecidas.

Conservación y estabilidad ^(1,6)

Suero / Plasma (heparina de litio)
- 8 horas a temperatura ambiente
- 3 días a 2-8°C
Plasma (fluoruro de sodio / oxalato de potasio)
- 2 días a temperatura ambiente
- 7 días a 2-8°C

►VALORES DE REFERENCIA ⁽⁶⁾

<i>Suero/plasma</i>	mg/dL	mmol/L			
Recien nacidos	30 – 60	1.7 – 3.3			
Niños	60 – 100	3.3 – 5.6			
Adultos 18-60 años	74 – 106	4.1 – 5.9			
Adultos 60-90 años	82 – 115	4.6 – 6.4			

Nota : Los valores anteriores son solo indicativos. Se recomienda que cada laboratorio establezca y mantenga sus propios valores de referencia en relación con la población destinataria.

PROCEDIMIENTO

Procedimiento manual

Longitud de onda : 505 nm
Trayectoria óptica : 1 cm
Ratio muestra/reactivo : 1:100
Temperatura: 37 °C

Leer contra blanco reactivo

	CALIBRACIÓN	PRUEBA
Reactivo R	1 000 µL	1 000 µL
E estándar/ Calibrator	10 µL	-
Muestra	-	10 µL

Mezclar y leer las absorbancias (A) después de una incubación de 10 minutos.

Procedimiento automático

Estos reactivos pueden ser utilizados en varios equipos. Para los equipos ELITech Selectra, las aplicaciones validadas están disponibles sobre pedido. Para el software Selectra TouchPro, use la aplicación incluida en el código de barras disponible al final de este inserto.

CÁLCULO

A Muestra _____ x n n = concentración del estándar/ calibrador
A Estándar / _____ calibrador

Factor de conversión: mg/dL x 0.0555 = mmol/L

CALIBRACIÓN

Para las referencias G^{PSL-0497/0507/0707}: ELICAL 2 o *el estándar Glucose Standard 100 mg/dL* son trazables al método de referencia DI-EM (Dilución Isotópica - Espectrometría de Masas)
Para las referencias G^{PSL-0250/0455/0500/0700} : ELICAL 2 es trazable al método de referencia DI-EM (Dilución Isotópica - Espectrometría de Masas).

Frecuencia de calibración : la frecuencia de calibración es específica para cada equipo (referirse al § DATOS DE RENDIMIENTO).

CONTROL DE CALIDAD

Es recomendado que sueros de control tales como ELITROL I y ELITROL II sean usados para monitorear el rendimiento de las pruebas.

Los controles deben realizarse :

- antes que las muestras del paciente sean evaluadas,
- por lo menos una vez al día,
- después de cada calibración,
- *yo* en acuerdo con el laboratorio y los requerimientos regulatorios.

Los resultados deben de encontrarse en el rango definido. Si los valores se encuentran fuera del mismo, cada laboratorio deberá tomar las medidas correctivas necesarias.

TRATAMIENTO DE LOS RESIDUOS

Todos los materiales de desecho deben eliminarse de acuerdo con los requisitos regulatorios locales, estatales y federales. (dirijase a la hoja de seguridad (SDS)).

RENDIMIENTO

El rendimiento fue obtenido en un Selectra ProM, siguiendo las recomendaciones técnicas del CLSI, bajo condiciones ambientales controladas.

- Rango analítico

20.0 – 400.0 mg/dL (1.11 - 22.20 mmol/L)
Las muestras que tengan concentraciones mayores deben diluirse 1:5 con una solución de NaCl 9 g/L y volver a analizarse.

PREPARAÇÃO

O reagente e o padrão estão prontos a usar.

DETERIORAÇÃO DO PRODUTO

- Esses produtos devem ser claros. Qualquer turidez seria sinal de deterioração do produto.

- Não use o produto se houver evidência visível de contaminação ou dano (por exemplo, partículas).
- Danos ao recipiente de produto podem afetar o desempenho do produto. Não use o produto se houver evidência física de deterioração (por exemplo, vazamentos ou recipiente perfurado).

AMOSTRAS

Amostras ^(1,6)

- Soro
- Plasma (heparina de lítio).
- Urina
- O uso de qualquer outro tipo de amostra deve ser validado pelo laboratório.

Aviso e precauções

- O soro deve ser separado das células o mais rápido possível. ^(1,2)

- As amostras devem estar isentas de hemólise ^(1,2)

- Após a coleta, as amostras de urina devem ser acidificadas com ácido clorídrico 6N até um pH <2 para evitar a precipitação de sal de magnésio. ⁽¹⁾

As amostras devem ser coletadas de acordo com as Boas Práticas de Laboratório e com as diretrizes apropriadas que podem estar em vigor.

Armazenamento e estabilidade

Soro/Plasma

- 7 dias a temperatura ambiente
- 7 dias a 2-8°C
- 1 ano a -20°C

Urina (acidificada)

- 3 dias a temperatura ambiente
- 3 dias a 2-8°C
- 1 ano a -20°C

VALORES DE REFERÊNCIAS ⁽⁷⁾

<i>Soro/plasma</i>	mg/dL	mmol/L
	1.5 - 2.6	0.63 - 1.05

<i>Urina (24h - coleta)</i>	mg/24h	mmol/24 h
	73 - 122	3.0 - 5.0
	mg/dL*	mmol/L*
	4.9 - 8.1	2.0 - 3.3

* para um volume urinário de 1,5 L em 24 horas

Observação: O intervalo citado deve servir apenas como guia. *Recomenda-se* que cada laboratório verifique esse intervalo ou estabeleça um intervalo de referência para a população pretendida.

PROCEDIMENTO

Procedimento manual
Comprimento de onda : 505 - 625 nm
Percurso óptico : 1 cm
Relação Amostra/Reagente : 1:100
Temperatura : 37 °C
As amostras de urina devem ser diluídas 1:5 com solução de NaCl 9 g/L antes da medição.
 Ler comparando com o branco de reagentes/ água destilada

	CALIBRAÇÃO	DOSAGEM
Reagente R	2 ml	2 ml
Padrão/Calibrador	20 µl	-
Amostra	-	20 µl

Misturar e ler as absorvâncias (A) após 5 minutos.

Procedimento automático

Estes reagentes podem ser utilizados em vários analisadores automáticos. Para os analisadores ELITech Selectra, as aplicações validadas estão disponíveis mediante solicitação. Com o Selectra TouchPro, utilize a aplicação incluída no código de barras disponível no final desde folheto.

As amostras de urina devem ser diluídas 1:5 com solução de NaCl 9 g/L antes da medição. Para usuários do software Selectra TouchPro, a diluição da urina é realizada automaticamente.

Informação de configuração importante :
O Reagente MAGNESIUM XB pode ser leve-
mente contaminado pela CHOLESTEROL SL em Selectra ProM e ProXL.
Para evitar a contaminação nestes equipamentos, programe as seguintes incompatibilidades :

Logiciel	Menu	Parametro
TouchPro	Incompatibilidades de Agulha	Incompatibilidade/ CHOLESTEROL - MAGNESIUM
Autres	Incompatibilidade de Agulha	CHOLESTEROL: MAGNESIUM

CÁLCULO

ΔA Amostra x n n = concentração do padrão/ calibrador

ΔA Padrão/ calibrador

Para o cálculo da concentração do magnésio na urina, multiplique o resultado pelo fator de diluição (5). Para usuários do software Selectra TouchPro, os resultados levam em consideração o fator de diluição.

Fator de conversão: mg/dL x 0.41 = mmol/L

CALIBRAÇÃO

Para referência MGXB-0600 : ELICAL 2 ou o padrão Magnesium XL são rastreadíveis relativamente ao método de referência de absorção atômica.
Para referência MGXB-0250 : ELICAL 2. o é rastreável relativamente ao método de referência de absorção atômica.

Frequência de calibração : A frequência de calibração é especifica a cada equipamento (consultar § DESEMPENHO).

CONTROLE DE QUALIDADE

Recomenda-se o uso de soros de controle de qualidade, como ELITROL I e ELITROL II, para monitorar o desempenho do ensaio.

Os controles devem ser executados:

- antes de analisar amostras de pacientes,
- pelo menos uma vez por dia,
- após cada calibração,
- e/ou de acordo com os requisitos laboratoriais e regulamentares.

Os resultados devem estar dentro dos intervalos definidos. Se os valores ficarem fora dos intervalos definidos, cada laboratório deve tomar as medidas corretvas necessárias.

TRATAMENTO DOS RESÍDUOS

O descarte de todo material residual deve estar de acordo com os requisitos regulamentares locais, estaduais e federais (consulte a Ficha de dados de segurança (SDS)).

DESEMPENHO

Os desempenhos foram obtidos no Selectra ProM, seguindo as recomendações técnicas do CLSI, sob condições ambientais controladas.

Precisão de medição

a) *Soro / plasma*
0.20 - 5.00 mg/dL (0.08 - 2.06 mmol/L)
As amostras com maiores concentrações devem ser diluídas 1:5 com solução de NaCl 9 g/L e analisadas novamente. Este procedimento estende a faixa de medição até 25 mg/dL (10.28 mmol/L).
Não relatar resultados fora do intervalo de medição.

Para utilizadores do Selectra TouchPro, a função de «diluir» realiza a diluição do amostras automaticamente. Os resultados são tomados em consideração na diluição.

b) *Urina*
1.0 - 20.0 mg/dL (0.41-8.23 mmol/L)
As amostras com maiores concentrações devem ser diluídas 1:5 com solução de NaCl 9 g/L e analisadas novamente. Este procedimento estende a faixa de medição até 100 mg/dL (41.14 mmol/L).

Não relatar resultados fora do intervalo de medição.

Para utilizadores do Selectra TouchPro, a função de «diluir» realiza a diluição do amostras automaticamente. Os resultados são tomados em consideração na diluição.

- **Limite de deteção (LoD) e limite de quantificação (LoQ)**

a) *Soro / plasma*
LoD = 0.03 mg/dL (0.01 mmol/L)
LoQ = 0.20 mg/dL (0.08 mmol/L)

b) *Urina*
LoD = 0.1 mg/dL (0.04 mmol/L)
LoQ = 1.0 mg/dL (0.41 mmol/L)

Precisão

Dados de imprecisão foram obtidos em 2 analisadores Selectra ProM ao longo de 20 dias (2 corridas por dia, testes realizados em duplicata).
Os resultados representativos são apresentados abaixo:

		Média		Intra-série	Total
	n	mg/dL	mmol/L	CV (%)	
Nível baixo	80	1.55	0.64	1.0	3.5
Nível médio	80	2.51	1.03	1.3	3.7
Nível elevado	80	3.84	1.58	0.7	3.0

		Média		Intra-série	Total
	n	mg/dL	mmol/L	CV (%)	
Nível baixo	80	1.2	0.49	2.9	9.2
Nível médio	80	5.0	2.06	0.7	4.0
Nível elevado	80	16.0	6.58	0.8	3.3

- **Correlação**
a) *Soro / plasma*
Foi realizado um estudo comparativo entre o reagente MAGNESIUM XB em um analisador Selectra ProM e um sistema similar disponível comercialmente em 102 amostras de soro humano.
As concentrações da amostra variaram de 0.27 para 4.99 mg/dL (0.11 - 2.05 mmol/L).
Os resultados são os seguintes:
Coeficiente de correlação: (r) = 0.996
Regressão linear: y = 1.041x - 0.01 mg/dL (0.00 mmol/L).

b) *Urina*
Foi realizado um estudo comparativo entre o reagente MAGNESIUM XB em um analisador Selectra ProM e um sistema similar disponível comercialmente em 80 amostras de urina humana.

As concentrações da amostra variaram de 1.7 para 19.6 mg/dL (0.70 - 8.06 mmol/L)..
Os resultados são os seguintes:
Coeficiente de correlação: (r) = 0.999
Regressão linear: y = 1.011x + 0.1 (0.00 mmol/L).

Limitações/Interferências

a) Soro / plasma

Estudos foram realizados para determinar o nível de interferência de diferentes compostos.
Os seguintes níveis de magnésium foram testados :
1.50 mg/dL e 3.90 mg/dL
Uma interferência não significativa é definida por uma recuperação ±±10% do valor inicial.
Triglicéridos: Nenhuma interferência significativa até 3000 mg/dL(33.90 mmol/L)
Bilirubina não conjugada: Nenhuma interferência significativa até 30.0 mg/dL (513 µmol/L)
Bilirubina conjugada: Nenhuma interferência significativa até 29.5 mg/dL (504 µmol/L)
Hemoglobina: Nenhuma interferência significativa até 500 mg/dL
Cálcio: Nenhuma interferência significativa até 20.0 mg/dL (4.99 mmol/L)
Ácido ascórbico: Nenhuma interferência significativa até 19.8 mg/dL.
Aceaminofeno: Nenhuma interferência significativa até 30 mg/dL.
Ácido acetilsalicílico: Nenhuma interferência significativa até 200 mg/dL.

- Em casos muito raros, as gamopatias monoclonais (mieloma múltiplo), em particular, tipo IgM (macroglobulinemia de Waldenstrom) podem causar resultados não confiáveis.⁽⁸⁾

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young. ⁽⁹⁻¹⁰⁾.

b) *Urina*
Estudos foram realizados para determinar o nível de interferência de diferentes compostos.
Os seguintes níveis de magnésio foram testados : 1.2 mg/dL e 15 mg/dL
Uma interferência não significativa é definida por uma recuperação ±±10% do valor inicial.

Bilirubina conjugada: : Nenhuma interferência significativa até 29.5 mg/dL (504 µmol/L)
Hemoglobina: Nenhuma interferência significativa até 500 mg/dL.
Cálcio : : Nenhuma interferência significativa até 60.0 mg/dL (14.97 mmol/L)
Ácido úrico: Nenhuma interferência significativa até 100 mg/dL (5.95 mmol/L).
Urea : : Nenhuma interferência significativa até 5000 mg/dL (832.50 mmol/L).
Ácido ascórbico: : Nenhuma interferência significativa até 19.8 mg/dL.
pH: : Nenhuma interferência significativa entre 2.5 à 6.0

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young. ⁽⁹⁻¹⁰⁾.

- **Estabilidade a bordo / frequência de calibração**
Estabilidade a bordo: 14 dias
Frequência de calibração: 7 dias
Recalibre quando os lotes de reagentes mudarem, quando os resultados do controle de qualidade estiverem fora da faixa estabelecida e após uma operação de manutenção.

Estes desempenhos foram obtidos utilizando o analisador ELITech Selectra ProM. Os resultados podem variar se um instrumento diferente ou um procedimento manual for usado. Os desempenhos de aplicações não validados pela ELITech não são garantidos e devem ser definidos pelo usuário.

DECLARAÇÃO DE INCIDENTE GRAVE
Notifique o fabricante (através do seu distribuidor) e a autoridade competente do Estado-Membro da união europeia em que o usuário e / ou o paciente está estabelecido, de qualquer incidente grave que tenha ocorrido em relação ao dispositivo.
Para outras jurisdições, a declaração de incidente grave deve estar de acordo com os requisitos regulamentares locais, estaduais e federais.

Ao relatar um incidente grave, você fornece informações que podem contribuir para a segurança de dispositivos médicos *in vitro*.

ASSISTÊNCIA TÉCNICA

Entre em contato com o seu distribuidor local ou com a ELITech Clinical Systems SAS. (CCsupport@elitechgroup.com).

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BIBLIOGRAFIA/BIBLIOGRAFIA
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SYMBOLES/SYMBOLS/ SIMBOLOS/SÍMBOLOS

- Les symboles utilisés sont décrits dans la norme ISO 15223-1 hormis ceux présentés ci-dessous.
- Symbols used are defined on ISO 15223-1 standard, except those presented below.

- Los símbolos utilizados son descritos en la norma ISO 15223-1 a la excepción de los presentados a continuación.

- Os símbolos utilizados são definidos na norma ISO 15223-1, exceto os apresentados abaixo.

CONT	Contient <p>Content</p> Contiene Conteúdo
R	Réactif <p>Reagent</p> Reactivo Reagente
Std	Standard <p>Standard</p> Estándar Padrão
	Modification par rapport à la version précédente <p>Modification from previous version</p> Modificación con respecto a la versión anterior Modificação relativamente à versão anterior
CE	Conformité Européenne <p>European Conformity</p> Conformidad Europea Conformidade Europeia



PIT-MGXB-4+v2 (10/2020)

Français - FR

USAGE PRÉVU

ELITech Clinical Systems MAGNESIUM XB est un réactif de diagnostic *in vitro*, destiné au dosage quantitatif du magnésium dans les échantillons de sérum, de plasma et d'urine humains sur des automates ou semi-automates.

Le standard est destiné à la calibration du réactif.
Ces dispositifs de diagnostic *in vitro* sont uniquement destinés aux professionnels.

SIGNIFICATION CLINIQUE ⁽¹⁻³⁾

Le magnésium est le quatrième cation les plus abondant dans le corps humain. Il est le cofacteur de nombreux systèmes enzymatiques dont les enzymes ATP-dépendantes et il joue également un rôle actif dans l'homéostasie minérale osseuse. Moins de 1% du magnésium total du corps humain est véhiculé par le sang dans lequel il se trouve en majorité sous forme d'ion libre, mais aussi lié à des protéines (principalement l'albumine) ou complexé avec divers anions. La magnésémie mesure le magnésium sanguin total mais seule la fraction libre est biologiquement active. Une hypomagnésémie peut être liée à des pertes rénales importantes (prise excessive de diurétiques), des désordres gastro-intestinaux (malabsorption, diarrhées), des maladies endocriniennes, plus rarement à un apport inadéquat. Une hypomagnésémie est souvent associée à une hypokaliémie et/ou une hypocalcémie. Une hypermagnésémie est rare et est presque toujours due à un apport excessif (traitement parentéral, prise excessive de traitements contenant du magnésium) ou à une insuffisance rénale. Le dosage du magnésium urinaire permet, en présence d'une hypomagnésémie, d'identifier une étiologie rénale. Dans la pratique clinique, le dosage du magnésium est indiqué pour l'aide au diagnostic des causes de concentrations anormales en calcium et/ou potassium ou de symptômes évoquant une hypo ou une hypermagnésémie, ainsi que pour le suivi de traitements à base de magnésium ou de calcium.

LIMITE D'UTILISATION

Une hypocalbinémie pouvant induire une pseudo-hypomagnésémie, tout dosage du magnésium sérique doit être analysé en regard de la protédimie et/ou de l'albuminémie.⁽⁴⁾
Le dosage de MAGNESIUM XB ne peut être utilisé seul pour diagnostiquer une maladie ou une pathologie spécifique.
Les résultats doivent toujours être confrontés aux résultats d'autres tests diagnostiques, aux examens cliniques, et à l'historique médical du patient.

MÉTHODE & PRINCIPE ⁽⁵⁾

Bleu de Xylidyl - Point Final

Le bleu de Xylidyl contenu dans le réactif se combine avec le magnésium de l'échantillon pour former un complexe rouge-pourpre. LEGTA sert à complexer le calcium et ainsi l'empêche d'interférer avec le test. L'augmentation d'absorbance à 505-510 nm et la diminution simultanée à 620-630 nm sont proportionnelles à la concentration de magnésium dans l'échantillon

COMPOSITION

Réactif : R
Tampon AMP, pH 11.2
Bleu de Xylidyl 120 µmol/L
Azide de sodium < 0,1 % (p/p)
Contient aussi de l'EGTA et des surfactants pour des performances optimales
AMP: 2-Amino-2-methyl-1-propanol

Standard: Std
Magnésium 2.0 mg/dL
823 µmol/L

MATÉRIELS REQUIS MAIS NON FOURNIS
-CALJ-0550 ELICAL 2
-CONT-0060 ELITROL I
-CONT-0160 ELITROL II
- Solution saline normale (NaCl 9 g/L).
- Automates ou semi-automates.
- Equipement général de laboratoire (ex. pipette).
- Ne pas utiliser de matériel ne figurant pas ci-dessus.

PRÉCAUTIONS D'EMPLOI ET MISES EN GARDE

- Le réactif R contient de l'azide de sodium qui peut réagir avec le plomb ou le cuivre et former des azides métalliques potentiellement explosifs. Lors de l'élimination de ces réactifs toujours rincer abondamment avec de l'eau pour éviter l'accumulation d'azides.
- Respecter les précautions d'usage et les bonnes pratiques de laboratoire.
- Utiliser du matériel de laboratoire propre ou à usage unique afin d'éviter toute contamination.
- Consulter la fiche de données de sécurité (FDS) pour une manipulation appropriée.

STABILITÉ

Stocker à 2-8 °C et à l'abri de la lumière. Ne pas congeler.
Ne pas utiliser après la date d'expiration indiquée sur les étiquettes des flacons.
Le standard doit être immédiatement et correctement refermé afin d'éviter toute contamination ou évaporation.
Stabilité à bord :
La stabilité à bord est spécifique à chaque automate. (Se référer au § PERFORMANCES).

PRÉPARATION

Le réactif et le standard sont prêts à l'emploi.



<i>MGXB-0250</i>
<i>MGXB-0600</i>

DÉTERIORATION DU PRODUIT
- Ces produits doivent être limpides. Tout trouble serait le signe d'une détérioration du produit.
- Ne pas utiliser le produit s'il y a des signes évidents de contamination ou de détérioration (ex : particules).
- Un flacon endommagé peut avoir un impact sur les performances du produit. Ne pas utiliser le produit si les flacons présentent des signes physiques de détérioration (par exemple, fuite, flacon percé).

ÉCHANTILLONS

Echantillons requis ^(1,6)

- Sérum
- Plasma (héparine de lithium)
- Urine
- L'utilisation de toute autre type d'échantillon doit être validée par le laboratoire.
Avertissements et précautions ^(1,2)
- Le sérum doit être séparé des cellules aussi rapidement que possible. ^(1,2)
- Les échantillons ne doivent pas être hémolysés ^(1,2)
- Après collecte, les urines doivent être acidifiées avec de l'acide chlorhydrique 6N à un pH < 2 pour empêcher le sel de magnésium de précipiter. ⁽¹⁾
Les échantillons doivent être prélevés selon les Bonnes Pratiques de Laboratoire et les guides appropriés qui sont mis en place.

Stockage et stabilité ⁽⁶⁾

Sérum/Plasma

- 7 jours à température ambiante
- 7 jours à 2-8°C
- 1 an à -20°C

Urine (acidifiée)

- 3 jours à température ambiante
- 3 jours à 2-8°C
- 1 an à -20°C

VALEURS DE RÉFÉRENCE ⁽⁷⁾

<i>Sérum/plasma</i>	mg/dL	mmol/L
	1.5 - 2.6	0.63 - 1.05

<i>Urine (recueil de 24h)</i>	mg/24h	mmol/24 h
	73 - 122	3.0 - 5.0
	mg/dL*	mmol/L*
	4.9 - 8.1	2.0 - 3.3

* pour un volume urinaire de 1.5 L par 24 heures

Remarque : *Les valeurs ci-dessus ne sont données qu'à titre indicatif. Il est recommandé à chaque laboratoire d'établir et de maintenir ses propres valeurs de référence par rapport à la population visée.*

PROCÉDURE

Procédure manuelle
Longueur d'onde : 505 - 625 nm
Trajet optique : 1 cm
Ratio échantillon/réactif : 1:100
Température : 37 °C
Les échantillons urinaires doivent être dilués au 1/5 dans une solution de NaCl 9 g/L avant la mesure.
Lire contre le blanc réactif/'eau distillé.

	CALIBRATION	DOSAGE
Réactif R	2 ml	2 ml
Standard/Calibrant	20 µl	-
Echantillon	-	20 µl

Mélanger et lire les absorbances (A) après 5 minutes d'incubation.

Procédure sur automate
Ces réactifs peuvent être utilisés sur différents automates. Pour les automates ELITech Selectra, les applications validées sont disponibles sur demande. Avec le logiciel Selectra TouchPro, utilisez l'application incluse dans le code barre disponible à la fin de cette notice.

Les échantillons urinaires doivent être dilués au 1/5 dans une solution de NaCl 9 g/L avant la mesure. Pour les utilisateurs du logiciel Selectra TouchPro, la dilution des urines est réalisée automatiquement.

Information importante de programmation:
Le réactif MAGNESIUM XB peut être faiblement contaminé par le réactif CHOLESTEROL SL sur les Selectra ProM

English - EN

INTENDED USE

ELITech Clinical Systems MAGNESIUM XB is an *in vitro* diagnostic reagent intended for the quantitative determination of magnesium in human serum, plasma and urine samples on analyzers or semi-automatic analyzers.

The standard is intended for the calibration of reagent. These *in vitro* diagnostic devices are for professional use only.

CLINICAL SIGNIFICANCE ⁽¹⁻³⁾

Magnesium is the fourth most abundant cation in the human body. It is the cofactor for many enzymatic systems including ATP-dependent enzymes and it also plays an active role in bone mineral homeostasis. Less than 1% of the total magnesium of the human body is carried by blood where it is mainly found as free ion, but also protein-bound (mainly albumin) or complexed with various anions. Magnesemia measures total magnesium but only free magnesium is biologically active. Hypomagnesemia can be due to important renal losses (excessive intake of diuretics), gastrointestinal disorders (malabsorption, diarrheas), endocrine disorders (less often due to inadequate intake. Hypomagnesemia is often associated with hypokalemia and/or hypocalcemia. Hypermagnesemia is rare and is caused almost always by excessive intake (parenteral therapy, excessive intake of magnesium-containing drugs) or renal insufficiency.

The measure of urinary magnesium allows, in presence of magnesium deficiency, to identify a kidney etiology. In clinical practice, measurement of magnesium is indicated to help diagnose the causes of abnormal calcium and/or potassium levels or symptoms suggesting hypo or hypermagnesemia as well as for monitoring magnesium- and/or calcium-based treatments.

LIMITATION OF USE

Because hypoalbuminemia may trigger pseudo-hypomagnesemia, serum magnesium results must be interpreted in regards of serum total protein and/or albumin levels.⁽⁴⁾

The quantitative assay of MAGNESIUM XB alone can not be used to diagnose a disease or a specific pathology. The results must be interpreted in conjunction with other diagnostic test results, clinical findings and the patient’s medical history.

METHOD & PRINCIPLE ⁽⁵⁾

Xylyldyl Blue - End Point

Xylyldyl blue in the reagent combines with the magnesium from the sample to form a red-purple chelate. EGTA is used to complex calcium and thus prevents it from interfering with the test.

The simultaneous increase in absorbance at 505-510 nm and decrease of the 620-630 nm absorbance are proportional to the magnesium concentration in the sample.

COMPOSITION

Reagent: R
AMP buffer, pH 11.2
Xylyldyl Blue 120 µmol/L
Sodium azide < 0.1 %(w/w)
Also contains EGTA and surfactants for optimal performance

AMP: 2-Amino-2-methyl-1-propanol

Standard: Std
Magnesium 2.0 mg/dL
823 µmol/L

MATERIALS REQUIRED BUT NOT PROVIDED

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Normal saline solution (NaCl 9 g/L).
- Analyzers or semi-automatic analyzers.
- General Laboratory equipment (e.g. pipette).
- Do not use materials that are not required as indicated above.

PRECAUTIONS FOR USE AND WARNINGS

- Reagent R contains sodium azide which may react with lead or copper plumbing to form potentially explosive metal azides. When disposing of these reagents always flush with copious amounts of water to prevent azide buildup.

- Take normal precautions and adhere to good laboratory practice.

- Use clean or single use laboratory equipment only to avoid contamination.
- Consult Safety Data Sheet (SDS) for a proper handling.

STABILITY

Store at 2-8 °C and protect from light. Do not freeze. Do not use after expiration dates indicated on the vial labels.

The standard should be immediately and tightly capped to prevent contamination and evaporation.

On-board stability :

The on-board stability is specific for each analyzer. (Refer to § PERFORMANCE DATA).

PREPARATION

The reagent and standard are ready to use.

PRODUCT DETERIORATION

- These products should be clear. Cloudiness would indicate deterioration.

- Do not use the product if there is visible evidence of contamination or damage (e.g. particle matter).
- Damage to the product container may impact on product performance. Do not use the product if there is physical evidence of deterioration (e.g. leakages or punctured container).

SAMPLES

Specimen ^(1,6)

- Serum.
- Plasma (lithium heparin).
- Urine.
- Using any other specimen type should be validated by the laboratory.

Warnings and precautions ^(1,2)
- Serum must be separated from cells as rapidly as possible.^(1,2)

- Samples must be free from hemolysis ^(1,2)
- After collection, urine specimens should be acidified with hydrochloric acid 6N to a pH < 2 to prevent magnesium salt precipitation. ⁽¹⁾

Samples should be collected in accordance with Good Laboratory Practice and appropriate guidelines that may be in place.

Storage and stability ⁽⁶⁾
Serum/plasma
- 7 days at room temperature.
- 7 days at 2-8°C.
- 1 year at -20°C.
Urine (acidified)
- 3 days at room temperature
- 3 days at 2-8°C
- 1 year at -20°C

REFERENCE VALUES ⁽⁷⁾

<i>Serum/plasma</i>	mg/dL	mmol/L
	1.5 - 2.6	0.63 - 1.05

<i>Urine (24h - collection)</i>	mg/24h	mmol/24 h
	73 - 122	3.0 - 5.0

	mg/dL*	mmol/L*
	4.9 - 8.1	2.0 - 3.3

* for a urinary volume of 1.5 L per 24 hours

Note : *The quoted range should serve as a guide only. It is recommended that each laboratory verifies this range or establishes a reference interval for the intended population.*

PROCEDURE

Manual Procedure

Wavelength : 505 - 625 nm
Optical path : 1 cm
Sample/ Reagent ratio : 1:100
Temperature : 37 °C

Urine samples must be diluted 1:5 with NaCl 9 g/L urine before measurement.

Read against reagent blank/distilled water.

	CALIBRATION	TEST
Reagent R	2 ml	2 ml
Standard/ Calibrator	20 µl	-
Sample	-	20 µl

Mix and read the absorbances (A) after an incubation of 5 minutes.

Automatic Procedure

These reagents may be used in several automatic analyzers. For ELITech Selectra Analyzers, validated applications are available on request. For Selectra TouchPro software, use the application included in the barcode available at the end of this insert.

Urine samples must be diluted 1:5 with NaCl 9 g/L solution before measurement. For users of Selectra TouchPro software, urine dilution is performed automatically.

Important set-up information:
MAGNESIUM XB reagent can be weakly contaminated by CHOLESTEROL SL on Selectra ProM and ProXL.
In order to avoid contamination on these instruments, program the following incompatibilities:

Software	Menu	Parameter
TouchPro	Probe incompatibilities	incompatibility/ CHOLESTEROL - MAGNESIUM
Other	Needle incompatibility	CHOLESTEROL: MAGNESIUM

CALCULATION

^{ΔA} Sample x n n = Calibrator/ standard
^{ΔA} Calibrator/ Standard concentration

For the calculation of magnesium concentration in urine, multiply the result by the dilution factor (5). For users of Selectra TouchPro software, the results take the dilution factor into account.

Conversion factor : mg/dL x 0.41 = mmol/L

CALIBRATION

For the reference MAGX-0600 : ELICAL 2 and Magnesium XL standard are traceable to the atomic absorption reference method
For the reference MAGX-0250 : ELICAL 2 is traceable to the atomic absorption reference method.

Calibration frequency : The calibration is specific for each analyzer. § PERFORMANCE DATA).

QUALITY CONTROL

It is recommended that quality control sera such as ELITROL I and ELITROL II be used to monitor the performance of the assay.

Controls have to be performed :
- prior to assaying patient samples,
- at least once per day,
- after every calibration,
- and/or in accordance with laboratory and regulatory requirements.

Results should be within the defined ranges. If values fall outside of the defined ranges, each laboratory should take necessary corrective measures.

WASTE MANAGEMENT

Disposal of all waste material should be in accordance with local, state and federal regulatory requirements (please refer to the Safety Data Sheet (SDS)).

PERFORMANCE DATA

Performances were obtained on Selectra ProM, following CLSI technical recommendations, under controlled environmental conditions.

Measuring range

a) Serum/Plasma
2.0 - 5.00 mg/dL (0.08 - 2.06 mmol/L)

Samples having greater concentrations should be diluted 1:5 with NaCl 9 g/L solution and re-assayed. This procedure extends the measuring range up to 25 mg/dL (10.28 mmol/L).
Do not report results outside this extended range.

For users with Selectra TouchPro software, the «dilute» function performs the sample dilution automatically. Results take the dilution into account.

b) Urine

1.0 - 20.0 mg/dL (0.41-8.23 mmol/L)

Samples having greater concentrations should be diluted 1:5 with NaCl 9 g/L solution and re-assayed. This procedure extends the measuring range up to 100 mg/dL (41.14 mmol/L).
Do not report results outside this extended range.

For users with Selectra TouchPro software, the «dilute» function performs the sample dilution automatically. Results take the dilution into account.

Limit of Detection (LOD) and Limit of Quantification (LoQ)

a) Serum/Plasma
LoD = 0.03 mg/dL (0.01 mmol/L)
LoQ = 0.20 mg/dL (0.08 mmol/L)

b) Urine

LoD = 0.1 mg/dL (0.04 mmol/L)
LoQ = 1.0 mg/dL (0.41 mmol/L)

Precision

Imprecision data has been obtained on 2 Selectra Pro analyzers over 20 days (2 runs per day, tests performed in duplicate).

Representative results are presented below.

a) Serum/Plasma

	Mean	Within-run	Total		
	n	mg/dL	mmol/L	CV (%)	
Low level	80	1.55	0.64	1.0	3.5
Medium level	80	2.51	1.03	1.3	3.7
High level	80	3.84	1.58	0.7	3.0

b) Urine

	Mean	Within-run	Total		
	n	mg/dL	mmol/L	CV (%)	
Low level	80	1.2	0.49	2.9	9.2
Medium level	80	5.0	2.06	0.7	4.0
High level	80	16.0	6.58	0.8	3.3

- Correlation

a) Serum/Plasma
A comparative study has been performed between MAGNESIUM XB reagent on a Selectra ProM analyzer and a similar commercially available system on 102 human serum samples. The sample concentrations ranged from 0.27 to 4.99 mg/dL (0.11 - 2.05 mmol/L). The results are as follows :
Correlation coefficient : (r) = 0.996
Linear regression: y = 1.041x - 0.01 mg/dL (0.00 mmol/L).

b) Urine
A comparative study has been performed between MAGNESIUM XB reagent on a Selectra ProM analyzer and a similar commercially available system on 80 human urine samples. The sample concentrations ranged from 1.7 to 19.6 mg/dL (0.70 - 8.06 mmol/L). The results are as follows :
Correlation coefficient : (r) = 0.999
Linear regression: y = 1.011x + 0.1 (0.00 mmol/L).

Limitations/Interferences

a) Serum/Plasma

- Studies have been performed to determine the level of interference from different compounds. The following analyte levels were tested: 1.50 mg/dL and 3.90 mg/dL

No significant interference is defined by a recovery ±10% of the initial value.
Triglycerides : No significant interference up to 3000 mg/dL(33.90 mmol/L).
Unconjugated bilirubin : No significant interference up to 30.0 mg/dL (513 µmol/L).
Conjugated bilirubin : No significant interference up to 29.5 mg/dL (504 µmol/L)
Hemoglobin : No significant interference up to 500 mg/dL.
Calcium : No significant interference up to 20.0 mg/dL (4.99 mmol/L).
Ascorbic acid : No significant interference up to 19.8 mg/dL.
Acetaminophen : No significant interference up to 30 mg/dL.
Acetylsalicylic acid : No significant interference up to 200 mg/dL

- In very rare cases, monoclonal gammopathies (multiple myeloma), in particular IgM type (Waldenstrom’s macroglobulinemia) can cause unreliable results.⁽⁸⁾

- Many other substances and drugs may interfere. Some of them are listed in reviews published by Young.⁽⁹⁻¹⁰⁾

b) Urine

- Studies have been performed to determine the level of interference from different compounds. The following magnesium levels were tested: 1.2 mg/dL and 15 mg/dL.

No significant interference is defined by a recovery ±10% of the initial value.
Conjugated bilirubin : Aucune interférence significative jusqu’à 29.5 mg/dL (504 µmol/L)
Hemoglobin : No significant interference up to 500 mg/dL.
Calcium : No significant interference up to 60.0 mg/dL (14.97 mmol/L)
Uric Acid : No significant interference up to100 mg/dL (5.95 mmol/L).
Urea : No significant interference up to 5000 mg/dL (832.50 mmol/L).
Ascorbic acid : No significant interference up to 19.8 mg/dL.
pH : No significant interference for pH values ranging between 2,5 à 6,0

- Many other substances and drugs may interfere. Some of them are listed in reviews published by Young.⁽⁹⁻¹⁰⁾

- On board stability/Calibration frequency On Board Stability: 14 days Calibration frequency: 7 days

Recalibrate when reagent lots change, when quality control results fall outside the established range and after a maintenance operation.

These performances have been obtained using ELITech Selectra ProM analyzer. Results may vary if a different instrument or a manual procedure is used. The performances of applications not validated by ELITech are not warranted and must be defined by the user.

DECLARATION OF SERIOUS INCIDENT

Please notify the manufacturer (through your distributor) and competent authority of the Member State of the european union in which the user and/or the patient is established, of any serious incident that has occurred in relation to the device. For other jurisdictions, the declaration of serious incident should be in accordance with local, state and federal regulatory requirements. By reporting a serious incident, you provide information that can contribute to the safety of *in vitro* medical devices.

TECHNICAL ASSISTANCE

Contact your local distributor or ELITech Clinical Systems SAS (CCsupport@elitechgroup.com).

Español - ES

USO PREVISTO

ELITech Clinical Systems MAGNESIUM XB es un reactivo de diagnóstico *in vitro* diseñado para la determinación cuantitativa de magnesio en muestras de suero, plasma y orina humanas en equipos automatizados o equipos semiautomáticos.

El estándar está diseñado para la calibración del reactivo.

Estos dispositivos de diagnóstico *in vitro* están destinados unicamente para los profesionales.

SIGNIFICADO CLÍNICO ⁽¹⁻³⁾

El magnesio es el cuarto catión más abundante en el cuerpo humano. Es el cofactor de muchos sistemas enzimáticos, incluidas las enzimas dependientes de ATP, y también juega un papel activo en la homeostasis mineral ósea. Menos del 1% del magnesio total en el cuerpo humano es transportado por la sangre en la que se encuentra principalmente en forma de un ion libre, pero también está vinculado a proteínas (principalmente albúmina) o forma un complejo con varios aniones. La magnesemia mide el magnesio sanguíneo total, pero solo la fracción libre es biológicamente activa. La hipomagnesemia puede estar relacionada con pérdidas renales significativas (ingesta excesiva de diuréticos), trastornos gastrointestinales (malabsorción, diarrea), enfermedades endocrinas, más raramente con una ingesta inadecuada. La hipomagnesemia a menudo se asocia con hipocalcemia y / o hipocalemia. La hipermagnesemia es rara y casi siempre se debe a una ingesta excesiva (tratamiento parenteral, ingesta excesiva de tratamientos que contienen magnesio) o insuficiencia renal. La dosificación de magnesio en orina permite, en presencia de hipomagnesemia, identificar una etiología renal. En la práctica clínica, la cuantificación de magnesio es indicada para ayudar en el diagnóstico de las causas de concentraciones anormales de calcio y / o potasio o de síntomas que sugieren hipo o hipermagnesemia, así como para el monitoreo de tratamientos basados en magnesio o calcio

LÍMITE DE UTILIZACIÓN

Debido a que la hypoalbuminemia puede desencadenar una pseudohipomagnesemia, los resultados de magnesio sérico deben interpretarse en relación con los niveles séricos de proteína total y / o albúmina.⁽⁴⁾ La cuantificación del MAGNESIUM XB no puede ser utilizado solo para diagnosticar una enfermedad o patología específica.

Los resultados siempre deben compararse con los resultados de otras pruebas de diagnóstico, exámenes clínicos y el historial médico del paciente.

MÉTODO & PRINCIPIO ⁽⁵⁾

Bleu de Xylydyl - Punto Final

El azul de xilidilo en el reactivo se combina con el magnesio de la muestra para formar un quelato rojo púrpura. El EGTA se usa para formar complejos de calcio y, por lo tanto, evita que interfiera con la prueba. El aumento de la absorbancia a 505-510 nm y la disminución simultáneo de la absorbancia 620-630 nm son proporcionales a la concentración de magnesio en la muestra.

COMPOSICIÓN

Reactivo : R
Tampón AMP, pH 11.2
Azul de Xilidilo 120 µmol/L
Azida sodíca < 0.1 %(p/p)
También contiene EGTA y tensioactivos para un rendimiento óptimo.

AMP: 2-amino-2-metil-1-propanol

Estándar : Std
Magnesio 2.0 mg/dL
823 µmol/L

MATERIALES REQUERIDOS PERO NO INCLUIDOS

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Solución salina normal (NaCl 9 g/L).
- Equipos automáticos o semiautomáticos.
- Equipamiento general de laboratorio (p. ej, pipeta).
- No utilice materiales que no se requieren, tal como se indica anteriormente.

PRECAUCIONES DE USO Y ADVERTENCIAS

- El reactivo R contiene azida sodíca que puede reaccionar con el plomo o el cobre de la tubería y formar potencialmente azidas metálicas explosivas. Cuando se elimine el reactivo enjuague con agua abundantemente para prevenir la acumulación de azidas.
- Tome las precauciones normales y respete las buenas prácticas de laboratorio.
- Para evitar contaminaciones utilizar equipo nuevo o completamente limpio.
- Consulte la Hoja de Datos de Seguridad (SDS) para un manejo adecuado.

ESTABILIDAD

Conservar a 2-8 °C y protegidos de la luz. No congelar.

No utilice después de la fecha de caducidad indicada en la etiqueta de los frascos.

El estándar debe cerrarse inmediatamente y correctamente para evitar contaminación y evaporación.

Estabilidad en el equipo:
La estabilidad es específica para cada equipo. (Referirse al § DATOS DE RENDIMIENTO).

PREPARACIÓN

El reactivo y el estándar están listos para su uso.

DETERIORACIÓN DEL PRODUCTO

- Los productos deben ser claros. Turbidez indicaría deterioro.

- No utilice el producto si este presenta signos evidentes de contaminación o deterioro (p. ej, partículas).

- Un frasco dañado puede tener un impacto en el rendimiento del producto. No utilice el producto si este tiene signos físicos de deterioro (p. ej, fugas, frasco perforado).

MUESTRAS

Muestras requeridas ^(1,6)

- Suero
- Plasma (heparina de litio).
- Orina.
- El uso de cualquier otro tipo de muestra debe ser validado por el laboratorio.

Advertencias y precauciones ^(1,2)

- El suero debe separarse de las células lo más rápido posible. ^(1,2)

- Las muestras deben estar libres de hemólisis ^(1,2)
- Después de la recolección, las muestras de orina deben acidificarse con ácido clorhídrico 6N a un pH <2 para evitar la precipitación de sal de magnesio. ⁽¹⁾
Las muestras deben de tomarse de acuerdo con las Buenas Prácticas de Laboratorio y las guías apropiadas establecidas.
Conservación y estabilidad ⁽⁶⁾
Suero/Plasma
- 7 días a temperatura ambiente.
- 7 días a 2-8 °C
- 1 año a -20 °C
Orina (acidificada)
- 3 días a temperatura ambiente.
- 3 días a 2-8 °C
- 1 año a -20 °C

VALORES DE REFERENCIA ⁽⁷⁾

<i>Suero/plasma</i>	mg/dL	mmol/L
	1.5 - 2.6	0.63 - 1.05
Orina		
<i>(recolectada de 24h)</i>	mg/24h	mmol/24 h
	73 - 122	3.0 - 5.0
	mg/dL* mmol/L*	
	4.9 - 8.1	2.0 - 3.3

* para un volumen de orina de 1.5 L por 24 horas

Nota : *Los valores anteriores son solo indicativos. Se recomienda que cada laboratorio establezca y mantenga sus propios valores de referencia en relación con la población destinataria.*

PROCEDIMIENTO

Procedimiento manual

Longitud de onda : 505 - 625 nm
Trayectoria óptica : 1 cm
Reliô muestra/reactivo : 1:100
Temperatura : 37 °C

Las muestras de orina deben diluirse 1:5 con una solución de NaCl 9 g/L antes de la medición Leer contra blanco reactivo / agua destilada.

Reactivo R	CALIBRACIÓN	PRUEBA
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▣ CALIBRAÇÃO

Para referências URSL_0427/0507 : ELICAL 2 ou o padrão Urea Standard 50 mg/dL são rastreáveis relativamente ao método de referência ID-MS (Diluição Isotópica por Espectrometria de Massa).

Para referências URSL_0250/0455/0420/0500 : ELICAL 2, o é rastreável relativamente ao método de referência ID-MS (Diluição Isotópica por Espectrometria de Massa).

Frequência de calibração : A frequência de calibração é específica a cada equipamento (consultar § DESEMPENHO).

CONTROLE DE QUALIDADE

Recomenda-se o uso de soros de controle de qualidade, como ELITROL I e ELITROL II, para monitorar o desempenho do ensaio.

Os controles devem ser executados:

- antes de analisar amostras de pacientes,
- pelo menos uma vez por dia,
- após cada calibração,
- e/ou de acordo com os requisitos laboratoriais e regulamentares.

Os resultados devem estar dentro dos intervalos definidos. Se os valores ficarem fora dos intervalos definidos, cada laboratório deve tomar as medidas corretivas necessárias.

TRATAMENTO DOS RESÍDUOS

O descarte de todo material residual deve estar de acordo com os requisitos regulamentares locais, estaduais e federais (consulte a Ficha de dados de segurança (SDS)).

DESEMPENHO

Os desempenhos foram obtidos no Selectra ProM, seguindo as recomendações técnicas do CLSI, sob condições ambientais controladas.

- Precisão de medição

a) *Soro / plasma*

10 - 300 mg/dL (1.67 - 49.95 mmol/L)

As amostras com maiores concentrações devem ser diluídas 1:5 com solução de NaCl 9 g/L e ensaiado novamente. Este procedimento estende a faixa de medição até 1 500 mg/dL (249.75 mmol/L).

Não relatar resultados fora do intervalo de medição.

Para utilizadores do Selectra TouchPro, a função de «diluir» realiza a diluição do amostras automaticamente. Os resultados são tomados em consideração na diluição.

b) *Urina*

200 - 6 000 mg/dL (33 - 999 mmol/L).

Não relate resultados fora da faixa de medição.

- **Limite de detecção (LoD) e limite de quantificação (LoQ)**

a) *Soro / plasma*

LoD = 1.5 mg/dL (0.25 mmol/L)

LoQ = 5.0 mg/dL (0.83 mmol/L)

b) *Urina*

LoD = 18 mg/dL (3 mmol/L)

LoQ = 200 mg/dL (33 mmol/L)

- Precisão

Dados de imprecisão foram obtidos em 2 analisadores Selectra ProM ao longo de 20 dias (2 corridas por dia, testes realizados em duplicata).

Os resultados representativos são apresentados abaixo :

a) *Soro / plasma*

	Média			Intra-série	Total
	n	mg/dL	mmol/L	CV (%)	
Nível 1	80	18.0	3.00	1.6	3.2
Nível 2	80	59.0	9.82	1.2	2.2
Nível 3	80	144.6	24.08	1.0	2.1

b) *Urina*

	Média			Intra-série	Total
	n	mg/dL	mmol/L	CV (%)	
Nível 1	80	482	80	1.7	3.8
Nível 2	80	1165	194	0.6	3.1
Nível 3	80	2587	431	0.4	3.6

- Correlação

a) *Soro / plasma*

Foi realizado um estudo comparativo entre o reagente UREA UV SL em um analisador Selectra ProM e um sistema similar disponível comercialmente em 98 amostras de soro humano.

As concentrações da amostra variaram de 12.5 para 285.5 mg/dL (2.08 - 47.54 mmol/L).

Os resultados são os seguintes:

Coefficiente de correlação: (r) = 1.000

Regressão linear: y = 0.993x - 0.1 mg/dL

(0.02 mmol/L)

▣ b) *Urina*

Foi realizado um estudo comparativo entre o reagente UREA UV SL em um analisador Selectra ProM e um sistema similar disponível comercialmente em 53 amostras de urinas humanas.

As concentrações da amostra variaram de 203 para 5 569 mg/dL (34 - 927 mmol/L).

Os resultados são os seguintes:

Coefficiente de correlação: (r) = 0.999

Regressão linear: y = 1.000x + 52 mg/dL

(9 mmol/L)

▣- Limitações/Interferências

a) *Soro / plasma*

Estudos foram realizados para determinar o nível de interferência de diferentes compostos.

Os seguintes níveis do ureia foram testados : 15.0 mg/dL e 60.1 mg/dL

Uma interferência não significativa é definida por uma recuperação \pm 10% do valor inicial.

Bilirrubina não conjugada: Nenhuma interferência significativa até 30.0 mg/dL (513 µmol/L).

Bilirrubina conjugada: Nenhuma interferência significativa até 29.5 mg/dL (505 µmol/L).

Turvação: Nenhuma interferência significativa até 614 mg/dL (6.94 mmol/L) equivalente de triglicéridos.

Hemoglobina: Nenhuma interferência significativa até 500 mg/dL.

Ácido ascórbico: Nenhuma interferência significativa até 20.0 mg/dL.

Metildopa : Nenhuma interferência significativa até 1.0 mg/dL.

- Em casos muito raros, as gamopatias monoclonais (mieloma múltiplo), em particular, tipo IgM (macroglbulinemia de Waldenstrom) podem causar resultados não confiáveis.⁽⁹⁾

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young.⁽⁶⁻⁷⁾.

b) *Urina*

Estudos foram realizados para determinar o nível de interferência de diferentes compostos.

Os seguintes níveis do ureia foram testados : 1 500 mg/dL e 3 000 mg/dL.

Uma interferência não significativa é definida por uma recuperação \pm 10% do valor inicial.

Bilirrubina conjugada: Nenhuma interferência significativa até 29.5 mg/dL (505 µmol/L).

Hemoglobina: Nenhuma interferência significativa até 500 mg/dL.

Ácido ascórbico : Nenhuma interferência signific cativa até 20.0 mg/dL.

Ácido úrico: Nenhuma interferência significativa até 120 mg/dL (7.14 mmol/L).

pH : Nenhuma interferência significativa para valores de pH variando entre 2.5 e 12.0.

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young.⁽⁶⁻⁷⁾.

- **Estabilidade a bordo / frequência de calibração**
Estabilidade a bordo: 14 dias
Frequência de calibração: 7 dias
Recalibre quando os lotes de reagentes mudarem, quando os resultados do controle de qualidade estiverem fora da faixa estabelecida e após uma operação de manutenção.

Estes desempenhos foram obtidos utilizando o analisador ELITech Selectra ProM. Os resultados podem variar se um instrumento diferente ou um procedimento manual for usado. Os desempenhos de aplicações não validados pela ELITech não são garantidos e devem ser definidos pelo usuário.

▣DECLARAÇÃO DE INCIDENTE GRAVE

Notifique o fabricante (através do seu distribuidor) e a autoridade competente do Estado-Membro da união europeia em que o usuário e / ou o paciente está estabelecido, de qualquer incidente grave que tenha ocorrido em relação ao dispositivo.

Para outras jurisdições, a declaração de incidente grave deve estar de acordo com os requisitos regulamentares locais, estaduais e federais.
Ao relatar um incidente grave, você fornece informações que podem contribuir para a segurança de dispositivos médicos *in vitro*.

▣ASSISTÊNCIA TÉCNICA

Entre em contato com o seu distribuidor local ou com a ELITech Clinical Systems SAS. (CCsupport@elitechgroup.com).

▣BIBLIOGRAPHIE/BIBLIOGRAPHY

BIBLIOGRAFIA/BIBLIOGRAFIA

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2. Wu, A.H.B., *Clinical guide to laboratory tests*, 4th Ed., (W.B. Saunders eds.), (2006), 1096.

3. Kaplan, J.M. & First, M.R., *Renal Function, Clinical Chemistry: Theory, Analysis, Correlation*, 5th Ed., Kaplan, L.A., Pesce, A.J., (Mosby Inc. eds.), (2010), 567 and appendix.

4. Bretaudière, J.P., *et al.*, *Clin. Chem.*, (1976), **22**, 1614.

5. Berth, M. & Delanghe, J. *Protein precipitation as a possible important pitfall in the clinical chemistry analysis of blood samples containing monoclonal immunoglobulins: 2 case reports and a review of literature*, *Acta Clin Belg.*, (2004), **59**, 263.

6. Young, D.S., *Effects of preanalytical variables on clinical laboratory tests*, 2nd Ed., AACC Press (1997).

7. Young, D.S., *Effects of drugs on clinical laboratory tests*, 4th Ed., AACC Press (1995).

SYMBOLES/SYMBOLS/ SÍMBOLOS/SÍMBOLOS

- Les symboles utilisés sont décrits dans la norme ISO 15223-1 hormis ceux présentés ci-dessous.

- Symbols used are defined on ISO 15223-1 standard, except those presented below.

- Los símbolos utilizados son descritos en la norma ISO 15223-1 a la excepción de los presentados a continuación.

- Os símbolos utilizados são definidos na norma ISO 15223-1, exceto os apresentados abaixo.

CONT	Contient Contient Contiene Conteúdo
R1	Réactif R1 Reagent R1 Reactivo R1 Reagente R1
R2	Réactif R2 Reagent R2 Reactivo R2 Reagente R2
Std	Standard Standard Estándar Padrão
☛	Modification par rapport à la version précédente Modification from previous version Modificación con respecto a la versión anterior Modificação relativamente à versão anterior
CE	Conformité Européenne European Conformity Conformidade Europeia Conformidade Europeia

Note/Nota

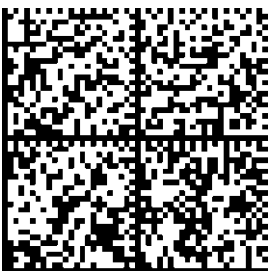
- Uniquement pour les réf. **URSL-0250/0455**, utilisées avec le logiciel Selectra TouchPro.

- Only for ref. **URSL-0250/0455**, used with Selectra TouchPro software.

- Únicamente para las ref. **URSL-0250/0455**, utilizadas con el software Selectra TouchPro.

- Somente para ref. **URSL-0250/0455**, usados com o Selectra TouchPro.

URSL



Urea 0
640 PIT-URSL



UREA UV SL

PIT-URSL-4-v23 (11/2020)

Français - FR

▣USAGE PRÉVU

ELITech Clinical Systems UREA UV SL est un réactif de diagnostic *in vitro*, destiné au dosage quantitatif de l'urée dans les échantillons de sérum, de plasma et d'urine humains sur des automates ou semi-automates.

Le standard est destiné à la calibration du réactif. Ces dispositifs de diagnostic *in vitro* sont uniquement destinés aux professionnels.

▣SIGNIFICATION CLINIQUE ⁽¹⁻³⁾

L'urée est le principal produit du catabolisme protéique. Elle provient du foie et est principalement excrétée par les reins.

La concentration en urée dans le sang peut être augmentée par de nombreux facteurs liés soit à des causes pré-rénales (augmentation du catabolisme protéique comme lors d'une hémorragie au niveau du tractus gastro-intestinal, un choc), soit à des causes rénales (maladies rénales aiguës ou chroniques) ou post-rénales (obstruction à l'écoulement urinaire). L'urémie est également augmentée en cas de régime à haute valeur protéique ou de déshydratation. Une diminution de la concentration sérique en urée peut s'observer pendant la grossesse ou avec une alimentation pauvre en protéines.

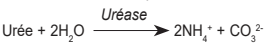
En pratique le dosage de l'urée dans le sérum est effectué pour aider au diagnostic des pathologies rénales, pour le suivi des traitements de certaines de ces pathologies ainsi que pour le suivi de la fonction rénale en cours de certains traitements pouvant altérer cette fonction. En raison des nombreuses causes non rénales de variation des taux sériques, l'urée est un moins bon marqueur de la fonction rénale que la créatinine.

▣LIMITE D'UTILISATION

Le dosage de l'urée ne peut être utilisé seul pour diagnostiquer une maladie ou une pathologie spécifique. Les résultats doivent toujours être confrontés aux résultats d'autres tests diagnostiques, aux examens cliniques, et à l'histoire médicale du patient.

▣MÉTHODE & PRINCIPE ⁽⁴⁾

Uréease/GIDH – Cinétique



GIDH = Glutamate déshydrogénase

▣COMPOSITION

Réactif 1 : R1

Tampou buffer, pH 7.60 (37 °C)

α-Cétoglutarate 9 mmol/L

Uréease \geq 8 100 U/L

GIDH \geq 1 350 U/L

Azide de sodium < 0.1 % (p/p)

Réactif 2 : R2

NADH 1.3 mmol/L

Azide de sodium < 0.1 % (p/p)

Standard: Std (Ref : URSL-0427/0507)

Urea 50 mg/dL

8.33 mmol/L

MATÉRIELS REQUIS MAIS NON FOURNIS

- CALI-0550 ELICAL 2

- CONT-0060 ELITROL I

- CONT-0160 ELITROL II

- Solution saline normale (NaCl 9 g/L).

- Automates ou semi-automates.

- Equipement général de laboratoire (ex. pipette).

- Ne pas utiliser de matériel ne figurant pas ci-dessus.

▣PRÉCAUTIONS D'EMPLOI ET MISES EN GARDE

- Consulter la fiche de données de sécurité (FDS) pour une manipulation appropriée.

- Les réactifs contiennent de l'azide de sodium qui peut réagir avec le plomb ou le cuivre et former des azides métalliques potentiellement explosifs. Lors de l'élimination de ces réactifs toujours rincer abondamment avec de l'eau pour éviter l'accumulation d'azides.

- Respecter les précautions d'usage et les bonnes pratiques de laboratoire.

- Utiliser du matériel de laboratoire propre ou à usage unique afin d'éviter toute contamination.

- Ne pas échanger les flacons réactifs de différents kits.

▣STABILITÉ

Stocké à 2-8 °C et à l'abri de la lumière. Ne pas congeler.

Ne pas utiliser après la date d'expiration indiquée sur les étiquettes des flacons.

Le standard doit être immédiatement et correctement réfermé afin d'éviter toute contamination ou évaporation.

Stabilité à bord : La stabilité à bord est spécifique à chaque automate. (Se référer au § PERFORMANCES).

PRÉPARATION

Le réactif et le standard sont prêts à l'emploi.

DÉTÉRIORATION DU PRODUIT

- Le produit doit être limpide. Tout trouble serait le signe d'une détérioration du produit.

- Ne pas utiliser le produit s'il y a des signes évidents de contamination ou de détérioration (ex : particules).

- Un flacon endommagé peut avoir un impact sur les performances du produit. Ne pas utiliser le produit si les flacons présentent des signes physiques de détérioration (par exemple, fuite, flacon percé).

URSL

☛

URSL-0427

URSL-0507

URSL-0250

URSL-0455

URSL-0420

URSL-0500

R1 4 x 50 mL + **R2** 2 x 26 mL + **Std** 1 x 5 mL

R1 5 x 100 mL + **R2** 1 x 127 mL + **Std** 1 x 5 mL

R1 8 x 20 mL + **R2** 8 x 5 mL

R1 4 x 44 mL + **R2** 4 x 11 mL

R1 4 x 50 mL + **R2** 2 x 26 mL

R1 5 x 100 mL + **R2** 1 x 127 mL



Les résultats doivent être dans les intervalles définis. Si les valeurs se situent en dehors des plages définies, chaque laboratoire devra prendre les mesures correctives nécessaires.

TRAITEMENT DES DÉCHETS

L'élimination de tous les déchets doit être effectuée conformément aux exigences réglementaires locales, d'état et fédérales (veuillez vous référer à la fiche de données de sécurité (FDS)).

PERFORMANCES

Les performances ont été obtenues sur l'automate Selectra ProM, en suivant les recommandations CLSI, dans des conditions environnementales contrôlées.

- Domaine de mesure

a) *Sérum/Plasma*

10 - 300 mg/dL (1.67 - 49.95 mmol/L).

Les échantillons ayant des concentrations supérieures devront être dilués au 1/5 dans une solution de NaCl 9 g/L et redosés. Cette procédure étend le domaine de mesure jusqu'à 1 500 mg/dL (249.75 mmol/L). Ne pas communiquer de résultats en dehors du domaine de mesure étendu.

Pour les utilisateurs du logiciel Selectra TouchPro, la fonction « diluer » réalise la dilution des échantillons automatiquement. Les résultats tiennent compte de la dilution.

b) *Urine*

200 - 6 000 mg/dL (33 - 999 mmol/L).

Ne pas communiquer de résultats en dehors du domaine de mesure.

- **Limite de Détection (LoD) et Limite de Quantification (LoQ)**

a) *Sérum/Plasma*

LoD = 1.5 mg/dL (0.25 mmol/L)

LoQ = 5.0 mg/dL (0.83 mmol/L)

b) *Urine*

LoD = 18 mg/dL (3 mmol/L)

LoQ = 200 mg/dL (33 mmol/L)

- Précision

Les données d'imprécision ont été obtenues sur 2 automates Selectra ProM sur 20 jours (2 routines par jour, tests effectués en double).

Des résultats représentatifs sont présentés ci dessous :

a) *Sérum/Plasma*

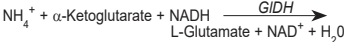
	Moyenne			Intra-série	Total
	n	mg/dL	mmol/L	CV (%)	
Niveau 1	80	18.0	3.00	1.6	3.2
Niveau 2	80	59.0	9.82	1.2	2.2
Niveau 3	80	144.6	24.08	1.0	2.1

b) *Urine*

	Moyenne			Intra-série	Total
	n	mg/dL	mmol/L	CV (%)	
Niveau 1	80	482	80	1.7	3.8
Niveau 2	80	1165	194	0.6	

⇨METHOD & PRINCIPLE ⁽⁴⁾

Urease/GIDH - Kínetic.



GIDH = Glutamate dehydrogenase

⇨COMPOSITION

Reagent 1 : R1
Tris buffer, pH 7.60 (37 °C)
α-Ketoglutarate 9 mmol/L
Urease ≥ 8100 U/L
GIDH ≥ 1 350 U/L
Sodium azide < 0.1% (w/w)

Reagent 2 : R2
NADH 1.3 mmol/L
Sodium azide < 0.1% (w/w)
Standard: Std (Ref : URSL-0427/0507)
Urea 50 mg/dL
8.33 mmol/L

MATERIALS REQUIRED BUT NOT PROVIDED

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Normal saline solution (NaCl 9 g/L).
- Analyzers or semi-automatic analyzers.
- General Laboratory equipment (e.g. pipette).
- Do not use materials that are not required as indicated above.

⇨PRECAUTIONS FOR USE AND WARNINGS

- Consult Safety Data Sheet (SDS) for a proper handling.
- The reagents contain sodium azide which may react with lead or copper plumbing to form potentially explosive metal azides. When disposing of these reagents always flush with copious amounts of water to prevent azide buildup.
- Take normal precautions and adhere to good laboratory practice.
- Use clean or single use laboratory equipment only to avoid contamination.
- Do not interchange reagent vials from different kits.

⇨STABILITY

Store at 2-8 °C and protect from light. Do not freeze.
Do not use after expiration dates indicated on the vial labels.
The standard should be immediately and tightly capped to prevent contamination and evaporation.
On board stability :
The on-board stability is specific for each analyzer. (Refer to § PERFORMANCE DATA).

PREPARATION

The reagent and standard are ready to use.

PRODUCT DETERIORATION

- The product should be clear. Cloudiness would indicate deterioration.
- Do not use the product if there is visible evidence of contamination or damage (e.g. particle matter).
- Damage to the product container may impact on product performance. Do not use the product if there is physical evidence of deterioration (e.g. leakages or punctured container).

⇨SAMPLES

Specimen ⁽³⁾
- Serum.
- Plasma (lithium heparin).
- Urine.
- Using any other specimen type should be validated by the laboratory.
Warnings and precautions
- Addition of thymol as preservative is not recommended as it inhibits urease activity ⁽³⁾
- Samples should be collected in accordance with Good Laboratory Practice and appropriate guidelines that may be in place.
Storage and stability ^(2,3)
Serum/plasma
- 24h at room temperature.
- 1 week at 2-8 °C.
- 3 months at -20 °C.
Urine
- 4 days at 2-8 °C provided bacterial contamination is avoided.

⇨REFERENCE VALUES ⁽²⁾

<i>Serum/plasma</i>	mg/dL	mmol/L
Children <1 year old	8.6 - 40.7	1.4 - 6.8
Children 1-18 years old	10.7 - 38.6	1.8 - 6.4
Adults (18 - 60 years)	12.9 - 42.9	2.14 - 7.14
Adults (60 - 90 years)	17.2 - 49.3	2.86 - 8.21
Adults (>90 years)	21.4 - 66.5	3.57 - 11.07

<i>Urine (24h - collection)</i>	g/24h	mol/24 h
Adults	26 - 43	0.43 - 0.71
	mg/dL*	mmol/L*
	1 700 - 2 900	290 - 470

* for a urinary volume of 1.5 L per 24 hours

Note : *The quoted range should serve as a guide only. It is recommended that each laboratory verifies this range or establishes a reference interval for the intended population.*

⇨PROCEDURE
Manual Procedure
Wavelength : 340 nm
Optical path : 1 cm
Sample/ Reagent ratio : 1:100
Temperature : 37 °C
Urine samples must be diluted 1:20 with NaCl 9 g/L solution before measurement.

Read against distilled water.

Working reagent (4 volumes of R1 + 1 volume of R2)	1000 µL
Sample	10 µL

Mix and after 30 seconds incubation, read absorbance at 30 seconds intervals during 90 seconds. Calculate the change of absorbances per minute (ΔA/min).

Automatic Procedure

These reagents may be used on several automatic analyzers. For ELITech Selectra Analyzers, validated applications are available on request. For Selectra TouchPro software, use the application included in the barcode available at the end of this insert.

Urine samples must be diluted 1:20 with NaCl 9 g/L solution before measurement. For users of Selectra TouchPro software, urine dilution is performed automatically.

⇨CALCULATION

$\frac{\Delta A}{\text{min. Sample}} \times n$ n = Calibrator/ standard
 $\frac{\Delta A}{\text{min. Calibrator/ Standard}}$ concentration

For the calculation of urea concentration in urine, multiply the result by the dilution factor (20). For users of Selectra TouchPro software, the results take the dilution factor into account.

Conversion factor : mg/dL x 0.1665 = mmol/L

⇨CALIBRATION

For the reference **URSL-0427/0507** : ELICAL 2 and Urea Standard 50 mg/dL are traceable to ID-MS (Isotope Dilution - Mass Spectrometry) reference method.
For the reference **URSL-0250/0455/0420/0500** : ELICAL 2 is traceable to ID-MS (Isotope Dilution - Mass Spectrometry) reference method.

Calibration frequency : The calibration is specific for each analyzer. (Refer to § PERFORMANCE DATA).

QUALITY CONTROL

It is recommended that quality control sera such as ELITROL I and ELITROL II be used to monitor the performance of the assay.
Controls have to be performed :
- prior to assaying patient samples,
- at least once per day,
- after every calibration,
- and/or in accordance with laboratory and regulatory requirements.
Results should be within the defined ranges. If values fall outside of the defined ranges, each laboratory should take necessary corrective measures.

WASTE MANAGEMENT

Disposal of all waste material should be in accordance with local, state and federal regulatory requirements (please refer to the Safety Data Sheet (SDS)).

PERFORMANCES

Performances were obtained on Selectra ProM, following CLSI technical recommendations, under controlled environmental conditions.

- Measuring range

a) Serum/Plasma
100 - 300 mg/dL (1.67 - 49.95 mmol/L)
Samples having greater concentrations should be diluted 1:5 with NaCl 9 g/L solution and re-assayed. This procedure extends the measuring range up to 1 500 mg/dL (249.75 mmol/L).
Do not report results outside this extended range.

For users with Selectra TouchPro software, the «dilute» function performs the sample dilution automatically. Results take the dilution into account.

b) Urine
200 - 6 000 mg/dL (33 - 999 mmol/L)
Do not report results outside the measuring range.

Limit of Detection (LoD) and Limit of Quantification (LoQ)

a) Serum/Plasma
LoD = 1.5 mg/dL (0.25 mmol/L)
LoQ = 5.0 mg/dL (0.83 mmol/L)

b) Urine
LoD = 18 mg/dL (3 mmol/L)
LoQ = 200 mg/dL (33 mmol/L)

- Precision

Imprecision data has been obtained on 2 Selectra ProM analyzers over 20 days (2 runs per day, tests performed in duplicate).

Representative results are presented below :

		Mean	Within-run	Total
	n	mg/dL	mmol/L	CV (%)
Level 1	80	18.0	3.00	1.6 3.2
Level 2	80	59.0	9.82	1.2 2.2
Level 3	80	144.6	24.08	1.0 2.1

b) Urine

		Mean	Within-run	Total
	n	mg/dL	mmol/L	CV (%)
Level 1	80	482	80	1.7 3.8
Level 2	80	1165	194	0.6 3.1
Level 3	80	2587	431	0.4 3.6

- Correlatíon

a) Serum/Plasma
A comparative study has been performed between UREA UV SL reagent on a Selectra ProM analyzer and a similar commercially available system on 98 human serum samples.
The sample concentrations ranged from 12.5 to 285.5 mg/dL (2.08 - 47.54 mmol/L).
The results are as follows :
Correlation coefficient : (r) = 1.000
Linear regression: y = 0.993x - 0.1 mg/dL (0.02 mmol/L)

⇨b) Urine

A comparative study has been performed between UREA UV SL reagent on a Selectra ProM analyzer and a similar commercially available system 53 human urine samples.
The sample concentrations ranged from 203 to 5 569 mg/dL (34 - 927 mmol/L).
The results are as follows :
Correlation coefficient : (r) = 0.999
Linear regression: y = 1.000x + 52 mg/dL (9 mmol/L)

⇨- Limitations/Interferences

a) Serum/Plasma
Studies have been performed to determine the level of interference from different compounds.
The following urea levels were tested: 15.0 mg/dL and 60.1 mg/dL.
No significant interference is defined by a recovery ≤±10% of the initial value.
Unconjugated Bilirubin: No significant interference up to 30.0 mg/dL (513 µmol/L).
Conjugated Bilirubin: No significant interference up to 29.5 mg/dL (505 µmol/L).
Turbidez: No significant interference up to 614 mg/dL (6.94 mmol/L) Triglycoeride equivalent.
Hemoglobin: No significant interference up 500 mg/dL.
Ascorbic acid: No significant interference up 20.0 mg/dL.
Methyl dopa: No significant interference up 1.0 mg/dL.

- In very rare cases, monoclonal gammopathies (multiple myeloma), in particular IgM type (Waldenström's macroglobulinemia) can cause unreliable results.⁽⁵⁾

- Many other substances and drugs may interfere. Some of them are listed in reviews published by Young.^(6,7)

b) Urine

- Studies have been performed to determine the level of interference from different compounds.
The following urea levels were tested: 1 500 mg/dL and 3 000 mg/dL.
No significant interference is defined by a recovery ≤±10% of the initial value.
Conjugated Bilirubin: No significant interference up to 29.5 mg/dL (505 µmol/L).
Hemoglobin: No significant interference up 500 mg/dL.
Ascorbic acid: No significant interference up 20.0 mg/dL.
Uric acid: No significant interference up 120 mg/dL (7.14 mmol/L).
pH: No significant interference for pH values ranging between 2.5 and 12.0.

- Many other substances and drugs may interfere. Some of them are listed in reviews published by Young.^(6,7)

- **On board stability/Calibration frequency**
On Board Stability: 14 days
Calibration frequency: 7 days
Recalibrate when reagent lots change, when quality control results fall outside the established range and after a maintenance operation.

These performances have been obtained using ELITech Selectra ProM analyzer. Results may vary if a different instrument or a manual procedure is used. The performances of applications not validated by ELITech are not warranted and must be defined by the user.

⇨DECLARATION OF SERIOUS INCIDENT

Please notify the manufacturer (through your distributor) and complete authority of the Member State of the european union in which the user and/or the patient is established, of any serious incident that has occurred in relation to the device. For other jurisdictions, the declaration of serious incident should be in accordance with local, state and federal regulatory requirements. By reporting a serious incident, you provide information that can contribute to the safety of *in vitro* medical devices.

⇨TECHNICAL ASSISTANCE

Contact your local distributor or ELITech Clinical Systems SAS (CCsupport@elitechgroup.com).

Español - ES

⇨USO PREVISTO

ELTech Clinical Systems UREA UV SL es un reactivo de diagnóstico *in vitro* diseñado para la determinación cuantitativa de la urea en muestras de suero, plasma y orina humanas en equipos automatizados o equipos semiautomáticos.
El estándar está diseñado para la calibración del reactivo.
Estos dispositivos de diagnóstico *in vitro* están destinados únicamente para los profesionales.

⇨SIGNIFICADO CLÍNICO ⁽¹⁻³⁾

La urea es el principal subproducto del catabolismo proteico. Se origina en el hígado y se excreta principalmente a través de los riñones.
Las concentraciones de urea en sangre pueden incrementarse por numerosos factores relacionados con causas prerrenales (aumento del catabolismo de las proteínas como en la hemorragia en el tracto gastrointestinal, shock) o causas renales (enfermedades renales agudas o crónicas) o causas post renales (obstrucción del flujo de orina). La uremia también se incrementa por la dieta alta en proteínas o la deshidratación. Se puede observar una disminución de la concentración sérica de urea durante el embarazo o con una dieta baja en proteínas.
En la práctica, la medición de la urea en suero se realiza para ayudar a diagnosticar enfermedades renales, para controlar los tratamientos contra estas patologías o para controlar la función renal durante los tratamientos que pueden afectar esta función. Debido a muchas causas no renales para la variación del nivel sérico, sin embargo la urea es un marcador menos bueno de la función renal que la creatinina.

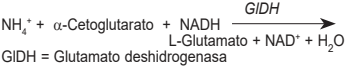
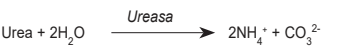
⇨LÍMITE DE UTILIZACIÓN

La cuantificación de la urea no puede ser utilizado solo para diagnosticar una enfermedad o patología específica.

Los resultados siempre deben compararse con los resultados de otras pruebas de diagnóstico, exámenes clínicos y el historial médico del paciente.

⇨MÉTODO & PRINCIPIO ⁽⁴⁾

Urease/GIDH – Cínetica.



⇨COMPOSICIÓN

Reactivo 1 : R1
Tampón Tris, pH 7.60 (37 °C) 9 mmol/L
α-Cetoglutarato ≥ 8 100 U/L
Ureasa ≥ 1 350 U/L
GIDH < 0.1 % (p/p)
Azida sodíca
Reactivo 2 : R2
NADH 1.3 mmol/L
Azida sodíca < 0.1 % (p/p)
Estándar : Std (Ref : URSL-0427/0507)
Urea 50 mg/dL
8.33 mmol/L

MATERIALES REQUERIDOS PERO NO INCLUIDOS

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Solución salina normal (NaCl 9 g/L).
- Equipos automatizados o equipos semiautomáticos.
- Equipamiento general de laboratorio (p. ej. pipeta).
- No utilice materiales que no se requieren, tal como se indica anteriormente.

⇨PRECAUCIONES DE USO Y ADVERTENCIAS

- Consulte la Hoja de Datos de Seguridad (SDS) para un manejo adecuado.

- Los reactivos contienen azida sodíca que puede reaccionar con el plomo o el cobre de la tubería y formar potencialmente azidas metálicas explosivas. Cuando se elimine el reactivo enjuague con agua abundantemente para prevenir la acumulación de azidas.
- Tome las precauciones normales y respete las buenas prácticas de laboratorio.
- Para evitar contaminaciones utilizar equipo nuevo o completamente limpio.
- No intercambie los frascos de reactivos de diferentes kits.

⇨ESTABILIDAD

Conservar a 2-8 °C y protegidos de la luz. No congelar.

No utilice después de la fecha de caducidad indicada en la etiqueta de los frascos.
El estándar debe cerrarse inmediatamente y correctamente para evitar contaminación y evaporación. Estabilidad en el equipo.

La estabilidad es específica para cada equipo. (Referirse al § DATOS DE RENDIMIENTO).

PREPARACIÓN

El reactivo y el estándar están listos para su uso.

DETERIORACION DEL PRODUCTO

- El producto debe ser claro. Turbidez indicaría deterioro.
- No utilice el producto si este presenta signos evidentes de contaminación o deterioro (p. ej partículas).
- Un frasco dañado puede tener un impacto en el rendimiento del producto. No utilice el producto si este tiene signos físicos de deterioro (p. ej, fugas, frasco perforado).
⇨MUESTRAS
Muestras requeridas ⁽³⁾
- Suero
- Plasma (heparina de litio).
- Orina.
- El uso de cualquier otro tipo de muestra debe ser validado por el laboratorio.

Advertencias y precauciones

- No se recomienda la adición de timol como conservador, ya que inhibe la actividad de la ureasa. ⁽⁶⁾
- Las muestras deben tomarse de acuerdo con las Buenas Prácticas de Laboratorio y las guías apropiadas establecidas.

Conservación y estabilidad ^(2,a)

Suero/Plasma
- 24h a temperatura ambiente.
- 1 semana a 2-8 °C
- 3 meses a -20 °C
Orina
- 4 días a 2-8 °C en ausencia de contaminación bacteriana.

⇨VALORES DE REFERENCIA ⁽²⁾
Suero/plasma

Niños <1 años	8.6 - 40.7	1.4 - 6.8
Niños 1-18 años	10.7 - 38.6	1.8 - 6.4
Adultos (18 - 60 años)	12.9 - 42.9	2.14 - 7.14
Adultos (60 - 90 años)	17.2 - 49.3	2.86 - 8.21
Adultos (> 90 años)	21.4 - 66.5	3.57 - 11.07

<i>Orina (recolectada de 24h)</i>	g/24h	mol/24 h
Adultos	26 - 43	0.43 - 0.71
	mg/dL*	mmol/L*
	1 700 - 2 900	290 - 470

* para un volumen de orina de 1.5 L por 24 horas

Nota : *Los valores anteriores son solo indicativos. Se recomienda que cada laboratorio establezca y mantenga sus propios valores de referencia en relación con la población destinataria.*

⇨PROCEDIMIENTO

Procedimiento manual
Longitud de onda : 340 nm
Traectoria óptica : 1 cm
Ratio muestra/reactivo : 1:100
Temperatura : 37 °C
Las muestras de orina deben diluirse 1:20 con una solución de NaCl 9 g/L antes de la medición
Leer contra agua destilada.

Reactivo de trabajo (4 volúmenes de R1 +1 volumen de R2)	1000 µL
Muestra	10 µL

Mezclar después de incubar 30 segundos, leer la absorbancia a intervalos de 30 segundos durante 90 segundos. Calcule el cambio de absorbancia por minuto (ΔA/min).

Procedimiento automático

Estos reactivos pueden ser utilizados en varios equipos. Para los equipos ELITech Selectra, las aplicaciones validadas están disponibles sobre pedido. Para el software Selectra TouchPro, use la aplicación incluida en el código de barras disponible al final de este inserto.

Las muestras de orina deben diluirse 1:20 con una solución de NaCl 9 g/L antes de la medición. Para los usuarios del software Selectra TouchPro, la dilución de orina se realiza automáticamente.

⇨CÁLCULO

$\frac{\Delta A}{\text{min. Muestra}} \times n$ n = concentración del estándar/calibrador

⇨b) Orina

Para el cálculo de la concentración de urea en orina, multiplique el resultado por el factor de dilución (20). Para los usuarios del software Selectra TouchPro, los resultados toman en cuenta el factor de dilución.

Factor de conversión: mg/dL x 0.1665 = mmol/L

⇨CALIBRACIÓN

Para las referencias **URSL-0427/0507** : ELICAL 2 o el estándar Urea Standard 50 mg/dL son trazables al método de referencia DI-EM (Dilución Isotópica - Espectrometría de Masas)
Para las referencias **URSL-0250/0455/0420/0500** : ELICAL 2 es trazable al método de referencia DI-EM (Dilución Isotópica - Espectrometría de Masas).

Frecuencia de calibración : la frecuencia de calibración es específica para cada equipo (referirse al § DATOS DE RENDIMIENTO).

CONTROL DE CALIDAD

Es recomendado que sueros de control tales como ELITROL I y ELITROL II sean usados para monitorear el rendimiento de las pruebas.
Los controles deben realizarse :
- antes que las muestras del paciente sean evaluadas, por lo menos una vez al día,
- después de cada calibración,
- y/o en acuerdo con el laboratorio y los requerimientos regulatorios.
Los resultados deben de encontrarse en el rango definido. Si los valores se encuentran fuera del mismo, cada laboratorio deberá tomar las medidas correctivas necesarias.

Los niveles siguientes de urea fueron probados: 15.0 mg/dL y 60.1 mg/dL.

- Estudios fueron llevados a cabo para determinar el nivel de interferencia de diferentes componentes.
No niveles siguientes de urea fueron probados: 1 500 mg/dL y 3 000 mg/dL.
Definimos una interferencia no significativa cuando se obtiene una recuperación de ≤±10% con respecto al valor inicial.
⇨MUESTRA
Muestras requeridas ⁽³⁾
- Suero
- Plasma (heparina de litio).
- Orina.
- El uso de cualquier otro tipo de muestra debe ser validado por el laboratorio.

⇨Rango analítico</



a) Soro, plasma

Turvação: Desvio positivo a partir de 300 mg/dL (3,39 mmol/L) equivalente de triglicéridos.

Hemoglobina: Desvio positivo a partir de 250 mg/dL (2,5 g/L).

Bilirrubina conjugada: Nenhuma interferência significativa até 25 mg/dL (427,6 µmol/L).

Bilirrubina não conjugada: Nenhuma interferência significativa até 36 mg/dL (615,8 µmol/L) em soro normal e desvio positivo a partir de 22,5 mg/dL (384,8 µmol/L) em soro patológico.

- Em casos muito raros, as gamopatias monoclonais (mieloma múltiplo), em particular, tipo IgM (macroglubulinemia de Waldenström) podem causar resultados não confiáveis.⁽⁶⁾

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young⁽⁸⁻¹⁰⁾.

- Os resultados deste teste só devem ser interpretados em conjunto com outros resultados de testes de diagnóstico, que constem no historial médico e clínico do paciente

b) Urina

Hemoglobina: Desvio positivo a partir de 3 g/L.

Bilirrubina conjugada : Nenhuma interferência significativa até 25 mg/dL (427,6 µmol/L).

- Muitas outras substâncias e drogas podem interferir. Alguns deles estão referenciados em análises publicadas por Young⁽⁸⁻¹⁰⁾.

- Os resultados deste teste só devem ser interpretados em conjunto com outros resultados de testes de diagnóstico, que constem no historico médico e clínico do paciente.

- Estabilidade a bordo / frequência de calibração
Estabilidade a bordo: 28 dias
Frequência de calibração: 7 dias
Uma nova calibração deve ser efetuada após cada mudança de lote de reagente, quando os resultados do(s) controle(s) de qualidade estiverem fora do intervalo estabelecido e após uma operação de manutenção.

☛ *Estes desempenhos foram obtidos utilizando o analisador ELITech Selectra E. Os resultados podem variar se um instrumento diferente ou um procedimento manual for usado.*
Os desempenhos de aplicações não validados pela ELITech não são garantidos e devem ser definidos pelo usuário.

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☛ SYMBOLES/SYMBOLS/ SÍMBOLOS/SÍMBOLOS

- Les symboles utilisés sont décrits dans la norme ISO-15223-1 hormis ceux présentés ci-dessous.
- Symbols used are defined on ISO-15223-1 standard, except those presented below.
- Los símbolos utilizados son descritos en la norma ISO-15223-1 a la excepción de los presentados a continuación.
- Os símbolos utilizados são definidos na norma ISO-15223-1, exceto os apresentados abaixo.

CONT	Contient Content Contiene Conteúdo
R	Réactif Reagent Reactivo Reagente
Std	Standard Standard Estándar Padrão
CE	Conformité Européenne European Conformity Conformidad Europea Conformidade Europeia

CHLO 260 VTL-CHLO 0

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CHLO 260 VTL-CHLO 0

VTL-CHLO-4-v14 (09/2019)

Français - FR

Code technique : EL

☛ USAGE PRÉVU

ELITech Clinical Systems CHLORIDE est un réactif de diagnostic *in vitro*, destiné au dosage quantitatif du chlorure dans les échantillons de sérum, de plasma et d'urine humains.

SIGNIFICATION CLINIQUE ⁽¹⁻³⁾

Le chlorure est le principal anion présent dans le fluide extracellulaire. Il assure une pression osmotique, un bilan hydrique et un équilibre anion-cation appropriés. Sa concentration est similaire à celle du sodium et subit les mêmes influences.

La détermination de la concentration plasmatique en Cl⁻ est utile pour le diagnostic différentiel des perturbations acido-basiques et est essentiel pour le calcul du trou anionique. Le dosage dans les urines est utile chez les patients avec une alcalose métabolique persistante ne recevant pas de diurétiques.

Une hypochlorémie est observée chez les individus souffrant de néphrite avec perte de sels couplée à une hyponatrémie, lors d'intoxication au bromure, de syndrome d'antiurdièse, d'expansion du fluide extracellulaire, d'alcalose métabolique, de sécrétions gastriques persistantes ou de vomissements prolongés.

Une hyperchlorémie apparaît lors de déshydratation, d'acidose tubulaire rénale, d'insuffisance rénale aiguë, d'acidose métabolique, de diabète insipide ou de prise extrêmement importante de sel.

MÉTHODE ⁽⁴⁾

Thiocyanate de mercure. Colorimétrie. Point final.

PRINCIPE ⁽⁴⁾

En présence de nitrate ferrique et de thiocyanate mercurique, les ions chlorures provoquent la formation de thiocyanate ferrique. L'intensité de coloration de ce complexe brun est proportionnelle à la concentration en chlorures :



☛ COMPOSITION

Réactif : R
Thiocyanate de mercure (II) 1,3 mmol/L
Nitrate ferrique (III) 30 mmol/L
Acide nitrique 29 mmol/L
Standard: Std (Ref : CHLO-0600)
Chlorures 100 mEq/L

☛ MATÉRIELS REQUIS MAIS NON FOURNIS

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Equipement général de laboratoire.
- Ne pas utiliser de matériel ne figurant pas ci-dessus.

☛ AVERTISSEMENTS ET PRÉCAUTIONS

- Ces dispositifs (réactif et standard) de diagnostic *in vitro* sont uniquement destinés aux professionnels.
- Le réactif R est classé comme dangereux :



ATTENTION : Provoque une irritation cutanée. Provoque une sévère irritation des yeux. Porter des gants de protection / un équipement de protection des yeux / du visage.

EN CAS DE CONTACT AVEC LES YEUX: rincer avec précaution à l'eau pendant plusieurs minutes. Enlever les lentilles de contact si la victime en porte et si elles peuvent être facilement enlevées. Continuer à rincer. Si l'irritation oculaire persiste: consulter un médecin.

- Pour plus d'information, se référer à la fiche de données de sécurité (FDS).
- Le standard doit être immédiatement et correctement réfermé afin d'éviter toute contamination ou évaporation.
- Respecter les précautions d'usage et les bonnes pratiques de laboratoire.
- Utiliser du matériel de laboratoire propre ou à usage unique afin d'éviter toute contamination.

☛ STABILITÉS

Stocké à 15-25 °C et à l'abri de la lumière. Ne pas congeler.
Ne pas utiliser après la date d'expiration indiquée sur les étiquettes des flacons.

Stabilité à bord :
La stabilité à bord est spécifique à chaque automate. (Se référer au § PERFORMANCES).

PRÉPARATION

Le réactif et le standard sont prêts à l'emploi.

VTL-CHLO-4-v14 (09/2019)

Français - FR

Code technique : EL

☛ DÉTÉRIORATION DU PRODUIT

- Le produit doit être limpide. Tout trouble serait le signe d'une détérioration du produit.
- Ne pas utiliser le produit s'il y a des signes évidents de détérioration biologique, chimique ou physique.
- Ne pas utiliser le produit si les dommages de l'emballage peuvent avoir un effet sur les performances du produit (fuites, flacon percé).

ÉCHANTILLONS ^(2,5)

Echantillons requis
- Sérum.
- Plasma recueilli sur héparine.
- Urine de 24h diluée au 1/2 avec de l'eau déminéralisée.
- Ne pas utiliser d'autres échantillons.

☛ Avertissements et précautions
- Selon les Bonnes Pratiques de Laboratoire, tout prélèvement devrait être réalisé avant l'administration de médicaments.
- Après prélèvement, tous les échantillons doivent être rapidement séparés des cellules afin d'éviter une rupture de l'équilibre ionique et des modifications du métabolisme et du pH.

Stockage et stabilité
Les ions chlorures présents dans les échantillons sont stables au moins 1 semaine à température ambiante, au réfrigérateur ou au congélateur

VALEURS DE RÉFÉRENCE ⁽⁵⁾

Sérum, plasma : 98 - 107 mEq/L
Urine, 24h : 110 - 250 mEq/24h

☛ *Remarque : Il est recommandé à chaque laboratoire d'établir et de maintenir ses propres valeurs de référence par rapport à la population visée. Les valeurs ci-dessus ne sont données qu'à titre indicatif.*

☛ PROCÉDURE

Procédure manuelle
Longueur d'onde : 505 nm
Trajet optique : 1 cm
Ratio échantillon/réactif : 1:100
Température: 37 °C
Lire contre le blanc réactif.

	CALIBRATION	DOSAGE
Réactif R	1 000 µL	1 000 µL
Calibrant/Standard	10 µL	-
Echantillon	-	10 µL

Mélanger et lire les absorbances (A) après 5 minutes d'incubation.

Procédure sur automate

Ces réactifs peuvent être utilisés sur différents automates. Pour les automates ELITech Selectra, les applications validées sont disponibles sur demande. Avec le logiciel Selectra TouchPro, utilisez l'application incluse dans le code barre disponible à la fin de cette notice.

CALCUL

$$\frac{A_{\text{Echantillon}}}{A_{\text{Standard/Calibrant}}} \times n \quad n = \text{concentration du standard/calibrant}$$

Facteur de conversion : 1 mEq/L = 1 mmol/L

Pour le dosage du chlorure dans les urines, tenir compte du facteur de dilution.

☛ CALIBRATION

Pour la référence CHLO-0600 : Pour la calibration utiliser soit le calibrant multiparamétrique ELICAL 2, soit le standard Chloride 100 mEq/L.
Pour la référence CHLO-0250 : Pour la calibration, utiliser le calibrant multiparamétrique ELICAL 2.

Le standard Chloride 100 mEq/L et le calibrant multiparamétrique ELICAL 2 sont traçables par rapport au matériau de référence SRM 909c (du National Institute of Standards and Technology).

Fréquence de calibration : La fréquence de calibration est spécifique à chaque automate (se référer au § PERFORMANCES).

☛ CONTRÔLE QUALITÉ

Pour vérifier l'exactitude des résultats, les sérums de contrôle ELITROL I et ELITROL II doivent être utilisés. Ces contrôles doivent être effectués et validés avant que les échantillons des patients soient testés. La fréquence de contrôle doit être au moins une fois par jour, après chaque calibration et doit être adaptés aux procédures de contrôle de qualité de chaque laboratoire et les exigences réglementaires.

Les résultats doivent être dans les intervalles définis. Si les valeurs se situent en dehors des plages définies,

chaque laboratoire doit prendre des mesures correctives. Les matériaux de contrôle qualité doivent être utilisés conformément aux directives locales.

TRAITEMENT DES DÉCHETS

L'élimination de tous les déchets doit être effectuée conformément aux exigences réglementaires locales, d'état et fédérales.

PERFORMANCES à 37 °C sur ELITech Clinical Systems Selectra E - Domaine de mesure

a) Sérum/plasma
Le réactif est linéaire de 10 à 130 mEq/L.

b) Urine
Le réactif est linéaire de 10 à 250 mEq/L.

- Limite de Détection ⁽⁶⁾
Déterminée selon le protocole recommandé par la SFBC, la limite de détection est égale à 2 mEq/L sur sérum/plasma et urine.

- Précision

a) Sérum/plasma

	N	Reproductibilité intrasérielle		Reproductibilité intersérielle	
		Moyenne (mEq/L)	CV (%)	Moyenne (mEq/L)	CV (%)
Niveau normal	20	97,2	0,9	96,3	1,0
Niveau Pathologique	20	113,2	0,9	111,4	1,3

b) Urine

	N	Reproductibilité intrasérielle		Reproductibilité intersérielle	
		Moyenne (mEq/L)	CV (%)	Moyenne (mEq/L)	CV (%)
Niveau bas	20	30,3	0,5	32,7	2,3
Niveau moyen	20	63,8	0,3	81,8	2,1
Niveau haut	20	151,8	0,3	145,2	2,0

- Corrélation

Sérum et plasma
Une étude comparative a été réalisée entre le réactif Chloride et un autre réactif Chlorure du commerce (méthode équivalente) sur 35 échantillons de sérum et plasma.

Les valeurs s'échelonnent de 50,3 à 132,5 mEq/L. Les paramètres de la droite de régression linéaire sont les suivants:
Coefficient de corrélation : r = 0,997
Droite de régression linéaire : y = 0,942 x + 3,3 mEq/L

- Limitations/Interférences ⁽⁶⁻⁷⁾
- Ne pas communiquer de résultats en dehors du domaine de mesure testé.

- Selon les recommandations de la SFBC, des tests ont été réalisés pour déterminer le niveau d'interférence de différents composés :

a) Sérum/plasma
Turbidité : Biais positif à partir de 300 mg/dL (3,39 mmol/L) équivalent Triglycérides.
Hémoglobine : Biais positif à partir de 250 mg/dL (2,5 g/L).
Bilirrubine conjugée : Aucune interférence significative jusqu'à 25 mg/dL (427,6 µmol/L).
Bilirrubine non-conjugée : Aucune interférence significative jusqu'à 36 mg/dL (615,8 µmol/L) sur sérum normal et biais positif à partir de 22,5 mg/dL (384,8 µmol/L) sur sérum pathologique.

- Dans des cas très rares, les gammopathies monoclonales (myélome multiple), en particulier de type IgM (Macroglubulinémie de Waldenström) peuvent être à l'origine de résultats peu fiables.⁽⁸⁾

- D'autres substances et médicaments peuvent interférer. Certains d'entre eux sont répertoriés dans les revues publiées par Young.⁽⁹⁻¹⁰⁾

- Pour le diagnostic, les résultats doivent toujours être confrontés aux résultats d'autres examens et aux données de l'anamnèse du patient.

b) Urine
Hémoglobine : Biais positif à partir de 3 g/L.
Bilirrubine conjugée : Aucune interférence significative jusqu'à 25 mg/dL (427,6 µmol/L).

- D'autres substances et médicaments peuvent interférer. Certains d'entre eux sont répertoriés dans les revues publiées par Young.⁽⁹⁻¹⁰⁾

- Pour le diagnostic, les résultats doivent toujours être confrontés aux résultats d'autres examens et aux données de l'anamnèse du patient.

- Stabilité à bord/ Fréquence de calibration

Stabilité à bord : 28 jours
Fréquence de calibration : 7 jours
Une nouvelle calibration doit être effectuée après chaque changement de lot de réactif, lorsque les résultats du ou des contrôles de qualité sont hors de l'intervalle établi, et après une opération de maintenance.

☛ *Ces performances ont été définies sur un automate ELITech Selectra E. Les résultats peuvent varier si le réactif est utilisé sur un automate différent ou en méthode manuelle.*

Les performances obtenues à partir d'applications non validées par ELITech ne peuvent être garanties et doivent être définies par l'utilisateur.

English - EN

☛ INTENDED USE

ELITech Clinical Systems CHLORIDE is an *in vitro* diagnostic reagent intended for the quantitative determination of chloride in human serum, plasma and urine samples.

CLINICAL SIGNIFICANCE ⁽¹⁻³⁾

Chloride ion is the most abundant anion in the extracellular fluid. It ensures appropriated osmotic pressure, hydrous distribution and anion-cation balance. Its concentration is similar to that of sodium and is influenced by the same factors. Determination of plasma Cl⁻ concentration is useful in the differential diagnoses of acid-base disturbances and is essential for calculation of the anion gap. Measure in urine is of clinical value with patients with persistent metabolic alkalosis who are not receiving diuretics.

Hypochloremia is observed in individuals with salt-losing nephritis coupled with hyponatremia and in cases such as bromide intoxication, SIADH (Secretion of AntiDiuretic Hormone) expansion of extracellular fluid, metabolic alkalosis or persistent gastric secretion and prolonged vomiting.

Hyperchloremia accompanies dehydration, RTA (Renal Tubular Acidosis), acute renal failure, metabolic acidosis, diabetes insipidus, extremely high intake of salt.

METHOD ⁽⁴⁾

Mercuric thiocyanate. Colorimetric. End point.

PRINCIPE ⁽⁴⁾

In presence of ferric nitrate and mercuric thiocyanate, chloride ions lead to ferric thiocyanate formation. The coloration intensity of this brown complex is proportional to the chloride concentration.



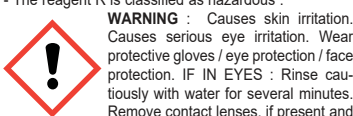
☛ COMPOSITION

Reagent : R
Mercury (II) thiocyanate 1.3 mmol/L
Ferric (III) nitrate 30 mmol/L
Nitric acid 29 mmol/L
Standard : Std (Ref : CHLO-0600)
Chloride 100 mEq/L

☛ MATERIALS REQUIRED BUT NOT PROVIDED

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- General Laboratory equipment.
- Do not use materials that are not required as indicated above.

☛ WARNINGS AND PRECAUTIONS
- These *in vitro* diagnostic devices (reagent and standard) are for professional use only.
- The reagent R is classified as hazardous :



WARNING : Causes skin irritation. Causes serious eye irritation. Wear protective gloves / eye protection / face protection. IF IN EYES : Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. If eye irritation persists: Get medical advice/attention.

- For more information, refer to the Safety Data Sheet (SDS).

- The standard should be immediately and tightly capped to prevent contamination and evaporation.

- Take normal precautions and adhere to good laboratory practice.

- Use clean or single use laboratory equipment only to avoid contamination.

Referințe:

GISL-0250
 GISL-0400
 GISL-0420

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
 R1 2 x 50 mL + R2 1 x 26 mL
 R1 4 x 50 mL + R2 2 x 26 mL



VTLRO-GISL-v9 (08/2020)_VTL-GISL-4-v9

SCOPUL UTILIZĂRII

ELITech Clinical Systems GAMMA GT PLUS SL este un reactiv de diagnostic *in vitro* destinat determinării cantitative a γ -GT din probele serul uman și plasmă.

SEMNIFICAȚIE CLINICĂ ⁽¹⁻³⁾

Gama glutamiltransferaza (γ -GT) este o enzimă legată de membrană prezentă în special în rinichi, pancreas, ficat și prostată. Această enzimă are un rol important în metabolismul glutationului și ia parte la transportul aminoacizilor în celule. Creșterea activității γ -GT este mai sensibilă decât fosfataza alcalină (ALP) în timpul unei afecțiuni a ficatului sau a căilor biliare. Cele mai mari creșteri sunt observate în cazurile de obstrucții biliare intrahepatice sau post-hepatice (ajungând la nivele de 5 până la 30 de ori mai mari decât normalul), neoplasme primare sau metastazice ale ficatului, bolile pancreatice (pancreatită, cancer...). Gama-glutamiltransferaza (γ -GT) este utilă ca un marker pentru cancerul pancreatic, cancerul de prostată și hematoame, deoarece nivelele reflectă remiterea și recurența.

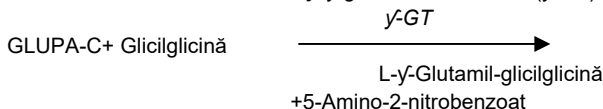
Creșteri mai moderate sunt observate în timpul hepatitei infecțioase, cirozei și steatozei hepatice. Alcool în ingestie cronică, unele medicamente precum antiepilepticele (fenobarbital, fenitoină) pot, de asemenea, crește rata γ -GT în ser.

METODĂ ⁽⁴⁾

Substrat Glupa-C – Standardizare conform metodei IFCC – Enzimatică. CINETICĂ.

PRINCIPIU ⁽⁴⁾

Determinarea cinetică a activității γ -glutamiltransferazei (γ -GT).



GLUPA-C: L- γ -Glutamil-3-carboxi-p-nitroanilidă.

Creșterea absorbantei la 405 nm datorită formării de 5-amino-2-nitrobenzoat este proporțională cu activitatea γ -GT.

COMPOZIȚIA
Reactiv 1: R1

Glicilglicină, pH 7,70 (37°C)	138	mmol/L
Azidă de sodiu	< 0,1	%

Reactiv 2: R2

GLUPA-C	23	mmol/L
Azidă de sodiu	< 0,1	%

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Soluție salină obișnuită (NaCl 9 g/L).
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest dispozitiv de diagnostic *in vitro* este destinat numai pentru uz profesional.
- Reactivii R1 și R2 conțin azidă de sodiu care poate reacționa cu plumbul sau instalațiile sanitare din cupru și poate forma azide metalice explozibile. În cazul aruncării acestor reactivi, spălați întotdeauna cu cantități mari de apă pentru a preveni formarea de azide.
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.
- Nu interschimbați fiolele de reactiv din truse diferite.
- Pentru mai multe informații, Fișa de date privind siguranța (SDS) este disponibilă la cerere pentru utilizatorul profesional.

STABILITATEA

A se depozita la 2-8°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor. (Consultați Ș DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactivi trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

PROBE ⁽⁵⁾
Specimen

- Ser sau plasmă heparinizată de litiu.
- A nu se utiliza alte specimene.

Avertisment și precauții

Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.

Depozitare și stabilitate

Serurile sunt stabile timp de 7 zile la 2-8°C și la temperatura camerei, și 1 an la -20°C.

VALORI DE REFERINȚĂ ⁽⁶⁾

	Bărbați	Femei
Ser, plasmă:	10-71 U/L	6-42 U/L



Referințe:

GISL-0250
 GISL-0400
 GISL-0420

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
 R1 2 x 50 mL + R2 1 x 26 mL
 R1 4 x 50 mL + R2 2 x 26 mL



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Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere

Lungime de undă: 405 nm

Temperatură: 37°C

Citiți pe reactivul martor.

Reactiv R1	220 µL
Proba	20 µL

Amestecați și așteptați 4 minute și 43 de secunde.

Reactiv R2	55 µL
-------------------	-------

Amestecați și așteptați 50 de secunde incubația, măsurati schimbarea absorbantei pe minut ($\Delta A/\text{min}$) timp de 159 de secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestei inserții.

CALCUL

$\Delta A \text{ Proba} \times n$ n=concentrație calibrator

$\Delta A \text{ Calibrator}$

Factor de conversie: U/L x 0,0167 = µkat/L

CALIBRARE

Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2. Valoarea sa este trasabilă conform metodei IFCC⁽⁴⁾.

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

CONTROLUL CALITĂȚII

Pentru a asigura calitatea adecvată, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca eșantioanele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare, și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform liniilor directe locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele locale și legale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems
Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁷⁾, intervalul de măsurare este între 15 și 1200 U/L (0,25 și 20,00 µkat/L), care depășesc 1 200 U/L trebuie să fie diluate 1:10 cu soluție NaCl

9g/L (salină obișnuită) și re-evaluate. Utilizarea acestei proceduri extinde intervalul de măsurare până la 12 000 U/L (200,00 µkat/L).

Pentru utilizatorii Selectra TouchPro, funcția „diluare” efectuează diluția automată a eșantionului. Rezultatele iau în considerare diluția.

Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁸⁾.

LoD= 4,4 U/L (0,07 µkat/L).

LoQ= 11,6 U/L (0,19 µkat/L).

Precizie

Determinată conform protocolului CLSI EP5-A2⁽⁹⁾.

	n	Medie		În interiorul ciclului	Total
		U/L	µkat/L	CV (%)	
Nivelul scăzut	80	39,7	0,66	1,7	3,0
Nivelul mediu	80	101,5	1,69	0,5	2,0
Nivelul înalt	80	525,9	8,77	0,2	1,9

Corelație

A fost efectuat un studiu comparativ între reactivul ELITech Clinical Systems și alt sistem concurent (metoda enzimatică standardizată IFCC) pe 94 de eșantioane de ser uman conform protocolului CLSI EP9-A2⁽¹⁰⁾.

Valorile au fost între 13,0 și 1169,7 U/L (între 0,22 și 19,50 µkat/L).

Parametrii regresii liniare sunt după cum urmează:

Coefficient de corelație: (r)=0,999

Regresie liniară: $y=0,900 x + 4,9$ U/L (0,08 µkat/L)

Limitări /interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Nu utilizați eșantioane vizibil turbide.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽¹¹⁾ al CLSI. Recuperarea este în intervalul ±10% din valoarea inițială a activității γ-GT de 100,00 și 500,00 U/L.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 µmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 µmol/L).

Hemoglobină: Nicio interferență semnificativă până la 500 mg/dL.

Trigliceride: Nicio interferență semnificativă până la 926 mg/dL (10,46 mmol/L).

Glucoză: Nicio interferență semnificativă până la 540 mg/dL (29,97 mmol/L).

Acid acetilsalicilic: Nicio interferență semnificativă până la 200,00 mg/dL.



Referințe:

GISL-0250
 GISL-0400
 GISL-0420

Compoziția trusei:

R1 8 x 20 mL + R2 8 x 5 mL
 R1 2 x 50 mL + R2 1 x 26 mL
 R1 4 x 50 mL + R2 2 x 26 mL



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Acetaminofen: Nicio interferență semnificativă până la 30,0 mg/dL.

Metil-dopa: Nicio interferență semnificativă până la 1,0 mg/dL.

Doxacilină HCl: Nicio interferență semnificativă până la 20 mg/dL.

- În cazuri foarte rare, gamopatiile monoclonale (mieloame multiple), în special de tipul IgM (macroglobulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹²⁾
-
- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ⁽¹³⁻¹⁴⁾
-
- Rezultatele acestui studiu trebuie interpretate doar în conjuncție cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- **Stabilitatea la bord/Frecvența calibrării**

Stabilitatea la bord: 14 de zile

Frecvența calibrării: 14 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit, și după o operație de întreținere.

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SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.


	Conținut
	Reactiv 1
	Reactiv 2
	Conformitate europeană

Notă

Doar pentru ref. **GISL-0250**, utilizată cu software-ul TouchPro.


 Gamma GT
 760

 1
 VTL-GISL

 Modificare față de versiunea precedentă.



Referințe:

PROB-0250 12 x 20 mL

PROB-0600 2 x 125 mL

PROB-0700 4 x 250 mL

Compoziția trusei:

R 12 x 20 mL

R 2 x 125 mL + Std 1 x 5 mL

R 4 x 250 mL + Std 1 x 5 mL



VTLRO-PROB-v12 (10/2020)_VTL-PROB-4-v12

SCOPUL UTILIZĂRII

ELITech Clinical Systems TOTAL PROTEIN PLUS este reactiv de diagnostic *in vitro* destinat determinării cantitative a proteinei totale din probele serul uman și plasmă.

SEMNIFICAȚIE CLINICĂ ^(1,2)

În plasma umană, albumina este prezentă în procent de 50-60% din proteinele totale: restul fracției conține în special globuline (α 1, α 2, β și γ). Majoritatea proteinelor plasmatică sunt sintetizate de ficat, cu excepția imunoglobulinelor. Creșterea volumului plasmatic (sindromul de reținere a sării, intoxicația cu apă...) sau reducerea sa (deshidratarea legată de vomă, diaree...) induc hipoproteinemie relativă, respectiv hiperproteinemie relativă.

Pentru un volum plasmatic normal, ratele anormale ale proteinelor totale apar doar în cazul bolii care afectează concentrația albuminei sau imunoglobulinelor.

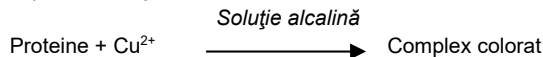
Astfel, insuficiența proteică severă (malabsorbția, maldigestia, insuficiența dietetică), bolile renale și hepatice, duc la hiperproteinemie. În cazul în care concentrația proteinelor totale este mai mică de 4g/dL, pot fi observate edemele. Hiperproteinemia poate fi observată, de exemplu, în cazul hiperimunoglobulinemiei (mielom multiplu, infecție).

METODĂ ⁽³⁾

Biuret. Punct final.

PRINCIPIU ⁽³⁾

Proteinele serice formează un complex colorat în prezența sării de cupru din soluția alcalină.

**COMPOZIȚIA****Reactiv: R**

Sulfat de cupru 6 mmol/L

Hidroxid de sodiu 490 mmol/L

Conține și ioduri de sodiu și tartrat de sodiu pentru performanță optimă.

Standard: Std. (Ref.: PROB-0600/0700)

Albumină 6 g/dL

Azidă de sodiu < 0.1 %

MATERIALE NECESARE DAR NEFURNIZATE

- CALI-0550 ELICAL 2
- CONT-0060 ELITROL I
- CONT-0160 ELITROL II
- Echipamente generale de laborator.
- Nu utilizați materiale care nu sunt necesare, după cum este indicat mai sus.

AVERTISMENTE ȘI PRECAUȚII

- Acest dispozitiv de diagnostic *in vitro* (Reactiv și Standardul) este destinat numai pentru uz profesional.
- Reactivul R este clasificat ca periculos.



ATENȚIE: Poate fi coroziv pentru metale. Provoacă iritarea pielii. Provoacă o iritare gravă a ochilor. Nociv pentru mediul acvatic cu efecte pe termen lung. Purtați mănuși de protecție/îmbrăcăminte de protecție/echipament de protecție a

ochilor/echipament de protecție a feței. Evitați dispersarea în mediu.

ÎN CAZ DE CONTACT CU OCHII: clătiți cu atenție cu apă timp de mai multe minute. Scoateți lentilele de contact, dacă este cazul și dacă acest lucru se poate face cu ușurință. Continuați să clătiți. Dacă iritarea ochilor persistă: consultați medicul. Absorbiți scurgerile de produs, pentru a nu afecta materialele din apropiere.

- Standardul conține azidă de sodiu care poate reacționa cu plumbul sau instalațiile din cupru pentru a forma potențiale azide metalice explozive. În momentul eliminării acestui standard, spălați întotdeauna cu apă din abundență pentru a preveni acumularea de azide.
- Standardul trebuie să fie imediat închis cu capacul pentru a preveni contaminarea și evaporarea.
- Pentru mai multe informații, consultați Fișa de date privind siguranța (SDS).
- Luați măsurile de precauție obișnuite și urmați buna practică de laborator.
- Utilizați doar echipamente de laborator curate sau de unică folosință pentru a evita contaminarea.

STABILITATEA

A se depozita la 2-25°C și a se proteja împotriva luminii. A nu se îngheța.

A nu se utiliza după datele de expirare indicate pe etichetele fiolelor.

Stabilitatea la bord:

Stabilitatea la bord este specifică pentru fiecare analizor.

(Consultați § DATE PRIVIND PERFORMANȚA).

PREGĂTIRE

Reactivii și Standardul sunt gata pentru utilizare.

DETERIORAREA PRODUSELOR

- Soluția de reactiv și standard trebuie să fie limpede. Aspectul tulbure indică deteriorarea.
- Nu utilizați produsul dacă există semne vizibile de deteriorare biologică, chimică sau fizică.
- Nu utilizați reactivul dacă deteriorările ambalajului ar putea avea un efect asupra performanței produsului (scurgeri, fiolă perforată).

TOTAL PROTEIN PLUS

Referințe:

PROB-0250 12 x 20 mL
 PROB-0600 2 x 125 mL
 PROB-0700 4 x 250 mL

Compoziția trusei:

R 12 x 20 mL
 R 2 x 125 mL + Std 1 x 5 mL
 R 4 x 250 mL + Std 1 x 5 mL



VTLRO-PROB-v12 (10/2020)_VTL-PROB-4-v12

PROBE (1,2,4)

Specimen

- Ser
- Plasmă heparinizată cu litiu.
- A nu se utiliza alte specimene.

☞ Avertismente și precauții

- Conform bunei practici de laborator, prelevarea trebuie efectuată înainte de administrarea de medicamente.
- Probele trebuie să fie libere din hemoliză și lipemie.

Depozitare și stabilitate

Probele sunt stabile timp de 7 zile la 2-8°C și cel puțin 2 luni la -20°C. Pentru o depozitare mai îndelungată, înghețați eșantioanele la -70°C.

VALORI DE REFERINȚĂ (1,2,4)

Ser: Pacienți în ambulatoriu Pacienți în repaus
 6,4-8,3 g/dL 6,0-7,8 g/dL
 64-83 g/L 60-78 g/L

Plasmă:

Concentrațiile plasmei sunt mărite de la 0,2 la 0,4 g/dL (de la 2 la 4 g/L), în comparație cu concentrațiile serului (fibrinogen).

Notă: Intervalul menționat ar trebui să servească doar ca un ghid. Se recomandă ca fiecare laborator să verifice acest interval sau să stabilească un interval de referință pentru populația țintă.

☞ PROCEDURĂ

Pentru Analizoarele Selectra ale ELITech Clinical Systems,

aplicațiile sunt disponibile la cerere

Lungime de undă 546 nm

Temperatură: 37°C

Citiți pe reactivul martor.

	MARTOR	CALIBRARE	TEST
Reactiv R	300 µL	300 µL	300 µL
Apă distilată	3 µL		
Calibrator		3 µL	
Eșantion			3 µL

Amestecați și citiți absorbanta (A) după o incubare de 11 minute și 30 secunde.

Cu software-ul Selectra TouchPro, utilizați aplicația inclusă în codul de bare disponibil la finalul acestui insert.

Informații importante privind setarea:

Reactivul MAGNESIUM XYLIDYL poate fi slab contaminat cu TOTAL PROTEIN PLUS pe Selectra ProM și ProXL.

Pentru a evita contaminarea pe aceste instrumente, programați următoarele incompatibilități:

Software	Meniu	Parametru
TouchPro	Incompatibilități sondă	Incompatibilitate/PROTEINA-MAGNEZIU
Altele	Incompatibilitate ace	PROTEINĂ: MAGNEZIU

☞ CALCUL

A Proba

_____ x n n = concentrație calibrator/standard

A Calibrator/

Standard

Factor de conversie: g/dL x 10 = g/L

CALIBRARE

Pentru referințele PROB-0600/PROB-0700: Pentru calibrare, trebuie utilizat calibratorul multiparametric ELICAL 2 sau Standardul proteină totală 6 g/dL.

Pentru referința PROB-0250: Pentru calibrare, utilizați calibratorul multiparametric ELICAL 2.

Valorile concentrației Standardului de proteina totala de 6 g/dL și calibratorului multiparametric ELICAL 2 sunt trasabile conform Materialului Standard de Referință 909c (al Institutului Național de Standarde și Tehnologie).

Frecvența de calibrare: Calibrarea este specifică pentru fiecare analizor. (Consultați § DATE PRIVIND PERFORMANȚA).

☞ CONTROLUL CALITĂȚII

Pentru a asigura calitatea adecvată, vor fi utilizate serurile de control precum ELITROL I și ELITROL II. Aceste controale trebuie efectuate și validate înainte ca probele pacienților să fie testate. Frecvența controlului trebuie să fie de cel puțin o dată pe zi, după fiecare calibrare și trebuie adaptată la procedurile de Controlul Calității fiecărui laborator și cerințele de reglementare. Rezultatele trebuie să fie în intervalele definite. Dacă valorile sunt în afara intervalelor definite, fiecare laborator trebuie să ia măsuri corective. Materialele pentru controlul calității trebuie utilizate conform reglementărilor locale.

MANAGEMENTUL DEȘEURILOR

Eliminarea tuturor deșeurilor trebuie să fie în conformitate cu cerințele de reglementare locale, statale și federale.

DATE DE PERFORMANȚĂ la 37°C privind Analizoarele Selectra ProM ale ELITech Clinical Systems
- Interval de măsurare

Determinat conform protocolului CLSI EP6-A⁽⁵⁾, intervalul de măsurare este între 0,20 și 12,0 g/dL (de la 2,0 la 120,0 g/L).

- Limita de detecție (LoD) și Limita de cuantificare (LoQ)

Determinată conform protocolului CLSI EP17-A⁽⁶⁾.

LoD= 0,03 g/dL (0,3 g/L).

LoQ= 0,10 g/dL (1,0 g/L).



Referințe:

PROB-0250 12 x 20 mL

PROB-0600 2 x 125 mL

PROB-0700 4 x 250 mL

Compoziția trusei:

R 12 x 20 mL

R 2 x 125 mL + Std 1 x 5 mL

R 4 x 250 mL + Std 1 x 5 mL



VTLRO-PROB-v12 (10/2020)_VTL-PROB-4-v12

- Precizie

 Determinată conform protocolului CLSI EP5-A2⁽⁷⁾.

	n	Medie		În	Total
		g/dL	g/L	interiorul ciclului	
		CV (%)			
Nivel scăzut	80	4,03	40,3	0,4	1,0
Nivel mediu	80	6,62	66,2	0,3	1,6
Nivel înalt	80	9,06	90,6	0,5	1,1

- Corelație

A fost efectuat un studiu comparativ între analizorul Selectra ProM ELITech Clinical Systems și un alt echipament al unui sistem aprobat de FDA (metoda Biuret) pe 100 probe de ser uman, conform protocolului CLSI EP9-A2⁽⁸⁾.

Concentrațiile probelor au fost între 0,27 și 11,25 g/dL (între 2,7 și 112,5 g/L).

Parametrii regresii liniare sunt după cum urmează:

Coeficient de corelație: (r)=0,997

Regresie liniară: $y=0,993x + 0,05$ g/dL
(0,5 g/L)

- Limitări și interferențe

- Nu raportați rezultatele în afara intervalului utilizabil.

- Au fost efectuate studii pentru a stabili nivelul interferenței din diferiți compuși conform protocolului CLSI EP7-A2⁽⁹⁾ al CLSL și recomandările SFBC. Recuperarea este în intervalul ±10% din valoarea inițială a concentrației proteinei totale de 4,00; 6,50 și 9,00 g/dL.

Bilirubină neconjugată: Nicio interferență semnificativă până la 30,0 mg/dL (513 μmol/L).

Bilirubină conjugată: Nicio interferență semnificativă până la 29,5 mg/dL (504 μmol/L).

Glucoză: Nicio interferență semnificativă până la 507 mg/dL (28,14 mmol/L).

Turbiditate: Nicio interferență semnificativă până la 263 mg/dL (2,97 mmol/L) echivalent trigliceride.

Hemoglobină: Nicio interferență semnificativă până la 300 mg/dL.

Dextran: Induce rezultate fals ridicate la concentrații terapeutice.

- În cazuri foarte rare, gamopatiile monoclonale (mielome multiple), în special de tipul IgM (macroglulinemia Waldenstrom) poate duce la rezultate nefiabile. ⁽¹¹⁾
- Multe alte substanțe și medicamente pot interfera. Unele dintre acestea sunt enumerate în reviste publicate de Young. ^(12,13)
- Rezultatele acestui studiu trebuie interpretate doar în corelație cu alte rezultate ale testelor de diagnosticare, constatările clinice și istoricul medical al pacientului.

- Stabilitatea la bord/Frecvența calibrării

Stabilitatea la bord: 14 zile

Frecvența calibrării: 14 zile

Recalibrați când loturile de reactiv se schimbă, când rezultatele controlului calității sunt în afara intervalului stabilit și după o operație de întreținere.

BIBLIOGRAFIE

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2. Tietz, N.W., *Clinical guide to laboratory tests*, 3rd Ed., (W.B. Saunders eds. Philadelphia USA), (1995), 518.
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8. *Method Comparison and Bias estimation Using Patient Samples; Approved Guideline - Second Edition*. CLSI (NCCLS) document EP9-A2 (2002), **22** (19).
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10. Vassault, A., et al., *Ann. Biol. Clin.*, (1986), **44**, 686.
11. Berth, M. & Delanghe, J. *Protein precipitation as a possible important pitfall in the clinical chemistry analysis of blood samples containing monoclonal immunoglobulins: 2 case reports and a review of literature*, *Acta Clin Belg.*, (2004), **59**, 263.
12. Young, D.S., *Effects of preanalytical variables on clinical laboratory tests*, 2nd Ed., AACC Press, (1997).
13. Young, D.S., *Effects of drugs on clinical laboratory tests*, 4th Ed., AACC Press, (1995).




TOTAL PROTEIN PLUS

Referințe:
PROB-0250 12 x 20 mL
PROB-0600 2 x 125 mL
PROB-0700 4 x 250 mL
Compoziția trusei:
R 12 x 20 mL
R 2 x 125 mL + Std 1 x 5 mL
R 4 x 250 mL + Std 1 x 5 mL


VTLRO-PROB-v12 (10/2020)_VTL-PROB-4-v12

SIMBOLURI

Simbolurile folosite sunt definite conform standardului ISO-15223-1 cu excepția celor prezentate mai jos.

CONT	Conținut
R	Reactiv
Std	Standard
CE	Conformitate europeană
	Modificare față de versiunea precedentă

NOTĂ IMPORTANTĂ

- Doar pentru ref. **PROB-0250**, utilizată cu software-ul Selectra TouchPro.
- Vezi **PROCEDURĂ**: Risc de contaminare


 Total Protein 0
 700 VTL-PROB


Declaration of Conformity

helena
Biosciences Europe

HL-7-0664DC DOI 2015/08 (1)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

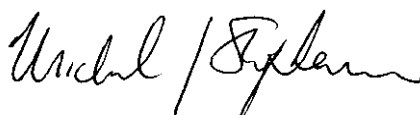
Product Code	Description	GMDN Classification Code
5267L	Thromboplastin L	55983

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

Signed:



Date: 06 Aug 2015

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Declaration of Conformity

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HL-7-0136DC DOI 2015/07 (6)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

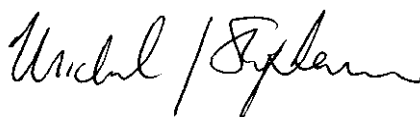
Product Code	Description	GMDN Classification Code
5185	Calibration Plasma	55995

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

Signed:



Date: 28 Jul 2015

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Declaration of Conformity

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HL-7-0512DC DOI 2015/08 (5)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

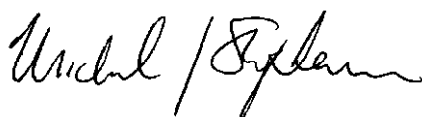
Product Code	Description	GMDN Classification Code
5556	Clauss Fibrinogen 50	55997

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

Signed:



Date: 12 Aug 2015

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HL-7-0229DC DOI 2015/08 (6)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

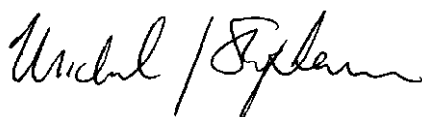
Product Code	Description	GMDN Classification Code
5392	Thrombin Time	55987

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

Signed:



Date: 06 Aug 2015

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HL-7-0137DC DOI 2015/07 (7)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

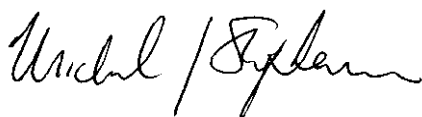
Product Code	Description	GMDN Classification Code
5186	Routine Control N	30590

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

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HL-7-0138DC DOI 2015/07 (7)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

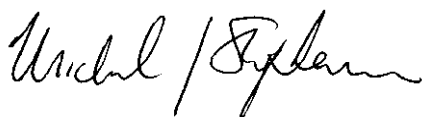
Product Code	Description	GMDN Classification Code
5187	Routine Control A	30590

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

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Declaration of Conformity

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HL-7-0135DC DOI 2015/07 (7)

In Application of the Council Directive 98/79/EC on the approximation of the laws of the Member States relating to *In Vitro* Diagnostic Medical Devices & CE marking.

Declaration of conformance to applicable sections of Annex I - Essential Requirements and Annex III (EC Declaration of Conformity) imposed by sections 2 to 5. The below listed products are not classified under Annex II Lists A or B. Access to the appropriate technical files will be made available to the appropriate body in the event this is required.

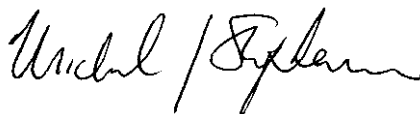
Product Code	Description	GMDN Classification Code
5183	Routine Control SA	30590

I, the undersigned declare that the devices registered against the above GMDN Classification Code conforms to the said Directives.

Full Name: M.J. Stephenson

Title: Managing Director

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Coagulation Control Plasmas

REF 5186
REF 5187
REF 5183
REF 5482

Routine Control N
Routine Control A
Routine Control SA
Routine Coagulation Control Set

Helena

Biosciences Europe



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Coagulation Control Plasmas

en

INTENDED PURPOSE

The Coagulation Control Plasmas kit is intended for use as a quality control material.
Routine Control N, Routine Control A and Routine Control SA are for use as normal, moderately prolonged and markedly prolonged controls for PT and aPTT assays. They are also assayed for Fibrinogen, TCT and ATIII, and are prepared from normal human plasma.

WARNINGS AND PRECAUTIONS

The reagents contained in this kit are for *in vitro* diagnostic use only. – DO NOT INGEST. Wear appropriate personal protective equipment when handling all kit components. Refer to the product safety declaration for the link to appropriate hazard and precautionary statements where applicable. Dispose of components in accordance with local regulations.
Blood products have been screened and found negative (unless otherwise stated on the kit box or vial) for the presence of: Hepatitis B Antigen (HbsAg) Hepatitis C Antibody (HCV Ab) HIV 2 antibody
However they should be handled with the same precautions as a human patient sample.

COMPOSITION

REF	Component	Content	Description
5186	Routine Control - N	10 x 1 mL	Prepared from pooled normal plasma.
5187	Routine Control - A	10 x 1 mL	Prepared from ascended human plasma.
5183	Routine Control - SA	10 x 1 mL	Prepared from ascended human plasma.
5482	Routine Coagulation Control Set:		
	Routine Control - N	4 x 1 mL	
	Routine Control - A	3 x 1 mL	
	Routine Control - SA	3 x 1 mL	

Each kit contains instructions for use.

Each kit contains 1 mL of buffered, lyophilised human plasma.
Reprepare the exact vial of the appropriate control with 1 mL of distilled or deionised water. Swirl gently. Allow to stand for 10 minutes for complete dissolution and mix well before use.

ITEMS REQUIRED BUT NOT PROVIDED

Coagulation Control Plasmas may be used when performing tests on any mechanical or photo-optical coagulation instrument in conjunction with suitable commercial reagents.

STORAGE SHELF-LIFE AND STABILITY

Unopened vials are stable until the given expiry date when stored under conditions indicated on the vial or kit label. The reconstituted controls are stable for 6 hours when kept at 2–8°C or 4 weeks at -20°C when flash frozen. Keep covered.

SAMPLE COLLECTION AND PREPARATION

Not applicable.

PROCEDURE

Each control should be treated in the same manner as the unknown specimen in accordance with the instructions outlined in each particular test protocol.

INTERPRETATION OF RESULTS

Routine Control N should give values within the laboratory normal range for PT, aPTT and fibrinogen assays. Routine Control A and Routine Control SA have been standardised to give prolonged and markedly prolonged PT and aPTT times respectively. Lot and Routine specific expected values are provided with each pack of controls.

LIMITATIONS

The results obtained with Coagulation Control Plasmas depend on several factors strongly associated with instrumentation. Types of reagent, diluent substrates and laboratory to laboratory variations. Each laboratory should establish an expected range for the particular instrument(s) used.

QUALITY CONTROL

Each laboratory should establish a quality control program. Normal and abnormal control plasmas should be tested prior to each batch of patient samples, to ensure satisfactory instrument and operator performance. If controls do not perform as expected, patient results should be considered invalid.

REFERENCE VALUES

Reference values can vary between laboratories depending on the techniques and systems in use. For this reason each laboratory should establish its own reference ranges.

PERFORMANCE CHARACTERISTICS

The following performance characteristics have been determined by Helena Biosciences Europe or their representatives using an opto-mechanical coagulation instrument. Each laboratory should establish its own performance data.

Reproducibility

Sample	n	Intra-assay precision	PT CV (%)	aPTT CV (%)
Routine Control N	5	2.83	1.01	1.01
Routine Control A	5	2.76	1.71	1.71
Routine Control SA	5	1.72	1.03	1.03

BIBLIOGRAPHY

- Kirkwood TBL, et al. (1977) Identification of Sources of Variation in Factor VIII Assay. *British Journal of Haematology*, 37:555-568.
- Goldaniga MD (1971) Reproducibility in Coagulation Assays. *AJCP* 55:561-564.
- Pallitt HA and Longbery JR (1979) A Precision Study of Coagulation Factor Assay Techniques. *AJCP* 59:231-235.

Plasmas de contrôle de coagulation

Fiche technique

fr

UTILISATION

Le kit Coagulation Control Plasmas est destiné à être utilisé comme produit de contrôle qualité.

Les contrôles Routine Control N, Routine Control A et Routine Control SA servent de témoins, normal, modérément prolongés et nettement prolongé dans les déterminations du PT et du TCA, Le fibrinogène, le TCT et l'ATIII ont été dosés et ils sont préparés à partir de plasma humain normal.

AVERTISSEMENTS ET PRECAUTIONS

Les réactifs du kit sont à usage diagnostique *in vitro* uniquement. – NE PAS INGESTER. Porter un équipement de protection individuelle appropriée lors de la manipulation de tous les composants du kit. Consulter la fiche de données de sécurité du produit pour obtenir les précautions de sécurité et les consignes de production et de stockage. Eliminer les déchets conformément aux réglementations locales.

Un déposé des produits sanguins a été réalisé et a donné un résultat négatif (sauf indication contraire sur la boîte du kit) ou sur le sérum quant à la présence de: Hépatite B Antigène (HbsAg) Hépatite C Anticorps anti-HCV Hépatite A Anticorps anti-VIH-2 Anticorps anti-VH-2
Cependant, ils doivent être manipulés avec les mêmes précautions que celles prises pour les échantillons patients humains.

COMPOSITION

REF	Composant	Contient	Description
5186	Routine Control - N	10 x 1 mL	Préparé à partir d'un pool de plasma normal.
5187	Routine Control - A	10 x 1 mL	Préparé à partir de plasma humain ascorbé.
5183	Routine Control - SA	10 x 1 mL	Préparées à partir de plasma humain ascorbé.
5482	Routine Coagulation Control Set:		
	Routine Control - N	4 x 1 mL	
	Routine Control - A	3 x 1 mL	
	Routine Control - SA	3 x 1 mL	

Chaque kit contient une fiche technique.

Chaque kit contient valeurs de référence spécifiques du lot.

Chaque flacon contient 1 mL de plasma humain tamponné lyophilisé.

Préparation Reconstituer chaque flacon du contrôle approprié avec 1 mL d'eau distillée ou déionisée. Agiter doucement. Attendre 10 minutes jusqu'à dissolution totale et bien mélanger avant d'utiliser.
MATÉRIEL NECESSAIRE NON FOURNI

Le Coagulation Control Plasmas peut être utilisé dans les analyses réalisées sur des instruments de coagulation mécanique ou photo-optiques avec les réactifs appropriés vendus dans le commerce.

CONSERVATION, DURÉE DE VIE UTILE ET STABILITÉ

Les flacons non ouverts sont stables jusqu'à la date de péremption indiquée s'ils sont conservés dans les conditions indiquées sur l'étiquette du kit ou du flacon. Une fois reconstruits, les contrôles sont stables 6 heures entre 2–8°C ou 4 semaines à -20°C en cas de congélation instantanée. Couvrir le produit.
PRÉLEVEMENT ET PRÉPARATION DES ÉCHANTILLONS

Non applicable.

PROCÉDURE

Chaque contrôle doit être traité de la même manière que l'échantillon à analyser en observant les instructions de chaque protocole spécifique.

INTERPRÉTATION DES RÉSULTATS

Le Routine Control N doit donner des valeurs se situant dans la plage normale du laboratoire pour le PT, le TCA et le Fibrinogène. Le Routine Control A et le Routine Control SA ont été standardisés pour donner des temps PT et TCA prolongés et très prolongés respectivement. Les valeurs prévues spécifiques du kit de l'instrument sur lequel le contrôle est utilisé doivent être prises en compte.

LIMITES

Les résultats obtenus avec le Coagulation Control Plasmas dépendent de plusieurs facteurs fortement corrélés avec l'instrument, le réactif, les substrats et les variations inter-laboratoires. Chaque laboratoire doit déterminer avec précaution pour chaque système instrument-réactif. Le laboratoire doit déterminer les plages prévues.

CONTRÔLE QUALITÉ

Chaque laboratoire doit établir un programme de contrôle qualité. Les plasmas de contrôle, normale et anormale, doivent être testés avant chaque lot de réactifs patients afin de s'assurer que l'instrument et l'opérateur offrent des performances satisfaisantes. Si les contrôles ne donnent pas les résultats prévus, les résultats du patient doivent être considérés comme non valides.

VALEURS DE RÉFÉRENCE

Les valeurs de référence peuvent varier d'un laboratoire à l'autre suivant les techniques et les systèmes utilisés. C'est pour cette raison qu'il appartient à chaque laboratoire de déterminer ses propres plages de référence.

CARACTÉRISTIQUES DE PERFORMANCES

Helena Biosciences Europe ou ses mandataires ont déterminé les caractéristiques de performance suivantes en utilisant un instrument de coagulation opto-mécanique. Chaque laboratoire doit établir ses propres données de performance.

Reproductibilité

Échantillon	n	Precision Intra-assay	PT CV (%)	TCA CV (%)
Routine Control N	5	2.83	1.01	1.01
Routine Control A	5	2.76	1.71	1.71
Routine Control SA	5	1.72	1.03	1.03

BIBLIOGRAPHIE

- Kirkwood TBL, et al. (1977) Identification of Sources of Variation in Factor VIII Assay. *British Journal of Haematology*, 37:555-568.
- Goldaniga MD (1971) Reproducibility in Coagulation Assays. *AJCP* 55:561-564.
- Pallitt HA and Longbery JR (1979) A Precision Study of Coagulation Factor Assay Techniques. *AJCP* 59:231-235.

Kontrollplasma für die Gerinnung

Anleitung

de

VERWENDUNGSWECK

Das Coagulation Control Plasma-Kit ist für die Qualitätskontrolle vorgesehen.

Routine Control N, Routine Control A und Routine Control SA sind als normale, mäßig verzögerte und stark verzögerte Kontrollen für PT und aPTT Tests geeignet. Sie sind auch auf Fibrinogen, TZ und ATIII getestet und werden aus normalem Humanplasma hergestellt.

WARNUNGSWEISE UND VORSICHTSMASSNAHMEN

Die in diesem Kit enthaltenen Reagenzien sind ausschließlich für die Verwendung von *in vitro*-Diagnose vorgesehen. NICHT NENNEN, KEIN VERWENDEN, KEIN VERKÄUFEN. Nur für diagnostische Zwecke verwenden. Handschuhe tragen und die Augen schützen. Bei Verdacht auf Exposition für PT und aPTT Tests geeignet. Sie sind auch auf Fibrinogen, TZ und ATIII getestet und werden aus normalem Humanplasma hergestellt.

Die Blutprodukte wurden untersucht und sind für folgende Gene ohne Befund (soweit nicht anderweitig auf der Verpackung oder den Reagenzienfläschchen angegeben): Hepatitis-B-Antikörper (HbsAg) Hepatitis-C-Antikörper (HCV-Ab) HIV-Antikörper 1 HIV-Antikörper 2

Sie sind jedoch mit den gleichen Vorkehrungen zu behandeln wie Proben von menschlichen Patienten.
ZUSAMMENSETZUNG

REF	Komponente	Inhalt	Beschreibung
5186	Routine Control - N	10 x 1 mL	Aus gepooltem Humanplasma hergestellt.
5187	Routine Control - A	10 x 1 mL	Ascorbiertem Humanplasma hergestellt.
5183	Routine Control - SA	10 x 1 mL	Ascorbiertem Humanplasma hergestellt.
5482	Routine Coagulation Control Set:		
	Routine Control - N	4 x 1 mL	
	Routine Control - A	3 x 1 mL	
	Routine Control - SA	3 x 1 mL	

Jedes Kit enthält eine Gebrauchsanweisung.

Jedes Kit enthält eine Gebrauchsanweisung.

Jedes Fläschchen enthält 1 mL gepuffertes, lyophilisiertes Humanplasma.
Vorbereitung Reconstituieren Sie jedes Fläschchen des Kontrollplasma mit 1 mL destilliertem oder entionisiertem Wasser, rekonsitulieren, Licht schützen und zum vollständigen Auflösen 10 Minuten stehen lassen und vor Gebrauch gut mischen.
LAGERUNG, HALTBARKEIT UND STABILITÄT

Coagulation Control Plasmas kann in Verbindung mit allen entsprechenden kommerziellen Reagenzien bei der Durchführung von Tests an mechanischen oder lichtoptischen Koagulometern verwendet werden.

ERFORDERLICHE, ABER NICHT MITGELIEFERTE ARTIKEL

Ungeöffnete Flaschen sind unter dem auf der Verpackung oder Fläschchen angegebenen Lagerbedingungen bis zum aufgedruckten Verfallsdatum stabil. Einmal wiederhergestellt, um verteilte oder stark verteilte PT und aPTT Zeiten zu ergeben. Chargen und Geräte spezifizieren Normwerte sind in jeder Packung mit Kontrollen enthalten.

PROBENTAHME UND VORBEREITUNG

ENTFALT

VORGEHENSWEISE

Jedes Kontrolle sollte gemäß den Anleitungen der einzelnen Testprotokolle wie unbekannte Probe behandelt werden.

INTERPRÉTATION DER ERGEBNISSE

Routine Control N sollte für PT, aPTT und Fibrinogen Tests Werte im Normalbereich ergeben. Routine Control A und Routine Control SA wurden standardisiert, um verteilte oder stark verteilte PT und aPTT Zeiten zu ergeben. Chargen und Geräte spezifizieren Normwerte sind in jeder Packung mit Kontrollen enthalten.

ENSICHERUNGSGEBEN

Die mit Coagulation Control Plasmas erhaltenen Resultate hängen von mehreren Faktoren ab, die stark mit dem Gerät, dem verwendeten Reagenzien, möglichen Substraten und Unterschieden zwischen den Labors in Verbindung stehen.
sollte daher für jedes Geräte-Reagenzien-System einen eigenen Normalwertbereich erstellen.

QUALITÄTSKONTROLLE

Jedes Labor muss für eine eigene Qualitätskontrolle sorgen. Normale und pathologische Kontrollplasmas müssen vor jeder Analyse getestet werden.
Die Kontrollen außerhalb des Normbereichs, sind die Patienten Ergebnisse nicht zu verwenden.

16/02/2021

LETTER OF DISTRIBUTION

To Whom It May Concern,

This letter is to serve notice that GBG-MLD SRL Global Biomarketing Group, located at str. Tighina, 65, of. 607 MD2001 Chisinau, Municipiul Chisinau, Moldova is authorized to distribute the Helena Biosciences Europe Haemostasis and Electrophoresis range of products in the whole Republic of Moldova territory. As such, GBG-MLD SRL Global Biomarketing Group is responsible for promotion, support, installation, and after-sales service for Helena Biosciences Europe products.

GBG-MLD SRL Global Biomarketing Group will maintain appropriate, up-to-date and accurate records to enable the immediate recall of any Products or batches of Products. These records shall include records of deliveries to customers (including batch numbers, expiry dates, delivery date, name and address of customer, telephone number, fax number and e-mail address). These records should be kept for a minimum of one year past the expiry of the product that has been delivered. This agreement is effective for a period of 3 years from the date of this letter, unless terminated by either party by giving 90 days notice, and can be extended through the mutual agreement of both parties based on sales performance.

Sincerely,

Dmitri Alexandrov

International Business Manager





CERTIFICATE

EC No 1434-IVDD-134/2019
Full Quality Assurance System

Directive 98/79/EC on in vitro diagnostic medical devices

Polish Centre for Testing and Certification certifies
that the quality assurance system in the organization:

Lorne Laboratories Ltd

**Unit 1 Cutbush Park Industrial Estate, Danehill,
Lower Earley, Berkshire RG6 4UT, United Kingdom**

for the design, manufacture and final inspection of in vitro diagnostic medical devices
List A

Products list in attachments: 1

complies with requirements of Annex IV excluding section 4 and 6 to Directive 98/79/EC (as amended)
implemented into Polish law, as evidenced by the audit conducted by the PCBC.

Validity of Certificate: from 10.04.2019 to 23.05.2023

The date of issue of the Certificate: 10.04.2019

The date of the first issue of the Certificate: 10.04.2019



Application No: 649/2019
Module: H7


mgr Anna Wyroba
Vice-President



Certificate No **1434-IVDD-134/2019**
Issued under the Contract No **MD-59/2019**
Bears the PCBC hologram.
Warsaw, 10.04.2019



ANNEX 1 TO CERTIFICATE
VALID ONLY WITH CERTIFICATE
No 1434-IVDD-134/2019

The products detailed below are covered under the scope of this certificate:

Name:	GMDN code:
Anti-A Monoclonal, 600010	52532
Anti-B Monoclonal, 610010	52538
Anti-A,B Monoclonal, 620010	46442
Anti-D Clone 1 Monoclonal, 730010	52647
Anti-D Clone 2 Monoclonal, 710010	52647
Anti-D Duoclone Monoclonal, 740010	52647
Anti-C Monoclonal, 690005	52546
Anti-E Monoclonal, 691005	52562
Anti-c Monoclonal, 692005	52547
Anti-e Monoclonal, 693005	52563
Anti-C+D+E Monoclonal, 700010	52550
Anti-K Monoclonal, 760010	52593




mgr Anna Wyroba
Vice-President



Annex 1 to certificate No. **1434-IVDD-134/2019**
Issued under the Contract No. **MD-59/2019**
Bears the PCBC hologram.
Warsaw, 10.04.2019

EC DECLARATION OF CONFORMITY

Lorne Laboratories Ltd declares that the following in vitro diagnostic reagent:

Product name	Catalogue number
ASO Latex kit	031100A

has been classified as non List A, non List B (Directive 98/79/EC, Annex II) and complies with the essential requirements and provisions of Directive 98/79/EC of the European Parliament and of the Council (also SI 2002 No.618 which transposes the requirements of Directive 98/79/EC).

and is in conformity with the national standards transposing harmonised standards:

- BS EN 980:2008
- BS EN ISO 13485:2012
- BS EN 13612:2002
- BS EN 13640:2002
- BS EN 13641:2002
- BS EN ISO 14971:2012
- BS EN ISO 18113, parts 1&2

The conformity assessment procedure performed was in accordance with Annex III of Directive 98/79/EC.

This declaration of conformity is issued under the sole responsibility of Lorne Laboratories Ltd and is valid from 13 April 2016.



Eddy Velthuis
Technical Director

EC DECLARATION OF CONFORMITY

Lorne Laboratories Ltd declares that the following in vitro diagnostic reagent:

Product name	Catalogue number
RF Latex kit	830100A

has been classified as non List A, non List B (Directive 98/79/EC, Annex II) and complies with the essential requirements and provisions of Directive 98/79/EC of the European Parliament and of the Council (also SI 2002 No.618 which transposes the requirements of Directive 98/79/EC).

and is in conformity with the national standards transposing harmonised standards:

- BS EN 980:2008
- BS EN ISO 13485:2012
- BS EN 13612:2002
- BS EN 13640:2002
- BS EN 13641:2002
- BS EN ISO 14971:2012
- BS EN ISO 18113, parts 1&2

The conformity assessment procedure performed was in accordance with Annex III of Directive 98/79/EC.

This declaration of conformity is issued under the sole responsibility of Lorne Laboratories Ltd and is valid from 13 April 2016.



Eddy Velthuis
Technical Director

EC DECLARATION OF CONFORMITY

Lorne Laboratories Ltd declares that the following in vitro diagnostic reagent:

Product name	Catalogue number
CRP Latex kit	850100A

has been classified as non List A, non List B (Directive 98/79/EC, Annex II) and complies with the essential requirements and provisions of Directive 98/79/EC of the European Parliament and of the Council (also SI 2002 No.618 which transposes the requirements of Directive 98/79/EC).

and is in conformity with the national standards transposing harmonised standards:

- BS EN 980:2008
- BS EN ISO 13485:2012
- BS EN 13612:2002
- BS EN 13640:2002
- BS EN 13641:2002
- BS EN ISO 14971:2012
- BS EN ISO 18113, parts 1&2

The conformity assessment procedure performed was in accordance with Annex III of Directive 98/79/EC.

This declaration of conformity is issued under the sole responsibility of Lorne Laboratories Ltd and is valid from 13 April 2016.



Eddy Velthuis
Technical Director



RAPID LATEX KIT
DIRECTIONS FOR USE

ASO Latex Kit: For Detection Of Anti-Streptolysin O (ASO) In Serum

SUMMARY

In acute streptococcal infections, the toxic immunogenic exoenzyme streptolysin O (ASO) is produced in response to streptolysin O antigens liberated by haemolytic streptococci of groups A, C and G. Information on extent and degree of infection can be obtained by measuring serum ASO levels. Elevated ASO levels have also been found in patients suffering from scarlet fever, acute rheumatoid arthritis, tonsillitis, and other streptococcal infections as well as in healthy carriers.

INTENDED PURPOSE

The reagent is a latex test reagent intended to be used to qualitatively and semi-quantitatively determine the presence or absence of Anti-Streptolysin O antibodies in the serum or plasma of patients when tested in accordance with the recommended techniques stated in this IFU.

PRINCIPLE

When used by the recommended techniques, latex particles in the reagent will agglutinate (clump) in the presence of anti-streptolysin O antibodies. No agglutination generally indicates the absence of anti-streptolysin O antibodies (see **Limitations**).

KIT DESCRIPTION

Lorne ASO Latex Kit is a serologic test for the detection of ASO antibodies. The reagents do not contain or consist of CMR substances, or endocrine disrupting substances or that could result in sensitisation or an allergic reaction by the user. All the reagents are supplied at optimal dilution for use with all recommended techniques without the need for further dilution or addition. For lot reference number and expiry date see **Vial Labels**.

STORAGE

Do not freeze. Reagent vials should be stored at 2 - 8°C on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity.

SPECIMEN COLLECTION

Specimens should be drawn with or without anticoagulant using an aseptic phlebotomy technique. If testing is delayed specimens can be stored at 2-8°C for 7 days or for up to 3 months at or below -20°C. Specimens must be free from bacterial contamination, fibrin, gross lipemia and gross haemolysis.

PRECAUTIONS

1. The kit is for *in vitro* diagnostic use only.
2. Do not use kit past expiration date (see **Vial and Box Labels**).
3. Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
4. The reagents in this kit have been processed to reduce the bio-burden, but are not supplied sterile. Once a vial has been opened the contents should remain viable up until the expiry date.
5. Materials used to produce the kit were tested at source and found to be negative for HIV 1+2 and HBsAg using approved microbiological tests. However no known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF KIT REAGENT AND DEALING WITH SPILLAGES

For information on disposal of kit reagent and decontamination of a spillage site see **Material Safety Data Sheets**, available on request.

CONTROLS AND ADVICE

1. ASO Positive and Negative Controls must be tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.

2. All the reagents must be allowed to reach 18-25°C before use.
3. Shake the reagents well before use to ensure homogeneity.
4. Do not interchange components between different kits.
5. Use of kit and interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of the country where the kit is in use.
6. The user must determine the suitability of the kit for use in other techniques.

KIT COMPONENTS SUPPLIED

- 1) ASO Latex Reagent (White cap, 5.0 mL): Latex particles coated with streptolysin O, pH, 8.2 containing a preservative.
- 2) ASO Positive Control (Red cap, 1.0 mL Red cap): Human serum with an ASO concentration > 200 IU/mL containing a preservative.
- 3) ASO Negative Control (Blue cap, 1.0 mL): Animal serum containing a preservative.
- 4) Pipette-Stirrers.
- 5) Disposable agglutination Slides.

REAGENTS AND MATERIALS REQUIRED BUT NOT SUPPLIED

- a) Small glass / plastic test tubes.
- b) Serological pipettes.
- c) Graduated container.
- d) 9 g/L saline solution.

RECOMMENDED QUALITATIVE TECHNIQUE

1. Allow the reagents and samples to reach room temperature. The sensitivity of the test may be reduced at low temperatures.
2. Place 50 µL of the sample and one drop of each Positive and Negative controls into separate circles on the slide test.
3. Swirl the ASO-latex reagent gently before using and add one drop (50 µL) next to the sample to be tested.
4. Mix the drops with a stirrer, spreading them over the entire surface of the circle. Use different stirrers for each sample.
5. Swirl the slide gently and after 2 minutes read the results macroscopically. False positive results could appear if the test is read after more than two minutes.

INTERPRETATION OF QUALITATIVE RESULTS

1. **Positive:** Agglutination of latex particles constitutes a positive result and within the accepted limitations of the test procedure, indicates a level of ASO in the specimen > 200 I.U/ml.
2. **Negative:** No visible agglutination of latex particles constitutes a negative result and within the accepted limitations of the test procedure, indicates a level of ASO in specimen < 200 I.U/ml.

RECOMMENDED SEMI-QUANTITATIVE TECHNIQUE

1. The semi-quantitative test can be performed in the same way as the quantitative technique using dilutions of the serum in 9 g/L saline solution.
2. Make doubling dilutions of specimen as follows:

Dilution	Serum	Saline
1/2	100 µl undiluted serum	100 µl
1/4	100 µl 1/2 diluted serum	100 µl
1/8	100 µl 1/4 diluted serum	100 µl

3. Test the specimen dilutions in the same way as for the quantitative technique above.
4. Agglutination of the sera indicates:

Dilution	ASO Levels (I.U. / ml)
1/2	400 (200 x 2)
1/4	800 (200 x 4)
1/8	1600 (200 x 8)

5. Normal levels of ASO in adults is < 200 I.U/ml.

RESULTS

The titre is expressed as the reciprocal of the highest dilution showing macroscopic agglutination, e.g. if this occurs in the 1/8 dilution, the titre is 1600.

INTERPRETATION OF SEMI-QUANTITATIVE RESULTS

Positive results may indicate an acute streptococcal infection. In which case the test should be repeated at weekly intervals to determine the progression of infection.

STABILITY OF THE REACTIONS

Slide tests should be interpreted immediately after the 2-minute period to avoid the possibility that a negative result may be incorrectly interpreted as positive due to drying of the reagent.

LIMITATIONS

1. False positive results may be obtained in conditions such as scarlet fever, acute rheumatoid arthritis, tonsillitis and other streptococcal infections as well as in healthy carriers.
2. Hemoglobin (≤ 10 g/L), bilirubin (≤ 20 mg/dL), lipemia (≤ 10 g/L), rheumatoid factors (≤ 300 IU/mL) do not interfere. Other substances may interfere⁷.
3. Early infections in children from 6 months to 5 years may cause false negative results.
4. A single ASO determination does not produce much information about the actual state of the disease. Titrations at biweekly intervals during 4 or 6 weeks are advisable to follow the disease evolution.
5. Clinical diagnosis should not be made on findings of a single test result, but should integrate both clinical and laboratory data.
6. False positive or false negative results may also occur due to:
 - a) Contamination of test materials
 - b) Improper storage of test materials or omission of reagents
 - c) Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS

1. The kit has been characterised by all the procedures mentioned in the **Recommended Techniques**.
2. Prior to release, each lot of Lorne ASO Latex Kit is tested by **Recommended Techniques** to ensure suitable reactivity.
3. The ASO-latex sensitivity is calibrated against the WHO 1st International Standard for ASO available from NIBSC.
4. Analytical sensitivity: 200 (± 50) IU/mL, under the described assay conditions.
5. Prozone effect: No prozone effect was detected up to 1500 IU/mL.
6. Diagnostic sensitivity: 98 %.
7. Diagnostic specificity: 97 %.

DISCLAIMER

1. The user is responsible for the performance of the kit by any method other than those mentioned in the **Recommended Techniques**.
2. Any deviations should be validated prior to use using established laboratory procedures.

BIBLIOGRAPHY

1. David S.Jacobs et al. Laboratory Test Handbook, 3rd edition, Lexi-Comp Inc, 1994.

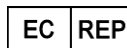
AVAILABLE KIT SIZES

Kit Size	Catalogue Number
100 Tests Per Kit	031100A



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RAPID LATEX KIT
DIRECTIONS FOR USE

RF Latex kit: For Detection Of Rheumatoid Factor (RF).

SUMMARY

Rheumatoid factors are a group of antibodies directed to determinants in the Fc portion of the IgG molecule. Although rheumatoid factors are found in a number of rheumatoid disorders, such as systemic lupus erythematosus (SLE) and Sjögren's syndrome, as well as in non-rheumatoid conditions, its central role lies in aiding in the diagnosis of rheumatoid arthritis.

INTENDED PURPOSE

The reagent is a latex test reagent intended to be used to qualitatively and semi-quantitatively determine the presence or absence of Rheumatoid Factors in the serum or plasma of patients when tested in accordance with the recommended techniques stated in this IFU.

PRINCIPLE

When used by recommended techniques, latex particles in the reagent will agglutinate (clump) in presence of rheumatoid factor (RF). No agglutination (no clumping) generally indicates absence of RF (see **Limitations**).

KIT DESCRIPTION

Lorne RF Latex Kit is for the detection of rheumatoid factor. The latex reagent is a suspension of polystyrene latex particles coated with human gamma globulins, which agglutinate in the presence of Rheumatoid Factor (RF). The reagents do not contain or consist of CMR substances, or endocrine disrupting substances or that could result in sensitisation or an allergic reaction by the user. All latex reagents are supplied at optimal dilution for use with all recommended techniques without the need for further dilution or addition. For lot reference number and expiry date see **Vial Labels**.

STORAGE

Do not freeze. Reagent vials should be stored at 2 - 8°C on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity.

SPECIMEN COLLECTION

Specimens should be drawn without anticoagulant using an aseptic phlebotomy technique. If testing is delayed, fresh serum can be stored at 2-8°C for 7 days or for up to 3 months at or below -20°C. Specimens must be free from bacterial contamination, fibrin, gross leucemia and gross haemolysis.

PRECAUTIONS

1. The kit is for *in vitro* diagnostic use only.
2. Do not use kit past expiration date (see **Vial and Box Labels**).
3. Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
4. All the reagents must be allowed to reach 18-25°C before use.
5. The reagents in this kit have been processed to reduce the bio-burden, but are not supplied sterile. Once a vial has been opened the contents should remain viable up until the expiry date.
6. Materials used to produce the kit were tested at source and found to be negative for HIV 1+2 and HBsAg using approved microbiological tests. However, no known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF KIT REAGENT AND DEALING WITH SPILLAGES

For information on disposal of kit reagent and decontamination of a spillage site see **Material Safety Data Sheets**, available on request.

CONTROLS AND ADVICE

1. It is recommended that the RF Positive and Negative Controls are tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.
2. All the reagents must be allowed to reach 18-25°C before use.
3. Shake the reagents well before use to ensure homogeneity.
4. Do not interchange components between different kits.
5. Use of kit and interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of country where kit is in use. The user must determine the suitability of the kit for use in other techniques.
6. Results obtained with a latex method do not compare with those obtained with the Rose Waaler test. Differences in the results between methods do not reflect differences in the ability to detect rheumatoid factors.

KIT COMPONENTS SUPPLIED

- 1) RF Latex Reagent (5 mL): Latex particles coated with human γ -globulin, pH, 8.2, and a preservative.
- 2) RF Positive Control (Red cap, 1 mL): Human serum with a RF concentration > 30 IU/mL and a preservative.
- 3) RF Negative Control (Blue cap, 1 mL): Animal serum and a preservative.
- 4) Pipette-Stirrers.
- 5) Reusable Agglutination Slide (18 each).

MATERIALS AND EQUIPMENT REQUIRED BUT NOT SUPPLIED

- 1) Glass Test Tubes (10 x 75 mm or 12 x 75 mm).
- 2) Pasteur and Graduated Pipettes.
- 3) Vortex mixer.
- 4) Mechanical rotator with adjustable speed of 80-100 rpm.

RECOMMENDED QUALITATIVE TECHNIQUE

1. Allow the reagents and samples to reach room temperature. The sensitivity of the test may be reduced at low temperatures.
2. Place 50 μ L of the sample and one drop of each Positive and Negative controls into separate circles on the slide test.
3. Shake the RF-latex reagent vigorously or use a vortex mixer before use and add one drop (50 μ L) next to the sample to be tested.
4. Mix the drops with a stirrer, spreading them over the entire surface of the circle. Use different stirrers for each sample.
5. Place the slide on a rotary shaker at 80-100 r.p.m. for 2 minutes. False positive results could appear if the test is read after more than two minutes.

INTERPRETATION OF QUALITATIVE RESULTS

1. **Positive:** Visible agglutination of latex particles constitutes a positive result and within the accepted limitations of the test procedure, indicates a level of RF in the specimen > 8 IU/ml.
2. **Negative:** No visible agglutination of latex particles in a milky liquid constitutes negative result and within accepted limitations of test procedure, indicates level of < 8 IU/ml RF in specimen.

RECOMMENDED SEMI-QUANTITATIVE TECHNIQUE

1. The semi-quantitative test can be performed in the same way as the qualitative test using dilutions of the serum.
2. Make doubling dilutions of serum specimen in 9 g/L saline as follows:

Dilution	Serum	Saline
1/2	100 μ l undiluted serum	100 μ l
1/4	100 μ l 1/2 diluted serum	100 μ l
1/8	100 μ l 1/4 diluted serum	100 μ l
1/16	100 μ l 1/8 diluted serum	100 μ l

3. Test the specimen dilutions in the same way as for the quantitative technique above.
4. Agglutination of the sera indicates:

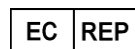
Dilution	RF Levels (I.U/mL)
1/2	16 (8 x 2)
1/4	32 (8 x 4)
1/8	64 (8 x 8)
1/16	128 (8 x 16)

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E-mail: info@lornelabs.com

5. Normal levels of RF in adults is < 8 IU/mL

RESULTS

The titre is expressed as the reciprocal of the highest dilution showing macroscopic agglutination: e.g. if this occurs in dilution 1/8, the titre is (8 x 8 IU/mL) = 64 IU/mL.



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Tower Street, Swatar, BKR 4013, Malta

STABILITY OF THE REACTIONS

Slide tests should be interpreted straight after the 2-minute rotation period to avoid the possibility that a negative result may be incorrectly interpreted as positive due to drying of the reagent.

LIMITATIONS

- Using a latex test system, positive results are not always found with every case of clinically defined rheumatoid arthritis, the number of positives reported using various types of latex reagent range from 70% to over 90%.
- The incidence of false positive results is about 3-5%. Individuals suffering from infectious mononucleosis, hepatitis, syphilis as well as elderly people may give positive results.
- Diagnosis should not be solely based on the results of latex method but also should be complemented with a Rose Waaler test along with the clinical examination.
- Haemoglobin (≤ 10 g/L), bilirubin (≤ 20 mg/dL) and lipaemia (≤ 10 g/L) do not interfere. Other substances may interfere⁹.
- False positive or false negative results may also occur due to:
 - Contamination of test materials
 - Improper storage of test materials or omission of reagents
 - Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS

- The kit has been characterised by all the procedures mentioned in the **Recommended Techniques**.
 - The RF latex sensitivity is calibrated against the RF International Calibrator from the WHO (WHO 64/2 Rheumatoid Arthritis Serum).
 - Analytical sensitivity: 8 (6-16) IU/mL, under the described assay conditions.
 - Prozone effect: No prozone effect was detected up to 1500 IU/mL.
 - Diagnostic sensitivity: 100 %.
 - Diagnostic specificity: 100 %.
- The diagnostic sensitivity and specificity have been obtained using 139 samples compared with the same method of a competitor.

DISCLAIMER

- The user is responsible for the performance of the kit by any method other than those mentioned in the **Recommended Techniques**.
- Any deviations should be validated prior to use using established laboratory procedures.

BIBLIOGRAPHY

- David S.Jacobs et al. Laboratory Test Handbook, 3rd edition, Lexi-Comp Inc, 1994.

AVAILABLE KIT SIZES

Kit Size	Catalogue Number
100 Tests Per Kit	830100A





RAPID LATEX KIT
DIRECTIONS FOR USE

CRP Latex Kit: For Detection Of C-Reactive Protein (CRP) In Serum.

SUMMARY

C-Reactive Protein (CRP) usually appears in serum of individuals in response to inflammatory conditions and tissue necrosis and disappears when causative conditions subside. It is routinely found in cases of bacterial infection, active rheumatic fever and many malignant diseases and is often seen in association with rheumatoid arthritis, viral infections and tuberculosis. CRP has also been detected in patients following blood transfusions and surgical operations as well as in patients with burns, pemphigus vulgaris and other bullous lesions.

INTENDED PURPOSE

The reagent is a latex test reagent intended to be used to qualitatively and semi-quantitatively determine the presence or absence of CRP in the serum or plasma of patients when tested in accordance with the recommended techniques stated in this IFU.

PRINCIPLE

When used by the recommended techniques, latex particles in the reagent will agglutinate (clump) in the presence of CRP. No agglutination (no clumping) generally indicates absence of CRP (see **Limitations**).

KIT DESCRIPTION

Lorne CRP Latex Test Kit is for the detection of CRP. Test reagent consists of latex particles coated with rabbit Anti-CRP (IgG). The reagents do not contain or consist of CMR substances, or endocrine disrupting substances or that could result in sensitisation or an allergic reaction by the user. All the latex reagents are supplied at optimal dilution for use with all recommended techniques without need for further dilution or addition. For lot reference number and expiry date see **Vial Labels**.

STORAGE

Do not freeze. Reagent vials should be stored at 2 - 8°C on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity.

SPECIMEN COLLECTION

Specimens should be drawn with or without anticoagulant using an aseptic phlebotomy technique. If testing is delayed specimens can be stored at 2-8°C for 7 days or for up to 3 months at or below -20°C. Specimens must be free from bacterial contamination, fibrin, gross lipaemia and gross haemolysis.

PRECAUTIONS

1. The kit is for *in vitro* diagnostic use only.
2. Do not use kit past expiration date (see **Vial and Box Label**).
3. Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
4. The reagents in this kit have been processed to reduce the bio-burden, but are not supplied sterile. Once a vial has been opened the contents should remain viable up until the expiry date.
5. Materials used to produce the kit were tested at source and found to be negative for HIV 1+2 and HBsAg using approved microbiological tests. However, no known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF KIT REAGENT AND DEALING WITH SPILLAGES

For information on disposal of kit reagent and decontamination of a spillage site see **Material Safety Data Sheets**, available on request.

CONTROLS AND ADVICE

1. It is recommended that the CRP Positive and Negative Controls are tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.
2. All the reagents must be allowed to reach 18-25°C before use.
3. Shake the reagents well before use to ensure homogeneity.
4. Do not interchange components between different kits.
5. Use of kit and interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of country where kit is in use. The user must determine the suitability of the kit for use in other techniques.

KIT COMPONENTS SUPPLIED

1. CRP Latex Reagent (White cap, 5 mL): Latex particles coated with goat IgG anti-human CRP, pH, 8.2, containing a preservative.
2. CRP Positive Control (Red cap, 1 mL): Human serum with a CRP concentration > 20 mg/L containing a preservative.
3. CRP Negative Control (Blue cap, 1 mL): Animal serum containing a preservative.
4. Pipette stirrers.
5. Disposable agglutination slide.

MATERIALS AND EQUIPMENT REQUIRED BUT NOT SUPPLIED

1. Serological Pipettes.
2. Mechanical rotator capable with adjustable speed of 80-100 rpm.
3. Vortex mixer.
4. Small Glass or Plastic Test Tubes.
5. Distilled or Deionised Water.
6. 9 g/L saline solution.

RECOMMENDED QUALITATIVE TECHNIQUE

1. Allow the reagents and samples to reach room temperature. The sensitivity of the test may be reduced at low temperatures.
2. Place 50 µL of the sample (Note 1) and one drop of each Positive and Negative controls into separate circles on the slide test.
3. Swirl the CRP Latex Reagent thoroughly but gently before use and add one drop (50 µL) next to the samples to be tested.
4. Mix the drops with a stirrer, spreading them over the entire surface of the circle. Use different stirrers for each sample.
5. Place the slide on a rotary shaker at 80-100 r.p.m. for 2 minutes. False positive results could appear if the test is read after more than two minutes.

INTERPRETATION OF QUALITATIVE RESULTS

1. **Positive:** Visible agglutination of latex particles constitutes a positive result and within the accepted limitations of the test procedure, indicates a level of CRP in the specimen > 6 mg/l.
2. **Negative:** No visible agglutination of latex particles constitutes a negative result and within the accepted limitations of the test procedure, indicates a level of CRP in the specimen < 6 mg/l.

RECOMMENDED SEMI-QUANTITATIVE TECHNIQUE

1. The semi-quantitative test can be performed in the same way as the qualitative test using dilutions of the serum.
2. Make doubling dilutions of the specimen using 9 g/L saline solution as follows:

Dilution	Serum	Saline
1/2	100 µl undiluted serum	100 µl
1/4	100 µl 1/2 diluted serum	100 µl
1/8	100 µl 1/4 diluted serum	100 µl
1/16	100 µl 1/8 diluted serum	100 µl

3. Test the specimen dilutions in the same way as for the quantitative technique above.
4. Agglutination of the sera indicates:

Dilution	CRP Levels (mg / l)
1/2	12 (6 x 2)
1/4	24 (6 x 4)
1/8	48 (6 x 8)
1/16	96 (6 x 16)



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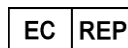
5. Normal levels of CRP in adults are < 6 mg/l.

NOTES

1. High CRP concentration samples may give false negative results (pro-zone effect). Re-test the sample again using a drop of 20 µl.

RESULTS

The titre is expressed as the reciprocal of the highest dilution showing macroscopic agglutination: e.g. if this occurs in dilution 1/8, the titre is 48.



Advena Ltd. Tower Business Centre, 2nd Flr.,
Tower Street, Swatar, BKR 4013, Malta

INTERPRETATION OF SEMI-QUANTITATIVE RESULTS

The elevation of CRP levels above normal indicates tissue damage, inflammation, or both with greater reliability. Regular monitoring of CRP levels is often used as a means of assessing disease activity and of guiding therapy. CRP determination is considered of greater practical significance than other indicators of inflammatory disease. Erythrocyte sedimentation rate (ESR) may become elevated as a result of non-inflammatory conditions. In these circumstances inflammatory disease may be excluded if CRP is absent.

STABILITY OF THE REACTIONS

Slide tests should be interpreted immediately after the 2-minute rotation period to avoid the possibility that a negative result may be incorrectly interpreted as positive due to drying of the reagent.

LIMITATIONS

- Reactions read beyond the two-minute interval may be invalid.
- The results obtained from this assay must be considered a part of the differential diagnosis and medical history of the patient.
- There is no relationship between the strength of reactivity and C-reactive protein levels.
- Hemoglobin (≤ 10 g/L), bilirubin (≤ 20 mg/dL) and lipemia (≤ 10 g/L), do not interfere. Rheumatoid factors (≥ 100 IU/mL), interfere. Other substances may interfere⁷.
- False positive or false negative results may also occur due to:
 - Contamination of test materials
 - Improper storage of test materials or omission of reagents
 - Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS

- The kit has been characterised by all the procedures mentioned in the **Recommended Techniques**.
- Prior to release, each lot of Lorne CRP Latex Test Kit is tested by **Recommended Techniques** to ensure suitable reactivity.
- The CRP-latex sensitivity is calibrated to the Reference Material ERM-DA 474/IFCC.
- Analytical sensitivity:** 6 (5-10) mg/L, under the described assay conditions.
- Prozone effect:** No prozone effect was detected up to 1600 mg/L (Note 1).
- Diagnostic sensitivity:** 95.6 %.
- Diagnostic specificity:** 96.2 %.

DISCLAIMER


- The user is responsible for the performance of the kit by any method other than those mentioned in the **Recommended Techniques**.
- Any deviations should be validated prior to use using established laboratory procedures.

BIBLIOGRAPHY

- David S.Jacobs et al. Laboratory Test Handbook, 3rd edition, Lexi-Comp Inc, 1994.

AVAILABLE KIT SIZES

Kit Size	Catalogue Number
100 Tests Per Kit	850100A

	Document Title: EU Declaration of Conformity
Date Effective: 02 Oct 2019	Document Number: DoCMiscellaneousLatex
DCN: 999	Revision Number: 01

Lorne Laboratories Ltd declares that the product family **Miscellaneous Latex** comprises of the following in vitro diagnostic reagents:

Product Name	Catalogue Number	GMDN Number
IM Latex kit	041050A	49688
LE Latex test kit	840050	63235
Strep Test kit	860050	43586
Staph Test kit	870050 and 870100	51659

have been classified as non List A, non List B (Directive 98/79/EC, Annex II) and comply with the essential requirements and provisions of the European Communities Council Directive 98/79/EC concerning In-Vitro Diagnostic Medical Devices as amended by Regulation (EC) 596/2009 and of the Council (also SI 2002 No.618 which transposes the requirements of Directive 98/79/EC).


The Intended Purpose of these products is to identify and quantitate specific antibodies in human sera following infection with pathogens which should only be carried out by suitably trained professionals.

The manufacturer of these products is Lorne Laboratories Ltd who is located at:

Address line 1: Unit 1 Cutbush Park Industrial Estate
Address line 2: Danehill
City: Lower Earley
County: Berkshire
Postal code: RG6 4UT
Country: United Kingdom
Telephone number: +44-(0)118 921 2264
Fax number: +44-(0)118 986 4518
Website: www.lornelabs.com
DUNS Number: 732670703

and are in conformity with the national standards transposing harmonised standards:

- BS EN ISO 13485:2016
- BS EN 13612:2002
- BS EN 13641:2002
- BS EN ISO 14971:2012
- BS EN ISO 15223-1:2016
- BS EN ISO 18113-2:2011
- BS EN ISO 23640:2015

	Document Title: EU Declaration of Conformity
Date Effective: 02 Oct 2019	Document Number: DoCMiscellaneousLatex
DCN: 999	Revision Number: 01

The conformity assessment procedure performed was in accordance with Annex III of Directive 98/79/EC.

Lorne's EU Authorised Representative is "Advena Limited" residing at Tower Business Centre, 2nd Floor, Tower Street, Swatar BKR 4013 Malta.

This declaration of conformity is issued under the sole responsibility of Lorne Laboratories Ltd and is valid from 02 October 2019.



Eddy Velthuis
Technical Director



**RAPID LATEX KIT
DIRECTIONS FOR USE**

LE Latex Test Kit: For Identification Of Anti-DNP.

SUMMARY

In LE (Lupus Erythematosis), autoantibodies directed against native deoxyribonucleic acid (DNA) and other nuclear constituents are produced. It is classed as the prototype of severe autoimmune diseases, involving a variety of tissues and associated with a wide range of antibodies in the circulation. Characteristics of the disease are antibodies against native DNA, nucleoprotein, denatured DNA and other extractable nuclear antigens. LE also affects a wide range of tissues. Organs affected are, in decreasing incidence, joints, skin, kidney, central nervous system, heart and lungs. One other important feature is the high frequency of the disease in women, approximately 3 to 4 times more frequent than in men. The high incidence of LE between monozygous twins (70-80%) and of close relatives (5-10%) indicates that LE may be a hereditary disease.

INTENDED PURPOSE

The reagent is a latex test reagent intended to be used to qualitatively and semi-quantitatively determine the presence or absence of antibodies against native DNA and other nuclear constituents in the serum or plasma of patients when tested in accordance with the recommended techniques stated in this IFU.

PRINCIPLE

When used by recommended techniques, latex particles in reagent will agglutinate (clump) in presence of Anti-DNA. No agglutination generally indicates the absence of Anti-DNA (see **Limitations**).

KIT DESCRIPTION

Lorne LE Rapid Latex Kit is for the identification of Anti-DNA. The test reagent consists of latex particles coated with DNA extracted from foetal calf thymus. The reagents do not contain or consist of CMR substances, or endocrine disrupting substances or that could result in sensitisation or an allergic reaction by the user. All the reagents are supplied at optimal dilution for use with all recommended techniques without the need for further dilution or addition. For lot reference number and expiry date see **Vial Labels**.

STORAGE

Do not freeze. Reagent vials should be stored at 2 - 8°C on receipt. Prolonged storage at temperatures outside this range may result in accelerated loss of reagent reactivity. Reagent will remain stable for up to 7 days when subjected to temperatures not exceeding 30°C.

SPECIMEN COLLECTION

Specimens should be drawn without anticoagulant, using an aseptic phlebotomy technique. If testing is delayed, store specimens at 2-8°C for up to 48 hours. For longer storage, remove serum from clot by centrifugation and freeze at or below -20°C. Avoid repeated freeze thawing of specimens. Do not use visibly haemolysed serum as this may cause false-positive reactions.

PRECAUTIONS

1. The kit is for *in vitro* diagnostic use only.
2. Do not use kit past expiration date (see **Vial and Box Labels**).
3. Protective clothing should be worn when handling the reagents, such as disposable gloves and a laboratory coat.
4. The reagents contain less than 0.1% sodium azide. Sodium azide may be toxic if ingested and may react with lead and copper plumbing to form explosive metal azides. On disposal flush away with large volumes of water.
5. The reagents in this kit have been processed to reduce the bio-burden, but are not supplied sterile. Once a vial has been opened the contents should remain viable up until the expiry date.
6. Materials used to produce the kit were tested at source and found to be negative for HTLV-1 and HBsAg using approved

microbiological tests. However, no known tests can guarantee that products derived from human or animal sources are free from infectious agents. Care must be taken in the use and disposal of each vial and its contents.

DISPOSAL OF KIT REAGENT AND DEALING WITH SPILLAGES

For information on disposal of kit reagent and decontamination of a spillage site see **Material Safety Data Sheets**, available on request.

CONTROLS AND ADVICE

1. It is recommended that LE Positive and Negative Controls be tested in parallel with each batch of tests. Tests must be considered invalid if controls do not show expected results.
2. All the reagents must be allowed to reach 18-25°C before use.
3. Shake the reagents well before use to ensure homogeneity.
4. Do not interchange components between different kits.
5. The reusable agglutination slide must be washed in a suitable mild disinfectant after use and then rinsed twice with deionised water to remove any residue.
6. The use of kit and interpretation of results must be carried out by properly trained and qualified personnel in accordance with the requirements of the country where the kit is in use.
7. The user must determine the suitability of the kit for use in other techniques.

KIT COMPONENTS SUPPLIED

- LE Latex Reagent (Yellow label).
- LE Positive Control (Red label/cap).
- LE Negative Control (Blue label/cap).
- Reusable agglutination slide.
- Pipette-Stirrers.

RECOMMENDED QUALITATIVE TECHNIQUE

1. Place in separate test circles of the same slide one drop of undiluted serum, one drop of positive control and one drop of negative control using the disposable pipettes provided.
2. Add one of LE Latex reagent next to each test circle.
3. Using the broad end of a pipette spread the latex reagent and specimen over entire area of the test circle.
4. Gently tilt agglutination slide backwards and forwards for 3 minutes whilst observing for agglutination.

INTERPRETATION OF QUALITATIVE RESULTS

1. **Positive:** Agglutination of latex reagent constitutes a positive result and within the accepted limitations of the test procedure, indicates the presence of Anti-DNA, LE positive.
2. **Negative:** No agglutination of latex reagent constitutes a negative result and within the accepted limitations of the test procedure, indicates the absence of Anti-DNA, LE negative.

RECOMMENDED SEMI-QUANTITATIVE TECHNIQUE

1. Using saline, dilute the specimen(s) 1:2, 1:4, 1:8, 1:16, 1:32 and 1:64.
2. Place one drop of each dilution on successive fields of the agglutination slide.
3. Add one drop of LE latex reagent to each test field and using stirrers spread the reaction mixture over the entire test field.
4. Rotate the slide for 3 minutes whilst observing for agglutination.

INTERPRETATION OF SEMI-QUANTITATIVE RESULTS

The serum LE antibody titre is the highest dilution of serum showing agglutination of the latex reagent, 3 minutes after mixing.

STABILITY OF THE REACTIONS

Slide tests should be interpreted immediately after the 3-minute rotation period to avoid the possibility that a negative result may be incorrectly interpreted as positive due to drying of the reagent.

LIMITATIONS

1. Freezing the LE Latex Reagent will cause it to agglutinate.
2. Intensity of agglutination is not necessarily indicative of relative LE titres; therefore screening reactions should not be graded.
3. Anti-DNA may be found in diseases other than LE. Low titres have been detected in rheumatoid arthritis, chronic hepatitis, periarteritis nodosa, dermatomyositis, scleroderma, atypical pneumonia, tuberculosis and lymphoma.
4. False positive or false negative results may also occur due to:
 - Contamination of test materials
 - Improper incubation time or temperature
 - Improper storage of test materials or omission of reagents
 - Deviation from the recommended techniques

SPECIFIC PERFORMANCE CHARACTERISTICS

1. The kit has been characterised by procedures mentioned in the **Recommended Techniques**.
2. Prior to release, each lot of Lorne LE Latex Test Kit is tested by **Recommended Techniques** to ensure suitable reactivity.
3. The LE latex test has been compared with a standard LE cell preparation test as well as a fluorescent ANA test. The three tests showed excellent agreement on serum from clinically active LE patients: LE latex 82% positive, LE cell prep 86% positive, ANA test 82% positive. Serum from clinically inactive LE patients: positive reactions, were LE latex 19%, ANA test 71%. Patients with connective tissue disease showed no positive reactions with the LE latex tests, but 17% and 50% positive reactions with the LE cell prep and ANA test, respectively.
4. Additional published studies have confirmed the sensitivity and specificity of the LE latex test.

DISCLAIMER

1. The user is responsible for the performance of the kit by any method other than those mentioned in the **Recommended Techniques**.
2. Any deviations should be validated prior to use using established laboratory procedures.

BIBLIOGRAPHY

1. David S.Jacobs et al. Laboratory Test Handbook, 3rd edition, Lexi-Comp Inc, 1994.

AVAILABLE KIT SIZES

Kit Size	Catalogue Number
50 Tests Per Kit	840050



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EC	REP	Advena Ltd. Tower Business Centre, 2 nd Flr., Tower Street, Swatar, BKR 4013, Malta
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MEDICA

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Fax 781 275 2731
www.medicacorp.com

Declaration of Conformity

Product Name:

EasyLyte and accessories per attachment

EasyElectrolytes and accessories per attachment

Model/Type:

EasyLyte Na/K, Na/K/Cl, Na/K/Li, Na/K/Cl/Li,
Na/K/Ca/pH, Na/K/Cl/Ca/Li

EasyElectrolytes Na/K/Cl, Na/K/Li

Manufacturer

 Medica Corporation
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Representative

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Fax: +31 70 346 7299

Means of Conformity

Medica Corporation declares that the products listed are covered by Annex III of Directive 98/79/EC. These products are self-certified since they are for professional use only and are not listed on Annex II, List A or Annex II, List B of Directive 98/79/EC. In addition, they are in conformity with the Annex I, "Essential Requirements" and provisions of council Directive 98/79/EC for In Vitro Diagnostic Medical Devices, Directive 2011/65/EU Restriction of Hazardous Substance in Electrical and Electronic Equipment, and the corresponding national laws of the Member States.

Place and Date: Bedford, Massachusetts, USA, September 27, 2018

Signature:



Name: Photios Makris, Ph.D.
Title: VP, Regulatory Affairs

EasyLyte Accessories

Catalog No.	Accessory	EDMA Code
2004	EasyLyte Na/K Analyzer	21 07 11 02
2014	EasyLyte Plus Na/K/Cl Analyzer	21 07 11 02
2015	EasyLyte Lithium Na/K/Li Analyzer	21 07 11 02
2016	EasyLyte Calcium Na/K/Ca/pH Analyzer	21 07 11 02
2021	EasyLyte Na/K/Cl/Li Analyzer	21 07 11 02
2030	EasyLyte EXPAND Analyzer, Na/K/Cl/Ca-Li	21 07 11 02
2070	EasyLyte EasySampler	21 07 11 02
2101	EasyLyte K+ Electrode	11 04 01 06
2102	EasyLyte Na+ Electrode	11 04 01 07
2113	EasyLyte Cl- Electrode	11 04 01 03
2106	EasyLyte Li+ Electrode	11 04 01 04
2150	EasyLyte Ca++ Electrode	11 04 01 02
2151	EasyLyte pH Electrode	11 70 31 02
2152	EasyLyte Disposable Reference Electrode	11 04 04 01
2103	EasyLyte Reference Electrode	11 04 04 01
2258	EasyLyte Membrane Assembly	21 07 11 02
2120	EasyLyte Na/K 800 ml Solutions Pack	11 04 04 02
2121	EasyLyte Na/K/Cl 800mL Solutions Pack	11 04 04 02
2122	EasyLyte Na/K/Li 800mL Solutions Pack	11 04 04 02
2123	EasyLyte Na/K/Ca/pH 800mL Solutions Pack	11 04 04 02
2028	EasyLyte Na/K/Cl/Li 400mL Solution Pack	11 04 04 02
2109	EasyLyte Na/K 400mL Solutions Pack	11 04 04 02
2112	EasyLyte Na/K/Cl 400mL Solutions Pack	11 04 04 02
2115	EasyLyte Na/K/Li 400mL Solutions Pack	11 04 04 02
2114	EasyLyte Na/K/Ca/pH 400mL Solutions Pack	11 04 04 02
2026	EasyLyte Na/K/Cl/Li 800mL Solution Pack	11 04 04 02
2124	EasyLyte Na/K/Cl/Ca-Li 800ml Solutions Pack	11 04 04 02
2814	EasyQC Bi-Level Quality Control Kit	11 50 02 04
2815	EasyQC Tri-Level Quality Control Kit	11 50 02 04
2843	EasyLyte Quality Control Sample Cups (60)	21 07 11 02
2118	Daily Cleaning Solution Kit	11 01 01 27
2598	EasyLyte Daily Cleaner Cup	21 07 11 02
2108	EasyLyte Solutions Valve	21 07 11 02
2107	EasyLyte Sample Probe	21 07 11 02
2257	EasyLyte Sample Detector	21 07 11 02

EasyLyte Accessories, continued

Catalog No.	Accessory	EDMA Code
2104	EasyLyte Tubing Kit	21 07 11 02
2100	EasyLyte Calcium Tubing Kit	21 07 11 02
2492	EasyLyte Internal Filling Solution (125mL)	11 04 04 90
2309	EasyLyte Wash Solution (50mL)	11 04 04 90
2111	EasyLyte Urine Diluent (500mL)	11 04 04 90
2577	EasyLyte Standard Solution, Urine (50mL)	11 04 04 90
2323	EasyLyte Probe Wipers (6)	21 07 11 02
2541	EasyLyte Printer Paper (3 rolls)	21 07 11 02
2595	EasyLyte EasySampler Sample Cups, 500uL (500)	21 07 11 02
2596	EasyLyte Sample Cups 2.0mL (500)	21 07 11 02
10745	Anti-Evaporation Caps (500)	21 07 11 02
2293	EasyLyte Capillary Tubes	21 07 11 02
2590	EasyLyte Capillary Adaptor Kit	21 07 11 02
2292	EasyLyte Capillary Adaptor Cleaning Kit	21 07 11 02
2578	EasyLyte Red Dye Test Solution (50mL)	11 30 01 11
2572	EasyLyte Troubleshooting Kit	21 07 11 02
2571	EasyLyte Troubleshooting Kit (Na/K/Ca/pH and Na/K/Cl/Li)	21 07 11 02
2105	EasyLyte Quarterly Operating Kit	21 07 11 02
2095	EasyLyte Maintenance Kit	21 07 11 02
2076	EasyLyte Sample Tray	21 07 11 02
2074	EasyLyte Sample Cup Retainer Ring	21 07 11 02
7118	Daily Rinse/Cleaning Solution Kit	11 01 01 27
2544	EasyLyte C Series Printer Paper (5 rolls)	21 07 11 02
2934	EasyLyte Barcode Reader Kit	21 07 11 02

EasyElectrolytes Accessories

Catalog No.	Accessory	EDMA Code
4002	EasyElectrolyte Na/K/Cl Analyzer	21 07 11 02
4003	EasyElectrolyte Na/K/Li Analyzer	21 07 11 02
4102	Reagent Module, Na/K/Cl	11 04 04 02
4103	Reagent Module, Na/K/Li	11 04 04 02
7205	EasyElectrolyte/EasyStat Na+ Electrode	11 04 01 07
7206	EasyElectrolyte/EasyStat K+ Electrode	11 04 01 06
4203	EasyElectrolyte Cl- Electrode	11 04 01 03
4204	EasyElectrolyte Li+ Electrode	11 04 01 04
6204	EasyElectrolyte/EasyStat/EasyBloodGas Reference Electrode	11 04 04 01
4207	EasyElectrolyte Spacer Electrode	11 04 01 90
4301	EasyElectrolyte Troubleshooting Kit	21 07 11 02
2118	Daily Cleaning Solution Kit	11 01 01 27
4402	EasyStat/EasyBloodGas/EasyElectrolyte Red Test Dye Solution	11 30 01 11
4403	EasyElectrolyte Urine Diluent	11 04 04 90
2814	Bi-Level Quality Control Kit	11 50 02 04
2815	Tri-Level Quality Control Kit	11 50 02 04
4405	EasyElectrolyte Na/K/Cl Demonstration Kit	21 07 11 02
4406	EasyElectrolyte Na/K/Li Demonstration Kit	21 07 11 02
4404	EasyElectrolyte Capillary Tube Kit	21 07 11 02
4306	EasyElectrolyte Sampler	21 07 11 02
6504	EasyBloodGas/EasyElectrolyte Pump Tube	21 07 11 02
6505	EasyStat/EasyBloodGas/EasyElectrolyte Printer Paper	21 07 11 02
4506	EasyElectrolyte Sensor Module	21 07 11 02
4507	EasyElectrolyte Valve Module	21 07 11 02
4508	EasyStat/EasyBloodGas/EasyElectrolyte Compression Plate	21 07 11 02
7302	Probe Wipers	21 07 11 02
4522	EasyElectrolyte Daily Cleaner Sample Cups	21 07 11 02
4539	EasyElectrolyte Sensor Module, Li+	21 07 11 02
6537	EasyElectrolyte/EasyStat/EasyBloodGas Serial Cable, 9-pin	21 07 11 02
6520	EasyElectrolyte/EasyStat/EasyBloodGas Barcode Reader Kit	21 07 11 02

EasyBloodGas™ analyzer
EasyLyte® analyzer

EasyElectrolytes® analyzer
EasyStat® analyzer

Training Certificate

This is to certify that

Mr. Sergiu Sorocovici

Of GBG-MLD S.R.L.

has completed training for the operation and service of the

EasyBloodGas™ analyzer, EasyElectrolytes® analyzer, EasyLyte® analyzer and EasyStat® analyzer

04/22/2016
DATE



Medica Corporation

David Hagopian
Director of Technical Support



**CERTIFICADO CE DE SISTEMA DE GARANTÍA DE CALIDAD TOTAL
de acuerdo con el Anexo IV (excepto punto 4) de la Directiva 98/79/CE**

**EC FULL QUALITY ASSURANCE SYSTEM CERTIFICATE
in accordance with Annex IV (except Section 4) of Directive 98/79/EC**

PRÓRROGA/EXTENSION — Fecha inicial/ Initial date: 11/12/2003

Fecha de última prórroga/ Last extension date: 27/11/2013

Certificado nº/Certificate no	Fecha de validez/Date of validity	ON nº/NB no
2003 12 0388 CT	Desde/From 27/11/2018 Hasta/To 18/11/2023	0318

A favor de /In favour of:

Fabricante/Manufacturer:

Nombre/Name: DIA. Pro Diagnostic Bioprobes S.r.l.

Dirección/Address: Via G. Carducci, 27 -20099- Sesto San Giovanni – Milano (Italy).

Representante autorizado ante la UE/Authorized EU representative:

Nombre/Name: Idem Dirección/Address: Idem

Para los productos/For the products:

Categoría/Category: Productos Sanitarios para Diagnóstico “In Vitro” / In Vitro Diagnostic Medical Devices

Grupo genérico/Generic group: Diagnóstico de enfermedades infecciosas / Diagnostic of infectious diseases

Tipo/Type: Especificados en Anexos de este Certificado/Specified in Annexes to this Certificate.

Elaborado en/In the facilities:

Dia. Pro Diagnostic Bioprobes S.r.l.

Via G. Carducci, 27 -20099- Sesto San Giovanni – Milano (Italy).

Este certificado debe ir acompañado por certificado de examen de diseño: SI
This certificate must be accompanied by design examination certificate: YES

Este certificado es consecuencia de la auditoria del Sistema Completo de Garantía de Calidad y del examen de la documentación técnica contenida en el expediente nº 2003 05 0240, y garantiza que los productos descritos cumplen los requisitos de la Directiva. / *This certificate is issued on the full quality assurance system audit, and the examination of the technical documentation contained in dossier nº 2003 05 02405, and guarantees that the described products fulfil the requirements of the Directive.*

Madrid, 26 de noviembre de 2018

DIRECTORA DE LA AGENCIA ESPAÑOLA DE MEDICAMENTOS Y PRODUCTOS SANITARIOS

Fdo. Mª Jesús Lamas Díaz

Firmado digitalmente por: Agencia Española de Medicamentos y Productos Sanitarios

Localizador: X9GVDEF5C3

Fecha de la firma: 26/11/2018

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CORREO ELECTRÓNICO

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ORGANISMO NOTIFICADO 0318



ANEXO N°/ANNEX NO: I
CERTIFICADO CE DE SISTEMA DE GARANTÍA DE CALIDAD TOTAL
de acuerdo con el Anexo IV (excepto punto 4) de la Directiva 98/79/CE
EC FULL QUALITY ASSURANCE SYSTEM CERTIFICATE
in accordance with Annex IV (except Section 4) of Directive 98/79/EC
PRÓRROGA/EXTENSION — Fecha inicial/ Initial date: 11/12/2003
Fecha de última prórroga/ Last extension date: 27/11/2013

Certificado n°/Certificate no	Fecha de validez/Date of validity	ON n°/NB no
2003 12 0388 CT	Desde/From 27/11/2018 Hasta/To 18/11/2023	0318

1.4. HBc IgM

- BCM.CE (96 tests) Descrito en el certificado / Described in the certificate 2004 03 0424 ED

1.5. HBe Ag & Ab

- HBE.CE (96 tests) Descrito en el certificado / Described in the certificate 2004 03 0425 ED

1.6. HBs Ag Confirmation

- SCONF.CE (20 tests) Descrito en el certificado / Described in the certificate 2006 11 0511 ED
- SCONF.CE.40 (40 tests)

1.7. HBs Ag one Version ULTRA

- SAG1ULTRA.CE (192 tests) Descrito en el certificado / Described in the certificate 2008 12 0588 ED
- SAG1ULTRA.CE.96 (96 tests)
- SAG1ULTRA.CE.480 (480 tests)
- SAG1ULTRA.CE.960 (960 tests)
- SAG1ULTRA.CE.DB (192 tests)

1.8. HCV Ab

- CVAB.CE (192 tests) Descrito en el certificado / Described in the certificate 2003 12 0392 ED
- CVAB.CE.96 (96 tests)
- CVAB.CE.480 (480 tests)
- CVAB.CE.960 (960 tests)
- CVAB.CE.DB (192 tests)

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1.9. HCV Ab Confirmation

- CCONF.CE (12 tests) Descrito en el certificado / *Described in the certificate*
2005 09 0485 ED

1.10. HCV IgM

- CVM.CE (96 tests) Descrito en el certificado / *Described in the certificate*
2007 09 0532 ED

1.11. HCV Ab (Format 20)

- CVAB.CE.EG (192 tests) Descrito en el certificado / *Described in the certificate*
- CVAB.CE.EG.96 (96 tests) 2015 10 0842 ED
- CVAB.CE.EG.480 (480 tests)
- CVAB.CE.EG.960 (960 tests)

1.12. HDV Ab

- DAB.CE (96 tests) Descrito en el certificado / *Described in the certificate*
2003 12 0393 ED

1.13. HDV Ag

- DAG.CE (96 tests) Descrito en el certificado / *Described in the certificate*
2003 12 0394 ED

1.14. HDV IgM

- DIM.CE (96 tests) Descrito en el certificado / *Described in the certificate*
2003 12 0395 ED

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1.15. HTLV I & II Ab

- HTLVAB.CE (192 tests) Descrito en el certificado/ *Described in the*
- HTLVAB.CE.96 (96 tests) *certificate* 2005 12 0493 ED
- HTLVAB.CE.480 (480 tests)
- HTLVAB.CE.960 (960 tests)

1.16. HTLV I & II Ab Version ULTRA

- HTLVABULTRA.CE (192 tests) Descrito en el certificado/ *Described in the*
- HTLVABULTRA.CE.96 (96 tests) *certificate* 2011 11 0775 ED
- HTLVABULTRA.CE.480 (480 tests)
- HTLVABULTRA.CE.960 (960 tests)
- HTLVABULTRA.CE.DB (192 tests)

1.17. HIV Ab & Ag

- IVCOMB.CE (192 tests) Descrito en el certificado/ *Described in the*
- IVCOMB.CE.96 (96 tests) *certificate* 2008 02 0539 ED
- IVCOMB.CE.480 (480 tests)
- IVCOMB.CE.960 (960 tests)
- IVCOMB.CE.DB (192 tests)

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2. Reactivos y productos reactivos para la determinación, confirmación y cuantificación de marcadores de infección en muestras humanas mediante técnicas de PCR en tiempo real/ Reagents and reactive products for the determination, confirmation and quantification of infection markers in human samples by Real-Time PCR [NANDO: IVD 0203]

2.1. HBV DNA Quantitation (QT)

- HBVDNAQT.CE (50 tests)
- HBVDNAQT.CE.25 (25 tests)
- HBVDNAQT.CE.100 (100 tests)
- HBVDNAQT.CE.150 (150 tests)

Descrito en el certificado / Described in the certificate 2012 09 0790 ED

2.2. HDV RNA Quantitation (QT)

- DRNA.CE (50 tests)
- DRNA.CE.25 (25 tests)
- DRNA.CE.100 (100 tests)
- DRNA.CE.150 (150 tests)

Descrito en el certificado / Described in the certificate 2009 11 0660 ED

3. Reactivos y productos reactivos para la determinación, confirmación y cuantificación de marcadores de infección en muestras humanas mediante ensayos de quimioluminiscencia (CLIA)/ Reagents and reactive products for the determination, confirmation and quantification of infection markers in human samples by Chemiluminescence Immunoassay (CLIA) [NANDO: IVD 0201; IVD 0202; IVD 0203]

3.1. DIA.CHEMILUX HCV Ab

- RACVAB.CE (100 tests)

Descrito en el certificado / Described in the certificate 2015 01 0834 ED

3.2. DIA.CHEMILUX HBs Ag

- RASAG.CE (100 tests)

Descrito en el certificado / Described in the certificate 2015 10 0841 ED

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3.3. DIA.CHEMILUX HIV Ab & Ag

- RAIVCOMB.CE (100 tests) Descrito en el certificado / *Described in the certificate* 2016 02 0844 ED

3.4. DIA.CHEMILUX HBc Ab

- RABCAB.CE (100 tests) Descrito en el certificado / *Described in the certificate* 2017 07 0863 ED

3.5. DIA.CHEMILUX HTLV I & II Ab

- RAHTLVAB.CE (100 tests) Descrito en el certificado / *Described in the certificate* 2018 11 0878 ED

Este certificado ampara todas las marcas de estos productos incluidas por el fabricante en su declaración de conformidad. / This certificate covers all trademarks of these products included by the manufacturer in his declaration of conformity.

Madrid, 26 de noviembre de 2018
DIRECTORA DE LA AGENCIA ESPAÑOLA DE MEDICAMENTOS Y PRODUCTOS SANITARIOS

 **agencia española de
medicamentos y
productos sanitarios**

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HBsAg_{one}

Version ULTRA

**Fourth generation Enzyme
Immunoassay (ELISA)
for the determination of
Hepatitis B surface Antigen or HBsAg
in human serum and plasma**

- for "in vitro" diagnostic use only -



DIA.PRO

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HBsAg One version ULTRA

A. INTENDED USE

Fourth generation Enzyme Immunoassay (ELISA) for the one-step determination of Hepatitis B surface Antigen or HBsAg in human plasma and sera.

The kit is intended for the screening of blood units, is able to detect HBsAg mutants and finds application in the follow-up of HBV-infected patients.

For "in vitro" diagnostic use only.

B. INTRODUCTION

The World Health Organization (WHO) defines Hepatitis B Virus infection as follows:

"Hepatitis B is one of the major diseases of mankind and is a serious global public health problem. Hepatitis means inflammation of the liver, and the most common cause is infection with one of 5 viruses, called hepatitis A,B,C,D, and E. All of these viruses can cause an acute disease with symptoms lasting several weeks including yellowing of the skin and eyes (jaundice); dark urine; extreme fatigue; nausea; vomiting and abdominal pain. It can take several months to a year to feel fit again. Hepatitis B virus can cause chronic infection in which the patient never gets rid of the virus and many years later develops cirrhosis of the liver or liver cancer.

HBV is the most serious type of viral hepatitis and the only type causing chronic hepatitis for which a vaccine is available. Hepatitis B virus is transmitted by contact with blood or body fluids of an infected person in the same way as human immunodeficiency virus (HIV), the virus that causes AIDS. However, HBV is 50 to 100 times more infectious than HIV. The main ways of getting infected with HBV are: (a) perinatal (from mother to baby at the birth); (b) child- to-child transmission; (c) unsafe injections and transfusions; (d) sexual contact.

Worldwide, most infections occur from infected mother to child, from child to child contact in household settings, and from reuse of un-sterilized needles and syringes. In many developing countries, almost all children become infected with the virus. In many industrialized countries (e.g. Western Europe and North America), the pattern of transmission is different. In these countries, mother-to-infant and child-to-child transmission accounted for up to one third of chronic infections before childhood hepatitis B vaccination programmes were implemented. However, the majority of infections in these countries are acquired during young adulthood by sexual activity, and injecting drug use. In addition, hepatitis B virus is the major infectious occupational hazard of health workers, and most health care workers have received hepatitis B vaccine.

Hepatitis B virus is not spread by contaminated food or water, and cannot be spread casually in the workplace. High rates of chronic HBV infection are also found in the southern parts of Eastern and Central Europe. In the Middle East and Indian sub-continent, about 5% are chronically infected. Infection is less common in Western Europe and North America, where less than 1% are chronically infected.

Young children who become infected with HBV are the most likely to develop chronic infection. About 90% of infants infected during the first year of life and 30% to 50% of children infected between 1 to 4 years of age develop chronic infection. The risk of death from HBV-related liver cancer or cirrhosis is approximately 25% for persons who become chronically infected during childhood. Chronic hepatitis B in some patients is treated with drugs called *interferon* or *lamivudine*, which can help some patients. Patients with cirrhosis are sometimes given liver transplants, with varying success. It is preferable to prevent this disease with vaccine than to try and cure it.

Hepatitis B vaccine has an outstanding record of safety and effectiveness. Since 1982, over one billion doses of hepatitis B vaccine have been used worldwide. The vaccine is given as a series of three intramuscular doses. Studies have shown that the vaccine is 95% effective in preventing children and adults from developing chronic infection if they have not yet been infected. In many countries where 8% to 15% of children used to become chronically infected with HBV, the rate of chronic infection has been reduced to less than 1% in immunized groups of children. Since 1991, WHO has called for all countries to add hepatitis B vaccine into their national immunization programs."

Hepatitis B surface Antigen or HBsAg is the most important protein of the envelope of Hepatitis B Virus, responsible for acute and chronic viral hepatitis.

The surface antigen contains the determinant "a", common to all the known viral subtypes, immunologically distinguished by two distinct subgroups (ay and ad).

The ability to detect HBsAg with high sensitive immunoassays in the last years has led to an understanding of its distribution and epidemiology worldwide and to radically decrease the risk of infection in transfusion.

C. PRINCIPLE OF THE TEST

A mix of mouse monoclonal antibodies specific to the determinants "a", "d" and "y" of HBsAg is fixed to the surface of microwells. Patient's serum/plasma is added to the microwell together with a second mix of mouse monoclonal antibodies, conjugated with Horseradish Peroxidase (HRP) and directed against a different epitope of the determinant "a" and against "preS".

The specific immunocomplex, formed in the presence of HBsAg in the sample, is captured by the solid phase.

At the end of the one-step incubation, microwells are washed to remove unbound serum proteins and HRP conjugate.

The chromogen/substrate is then added and, in the presence of captured HBsAg immunocomplex, the colorless substrate is hydrolyzed by the bound HRP conjugate to a colored end-product. After blocking the enzymatic reaction, its optical density is measured by an ELISA reader.

The color intensity is proportional to the amount of HBsAg present in the sample.

The version ULTRA is particularly suitable for automated screenings and is able to detect "s" mutants.

D. COMPONENTS

The standard configuration contains reagents to perform 192 tests and is made of the following components:

1. Microplate MICROPLATE

n° 2. 12 strips of 8 breakable wells coated with anti HBsAg, affinity purified mouse monoclonal antibodies, specific to "a", "y" and "d" determinants, and sealed into a bag with desiccant.

2. Negative Control CONTROL -

1x4.0ml/vial. Ready to use control. It contains goat serum, 10 mM phosphate buffer pH 7.4+/-0.1, 0.09% Na-azide and 0.1% Kathon GC as preservatives. The negative control is pale yellow color coded.

3. Positive Control CONTROL +

1x4.0ml/vial. Ready to use control. It contains goat serum, non infectious recombinant HBsAg, 10 mM phosphate buffer pH 7.4+/-0.1, 0.02% gentamicine sulphate and 0.1% Kathon GC as preservatives. The positive control is color coded green.

4. Calibrator CAL ...

n° 2 vials. Lyophilized calibrator. To be dissolved with EIA grade water as reported in the label. Contains fetal bovine serum, non infectious recombinant HBsAg at 0.5 IU/ml (2nd WHO international standard for HBsAg, NIBSC code 00/588), 10 mM phosphate buffer pH 7.4+/-0.1, 0.02% gentamicine sulphate and 0.1% Kathon GC as preservatives.

Note: The volume necessary to dissolve the content of the vial may vary from lot to lot. Please use the right volume reported on the label .

5. Wash buffer concentrate WASHBUF 20X

2x60ml/bottle. 20X concentrated solution. Once diluted, the wash solution contains 10 mM phosphate buffer pH 7.0+/-0.2, 0.05% Tween 20 and 0.1% Kathon GC.

6. Enzyme Conjugate Diluent CONJ DIL

2x16ml/vial. Ready to use and pink/red color coded reagent. It contains 10 mM Tris buffer pH 6.8+/-0.1, 1% normal mouse serum, 5% BSA, 0.1% Kathon GC and 0.02% gentamicine sulphate as preservatives. The solution is normally opalescent.

7. Enzyme Conjugate CONJ 20X

2x1ml/vial. 20X concentrated reagent. It contains Horseradish Peroxidase (HRP) labeled mouse monoclonal antibodies to HBsAg, determinant "a" and "preS", 10 mM Tris buffer pH 6.8+/-0.1, 5% BSA, 0.1% Kathon GC and 0.02% gentamicine sulphate as preservatives.

8. Chromogen/Substrate SUBS TMB

2x25ml/bottle. It contains a 50 mM citrate-phosphate buffered solution at pH 3.5-3.8, 4% dimethylsulphoxide, 0.03% tetra-methyl-benzidine (TMB) and 0.02% hydrogen peroxide (H₂O₂).

Note: To be stored protected from light as sensitive to strong illumination.

9. Sulphuric Acid H₂SO₄ 0.3 M

1x25ml/bottle. It contains 0.3 M H₂SO₄ solution.

Note: Attention: Irritant (H315; H319; P280; P302+P352; P332+P313; P305+P351+P338; P337+P313; P362+P363)

10. Plate sealing foils n° 4

11. Package insert

Important note:

Only upon specific request, Dia.Pro can supply reagents for 96, 480, 960 tests, as reported below:

	N°1	N°5	N°10
Microplates	N°1	N°5	N°10
Negative Control	1x2ml/vial	1x10ml/vial	1x20ml/vial
Positive Control	1x2ml/vial	1x10ml/vial	1x20ml/vial
Calibrator	N° 1 vial	N° 5 vials	N° 10 vials
Wash buffer concentrate	1x60ml/vial	5x60ml/vial	4x150ml/vial
Enzyme conjugate	1x0.8ml/vial	1x4ml/vial	2x4ml/vial
Conjugate Diluent	1x16ml/vial	2x40ml/vial	2x80ml/vial
Chromogen/Substrate	1x25ml/vial	3x42ml/vial	2x125ml/vial
Sulphuric Acid	1x15ml/vial	2x40ml/vial	2x80ml/vial
Plate sealing foils	N° 2	N° 10	N° 20
Package insert	N° 1	N° 1	N° 1
Number of tests	96	480	960
Code SAGIULTRA.CE	96	480	960

E. MATERIALS REQUIRED BUT NOT PROVIDED

1. Calibrated Micropipettes (150ul, 100ul and 50ul) and disposable plastic tips.
2. EIA grade water (double distilled or deionised, charcoal treated to remove oxidizing chemicals used as disinfectants).
3. Timer with 60 minute range or higher.
4. Absorbent paper tissues.
5. Calibrated ELISA microplate thermostatic incubator (dry or wet), capable to provide shaking at 1300 rpm+/-150, set at +37°C.
6. Calibrated ELISA microwell reader with 450nm (reading) and with 620-630nm (blanking) filters.
7. Calibrated ELISA microplate washer.
8. Vortex or similar mixing tools.

F. WARNINGS AND PRECAUTIONS

1. The kit has to be used by skilled and properly trained technical personnel only, under the supervision of a medical doctor responsible of the laboratory.
2. When the kit is used for the screening of blood units and blood components, it has to be used in a laboratory certified and qualified by the national authority in that field (Ministry of Health or similar entity) to carry out this type of analysis.
3. All the personnel involved in performing the assay have to wear protective laboratory clothes, talc-free gloves and glasses. The use of any sharp (needles) or cutting (blades) devices should be avoided. All the personnel involved should be trained in biosafety procedures, as recommended by the Center for Disease Control, Atlanta, U.S. and reported in the National Institute of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
4. All the personnel involved in sample handling should be vaccinated for HBV and HAV, for which vaccines are available, safe and effective.
5. The laboratory environment should be controlled so as to avoid contaminants such as dust or air-borne microbial agents, when opening kit vials and microplates and when performing the test. Protect the Chromogen (TMB) from strong light and avoid vibration of the bench surface where the test is undertaken.
6. Upon receipt, store the kit at 2.8°C into a temperature controlled refrigerator or cold room.
7. Do not interchange components between different lots of the kits. It is recommended that components between two kits of the same lot should not be interchanged.
8. Check that the reagents are clear and do not contain visible heavy particles or aggregates. If not, advise the laboratory supervisor to initiate the necessary procedures for kit replacement.
9. Avoid cross-contamination between serum/plasma samples by using disposable tips and changing them after each sample.
10. Avoid cross-contamination between kit reagents by using disposable tips and changing them between the use of each one.
11. Do not use the kit after the expiration date stated on the external container and internal (vials) labels. A study conducted on an opened kit has not pointed out any relevant loss of activity up to 6 re-use of the device and up to 6 months.
12. Treat all specimens as potentially infective. All human serum specimens should be handled at Biosafety Level 2, as recommended by the Center for Disease Control, Atlanta, U.S. in compliance with what reported in the Institutes of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
13. The use of disposable plastic-ware is recommended in the preparation of the liquid components or in transferring components into automated workstations, in order to avoid cross contamination.
14. Waste produced during the use of the kit has to be discarded in compliance with national directives and laws concerning laboratory waste of chemical and biological substances. In particular, liquid waste generated from the washing procedure, from residuals of controls and from samples has to be treated as potentially infective material and inactivated before waste. Suggested procedures of inactivation are treatment with a 10% final concentration of household bleach for 16-18 hrs or heat inactivation by autoclave at 121°C for 20 min..
15. Accidental spills from samples and operations have to be adsorbed with paper tissues soaked with household bleach and then with water. Tissues should then be discarded in proper containers designated for laboratory/hospital waste.
16. The Stop Solution is an irritant. In case of spills, wash the surface with plenty of water
17. Other waste materials generated from the use of the kit (example: tips used for samples and controls, used microplates) should be handled as potentially infective and disposed according to national directives and laws concerning laboratory wastes.

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G. SPECIMEN: PREPARATION AND WARNINGS

1. Blood is drawn aseptically by venepuncture and plasma or serum is prepared using standard techniques of preparation of samples for clinical laboratory analysis. No influence has been observed in the preparation of the sample with citrate, EDTA and heparin.
2. Avoid any addition of preservatives to samples; especially sodium azide as this chemical would affect the enzymatic activity of the conjugate, generating false negative results.
3. Samples have to be clearly identified with codes or names in order to avoid misinterpretation of results. When the kit is used for the screening of blood units, bar code labeling and electronic reading is strongly recommended.
4. Haemolysed (red) and lipemic ("milky") samples have to be discarded as they could generate false results. Samples containing residues of fibrin or heavy particles or microbial filaments and bodies should be discarded as well as they could give rise to false positive results. Specimens with an altered pathway of coagulation, presenting particles after blood collection and preparation of serum/plasma as those coming from hemodialized patients, could give origin to false positive results.
5. Sera and plasma can be stored at +2°..8°C for up to seven days after collection. For longer storage periods, samples can be stored frozen at -20°C for several months. Any frozen sample should not be frozen/thawed more than once as this may generate particles that could affect the test result. If some turbidity is present or presence of microparticles is suspected after thawing, filter the sample on a disposable 0.2-0.8µ filter to clean it up for testing or use the two-steps alternative method.

H. PREPARATION OF COMPONENTS AND WARNINGS

A study conducted on an opened kit has not pointed out any relevant loss of activity up to 6 re-uses of the device and up to 6 months.

1. Microplates:

Allow the microplate to reach room temperature (about 1 hr) before opening the container. Check that the desiccant has not turned green, indicating a defect in conservation. In this case, call Dia.Pro's customer service. Unused strips have to be placed back inside the aluminum pouch, with the desiccant supplied, firmly zipped and stored at +2°..8°C. After first opening, remaining strips are stable until the humidity indicator inside the desiccant bag turns from yellow to green.

2. Negative Control:

Ready to use. Mix well on vortex before use.

3. Positive Control:

Ready to use. Mix well on vortex before use. The positive control does not contain any infective HBV as it is composed of recombinant synthetic HBsAg.

4. Calibrator:

Add the volume of ELISA grade water, reported on the label, to the lyophilized powder; let fully dissolve and then gently mix on vortex. The solution is not stable. Store the Calibrator frozen in aliquots at -20°C.

5. Wash buffer concentrate:

The 20x concentrated solution has to be diluted with EIA grade water up to 1200 ml and mixed gently end-over-end before use. As some salt crystals may be present into the vial, take care to dissolve all the content when preparing the solution. In the preparation avoid foaming as the presence of bubbles could give origin to a bad washing efficiency.

Note: Once diluted, the wash solution is stable for 1 week at +2°..8°C.

6. Enzyme conjugate:

The working solution is prepared by diluting the 20X concentrated reagent into the Conjugate Mix well on vortex before use.

Avoid any contamination of the liquid with oxidizing chemicals, dust or microbes. If this component has to be transferred, use only plastic sterile disposable containers.

Important note: The working solution is not stable. Prepare only the volume necessary for the work of the day. As an example when the kit is used in combination with other instruments or manually, dilute 0.1 ml 20X Conjugate with 1.9 ml Conjugate Diluent into a disposable plastic vial and mix carefully before use.

7. Chromogen/Substrate:

Ready to use. Mix well by end-over-end mixing.

Avoid contamination of the liquid with oxidizing chemicals, air-driven dust or microbes. Do not expose to strong light, oxidizing agents and metallic surfaces.

If this component has to be transferred use only plastic, and if possible, sterile disposable container.

8. Sulphuric Acid:

Ready to use. Mix well by end-over-end mixing.

Attention: Irritant (H315; H319; P280; P302+P352; P332+P313; P305+P351+P338; P337+P313; P362+P363).

Legenda:

Warning H statements:

H315 – Causes skin irritation.

H319 – Causes serious eye irritation.

Precautionary P statements:

P280 – Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352 – IF ON SKIN: Wash with plenty of soap and water.

P332 + P313 – If skin irritation occurs: Get medical advice/attention.

P305 + P351 + P338 – IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P337 + P313 – If eye irritation persists: Get medical advice/attention.

P362 + P363 – Take off contaminated clothing and wash it before reuse.

I. INSTRUMENTS AND TOOLS USED IN COMBINATION WITH THE KIT

1. **Micropipettes** have to be calibrated to deliver the correct volume required by the assay and must be submitted to regular decontamination (70% ethanol, 10% solution of bleach, hospital grade disinfectants) of those parts that could accidentally come in contact with the sample or the components of the kit. They should also be regularly maintained in order to show a precision of 1% and a trueness of $\pm 2\%$.
2. The **ELISA incubator** has to be set at +37°C (tolerance of $\pm 1^\circ\text{C}$) and regularly checked to ensure the correct temperature is maintained. Both dry incubators and water baths are suitable for the incubations, provided that the instrument is validated for the incubation of ELISA tests.
3. In case of **shaking** during incubations, the instrument has to ensure 350 rpm ± 150 . Amplitude of shaking is very important as a wrong one could give origin to splashes and therefore to some false positive result.
4. The **ELISA washer** is extremely important to the overall performances of the assay. The washer must be carefully validated and correctly optimized using the kit controls/calibrator and reference panels, before using the kit for routine laboratory tests. Usually 4-5 washing cycles (aspiration + dispensation of 350µl/well of washing solution = 1 cycle) are sufficient to ensure that the assay performs as expected. A soaking time of 20-30 seconds between cycles is suggested. In order to set correctly their number, it

is recommended to run an assay with the kit controls/calibrator and well-characterized negative and positive reference samples, and check to match the values reported below in the section "Internal Quality Control". Regular calibration of the volumes delivered and maintenance (decontamination and cleaning of needles) of the washer has to be carried out according to the instructions of the manufacturer.

5. **Incubation times** have a tolerance of $\pm 5\%$.
6. The **microplate reader** has to be equipped with a reading filter of 450nm and with a second filter (620-630nm, strongly recommended) for blanking purposes. Its standard performances should be (a) bandwidth ≤ 10 nm; (b) absorbance range from 0 to ≥ 2.0 ; (c) linearity to ≥ 2.0 ; (d) repeatability $\geq 1\%$. Blanking is carried out on the well identified in the section "Assay Procedure". The optical system of the reader has to be calibrated regularly to ensure that the correct optical density is measured. It should be regularly maintained according to the manufacturer's instructions.
7. When using **ELISA automated workstations**, all critical steps (dispensation, incubation, washing, reading, shaking, data handling, etc.) have to be carefully set, calibrated, controlled and regularly serviced in order to match the values reported in the sections "Internal Quality Control". The assay protocol has to be installed in the operating system of the unit and validated by checking full matching the declared performances of the kit. In addition, the liquid handling part of the station (dispensation and washing) has to be validated and correctly set paying particular attention to avoid carry over by the needles used for dispensing samples and for washing. The carry over effect must be studied and controlled to minimize the possibility of contamination of adjacent wells due to strongly reactive samples, leading to false positive results. The use of ELISA automated work stations is recommended for blood screening and when the number of samples to be tested exceed 20-30 units per run.
8. When using automatic devices, in case the vial holder of the instrument does not fit with the vials supplied in the kit, transfer the solution into appropriate containers and label them with the same label peeled out from the original vial. This operation is important in order to avoid mismatching contents of vials, when transferring them. When the test is over, return the secondary labeled containers to $2..8^{\circ}\text{C}$, firmly capped.
9. **Dia.Pro's customer service** offers support to the user in the setting and checking of instruments used in combination with the kit, in order to assure full compliance with the essential requirements of the assay. Support is also provided for the installation of new instruments to be used in combination with the kit.

L. PRE ASSAY CONTROLS AND OPERATIONS

1. Check the expiration date of the kit printed on the external label of the kit box. Do not use if expired.
2. Check that the liquid components are not contaminated by naked-eye visible particles or aggregates. Check that the Chromogen/Substrate is colorless or pale blue. Check that no breakage occurred in transportation and no spillage of liquid is present inside the box. Check that the aluminum pouch, containing the microplate, is not punctured or damaged.
3. Dilute all the content of the 20x concentrated Wash Solution as described above.
4. Dilute the 20X concentrated Enzyme Conjugate with its Diluent as reported.
5. Dissolve the Calibrator as described above.
6. Allow all the other components to reach room temperature (about 1 hr) and then mix as described.
7. Set the ELISA incubator at $+37^{\circ}\text{C}$ and prepare the ELISA washer by priming with the diluted washing solution,

according to the manufacturers instructions. Set the right number of washing cycles as found in the validation of the instrument for its use with the kit.

8. Check that the ELISA reader has been turned on at least 20 minutes before reading.
9. If using an automated workstation, turn it on, check settings and be sure to use the right assay protocol.
10. Check that the micropipettes are set to the required volume.
11. Check that all the other equipment is available and ready to use.
12. In case of problems, do not proceed further with the test and advise the supervisor.

M. ASSAY PROCEDURE

The assay has to be carried out according to what reported below, taking care to maintain the same incubation time for all the samples in testing.

Automated assay:

In case the test is carried out automatically with an ELISA system, we suggest to make the instrument dispense first 150 ul controls & calibrator, then all the samples and finally 100 ul diluted Enzyme Conjugate.

For the pre-washing step (point 1 of the assay procedure) and all the next operations follow the operative instructions reported below for the Manual Assay.

It is strongly recommended to check that the time lap between the dispensation of the first and the last sample will be calculated by the instrument and taken into consideration by delaying the first washing operation accordingly.

Manual Assay:

1. Place the required number of strips in the plastic holder and wash them once to hydrate wells. Carefully identify the wells for controls, calibrator and samples.

Important note: *Pre washing (1 cycle: dispensation of 350ul/well of washing solution+ aspiration) is fundamental to obtain reliable and specific results both in the manual and in the automatic procedures. Do not omit it !*

2. Leave the A1 well empty for blanking purposes.
3. Pipette 150ul of the Negative Control in triplicate, 150ul of the Calibrator in duplicate and then 150ul of the Positive Control in single followed by 150ul of each of the samples.
4. Check for the presence of samples in wells by naked eye (there is a marked color difference between empty and full wells) or by reading at 450/620nm. (samples show OD values higher than 0.100).
5. Dispense 100ul diluted Enzymatic Conjugate in all wells, except for A1, used for blanking operations.

Important note: *Be careful not to touch the inner surface of the well with the pipette tip when the conjugate is dispensed. Contamination might occur.*

6. Following addition of the conjugate, check that the color of the samples have changed from yellowish to pink/red and then incubate the microplate for **120 min at $+37^{\circ}\text{C}$** .

Important notes:

- a. *Strips have to be sealed with the adhesive sealing foil, only when the test is performed manually. Do not cover strips when using ELISA automatic instruments.*
- b. *If the procedure is carried out on shaking, be sure to deliver the rpm reported for in Section I.3 as otherwise intra-well contamination could occur.*
7. When the first incubation is over, wash the microwells as previously described (section I.4)

8. Pipette 200 µl Chromogen/Substrate into all the wells, A1 included.

Important note: Do not expose to strong direct light as a high background might be generated.

9. Incubate the microplate protected from light at **18-24°C for 30 min**. Wells dispensed with the positive control, the calibrator and positive samples will turn from clear to blue.
10. Pipette 100 µl Sulphuric Acid into all the wells to stop the enzymatic reaction, using the same pipetting sequence as in step 8. Addition of the acid solution will turn the positive control, the calibrator and positive samples from blue to yellow/brown.
11. Measure the color intensity of the solution in each well, as described in section I.6 using a 450nm filter (reading) and a 620-630nm filter (background subtraction, strongly recommended), blanking the instrument on A1.

Important general notes:

- If the second filter is not available, ensure that no fingerprints or dust are present on the external bottom of the microwell before reading at 450nm. They could generate false positive results on reading.
- Reading should ideally be performed immediately after the addition of the acid solution but definitely no longer than 20 minutes afterwards. Some self-oxidation of the chromogen can occur leading to a higher background.
- When samples to be tested are not surely clean or have been stored frozen, the assay procedure reported below is recommended as long as it is far less sensitive to interferences due to hemolysis, hyperlipaemia, bacterial contamination and fibrin microparticles. The assay is carried out in two-steps at +37°C on shaking at 350 rpm ±150 as follows:
 - dispense 100 µl of controls, calibrator and samples
 - incubate 60 min at +37°C on shaking
 - wash according to instructions (section I.4)
 - dispense 100 µl diluted enzyme tracer
 - incubate 30 min at +37°C on shaking
 - wash
 - dispense 100 µl TMB&H₂O₂ mix
 - incubate 30 min at r.t. on shaking
 - stop and read

In this procedure the pre-wash can be omitted. This method shows performances similar to the standard one and therefore can be used in alternative.
- The Calibrator (CAL) does not affect the cut-off calculation and therefore the test results calculation. The Calibrator may be used only when a laboratory internal quality control is required by the management.

N. ASSAY SCHEME

Operations	Procedure
Pre-Washing step	n° 1 cycle
Controls&Calibrator&samples	150 µl
Diluted Enzyme Conjugate	100 µl
1st incubation	120 min
Temperature	+37°C
Washing steps	n° 4-5
Chromogen/Substrate	200µl
2nd incubation	30 min
Temperature	room
Sulphuric Acid	100 µl
Reading OD	450nm

An example of dispensation scheme is reported in the following section:

Microplate

	1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	S2										
B	NC	S3										
C	NC	S4										
D	NC	S5										
E	CAL	S6										
F	CAL	S7										
G	PC	S8										
H	S1	S9										

Legenda: BLK = Blank NC = Negative Control
CAL = Calibrator PC = Positive Control S = Sample

O. INTERNAL QUALITY CONTROL

A check is performed on the controls/calibrator any time the kit is used in order to verify whether the expected OD450nm or S/Co values have been matched in the analysis.

Ensure that the following results are met:

Parameter	Requirements
Blank well	< 0.100 OD450nm value
Negative Control (NC)	< 0.050 mean OD450nm value after blanking
Calibrator 0.5 IU/ml	S/Co ≥ 2
Positive Control	> 1.000 OD450nm value

If the results of the test match the requirements stated above, proceed to the next section.

If they do not, do not proceed any further and perform the following checks:

Problem	Check
Blank well > 0.100 OD450nm	1. that the Chromogen/Substrate solution has not become contaminated during the assay
Negative Control (NC) > 0.050 OD450nm after blanking	1. that the washing procedure and the washer settings are as validated in the pre qualification study; 2. that the proper washing solution has been used and the washer has been primed with it before use; 3. that no mistake has been done in the assay procedure (dispensation of positive control instead of the negative one); 4. that no contamination of the negative control or of the wells where the control was dispensed has occurred due to spills of positive samples or of the enzyme conjugate; 5. that micropipettes have not become contaminated with positive samples or with the enzyme conjugate 6. that the washer needles are not blocked or partially obstructed.

Calibrator S/Co < 2	<ol style="list-style-type: none"> 1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution (ex.: dispensation of negative control instead of calibrator) 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the calibrator has occurred.
Positive Control < 1.000 OD450nm	<ol style="list-style-type: none"> 1. that the procedure has been correctly performed; 2. that no mistake has occurred during the distribution of the control (dispensation of negative control instead of positive control. In this case, the negative control will have an OD450nm value > 0.050). 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the positive control has occurred.

If any of the above problems have occurred, report the problem to the supervisor for further actions.

P. CALCULATION OF THE CUT-OFF

The test results are calculated by means of a cut-off value determined on the mean OD450nm value of the negative control (NC) with the following formula:

$$NC + 0.050 = \text{Cut-Off (Co)}$$

The value found for the test is used for the interpretation of results as described in the next paragraph.

Important note: When the calculation of results is performed by the operating system of an ELISA automated work station, ensure that the proper formulation is used to calculate the cut-off value and generate the correct interpretation of results.

Q. INTERPRETATION OF RESULTS

Test results are interpreted as a ratio of the sample OD450nm (S) and the Cut-Off value (Co), mathematically S/Co, according to the following table:

S/Co	Interpretation
< 0.9	Negative
0.9 – 1.1	Equivocal
> 1.1	Positive

A negative result indicates that the patient is not infected by HBV and that the blood unit may be transfused.

Any patient showing an equivocal result should be retested on a second sample taken 1-2 weeks after the initial sample; the blood unit should not be transfused.

A positive result is indicative of HBV infection and therefore the patient should be treated accordingly or the blood unit should be discarded.

Important notes:

1. Interpretation of results should be done under the supervision of the laboratory supervisor to reduce the risk of judgment errors and misinterpretations.
2. Any positive result must be confirmed first by repeating the test on the sample, after having filtered it on 0.2-0.8 µ filter to remove any microparticles interference. Then, if still

positive, the sample has to be submitted to a confirmation test before a diagnosis of viral hepatitis is released.

3. When test results are transmitted from the laboratory to another department, attention must be paid to avoid erroneous data transfer.
4. Diagnosis of viral hepatitis infection has to be taken and released to the patient by a suitably qualified medical doctor.

An example of calculation is reported below.

The following data must not be used instead of real figures obtained by the user.

Negative Control: 0.012 – 0.008 – 0.010 OD450nm
Mean Value: 0.010 OD450nm
Lower than 0.050 – Accepted
Positive Control: 2.489 OD450nm
Higher than 1.000 – Accepted
Cut-Off = 0.010+0.050 = 0.060
Calibrator: 0.350 - 0.370 OD450nm
Mean value: 0.360 OD450nm S/Co = 6.0
S/Co higher than 2.0 – Accepted
Sample 1: 0.028 OD450nm
Sample 2: 1.690 OD450nm
Sample 1 S/Co < 0.9 = negative
Sample 2 S/Co > 1.1 = positive

R. PERFORMANCE CHARACTERISTICS

Evaluation of Performances has been conducted in accordance to what reported in the Common Technical Specifications or CTS (art. 5, Chapter 3 of IVD Directive 98/79/EC). Version ULTRA proved to be at least equivalent to the original design in a study conducted for the validation of the new version.

1. Analytical Sensitivity

The limit of detection of the assay has been calculated on the 2nd WHO international standard, NIBSC code 00/588.

In the following table, results are given for three lots (P1, P2 and P3) of the version ULTRA in comparison with the reference device (Ref.):

WHO IU/ml	Lot # P1 S/Co	Lot # P2 S/Co	Lot # P3 S/Co	Ref. S/Co
0.4	4.6	4.8	4.6	4.6
0.2	2.3	2.4	2.4	2.4
0.1	1.4	1.4	1.5	1.2
0.05	0.8	0.8	1.0	0.7
0.025	0.6	0.6	0.6	0.4
FCS (NC)	0.3	0.2	0.3	0.1

The assay shows an Analytical Sensitivity better than 0.1 WHO IU/ml of HBsAg.

In addition two panels of sensitivity supplied by EFS, France, and by SFTS, France, were tested and gave in the best conditions the following results:

Panel EFS Ag HBs HB1-HB6 lot n° 04

Sample ID	Characteristics	ng/ml	S/Co
HB1	diluent	/	0,2
HB2	adw2+ayw3	0,05	0,6
HB3	adw2+ayw3	0,1	1,0
HB4	adw2+ayw3	0,2	1,8
HB5	adw2+ayw3	0,3	2,4
HB6	adw2+ayw3	0,5	4,2

Sensitivity panel SFTS, France, Ag HBs 2005

Sample ID	Characteristics	ng/ml	S/Co
171	Adw2 + ayw3	2.21 ± 0.15	15,4
172	Adw2 + ayw3	1.18 ± 0.10	8,7
173	Adw2 + ayw3	1.02 ± 0.05	6,1
174	Adw2 + ayw3	0.64 ± 0.04	4,0
175	Adw2 + ayw3	0.49 ± 0.03	3,4
176	Adw2 + ayw3	0.39 ± 0.02	2,6
177	Adw2 + ayw3	0.25 ± 0.02	2,0
178	Adw2 + ayw3	0.11 ± 0.02	1,3
179	Adw2 + ayw3	0.06 ± 0.01	0,9
180	Adw2 + ayw3	0.03 ± 0.01	0,8
181	Adw2	0.5 – 1.0	4,7
182	Adw4	0.5 – 1.0	3,6
183	Adr	0.5 – 1.0	4,5
184	Ayw1	0.5 – 1.0	5,1
185	Ayw2	0.5 – 1.0	6,4
186	Ayw3	0.5 – 1.0	7,3
187	Ayw3	0.5 – 1.0	5,8
188	Ayw4	0.5 – 1.0	6,9
189	Ayr	0.5 – 1.0	6,1
190	diluent	/	0,6

The panel # 808, supplied by Boston Biomedical Inc., USA, was also tested to define the limit of sensitivity. Results in the best conditions are as follows :

BBI panel PHA 808

Sample ID	Characteristics	ng/ml	S/Co
01	ad	2,49	10,2
02	ad	1,17	4,8
03	ad	1,02	4,3
04	ad	0,96	3,8
05	ad	0,69	2,9
06	ad	0,50	2,2
07	ad	0,41	1,5
08	ad	0,37	1,3
09	ad	0,30	1,2
10	ad	0,23	1,0
11	ay	2,51	11,2
12	ay	1,26	5,9
13	ay	0,97	4,1
14	ay	0,77	3,7
15	ay	0,63	2,0
16	ay	0,48	2,4
17	ay	0,42	2,0
18	ay	0,33	1,8
19	ay	0,23	1,6
20	ay	0,13	1,1
21	negative	/	0,6

2. Diagnostic Sensitivity:

The diagnostic sensitivity was tested according to what required by Common Technical Specifications (CTS) of the directive 98/79/EC on IVD for HBsAg testing.

Positive samples, including HBsAg subtypes and a panel of "s" mutants from most frequent mutations, were collected from different HBV pathologies (acute, a-symptomatic and chronic hepatitis B) or produced synthetically, and were detected positive in the assay.

All the HBsAg known subtypes, "ay" and "ad", and isoforms "w" and "r", supplied by CNTS, France, were tested in the assay and determined positive by the kit as expected.

An overall value of 100% has been found in a study conducted on a total number of more than 400 samples positive with the original reference IVD code SAG1.CE, CE marked.

A total of 30 sero-conversions were studied, most of them produced by Boston Biomedica Inc., USA.

Results obtained by examining eight panels supplied by Boston Biomedica Inc., USA, are reported below for the version ULTRA in comparison with the reference device code SAG1.CE.

Panel ID	1 st sample positive	HBsAg subtype	HBsAg ng/ml	Version ULTRA S/Co	Ref. device S/Co
PHM 906	02	ad	0.5	3.7	1.4
PHM 907 (M)	06	ay	1.0	4.4	2.9
PHM 909	04	ad	0.3	1.2	0.8
PHM 914	04	ad	0.5	1.1	1.1
PHM 918	02	ad	0.1	1.8	0.5
PHM 923	03	ay	< 0.2	2.2	1.2
PHM 925	03	Ind.	n.d.	1.4	0.9
PHM 934	01	ad	n.d.	1.0	0.8

3. Diagnostic Specificity:

It is defined as the probability of the assay of scoring negative in the absence of specific analyte. In addition to the first study, where more than 5000 negative samples from blood donors (two blood centers), classified negative with a CE marked device in use at the laboratory of collection were examined, the diagnostic specificity was recently assessed by testing a total of 2288 negative blood donors on seven different lots. A value of specificity of 100% was found.

Both plasma, derived with different standard techniques of preparation (citrate, EDTA and heparin), and sera have been used to determine the specificity.

No false reactivity due to the method of specimen preparation has been observed.

Frozen specimens have also been tested to check whether samples freezing interferes with the performance of the test. No interference was observed on clean and particle free samples.

Samples derived from patients with different viral (HCV, HAV) and non viral pathologies of the liver that may interfere with the test were examined. No cross reaction were observed.

4. Precision:

It has been calculated for the version ULTRA on two samples examined in 16 replicates in 3 different runs for three lots.

Results are reported in the following tables:

Average values	Negative Sample	Calibrator 0.5 IU/ml
Total n = 144	0.026	0.332
Std.Deviation	0.004	0.027
CV %	16%	8%

The variability shown in the tables did not result in sample misclassification.

S. LIMITATIONS

Repeatable false positive results were assessed on freshly collected specimens in less than 0.1% of the normal population, mostly due to high titers Heterophilic Anti Mouse Antibodies (HAMA).

Interferences in fresh samples were also observed when they were not particles-free or were badly collected (see chapter G).

Old or frozen samples, presenting fibrin clots, cryoglobulins, lipid-containing micelles or microparticles after storage or thawing, can generate false positive results.

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All the IVD Products manufactured by the company are under the control of a certified Quality Management System approved by an EC Notified Body. Each lot is submitted to a quality control and released into the market only if conforming with the EC technical specifications and acceptance criteria.

Manufacturer:
Dia.Pro Diagnostic Bioprobes S.r.l.
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CE
0318

HBsAb

**Enzyme Immunoassay for
qualitative/quantitative determination of
antibodies to Hepatitis B surface Antigen
in human serum and plasma**

- for "in vitro" diagnostic use only -



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REF SAB.CE
96 Tests

HBs Ab

A. INTENDED USE

Enzyme ImmunoAssay (ELISA) for both the quantitative and qualitative determination of antibodies to the Surface Antigen of Hepatitis B Virus in human plasma and sera.
For "in vitro" diagnostic use only.

B. INTRODUCTION

The World Health Organization (WHO) defines Hepatitis B Virus infection as follows:

"Hepatitis B is one of the major diseases of mankind and is a serious global public health problem. Hepatitis means inflammation of the liver, and the most common cause is infection with one of 5 viruses, called hepatitis A,B,C,D, and E. All of these viruses can cause an acute disease with symptoms lasting several weeks including yellowing of the skin and eyes (jaundice); dark urine; extreme fatigue; nausea; vomiting and abdominal pain. It can take several months to a year to feel fit again. Hepatitis B virus can cause chronic infection in which the patient never gets rid of the virus and many years later develops cirrhosis of the liver or liver cancer.

HBV is the most serious type of viral hepatitis and the only type causing chronic hepatitis for which a vaccine is available. Hepatitis B virus is transmitted by contact with blood or body fluids of an infected person in the same way as human immunodeficiency virus (HIV), the virus that causes AIDS. However, HBV is 50 to 100 times more infectious than HIV. The main ways of getting infected with HBV are: (a) perinatal (from mother to baby at the birth); (b) child-to-child transmission; (c) unsafe injections and transfusions; (d) sexual contact.

Worldwide, most infections occur from infected mother to child, from child to child contact in household settings, and from reuse of un-sterilized needles and syringes. In many developing countries, almost all children become infected with the virus. In many industrialized countries (e.g. Western Europe and North America), the pattern of transmission is different. In these countries, mother-to-infant and child-to-child transmission accounted for up to one third of chronic infections before childhood hepatitis B vaccination programmes were implemented. However, the majority of infections in these countries are acquired during young adulthood by sexual activity, and injecting drug use. In addition, hepatitis B virus is the major infectious occupational hazard of health workers, and most health care workers have received hepatitis B vaccine.

Hepatitis B virus is not spread by contaminated food or water, and cannot be spread casually in the workplace. High rates of chronic HBV infection are also found in the southern parts of Eastern and Central Europe. In the Middle East and Indian sub-continent, about 5% are chronically infected. Infection is less common in Western Europe and North America, where less than 1% are chronically infected.

Young children who become infected with HBV are the most likely to develop chronic infection. About 90% of infants infected during the first year of life and 30% to 50% of children infected between 1 to 4 years of age develop chronic infection. The risk of death from HBV-related liver cancer or

cirrhosis is approximately 25% for persons who become chronically infected during childhood.

Chronic hepatitis B in some patients is treated with drugs called *interferon or lamivudine*, which can help some patients. Patients with cirrhosis are sometimes given liver transplants, with varying success. It is preferable to prevent this disease with vaccine than to try and cure it.

Hepatitis B vaccine has an outstanding record of safety and effectiveness. Since 1982, over one billion doses of hepatitis B vaccine have been used worldwide. The vaccine is given as a series of three intramuscular doses. Studies have shown that the vaccine is 95% effective in preventing children and adults from developing chronic infection if they have not yet been infected. In many countries where 8% to 15% of children used to become chronically infected with HBV, the rate of chronic infection has been reduced to less than 1% in immunized groups of children. Since 1991, WHO has called for all countries to add hepatitis B vaccine into their national immunization programmes."

Hepatitis B surface Antigen (HBsAg) is the major structural polypeptide of the envelope of the Hepatitis B Virus (HBV).

This antigen is composed mainly of the type common determinant "a" and the type specific determinants "d" and "y", present only on the specific serotypes.

Upon infection, a strong immunological response develops firstly against the type specific determinants and in a second time against the "a" determinant.

Anti "a" antibodies are however recognised to be most effective in the neutralisation of the virus, protecting the patient from other infections and leading it to convalescence.

The detection of HBsAb has become important for the follow up of patients infected by HBV and the monitoring of recipients upon vaccination with synthetic and natural HBsAg.

C. PRINCIPLE OF THE TEST

Microplates are coated with a preparation of highly purified HBsAg that in the first incubation with sample specifically captures anti HBsAg antibodies to the solid phase.

After washing, captured antibodies are detected by an HBsAg, labelled with peroxidase (HRP), that specifically binds the second available binding site of these antibodies.

The enzyme specifically bound to wells, by acting on the substrate/chromogen mixture, generates an optical signal that is proportional to the amount of HBsAb in the sample and can be detected by an ELISA reader.

The amount of antibodies may be quantitated by means of a standard curve calibrated against the W.H.O reference preparation.

Samples are pre treated in the well with an specimen diluent able to block interference present in vaccinated individuals.

D. COMPONENTS

Each kit contains sufficient reagents to perform 96 tests.

1. Microplate: MICROPLATE

8x12 microwell strips coated with purified heat-inactivated HBsAg of both subtypes (ad and ay) from human origin and sealed into a bag with desiccant.

Allow the microplate to reach room temperature before opening; reseal unused strips in the bag with desiccant and store at 4°C.

2. Calibration Curve: **CAL N° ...**

5x2.0 ml/vial. Ready to use and colour coded standard curve, derived from HBsAg positive plasma titrated on WHO standard for anti HBsAg (1st reference preparation 1977, lot 17-2-77), ranging: CAL1 = 0 mIU/ml // CAL2 = 10 mIU/ml // CAL3 = 50 mIU/ml // CAL4 = 100 mIU/ml // CAL5 = 250 mIU/ml. Contains human serum proteins, 5% BSA, 10 mM phosphate buffer pH 7.4+/-0.1, 0.09% sodium azide and 0.1% Kathon GC as preservatives. Standards are blue coloured.

3. Wash buffer concentrate: **WASHBUF 20X**

1x60ml/bottle. 20x concentrated solution. Once diluted, the wash solution contains 10 mM phosphate buffer pH 7.0+/-0.2, 0.05% Tween 20 and 0.1% Kathon GC.

4. Enzyme conjugate : **CONJ**

1x16.0 ml/vial. Ready-to-use solution and red color coded. It contains inactivated purified HBsAg of both subtypes ad and ay, labelled with HRP, 5% BSA, 10 mM Tris buffer pH 6.8+/-0.1, 0.3 mg/ml gentamicine sulphate and 0.1% Kathon GC as preservatives.

5. Chromogen/Substrate: **SUBS TMB**

1x16ml/vial. Contains a 50 mM citrate-phosphate buffered solution at pH 3.5-3.8, 4% dimethylsulphoxide, 0.03% tetramethyl-benzidine (TMB) and 0.02% hydrogen peroxide (H₂O₂).
Note: To be stored protected from light as sensitive to strong illumination.

6. Sulphuric Acid: **H₂SO₄ 0.3 M**

1x15ml/vial. Contains 0.3 M H₂SO₄ solution. Attention: Irritant (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

7. Specimen Diluent: **DILSPE**

1x8ml. 10 mM Tris Buffered solution pH 7.4 +/-0.1, suggested to be used in the follow up of vaccination. It contains 0.09% sodium azide as preservatives.

8. Control Serum: **CONTROL ...ml**

1 vial. Lyophilized. Contains fetal bovine serum proteins, human anti HBsAg antibodies calibrated at 50 ± 10% WHO mIU/ml. 0.3 mg/ml gentamicine sulphate and 0.1% Kathon GC as preservatives.

9. Plate sealing foil n° 2

10. Package insert n° 1

E. MATERIALS REQUIRED BUT NOT PROVIDED

1. Calibrated Micropipettes (100ul and 50ul) and disposable plastic tips.
2. EIA grade water (double distilled or deionised, charcoal treated to remove oxidizing chemicals used as disinfectants).
3. Timer with 60 minute range or higher.
4. Absorbent paper tissues.
5. Calibrated ELISA microplate thermostatic incubator (dry or wet), set at +37°C (+/-1°C tolerance)..
6. Calibrated ELISA microwell reader with 450nm (reading) and with 620-630nm (blanking, strongly recommended) filters.
7. Calibrated ELISA microplate washer.
8. Vortex or similar mixing tools.

F. WARNINGS AND PRECAUTIONS

1. The kit has to be used by skilled and properly trained technical personnel only, under the supervision of a medical doctor responsible of the laboratory.

2. All the personnel involved in performing the assay have to wear protective laboratory clothes, talc-free gloves and glasses. The use of any sharp (needles) or cutting (blades) devices should be avoided. All the personnel involved should be trained in biosafety procedures, as recommended by the Center for Disease Control, Atlanta, U.S. and reported in the National Institute of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.

3. All the personnel involved in sample handling should be vaccinated for HBV and HAV, for which vaccines are available, safe and effective.

4. The laboratory environment should be controlled so as to avoid contaminants such as dust or air-borne microbial agents, when opening kit vials and microplates and when performing the test. Protect the Chromogen (TMB) from strong light and avoid vibration of the bench surface where the test is undertaken.

5. Upon receipt, store the kit at 2..8°C into a temperature controlled refrigerator or cold room.

6. Do not interchange components between different lots of the kits. It is recommended that components between two kits of the same lot should not be interchanged.

7. Check that the reagents are clear and do not contain visible heavy particles or aggregates. If not, advise the laboratory supervisor to initiate the necessary procedures for kit replacement.

8. Avoid cross-contamination between serum/plasma samples by using disposable tips and changing them after each sample.

9. Avoid cross-contamination between kit reagents by using disposable tips and changing them between the use of each one.

10. Do not use the kit after the expiration date stated on the external container and internal (vials) labels. A study conducted on an opened kit did not pointed out any relevant loss of activity up to six 6 uses of the device and up to 6 months.

11. Treat all specimens as potentially infective. All human serum specimens should be handled at Biosafety Level 2, as recommended by the Center for Disease Control, Atlanta, U.S. in compliance with what reported in the Institutes of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.

12. The use of disposable plastic-ware is recommended in the preparation of the liquid components or in transferring components into automated workstations, in order to avoid cross contamination.

13. Waste produced during the use of the kit has to be discarded in compliance with national directives and laws concerning laboratory waste of chemical and biological substances. In particular, liquid waste generated from the washing procedure, from residuals of controls and from samples has to be treated as potentially infective material and inactivated before waste. Suggested procedures of inactivation are treatment with a 10% final concentration of household bleach for 16-18 hrs or heat inactivation by autoclave at 121°C for 20 min..

14. Accidental spills from samples and operations have to be adsorbed with paper tissues soaked with household bleach and then with water. Tissues should then be discarded in proper containers designated for laboratory/hospital waste.

15. The Sulphuric Acid is an irritant. In case of spills, wash the surface with plenty of water

16. Other waste materials generated from the use of the kit (example: tips used for samples and controls, used microplates) should be handled as potentially infective and disposed according to national directives and laws concerning laboratory wastes.

G. SPECIMEN: PREPARATION AND WARNINGS

1. Blood is drawn aseptically by venipuncture and plasma or serum is prepared using standard techniques of preparation of samples for clinical laboratory analysis. No influence has been observed in the preparation of the sample with citrate, EDTA and heparin.

2. Samples have to be clearly identified with codes or names in order to avoid misinterpretation of results. Bar code labeling and electronic reading is strongly recommended.

3. Haemolysed ("red") and visibly hyperlipemic ("milky") samples have to be discarded as they could generate false results. Samples containing residues of fibrin or heavy particles or microbial filaments and bodies should be discarded as they could give rise to false results.

4. Sera and plasma can be stored at +2°.8°C for up to five days after collection. For longer storage periods, samples can be stored frozen at -20°C for several months. Any frozen samples should not be freeze/thawed more than once as this may generate particles that could affect the test result.

5. If particles are present, centrifuge at 2.000 rpm for 20 min or filter using 0.2-0.8µ filters to clean up the sample for testing.

6. Samples whose anti-HBsAg antibody concentration is expected to be higher than 250 mIU/ml should be diluted before use either 1:10 or 1:100 in the Calibrator 0 mIU/ml. Dilutions have to be done in clean disposable tubes by diluting 50 µl of each specimen with 450 µl of Cal 0 (1:10). Then 50 µl of the 1:10 dilution are diluted with 450 µl of the Cal 0 (1:100). Mix tubes thoroughly on vortex when preparing the diluted samples.

H. PREPARATION OF COMPONENTS AND WARNINGS

1. Microplate:

Allow the microplate to reach room temperature (about 1 hr) before opening the container. Check that the desiccant has not turned green, indicating a defect in conservation.

In this case, call Dia.Pro's customer service.

Unused strips have to be placed back into the aluminum pouch, with the desiccant supplied, firmly zipped and stored at +2°-8°C. After first opening, remaining strips are stable until the humidity indicator inside the desiccant bag turns from yellow to green.

2. Calibration Curve

Ready to use. Mix well on vortex before use.

3. Control Serum

Add the volume of ELISA grade water, reported on the label, to the lyophilised powder; let fully dissolve and then gently mix on vortex.

Note: *The control after dissolution is not stable. Store frozen in aliquots at -20°C.*

4. Wash buffer concentrate:

The whole content of the concentrated solution has to be diluted 20x with bidistilled water and mixed gently end-over-end before use. During preparation avoid foaming as the presence of bubbles could impact on the efficiency of the washing cycles.

Note: *Once diluted, the wash solution is stable for 1 week at +2..8° C.*

5. Enzyme conjugate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidising chemicals, dust or microbes. If this component has to be transferred, use only plastic, and if possible, sterile disposable containers.

6. Specimen Diluent:

Ready to use. Mix well on vortex before use.

7. Chromogen/Substrate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidising chemicals, air-driven dust or microbes. Do not expose to strong light, oxidising agents and metallic surfaces.

If this component has to be transferred use only plastic, and if possible, sterile disposable container

8. Sulphuric Acid:

Ready to use. Mix well on vortex before use.

Attention: Irritant (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Legenda:

Warning **H statements:**

H315 – Causes skin irritation.

H319 – Causes serious eye irritation.

Precautionary **P statements:**

P280 – Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352 – IF ON SKIN: Wash with plenty of soap and water.

P332 + P313 – If skin irritation occurs: Get medical advice/attention.

P305 + P351 + P338 – IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P337 + P313 – If eye irritation persists: Get medical advice/attention.

P362 + P363 – Take off contaminated clothing and wash it before reuse.

I. INSTRUMENTS AND TOOLS USED IN COMBINATION WITH THE KIT

1. Micropipettes have to be calibrated to deliver the correct volume required by the assay and must be submitted to regular decontamination (70% ethanol, 10% solution of bleach, hospital grade disinfectants) of those parts that could accidentally come in contact with the sample or the components of the kit. They should also be regularly maintained in order to show a precision of 1% and a trueness of ±2%.

2. The ELISA incubator has to be set at +37°C (tolerance of ±1°C) and regularly checked to ensure the correct temperature is maintained. Both dry incubators and water baths are suitable for the incubations, provided that the instrument is validated for the incubation of ELISA tests.

3. The ELISA washer is extremely important to the overall performances of the assay. The washer must be carefully validated and correctly optimized using the kit controls/calibrator and reference panels, before using the kit for routine laboratory tests. Usually 4-5 washing cycles (aspiration + dispensation of 350µl/well of washing solution = 1 cycle) are sufficient to ensure that the assay performs as expected. A soaking time of 20-30 seconds between cycles is suggested. In order to set correctly their number, it is recommended to run an assay with the kit controls/calibrator and well-characterized negative and positive reference samples, and check to match the values reported below in the sections "Validation of Test" and "Assay Performances". Regular calibration of the volumes delivered and maintenance (decontamination and cleaning of needles) of the washer has to be carried out according to the instructions of the manufacturer.

4. Incubation times have a tolerance of ±5%.

5. The ELISA microplate reader has to be equipped with a reading filter of 450nm and with a second filter (620-630nm, strongly recommended) for blanking purposes. Its standard performances should be (a) bandwidth ≤ 10 nm; (b) absorbance range from 0 to ≥ 2.0; (c) linearity to ≥ 2.0; repeatability ≥ 1%. Blanking is carried out on the well identified in the section "Assay Procedure". The optical system of the reader has to be calibrated regularly to ensure that the correct optical density is measured. It should be regularly maintained according to the manufacturer 's instructions.

6. When using an ELISA automated workstation, all critical steps (dispensation, incubation, washing, reading, shaking, data handling) have to be carefully set, calibrated, controlled and regularly serviced in order to match the

values reported in the sections "Validation of Test" and "Assay Performances". The assay protocol has to be installed in the operating system of the unit and validated as for the washer and the reader. In addition, the liquid handling part of the station (dispensation and washing) has to be validated and correctly set. Particular attention must be paid to avoid carry over by the needles used for dispensing samples and for washing. This must be studied and controlled to minimize the possibility of contamination of adjacent wells due to strongly reactive samples, leading to false positive results. The use of ELISA automated work stations is recommended for blood screening and when the number of samples to be tested exceed 20-30 units per run.

7. Dia.Pro's customer service offers support to the user in the setting and checking of instruments used in combination with the kit, in order to assure full compliance with the requirements described. Support is also provided for the installation of new instruments to be used with the kit.

L. PRE ASSAY CONTROLS AND OPERATIONS

1. Check the expiration date of the kit printed on the external label of the kit box. Do not use if expired.
2. Check that the liquid components are not contaminated by naked-eye visible particles or aggregates. Check that the Chromogen/Substrate is colorless or pale blue by aspirating a small volume of it with a sterile transparent plastic pipette. Check that no breakage occurred in transportation and no spillage of liquid is present inside the box. Check that the aluminum pouch, containing the microplate, is not punctured or damaged.
3. Dilute all the content of the 20x concentrated Wash Solution as described above.
4. Dissolve the Control Serum as described above.
5. Allow all the other components to reach room temperature (about 1 hr) and then mix as described.
6. Set the ELISA incubator at +37°C and prepare the ELISA washer by priming with the diluted washing solution, according to the manufacturers instructions. Set the right number of washing cycles as found in the validation of the instrument for its use with the kit.
7. Check that the ELISA reader has been turned on at least 20 minutes before reading.
8. If using an automated workstation, turn it on, check settings and be sure to use the right assay protocol.
9. Check that the micropipettes are set to the required volume.
10. Check that all the other equipments are available and ready to use.

In case of problems, do not proceed further with the test and advise the supervisor.

M. ASSAY PROCEDURE

The assay has to be carried out according to what reported below, taking care to maintain the same incubation time for all the samples in testing.

Two procedures can be carried out with the device according to the request of the clinician.

M.1 Quantitative analysis

1. Place the required number of strips in the microplate holder. Leave A1 and B1 wells empty for the operation of blanking. Store the other strips into the bag in presence of the desiccant at 2..8°C, sealed. Then Dispense in all the wells to be used for the test, except for A1 and B1, 50µl of the Specimen Diluent.

Important note: This additive is added before distributing samples and controls into specific wells and is particularly

intended for blocking some substances present in people undergoing vaccination and capable to mask antibodies.

2. Pipette 100µl of all the Calibrators, 100µl of Control Serum in duplicate and then 100ul of samples. The Control Serum is used to verify that the whole analytical system works as expected. Check that Calibrators, Control Serum and samples have been correctly added. Then incubate the microplate at **+37°C for 60 min.**

Important note: Strips have to be sealed with the adhesive sealing foil only when the test is performed manually. Do not cover strips when using ELISA automatic instruments.

3. Wash the microplate as reported in section I.3.
4. In all the wells except A1 and B1, pipette 100 µl Enzyme Conjugate. Check that the reagent has been correctly added. Incubate the microplate at **+37°C for 60 minutes.**

Important note:

- 1) *Be careful not to touch the inner surface of the well with the pipette tip when dispensing the Enzyme Conjugate. Contamination might occur.*
- 2) *Mix thoroughly the Enzyme Conjugate on vortex before use.*

5. Wash the microplate as described.
6. Pipette 100µl TMB/H₂O₂ mixture in each well, the blank wells included. Check that the reagent has been correctly added. Then incubate the microplate at **room temperature for 20 minutes.**

Important note: Do not expose to strong direct light as a high background might be generated.

7. Stop the enzymatic reaction by pipette 100µl Sulphuric Acid into each well and using the same pipetting sequence as in step 6. Then measure the colour intensity with a microplate reader at 450nm (reading) and at 620-630nm (blanking, strongly recommended), blanking the instrument on A1 and B1 wells.

M.2 Qualitative analysis

1. Place the required number of strips in the microplate holder. Leave A1 well empty for the operation of blanking. Store the other strips into the bag in presence of the desiccant at 2..8°C, sealed.
2. Dispense 50 ul Specimen Diluent in all the wells, except for the blank A1. Then pipette 100µl of the Calibrator 0 mIU/ml in duplicate, 100µl of the Calibrator 10 mIU/ml in duplicate, 100µl of the Calibrator 250 mIU/ml in single, and then 100ul of samples. Check that Calibrators and samples have been correctly added. Then incubate the microplate at **+37°C for 60 min.**
3. Wash the microplate as reported in section I.3.
4. In all the wells except A1, pipette 100 µl Enzyme Conjugate. Check that the reagent has been correctly added. Incubate the microplate at **+37°C for 60 minutes.**

Important note:

- 3) *Be careful not to touch the inner surface of the well with the pipette tip when dispensing the Enzyme Conjugate. Contamination might occur.*
- 4) *Mix thoroughly the Enzyme Conjugate on vortex before use.*

5. Wash the microplate as described.
6. Pipette 100µl TMB/H₂O₂ mixture in each well, the blank wells included. Check that the reagent has been correctly added. Then incubate the microplate at **room temperature for 20 minutes.**

Important note: Do not expose to strong direct light as a high background might be generated.

7. Stop the enzymatic reaction by pipette 100µl Sulphuric Acid into each well and using the same pipetting sequence as in step 6. Then measure the colour intensity with a microplate reader at 450nm (reading) and at 620-630nm (blinking, strongly recommended), blanking the instrument on A1 and B1 wells.

Important general notes:

1. If the second filter is not available, ensure that no finger prints are present on the bottom of the microwell before reading at 450nm. Finger prints could generate false positive results on reading.
2. Reading has should ideally be performed immediately after the addition of the Stop Solution but definitely no longer than 20 minutes afterwards. Some self oxidation of the chromogen can occur leading to a higher background.
3. The Control Serum (CS) does not affect the cut-off calculation and therefore the test results calculation. The Control Serum may be used only when a laboratory internal quality control is required by the management.

N. ASSAY SCHEME (standard procedure)

Specimen Diluent	50 ul
Calibrators	100 ul
Control Serum	100 ul
Samples	100 ul
1st incubation	60 min
Temperature	+37°C
Wash step	4-5 cycles
Enzyme Conjugate	100 ul
2nd incubation	60 min
Temperature	+37°C
Wash step	4-5 cycles
TMB/H2O2 mix	100 ul
3rd incubation	20 min
Temperature	r.t.
Sulphuric Acid	100 ul
Reading OD	450nm & 620nm

An example of dispensation scheme in quantitative assays is reported below:

		Microplate											
		1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	CAL4	S3										
B	BLK	CAL4	S4										
C	CAL1	CAL5	S5										
D	CAL1	CAL5	S6										
E	CAL2	CS	S7										
F	CAL2	CS	S8										
G	CAL3	S1	S9										
H	CAL3	S2	S10										

Legenda: BLK = Blank // CAL = Calibrators // CS = Control Serum // S = Sample

An example of dispensation scheme in qualitative assays is reported below:

		Microplate											
		1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	S3	S11										
B	CAL1	S4	S12										
C	CAL1	S5	S13										
D	CAL2	S6	S14										
E	CAL2	S7	S15										
F	CAL5	S8	S16										
G	S1	S9	S17										
H	S2	S10	S18										

Legenda: BLK = Blank // CAL = Calibrators // S = Sample

O. INTERNAL QUALITY CONTROL

A validation check is carried out on the controls any time the kit is used in order to verify whether the performances of the assay are as qualified.

Control that the following data are matched:

Parameters	Requirements
Blank well	< 0.100 OD450nm
Calibrator 0 WHO mIU/ml	< 0.200 OD450nm after blanking
Calibrator 10 WHO mIU/ml	OD450nm higher than the OD450nm of the Calibrator 0 mIU/ml + 0.100
Calibrator 250 WHO mIU/ml	> 1.500 OD450nm
Control Serum	OD450nm = OD450nm CAL 50 mIU/ml ± 10%
Coefficient of variation	< 30% for the Calibrator 0 mIU/ml

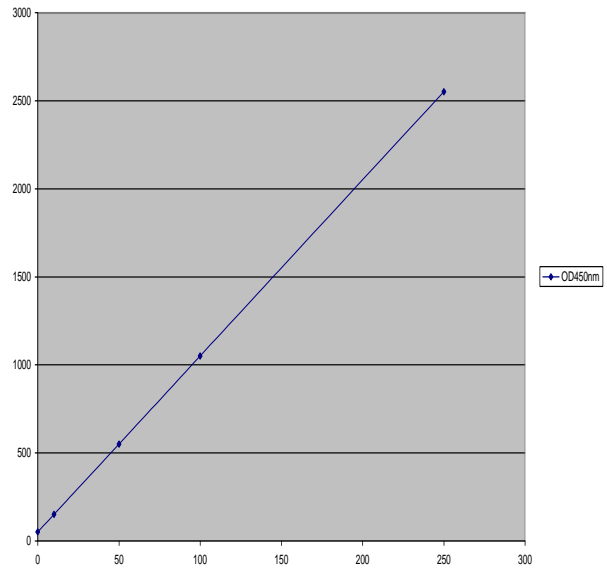
If the results of the test match the requirements stated above, proceed to the next section.

If they do not, do not proceed any further and perform the following checks:

Problem	Check
Blank well > 0.100 OD450nm	1. that the Chromogen/Substrate solution has not become contaminated during the assay
Calibrator 0 mIU/ml > 0.200	1. that the washing procedure and the washer settings are as validated in the pre qualification study;
coefficient of variation > 30%	2. that the proper washing solution has been used and the washer has been primed with it before use;
	3. that no mistake has been done in the assay procedure when the dispensation of standards is carried out;
	4. that no contamination of the Cal 0 mIU/ml or of the wells where it was dispensed has occurred due to positive samples, to spills or to the enzyme conjugate;
	5. that micropipettes have not become contaminated with positive samples or with the enzyme conjugate
	6. that the washer needles are not blocked or partially obstructed.

Calibrator 10 mIU/ml OD450nm < Cal 0 + 0.100	1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution (ex.: dispensation of a wrong calibrator); 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the standard has occurred.
Calibrator 250 mIU/ml OD450nm < 1.500 OD450nm	1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution; 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the standard has occurred.
Control Serum Different from expected value	First verify that: 1. the procedure has been correctly performed; 2. no mistake has occurred during its distribution (ex.: dispensation of a wrong sample); 3. the washing procedure and the washer settings are correct; 4. no external contamination of the standard has occurred. 5. the Control Serum has been dissolved with the right volume reported on the label. If a mistake has been pointed out, the assay has to be repeated after eliminating the reason of this error. If no mistake has been found, proceed as follows: a) a value up to +/-20% is obtained: the overall Precision of the laboratory might not enable the test to match the expected value +/-10%. Report the problem to the Supervisor for acceptance or refusal of this result. b) a value higher than +/-20% is obtained: in this case the test is invalid and the DiaPro's customer service has to be called.

Example of Calibration Curve :



Important Note:

Do not use the calibration curve above to make calculations.

P.2 Qualitative method

In the qualitative method, calculate the mean OD450nm values for the Calibrators 0 and 10 mIU/ml and then check that the assay is valid.

Example of calculation:

The following data must not be used instead of real figures obtained by the user.

Calibrator 0 mIU/ml: 0.020 – 0.024 OD450nm
 Mean Value: 0.022 OD450nm
 Lower than 0.200 – Accepted

Calibrator 10 mIU/ml: 0.250 – 0.270 OD450nm
 Mean Value: 0.260 OD450nm
 Higher than Cal 0 + 0.100 – Accepted

Calibrator 250 mIU/ml: 2.845 OD450nm
 Higher than 1.500 – Accepted

P. RESULTS

P.1 Quantitative method

If the test turns out to be valid, use for the quantitative method an approved curve fitting program to draw the calibration curve from the values obtained by reading at 450nm (4-parameters interpolation is suggested).

Then on the calibration curve calculate the concentration of anti HBsAg antibody in samples.

An example of Calibration curve is reported in the next page.

Q. INTERPRETATION OF RESULTS

Samples with a concentration lower than 10 WHO mIU/ml are considered negative for anti HBsAg antibody by most of the international medical literature.

Samples with a concentration higher than 10 WHO mIU/ml are considered positive for anti HBsAg antibody.

In the follow up of vaccination recipients, however, the value of 20 WHO mIU/ml is usually accepted by the medical literature as the minimum concentration at which the patient is considered clinically protected against HBV infection.

Important notes:

1. Interpretation of results should be done under the supervision of the laboratory supervisor to reduce the risk of judgement errors and misinterpretations.
2. When test results are transmitted from the laboratory to another facility, attention must be paid to avoid erroneous data transfer.
3. Diagnosis has to be done and released to the patient by a suitably qualified medical doctor.

R. PERFORMANCES

Evaluation of Performances has been conducted in accordance to what reported in the Common Technical Specifications or CTS (art. 5, Chapter 3 of IVD Directive 98/79/EC).

1. LIMIT OF DETECTION:

The limit of detection of the assay has been calculated by means of the HBsAb international preparation supplied by CLB on behalf of WHO (1st reference preparation 1977, lot 17-2-77), on which Calibration Curve has been calibrated. HBV negative serum was used as diluent, as recommended by the supplier. Results of Quality Control are given in the following table:

WHO mIU/ml	SAB.CE Lot # 1002	SAB.CE Lot # 1001	SAB.CE Lot # 1002/2
50	0.933	0.812	0.846
10	0.219	0.192	0.194
5	0.110	0.096	0.104
2.5	0.057	0.058	0.067
Std 0	0.021	0.015	0.023

2. DIAGNOSTIC SPECIFICITY AND SENSITIVITY

A Performance Evaluation has been conducted on a total number of more than 700 samples.

2.1 Diagnostic Specificity

It is defined as the probability of the assay of scoring negative in the absence of specific analyte.

More than 500 negative specimens were tested, internally and externally, against a European company.

A diagnostic specificity of 98.8% was assessed. .

Moreover, diagnostic specificity was assessed by testing 113 potentially interfering specimens (other infectious diseases, patients affected by non viral hepatic diseases, dialysis patients, pregnant women, hemolized, lipemic, etc.) against the European company. A value of specificity of 100% was assessed.

Finally, both human plasma, derived with different standard techniques of preparation (citrate, EDTA and heparin), and human sera have been used to determine the specificity.

No false reactivity due to the method of specimen preparation has been observed.

2.2 Diagnostic Sensitivity

It defined as the probability of the assay of scoring positive in the presence of specific analyte.

106 vaccinated patients were evaluated providing a diagnostic sensitivity of 100%.

More than 100 HBV naturally infected patients were tested, internally and externally, against the European company; a diagnostic sensitivity of 100% was found.

3. PRECISION:

The mean values obtained from a study conducted on three samples of different anti-HBsAg reactivity, examined in 16 replicates in three separate runs is reported below:

SAB.CE: lot # 1202

Calibrator 0 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.038	0.038	0.039	0.039
Std.Deviation	0.003	0.004	0.005	0.004
CV %	8.8	9.5	11.8	10.0

Calibrator 10 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.250	0.243	0.244	0.246
Std.Deviation	0.020	0.023	0.017	0.020
CV %	8.0	9.3	7.0	8.1

Calibrator 250 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.998	3.000	3.259	3.085
Std.Deviation	0.152	0.151	0.158	0.153
CV %	5.1	5.0	4.8	5.0

SAB.CE: lot # 1002

Calibrator 0 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.048	0.048	0.050	0.049
Std.Deviation	0.005	0.004	0.006	0.005
CV %	9.4	8.4	11.5	9.8

Calibrator 10 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.249	0.252	0.242	0.248
Std.Deviation	0.021	0.020	0.023	0.021
CV %	8.3	7.9	9.6	8.6

Calibrator 250 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	3.544	3.653	3.612	3.603
Std.Deviation	0.153	0.176	0.138	0.156
CV %	4.3	4.8	3.8	4.3

SAB.CE: lot # 1002/2

Calibrator 0 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.050	0.051	0.050	0.050
Std.Deviation	0.005	0.006	0.006	0.005
CV %	10.0	10.9	11.9	10.9

Calibrator 10 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.226	0.238	0.239	0.234
Std.Deviation	0.015	0.017	0.018	0.016
CV %	6.5	7.0	7.5	7.0

Calibrator 250 mIU/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	3.526	3.457	3.499	3.494
Std.Deviation	0.137	0.143	0.162	0.147
CV %	3.9	4.1	4.6	4.2

The variability shown in the tables did not result in sample misclassification.

4. ACCURACY

The assay accuracy has been checked by the dilution and recovery tests. Any "hook effect", underestimation likely to happen at high doses of analyte, was ruled out up to 10.000 mIU/ml.

S. LIMITATIONS OF THE PROCEDURE

Bacterial contamination or heat inactivation of the specimen may affect the absorbance values of the samples with consequent alteration of the level of the analyte.

This test is suitable only for testing single samples and not pooled ones.

Diagnosis of an infectious disease should not be established on the basis of a single test result. The patient's clinical history, symptomatology, as well as other diagnostic data should be considered.

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All the IVD Products manufactured by the company are under the control of a certified Quality Management System approved by an EC Notified Body. Each lot is submitted to a quality control and released into the market only if conforming with the EC technical specifications and acceptance criteria.

Manufacturer:
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0318

HDV Ab

**Competitive Enzyme Immunoassay
for the qualitative determination of
antibodies to Hepatitis Delta Virus
in human serum and plasma**

- for "in vitro" diagnostic use only -



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REF DAB.CE
96 Tests

HDV Ab

A. INTENDED USE

Competitive Enzyme ImmunoAssay (ELISA) for the qualitative determination of antibodies to Hepatitis Delta Virus or HDV in human plasma and sera with a "two-steps" methodology.

The kit is used for the follow-up of patients infected by HDV.

For "in vitro" diagnostic use only.

B. INTRODUCTION

The Hepatitis Delta Virus or HDV is a RNA defective virus composed of a core presenting the delta-specific antigen, encapsulated by HBsAg, that requires the helper function of HBV to support its replication.

Infection by HDV occurs in the presence of acute or chronic HBV infection. When acute delta and acute HBV simultaneously occur, the illness becomes severe and clinical and biochemical features may be indistinguishable from those of HBV infection alone. In contrast, a patient with chronic HBV infection can support HDV replication indefinitely, usually with a less severe illness appearing as a clinical exacerbation.

The determination of HDV specific serological markers (HDV Ag, HDV Ab, HDV IgM and HDV IgG) represents in these cases an important tool to the clinician for the classification of the etiological agent, for the follow up of infected patients and their treatment. The detection of HDV total antibodies allows the classification of the illness and the monitoring of the seroconversion event.

C. PRINCIPLE OF THE TEST

Anti-HDV antibodies, if present in the sample, compete with a virus-specific polyclonal IgG, labeled with peroxidase (HRP), for a fixed amount of rec-HDV coated on the microplate. The test is carried out with a two steps incubation competitive system. First the sample is added to the plate and specific anti HDV antibodies bind to the adsorbed antigen. After washing, an enzyme conjugated antibody to HDV is added and binds to the free portion of the antigen coated. After washing a chromogen/substrate mixture is dispensed. The concentration of the bound enzyme on the solid phase becomes inversely proportional to the amount of anti-HDV antibodies in the sample and its activity is detected by the added chromogen/substrate. The concentration of HDV-specific antibodies in the sample is determined by means of a cut-off value that allows for the semi quantitative detection of anti-HDV antibodies.

D. COMPONENTS

Each kit contains sufficient reagents to perform 96 tests.

1. Microplate: **MICROPLATE**

8x12 microwell strips coated with recombinant HDV-specific antigen and sealed into a bag with desiccant. Allow the microplate to reach room temperature before opening; reseal unused strips in the bag with desiccant and store at 4°C.

2. Negative Control: **CONTROL -**

1x2.0ml/vial. Ready to use. Contains goat serum proteins, 100 mM Tris-HCl buffer pH 7.4 +/-0.1, 0.09% Sodium Azide and 0.045% ProClin 300 as preservatives. The negative control is colour coded pale yellow.

3. Positive Control: **CONTROL +**

1x2.0ml/vial. Ready to use. Contains goat serum proteins, high titer anti HDV antibodies, 100 mM Tris-HCl buffer pH 7.4 +/-0.1, 0.09% Sodium Azide and 0.045% ProClin 300 as preservatives. The positive control is colour coded green.

4. Calibrator: **CAL**

n° 1 vial. Lyophilised. To be dissolved with EIA grade water as reported in the label. Contains bovine serum proteins, low titer human antibodies to HDV, 0.2 mg/ml gentamicine sulphate and 0.045% ProClin 300 as preservatives.

Note: The volume necessary to dissolve the content of the vial may vary from lot to lot. Please use the right volume reported on the label.

5. Wash buffer concentrate: **WASHBUF 20X**

1x60ml/bottle. 20x concentrated solution.

Once diluted, the wash solution contains 10 mM phosphate buffer pH 7.0+/-0.2, 0.05% Tween 20 and 0.045% ProClin 300.

6. Enzyme conjugate: **CONJ**

1x16ml/vial. Ready-to-use solution. Contains 5% bovine serum albumine, 10 mM tris buffer pH 6.8 +/-0.1, Horseradish peroxidase conjugated antibody to HDV in presence of 0.2 mg/ml gentamicine sulphate and 0.045% ProClin 300 as preservatives. The component is colour coded red.

7. Chromogen/Substrate: **SUBS TMB**

1x16ml/vial. Contains a 50 mM citrate-phosphate buffered solution at pH 3.5-3.8, 4% DMSO, 0.03% tetra-methyl-benzidine or TMB and 0.02% hydrogen peroxide of H₂O₂.

Note: To be stored protected from light as sensitive to strong illumination.

8. Sulphuric Acid: **H₂SO₄ 0.3 M**

1x15ml/vial. Contains 0.3 M H₂SO₄ solution.

Attention: Irritant (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Plate sealers n° 2

Instructions for Use n° 1

E. MATERIALS REQUIRED BUT NOT PROVIDED

1. Calibrated Micropipettes in the range 10-1000 ul and disposable plastic tips.
2. EIA grade water (double distilled or deionized, charcoal treated to remove oxidizing chemicals used as disinfectants).
3. Timer with 60 minute range or higher.
4. Absorbent paper tissues.
5. Calibrated ELISA microplate thermostatic incubator (dry or wet) set at +37°C.
6. Calibrated ELISA microwell reader with 450nm (reading) and with 620-630nm (blinking) filters.
7. Calibrated ELISA microplate washer.
8. Vortex or similar mixing tools.

F. WARNINGS AND PRECAUTIONS

1. The kit has to be used by skilled and properly trained technical personnel only, under the supervision of a medical doctor responsible of the laboratory.
2. All the personnel involved in performing the assay have to wear protective laboratory clothes, talc-free gloves and glasses. The use of any sharp (needles) or cutting (blades) devices should be avoided. All the personnel involved should be trained in biosafety procedures, as recommended by the Center for Disease Control, Atlanta, U.S. and reported in the National Institute of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
3. All the personnel involved in sample handling should be vaccinated for HBV and HAV, for which vaccines are available, safe and effective.

4. The laboratory environment should be controlled so as to avoid contaminants such as dust or air-borne microbial agents, when opening kit vials and microplates and when performing the test. Protect the Chromogen/Substrate (TMB/H₂O₂) from strong light and avoid vibration of the bench surface where the test is undertaken.
5. Upon receipt, store the kit at +2..8°C into a temperature controlled refrigerator or cold room.
6. Do not interchange components between different lots of the kits. It is recommended that components between two kits of the same lot should not be interchanged.
7. Check that the reagents are clear and do not contain visible heavy particles or aggregates. If not, advise the laboratory supervisor to initiate the necessary procedures.
8. Avoid cross-contamination between serum/plasma samples by using disposable tips and changing them after each sample. Do not reuse disposable tips.
9. Avoid cross-contamination between kit reagents by using disposable tips and changing them between the use of each one. Do not reuse disposable tips.
10. Do not use the kit after the expiration date stated on external (primary container) and internal (vials) labels.
11. Treat all specimens as potentially infective. All human serum specimens should be handled at Biosafety Level 2, as recommended by the Center for Disease Control, Atlanta, U.S. in compliance with what reported in the Institutes of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
12. The use of disposable plastic labware is recommended in the preparation of the washing solution or in transferring components into other containers of automated workstations, in order to avoid contamination.
13. Waste produced during the use of the kit has to be discarded in compliance with national directives and laws concerning laboratory waste of chemical and biological substances. In particular, liquid waste generated from the washing procedure, from residuals of controls and from samples has to be treated as potentially infective material and inactivated. Suggested procedures of inactivation are treatment with a 10% final concentration of household bleach for 16-18 hrs or heat inactivation by autoclave at 121°C for 20 min..
14. Accidental spills have to be adsorbed with paper tissues soaked with household bleach and then with water. Tissues should then be discarded in proper containers designated for laboratory/hospital waste.
15. The Sulphuric Acid is an irritant. In case of spills, wash the surface with plenty of water.
16. Other waste materials generated from the use of the kit (example: tips used for samples and controls, used microplates) should be handled as potentially infective and disposed according to national directives and laws concerning laboratory wastes.

G. SPECIMEN: PREPARATION AND RECOMMENDATIONS

1. Blood is drawn aseptically by venepuncture and plasma or serum is prepared using standard techniques of preparation of samples for clinical laboratory analysis. No influence has been observed in the preparation of the sample with citrate, EDTA and heparin.
2. Avoid any addition of preservatives to samples; especially sodium azide as this chemical would affect the enzymatic activity of the conjugate.
3. Samples have to be clearly identified with codes or names in order to avoid misinterpretation of results. When the kit is used for the screening of blood units, bar code labeling and electronic reading is strongly recommended.
4. Haemolysed (red) and visibly hyperlipemic ("milky") samples have to be discarded as they could generate false results. Samples containing residues of fibrin or heavy particles or microbial filaments and bodies should be discarded as they could give rise to false results.

5. Sera and plasma can be stored at +2°...+8°C in primary collection tubes for up to five days after collection. Do not freeze primary tubes of collection. For longer storage periods, sera and plasma samples, carefully removed from the primary collection tube, can be stored frozen at -20°C for at least 12 months. Any frozen samples should not be frozen/thawed more than once as this may generate particles that could affect the test result.
6. If particles are present, centrifuge at 2.000 rpm for 20 min or filter using 0.2-0.8µ filters to clean up the sample for testing.

H. PREPARATION OF COMPONENTS AND WARNINGS

A study conducted on an opened kit has not pointed out any relevant loss of activity up to 6 re-uses of the device and up to 3 months.

1. Antigen coated microwells:

Allow the microplate to reach room temperature (about 1 hr) before opening the container. Check that the desiccant has not turned dark green, indicating a defect in manufacturing.

In this case, call Dia.Pro's customer service.

Unused strips have to be placed back into the aluminium pouch, with the desiccant supplied, firmly zipped and stored at +2°-8°C. When opened the first time, unused strips are stable until the humidity indicator inside the desiccant bag turns from yellow to green.

2. Negative Control:

Ready to use. Mix well on vortex before use.

3. Positive Control:

Ready to use. Mix well on vortex before use.

4. Calibrator:

Low positive control. Add precisely the volume of EIA grade water, reported on its label, to the lyophilized powder; let fully dissolve and then gently mix on vortex.

Note: *The dissolved calibrator is not stable. Store it frozen in aliquots at -20°C. When thawed do not freeze again; discard it.*

5. Wash buffer concentrate:

The whole content of the 20x concentrated solution has to be diluted with EIA grade water up to 1200 ml and mixed gently end-over-end before use. During preparation avoid foaming as the presence of bubbles could impact on the efficiency of the washing cycles.

Note: *Once diluted, the wash solution is stable for 1 week at +2..8° C.*

6. Enzyme conjugate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, dust or microbes. If this component has to be transferred, use only plastic, and if possible, sterile disposable containers.

7. Chromogen/Substrate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, air-driven dust or microbes. Do not expose to strong light, oxidizing agents and metallic surfaces.

If this component has to be transferred use only plastic, and if possible, sterile disposable container

8. Sulphuric Acid:

Ready to use. Mix well on vortex before use.

Attention: Irritant (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Legenda:

Warning H statements:

H315 – Causes skin irritation.

H319 – Causes serious eye irritation.

Precautionary P statements:

P280 – Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352 – IF ON SKIN: Wash with plenty of soap and water.

P332 + P313 – If skin irritation occurs: Get medical advice/attention.

P305 + P351 + P338 – IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P337 + P313 – If eye irritation persists: Get medical advice/attention.

P362 + P363 – Take off contaminated clothing and wash it before reuse.

I. INSTRUMENTS AND TOOLS USED IN COMBINATION WITH THE KIT

1. Micropipettes have to be calibrated to deliver the correct volume required by the assay and must be submitted to regular decontamination (70% ethanol, 10% solution of bleach, hospital grade disinfectants) of those parts that could accidentally come in contact with the sample or the components of the kit. They should also be regularly maintained in order to show a precision of 1% and a trueness of $\pm 2\%$.
2. The ELISA incubator has to be set at $+37^{\circ}\text{C}$ (tolerance of $\pm 0.5^{\circ}\text{C}$) and regularly checked to ensure the correct temperature is maintained. Both dry incubators and water baths are suitable for the incubations, provided that the instrument is validated for the incubation of ELISA tests.
3. The **ELISA washer** is extremely important to the overall performances of the assay. The washer must be carefully validated in advance, checked for the delivery of the right dispensation volume and regularly submitted to maintenance according to the manufacturer's instructions for use. In particular the washer, at the end of the daily workload, has to be extensively cleaned out of salts with deionized water. Before use, the washer has to be extensively primed with the diluted Washing Solution. The instrument weekly has to be submitted to decontamination according to its manual (NaOH 0.1 M decontamination suggested).
5 washing cycles (aspiration + dispensation of 350ul/well of washing solution + 20 sec soaking = 1 cycle) are sufficient to ensure the assay with the declared performances. If soaking is not possible add one more cycle of washing. An incorrect washing cycle or salt-blocked needles are the major cause of false positive reactions.
4. Incubation times have a tolerance of $\pm 5\%$.
5. The ELISA microplate reader has to be equipped with a reading filter of 450nm and with a second filter of 620-630nm, mandatory for blanking purposes. Its standard performances should be (a) bandwidth ≤ 10 nm; (b) absorbance range from 0 to 4; (c) linearity to 4; repeatability $\geq 1\%$. Blanking is carried out on the well identified in the section "Assay Procedure". The optical system of the reader has to be calibrated regularly to ensure that the correct optical density is measured. It should be regularly maintained according to the manufacturer 's instructions.
6. When using an ELISA automated work station, all critical steps (dispensation, incubation, washing, reading, shaking, data handling) have to be carefully set, calibrated, controlled and regularly serviced in order to match the values reported in the sections "Validation of Test" and "Assay Performances". The assay protocol has to be installed in the operating system of the unit and validated as for the washer and the reader. In addition, the liquid handling part of the station (dispensation and washing) has to be validated and correctly set. Particular attention must be paid to avoid carry over by the needles used for

dispensing samples and for washing. This must be studied and controlled to minimize the possibility of contamination of adjacent wells due to strongly reactive samples, leading to false positive results. The use of ELISA automated work stations is recommended for blood screening and when the number of samples to be tested exceed 20-30 units per run.

7. Dia.Pro's customer service offers support to the user in the setting and checking of instruments used in combination with the kit, in order to assure full compliance with the requirements described. Support is also provided for the installation of new instruments to be used with the kit.

L. PRE ASSAY CONTROLS AND OPERATIONS

1. Check the expiration date of the kit printed on the external label (primary container). Do not use if expired.
2. Check that the liquid components are not contaminated by visible particles or aggregates. Check that the Chromogen/Substrate is colorless or pale blue by aspirating a small volume of it with a sterile plastic pipette. Check that no breakage occurred in transportation and no spillage of liquid is present inside the box (primary container). Check that the aluminum pouch, containing the microplate, is not punctured or damaged.
3. Dilute all the content of the 20x concentrated Wash Solution as described above.
4. Dissolve the Calibrator as described above and gently mix.
5. Allow all the other components to reach room temperature (about 1 hr) and then mix gently on vortex all liquid reagents.
6. Set the ELISA incubator at $+37^{\circ}\text{C}$ and prepare the ELISA washer by priming with the diluted washing solution, according to the manufacturers instructions. Set the right number of washing cycles as reported in the specific section.
7. Check that the ELISA reader is turned on or ensure it will be turned on at least 20 minutes before reading.
8. If using an automated work station, turn on, check settings and be sure to use the right assay protocol.
9. Check that the micropipettes are set to the required volume.
10. Check that all the other equipment is available and ready to use.
11. In case of problems, do not proceed further with the test and advise the supervisor.

M. ASSAY PROCEDURE

The assay has to be carried out according to what reported below, taking care to maintain the same incubation time for all the samples in testing.

1. Place the required number of strips in the microplate holder. Leave A1 well empty for the operation of blanking. Store the other strips into the bag in presence of the desiccant at $+2.8^{\circ}\text{C}$, sealed.
2. Pipette 100 μl of Negative Control in triplicate, 100 μl Positive Control in single and then 100 μl of samples. Check that controls and samples have been correctly added. Then incubate the microplate at **$+37^{\circ}\text{C}$ for 60 min.**
3. Wash the microplate as reported in section I.3.
4. In all the wells except A1, pipette 100 μl Enzyme Conjugate. Check that the reagent has been correctly added. Then incubate the microplate at **$+37^{\circ}\text{C}$ for 60 min.**

Important note: Be careful not to touch the inner surface of the well with the pipette tip when dispensing the Enzyme Conjugate. Contamination might occur.

5. Wash the microplate as described.

6. Pipette 100 µl TMB/H₂O₂ mixture in each well, the blank wells included. Check that the reagent has been correctly added. Then incubate the microplate at **room temperature for 20 min**.

Important note: Do not expose to strong direct light as a high background might be generated.

7. Pipette 100 µl Sulphuric Acid into all the wells using the same pipetting sequence as in step n° 6 to stop the enzymatic reaction. Addition of the stop solution will turn the negative control and negative samples from blue to yellow.

8. Measure the colour intensity of the solution in each well, as described in section I.5 using a 450nm filter (reading) and a 620-630nm filter (background subtraction, mandatory), blanking the instrument on A1.

Important notes:

1. Ensure that no finger prints are present on the bottom of the microwell before reading. Finger prints could generate false positive results on reading.
2. Reading has should ideally be performed immediately after the addition of the Stop Solution but definitely no longer than 20 minutes afterwards. Some self oxidation of the chromogen can occur leading to a higher background.
3. The use of the Calibrator, a low positive control, is not mandatory for the assay as the CAL does not enter into the cut-off calculation. The CAL may be used as a low titer positive control when a laboratory internal quality verification is required by the management. When used for such purpose, dispense 100 ul of it, possibly in duplicate.

N. ASSAY SCHEME

Controls/Calibrator	100 ul
Samples	100 ul
1st incubation	60 min
Temperature	+37°C
Washing step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
Enzyme Conjugate	100 ul
2nd incubation	60 min
Temperature	+37°C
Washing step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
TMB/H ₂ O ₂ mix	100 ul
3rd incubation	20 min
Temperature	r.t.
Sulphuric Acid	100 ul
Reading OD	450nm / 620-630nm

An example of dispensation scheme (including CAL) is reported in the table below:

Microplate

	1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	S2										
B	NC	S3										
C	NC	S4										
D	NC	S5										
E	CAL	S6										
F	CAL	S7										
G	PC	S8										
H	S1	S9										

Legenda: BLK = Blank NC = Negative Control
CAL = Calibrator PC = Positive Control S = Sample

O. INTERNAL QUALITY CONTROL

A check is performed on the negative and positive controls any time, and on the Calibrator in addition when the kit is used for the first time, in order to verify whether the expected OD_{450nm} / 620-630nm or Co/S values have been matched in the analysis. Ensure that the following parameters are met:

Parameter	Requirements
Blank well	< 0.100 OD _{450nm} value
Negative Control (NC)	> 1.000 OD _{450nm} after blanking If lower carefully control the washing procedure and decrease the number of cycles or the soaking time coefficient of variation < 30%
Positive Control (PC)	OD _{450 nm} < NC/10
Calibrator (CAL)	PC ≤ OD _{450nm} < (NC+PC)/5

If the results of the test match the requirements stated above, proceed to the next section.

If they don't, do not proceed any further and perform the following checks:

Problem	Check
Blank well > 0.100 OD _{450nm}	that the Chromogen/Substrate solution has not become contaminated during the assay
Negative Control (NC) < 1.000 OD _{450nm} after blanking coefficient of variation > 30%	1. that the washing procedure and the washer settings are as validated in the pre qualification study; 2. that the proper washing solution has been used and the washer has been primed with it before use; 3. that no mistake has been done in the assay procedure (dispensation of positive control instead of negative control); 4. that no contamination of the negative control or of the wells where the control was dispensed has occurred due to positive samples, to spills or to the enzyme conjugate; 5. that micropipettes have not become contaminated with positive samples or with the enzyme conjugate; 6. that the washer needles are not blocked or partially obstructed.

Calibrator OD450nm Outside the range	<ol style="list-style-type: none"> 1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution (ex.: dispensation of negative control instead of Calibrator); 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the calibrator has occurred.
Positive Control OD450nm > NC/10	<ol style="list-style-type: none"> 1. that the procedure has been correctly performed; 2. that no mistake has occurred during the distribution of the control (dispensation of negative control instead of positive control). 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the positive control has occurred.

If any of the above problems have occurred, report the problem to the supervisor for further actions.

Important note:

The analysis must be done proceeding as the reading step described in the section M, point 8.

P. RESULTS

The results are calculated by means of a cut-off value determined with the following formula:

$$\text{Cut-Off} = (\text{NC} + \text{PC}) / 5$$

Important note: When the calculation of results is performed by the operating system of an ELISA automated work station, ensure that the proper formulation is used to calculate the cut-off value and generate the correct interpretation of results.

Q. INTERPRETATION OF RESULTS

Results are interpreted as ratio between the cut-off value and the sample OD450nm / 620-630nm or Co/S. Results are interpreted according to the following table:

Co/S	Interpretation
< 0.9	Negative
0.9 – 1.1	Equivocal
> 1.1	Positive

A negative result indicates that the patient has not been infected by HDV.

Any patient showing an equivocal result should be re-tested on a second sample taken 1-2 weeks after the initial sample.

A positive result is indicative of HDV infection and therefore the patient should be treated accordingly.

Important notes:

1. Interpretation of results should be done under the supervision of the laboratory supervisor to reduce the risk of judgement errors and misinterpretations.
2. When test results are transmitted from the laboratory to another facility, attention must be paid to avoid erroneous data transfer.
3. Diagnosis of viral hepatitis infection has to be taken by and released to the patient by a suitably qualified medical doctor.

An example of calculation is reported below (data obtained proceeding as the the reading step described in the section M, point 8).

The following data must not be used instead of real figures obtained by the user.

Negative Control: 2.100 – 2.200 – 2.000 OD450nm

Mean Value: 2.100 OD450nm

Higher than 1.000 – Accepted

Positive Control: 0.100 OD450nm

Lower than NC/10 – Accepted

$$\text{Cut-Off} = (2.100 + 0.100) / 5 = 0.440$$

Calibrator: 0.300-0.260 OD450nm

Mean value: 0.280 OD450nm

Within the range $\text{PC} \leq \text{OD450nm} < (\text{NC} + \text{PC})/5$ – Accepted

Sample 1: 0.020 OD450nm

Sample 2: 1.900 OD450nm

Sample 1 Co/S > 1.1 positive

Sample 2 Co/S < 0.9 negative

R. PERFORMANCES

Evaluation of Performances has been conducted in accordance to what reported in the Common Technical Specifications or CTS (art. 5, Chapter 3 of IVD Directive 98/79/EC)

1. LIMIT OF DETECTION:

In absence of an international standard, the sensitivity of the assay has been calculated by means of the product named Accurun n° 127 supplied by Boston Biomedica Inc. – USA .

The table below reports the OD450nm shown by this preparation when diluted in Fetal Calf Serum to prepare a limiting dilution curve, in three different lots.

Co/S values

Accurun # 127	DAB.CE	Lot # 1102	DAB.CE	Lot # 0103	DAB.CE	Lot # 0403
	OD450 nm	Co/S value	OD450 nm	Co/S value	OD450 nm	Co/S value
1x	0.171	3.0	0.163	2.9	0.156	2.8
2x	0.187	2.7	0.176	2.6	0.179	2.5
4x	0.230	2.2	0.220	2.1	0.202	2.2
8x	0.298	1.7	0.285	1.6	0.271	1.6
16x	0.417	1.2	0.405	1.1	0.402	1.1
32x	0.514	1.0	0.490	0.9	0.482	0.9
64x	0.717	0.7	0.700	0.7	0.705	0.6
128x	1.063	0.5	1.006	0.5	1.015	0.4
CTRL (-)	2.484	////////	2.261	////////	2.114	////////

2. DIAGNOSTIC SPECIFICITY AND SENSITIVITY

The diagnostic performances were evaluated in a clinical trial conducted by the Department of Gastro-Hepatology, Prof. M.Rizzetto, S.Giovanni Battista hospital, Torino, Italy, on more than 400 samples against a reference kit.

Negative, positive and potentially interfering samples were examined in the trial.

Both plasma, derived with different standard techniques of preparation (citrate, EDTA and heparin), and sera have been used to determine the specificity. No false reactivity due to the method of specimen preparation has been observed.

Results are briefly reported in the tables below:

Sensitivity	> 98 %
Specificity	> 98 %

3. PRECISION

The mean values obtained from a study conducted on two samples of different anti-HDV antibody reactivity, examined in 16 replicates in three separate runs for three lots of product, is reported below:

DAB.CE: lot #1102

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.342	2.428	2.433	2.401
Std.Deviation	0.113	0.106	0.122	0.114
CV %	4.8	4.4	5.0	4.7

Calibrator (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.298	0.289	0.286	0.291
Std.Deviation	0.023	0.027	0.026	0.025
CV %	7.7	9.3	9.1	8.7
Co/S	1.6	1.7	1.7	1.7

DAB.CE: lot #0103

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.208	2.237	2.246	2.230
Std.Deviation	0.105	0.108	0.108	0.107
CV %	4.7	4.8	4.8	4.8

Calibrator (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.269	0.277	0.266	0.271
Std.Deviation	0.026	0.024	0.025	0.025
CV %	9.8	8.5	9.5	9.3
Co/S	1.7	1.7	1.7	1.7

DAB.CE: lot # 0403

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.246	2.221	2.182	2.216
Std.Deviation	0.097	0.103	0.118	0.106
CV %	4.3	4.6	5.4	4.8

Calibrator (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.286	0.273	0.280	0.280
Std.Deviation	0.027	0.023	0.026	0.025
CV %	9.3	8.5	9.1	9.0
Co/S	1.6	1.7	1.6	1.6

The variability shown in the tables did not result in sample misclassification.

Important note:

The performance data have been obtained proceeding as the reading step described in the section M, point 8.

S. LIMITATIONS

Bacterial contamination or heat inactivation of the specimen may affect the absorbance values of the samples with consequent alteration of the level of the analyte.

This test is suitable only for testing single samples and not pooled ones.

Diagnosis of an infectious disease should not be established on the basis of a single test result. The patient's clinical history, symptomatology, as well as other diagnostic data should be considered.

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All the IVD Products manufactured by the company are under the control of a certified Quality Management System approved by an EC Notified Body. Each lot is submitted to a quality control and released into the market only if conforming with the EC technical specifications and acceptance criteria.

Manufacturer:
Dia.Pro Diagnostic Bioprobes Srl
Via G. Carducci n° 27 – Sesto San Giovanni (MI) – Italy



0318

HDV Ab

**Ensayo inmunoenzimático competitivo
para la determinación cualitativa de
anticuerpos frente al Virus de la
Hepatitis Delta
en plasma y suero humanos**

Uso exclusivo para diagnóstico "in vitro"



DIA.PRO

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REF DAB.CE
96 pruebas

HDV Ab

A. OBJETIVO DEL EQUIPO.

Ensayo inmunoenzimático competitivo (ELISA) para la determinación cualitativa de anticuerpos frente al Virus de la Hepatitis Delta (HDV) en plasma y suero humanos con una metodología de "dos pasos".

El equipo ha sido desarrollado para el seguimiento de pacientes infectados con HDV.

Uso exclusivo para diagnóstico "in vitro".

B. INTRODUCCIÓN.

El Virus de la Hepatitis Delta es un virus ARN defectivo. Se compone de un núcleo con los antígenos delta específicos, y está encapsulado por el HBsAg. Para su replicación necesita ayuda funcional de HBV.

La infección por HDV ocurre en presencia de una infección aguda o crónica por HBV. Cuando se presenta simultáneamente la infección aguda por los dos virus, la enfermedad es grave y el cuadro clínico, así como las características bioquímicas son prácticamente indistinguibles de una infección por HBV. Sin embargo, una persona infectada por HBV de forma crónica puede soportar indefinidamente la replicación por HDV, normalmente la enfermedad es menos severa y aparece como exacerbación clínica.

La determinación de los marcadores serológicos específicos de HDV (HDV Ag, HDV Ab, HDV IgM y HDV IgG) representa una herramienta importante para los clínicos en la clasificación del agente etiológico, en el seguimiento de los pacientes así como en el tratamiento.

La detección de anticuerpos totales permite la clasificación de la enfermedad y el seguimiento de la seroconversión.

C. PRINCIPIOS DEL ENSAYO.

El ensayo es de tipo competitivo, donde los anticuerpos anti-HDV de la muestra compiten con un anticuerpo policlonal (IgG) específico para el virus y conjugado con peroxidasa (HRP), por el antígeno recombinante-HDV de la fase sólida.

El ensayo se realiza mediante un sistema de dos pasos con incubación competitiva. La muestra se añade a la placa y los anticuerpos específicos anti-HDV se combinan con el antígeno de la fase sólida. Después del lavado, se añade un anticuerpo conjugado con peroxidasa (HRP) que se une al antígeno libre en la placa. Previo lavado, se añade el substrato cromogénico.

La concentración de la enzima conjugada, unida a la fase sólida es inversamente proporcional a la cantidad de anticuerpos al HDV presentes en la muestra y su actividad se detecta por la adición del substrato cromogénico.

La concentración de anticuerpos específicos al HDV en la muestra se determina de manera semicuantitativa a través del cálculo de un valor de corte.

D. COMPONENTES.

Cada equipo contiene reactivos suficientes para realizar 96 pruebas.

1. Microplaca: **MICROPLATE**

12 tiras de 8 pocillos recubiertos con antígeno recombinante específico de HDV, en bolsas selladas con desecante. Se deben poner las placas a temperatura ambiente antes de abrirlas, sellar las tiras sobrantes en la bolsa con el desecante y almacenar a 4°C.

2. Control Negativo: **CONTROL -**

1x2.0ml/vial. Listo para el uso. Contiene proteínas del suero de cabra, tampón Tris-HCl 100 mM pH 7.4 +/-0.1, además de azida sódica 0.09% y ProClin 300 0.045% como conservantes. El control negativo está codificado con el color amarillo pálido.

3. Control Positivo: **CONTROL +**

1x2.0ml/vial. Listo para el uso. Contiene proteínas del suero de cabra, alto título de anticuerpos anti-HDV, tampón Tris-HCl 100 mM pH 7.4 +/-0.1, además de azida sódica 0.09% y ProClin 300 0.045% como conservantes. El control positivo está codificado con el color verde.

4. Calibrador: **CAL ...**

n° 1 vial. Liofilizado. Para disolver en agua calidad EIA como se indica en la etiqueta. Contiene suero bovino fetal, bajo título de anticuerpos humanos al HDV, además de sulfato de gentamicina 0.02 mg/ml y ProClin 300 0.045% como conservantes.

Nota: El volumen necesario para disolver el contenido del frasco, varía en cada lote. Se recomienda usar el volumen indicado en la etiqueta.

5. Tampón de Lavado Concentrado: **WASHBUF 20X**

1x60ml/botella. Solución concentrada 20x.

Una vez diluida, la solución de lavado contiene tampón fosfato 10 mM a pH 7.0 +/- 0.2, Tween 20 al 0.05% y ProClin 300 al 0.045%

6. Conjugado **CONJ**

1x16ml/vial. Solución lista para el uso. Contiene 5% de albúmina de suero bovino, tampón Tris 10mM a pH 6.8 +/- 0.1, anticuerpo anti-HDV conjugado con peroxidasa (HRP) en presencia de 0.2 mg/ml de sulfato de gentamicina y ProClin 300 0.045% como conservante. El conjugado está codificado con el color rojo.

7. Cromógeno/Substrato **SUBS TMB**

1x16ml/vial. Contiene una solución tamponada citrato-fosfato 50mM pH 3.5-3.8, tetra-metil-benzidina (TMB) 0.03% y peróxido de hidrógeno (H₂O₂) 0.02% así como dimetilsulfóxido 4%.

Nota: Evitar la exposición a la luz, ya que la sustancia es fotosensible.

8. Ácido Sulfúrico: **H₂SO₄ 0.3M**

1x15ml/vial. Contiene solución de H₂SO₄ 0.3M

Atención: Irritante (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Sellador adhesivo, n° 2

Manual de instrucciones, n° 1

E. MATERIALES NECESARIOS NO SUMINISTRADOS.

1. Micropipetas calibradas (10-1000 µl) y puntas plásticas desechables.
2. Agua de calidad EIA (Bidestilada o desionizada, tratada con carbón para remover químicos oxidantes usados como desinfectantes).
3. *Timer* con un rango de 60 minutos como mínimo.
4. Papel absorbente.
5. Incubador termostático de microplacas ELISA, calibrado (en seco o húmedo) fijo a 37°C.
6. Lector calibrado de microplacas de ELISA con filtros de 450nm (lectura) y de 620-630 nm.
7. Lavador calibrado de microplacas ELISA.
8. Vórtex o similar.

F. ADVERTENCIAS Y PRECAUCIONES.

1. El equipo debe ser usado por personal técnico adecuadamente entrenado, bajo la supervisión de un doctor responsable del laboratorio.
2. Todas las personas encargadas de la realización de las pruebas deben llevar las ropas protectoras adecuadas de laboratorio, guantes y gafas. Evitar el uso de objetos cortantes (cuchillas) o punzantes (agujas). El personal

debe ser adiestrado en procedimientos de bioseguridad, según ha sido recomendado por el Centro de Control de Enfermedades de Atlanta, Estados Unidos, y publicado por el Instituto Nacional de Salud: "Biosafety in Microbiological and Biomedical Laboratories", ed.1984.

3. Todo el personal involucrado en el manejo de muestras debe estar vacunado contra HBV y HAV, para lo cual existen vacunas disponibles, seguras y eficaces.
4. Se debe controlar el ambiente del laboratorio para evitar la contaminación de los componentes con polvo o agentes microbianos cuando se abran los equipos, así como durante la realización del ensayo. Evitar la exposición del substrato (TMB/H₂O₂) a la luz y las vibraciones de la mesa de trabajo durante el ensayo.
5. Conservar el equipo a temperaturas entre 2-8 °C, en un refrigerador con temperatura regulada o en cámara fría.
6. No intercambiar reactivos de diferentes lotes ni tampoco de diferentes equipos.
7. Comprobar que los reactivos no contienen precipitados ni agregados en el momento del uso. De darse el caso, informar al responsable para realizar el procedimiento pertinente.
8. Evitar contaminación cruzada entre muestras de suero/plasma usando puntas desechables y cambiándolas después de cada uso. No reutilizar puntas desechables.
9. Evitar contaminación cruzada entre los reactivos del equipo usando puntas desechables y cambiándolas después de cada uso. No reutilizar puntas desechables.
10. No usar el producto después de la fecha de caducidad indicada en el equipo e internamente en los reactivos.
11. Tratar todas las muestras como potencialmente infecciosas. Las muestras de suero humano deben ser manipuladas al nivel 2 de bioseguridad, según ha sido recomendado por el Centro de Control de Enfermedades de Atlanta, Estados Unidos y publicado por el Instituto Nacional de Salud: "Biosafety in Microbiological and Biomedical Laboratories", ed.1984.
12. Se recomienda el uso de material plástico desechable para la preparación de las soluciones de lavado y para la transferencia de los reactivos a los diferentes equipos automatizados a fin de evitar contaminaciones.
13. Los desechos producidos durante el uso del equipo deben de ser eliminados según lo establecido por las directivas nacionales y las leyes relacionadas con el tratamiento de los residuos químicos y biológicos de laboratorio. En particular, los desechos líquidos provenientes del proceso de lavado deben ser tratados como potencialmente infecciosos y deben ser inactivados. Se recomienda la inactivación con lejía al 10% de 16 a 18 horas o el uso de la autoclave a 121°C por 20 minutos.
14. En caso de derrame accidental de algún producto, se debe utilizar papel absorbente embebido en lejía y posteriormente en agua. El papel debe eliminarse en contenedores designados para este fin en hospitales y laboratorios.
15. El ácido sulfúrico es irritante. En caso de derrame, se debe lavar la superficie con abundante agua.
16. Otros materiales de desecho generados durante la utilización del equipo (por ejemplo: puntas usadas en la manipulación de las muestras y controles, microplacas usadas) deben ser manipuladas como fuentes potenciales de infección de acuerdo a las directivas nacionales y leyes para el tratamiento de residuos de laboratorio.

G. MUESTRA: PREPARACIÓN Y RECOMENDACIONES.

1. Extraer la sangre asépticamente por punción venosa y preparar el suero o plasma según las técnicas estándar de los laboratorios de análisis clínico. No se ha detectado que el tratamiento con citrato, EDTA o heparina afecte las muestras.
2. Evitar el uso de conservantes, en particular azida sódica, ya que pudiera afectar la actividad enzimática del conjugado.

3. Las muestras deben estar identificadas claramente mediante código de barras o nombres, a fin de evitar errores en los resultados. Cuando el equipo se emplea para el pesquiseaje en unidades de sangre, se recomienda el uso del código de barras.
4. Las muestras hemolizadas (color rojo) o hiperlipémicas (aspecto lechoso) deben ser descartadas para evitar falsos resultados, al igual que aquellas donde se observe la presencia de precipitados, restos de fibrina o filamentos microbianos.
5. El suero y el plasma pueden conservarse a una temperatura entre +2° y +8°C en tubos de recolección principales hasta cinco días después de la extracción. No congelar tubos de recolección principales. Para periodos de almacenamiento más prolongados, las muestras de plasma o suero, retiradas cuidadosamente del tubo de extracción principal, pueden almacenarse congeladas a -20°C durante al menos 12 meses. Evitar congelar/descongelar cada muestra más de una vez, ya que pueden generarse partículas que podrían afectar al resultado de la prueba.
6. Si hay presencia de agregados, la muestra se puede aclarar mediante centrifugación a 2000 rpm durante 20 minutos o por filtración con un filtro de 0,2-0,8 micras.

H. PREPARACIÓN DE LOS COMPONENTES Y PRECAUCIONES.

Según estudios realizados, no se ha detectado pérdida relevante de actividad en equipos abiertos, en uso por un período de hasta 3 meses.

1. Microplacas:

Dejar la microplaca a temperatura ambiente (aprox. 1 hora) antes de abrir el envase. Compruebe que el desecante no esté de un color verde oscuro, lo que indicaría un defecto de fabricación. De ser así, debe solicitar el servicio de Dia.Pro: atención al cliente.

Las tiras de pocillos no utilizadas, deben guardarse herméticamente cerradas en la bolsa de aluminio con el desecante a 2-8°C. Una vez abierto el envase, las tiras sobrantes, se mantienen estables hasta que el indicador de humedad dentro de la bolsa del desecante cambie de amarillo a verde.

2. Control Negativo:

Listo para el uso. Mezclar bien con la ayuda de un vórtex, antes de usar.

3. Control Positivo:

Listo para el uso. Mezclar bien con la ayuda de un vórtex, antes de usar.

4. Calibrador:

Control positivo bajo. Añadir de manera precisa al polvo liofilizado el volumen de agua de calidad EIA indicado en la etiqueta. Dejar disolver totalmente y mezclar suavemente en el vórtex.

Note: Una vez reconstituida, la solución no es estable. Se recomienda mantenerla congelada en alícuotas a -20°C. Cuando se descongele, descartar el agua en lugar de congelarla nuevamente.

5. Solución de Lavado Concentrada:

Todo el contenido de la solución concentrada 20x debe diluirse con agua bidestilada hasta 1200ml y mezclarse suavemente antes de usarse. Durante la preparación evitar la formación de espuma y burbujas, lo que podría influir en la eficiencia de los ciclos de lavado.

Nota: Una vez diluida, la solución es estable por una semana a temperaturas entre +2 y 8°C.

6. Conjugado:

Listo para el uso. Mezclar bien con un vórtex antes de usar. Evitar posible contaminación del líquido con oxidantes químicos, polvo o microbios. En caso de que deba transferirse el reactivo, usar contenedores de plástico, estériles y desechables, siempre que sea posible.

7. Cromógeno/ Substrato:

Listo para el uso. Mezclar bien con un vórtex antes de usar. Evitar posible contaminación del líquido con oxidantes químicos, polvo o microbios. Evitar la exposición a la luz, agentes oxidantes y superficies metálicas. En caso de que deba transferirse el reactivo, usar contenedores de plástico, estériles y desechables, siempre que sea posible.

8. Ácido Sulfúrico:

Listo para el uso. Mezclar bien con un vórtex antes de usar. Atención: Irritante (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Leyenda:

Indicación de peligro, **Frases H**

H315 – Provoca irritación cutánea.

H319 – Provoca irritación ocular grave.

Consejo de prudencia, **Frases P**

P280 – Llevar guantes/prendas/gafas/máscara de protección.

P302 + P352 – EN CASO DE CONTACTO CON LA PIEL: Lavar con agua y jabón abundantes.

P332 + P313 – En caso de irritación cutánea: Consultar a un médico.

P305 + P351 + P338 – EN CASO DE CONTACTO CON LOS OJOS: Aclarar cuidadosamente con agua durante varios minutos. Quitar las lentes de contacto, si lleva y resulta fácil. Seguir aclarando.

P337 + P313 – Si persiste la irritación ocular: Consultar a un médico.

P362 + P363 – Quitarse las prendas contaminadas y lavarlas antes de volver a usarlas.

I. INSTRUMENTOS Y EQUIPAMIENTO UTILIZADOS EN COMBINACIÓN CON EL EQUIPO.

- Las micropipetas deben ser calibradas para dispensar correctamente el volumen requerido en el ensayo y sometidas a una descontaminación periódica de las partes que pudieran entrar accidentalmente en contacto con la muestra o los reactivos (etanol 70%, lejía 10%, de calidad de los desinfectantes hospitalarios). Deben además, ser regularmente revisadas para mantener una precisión del 1% y una confiabilidad de +/- 2%.
- La incubadora de ELISA debe ser ajustada a 37°C (+/- 0.5°C) y controlada periódicamente para mantener la temperatura correcta. Pueden emplearse incubadoras secas o baños de agua siempre que estén validados para la incubación de pruebas de ELISA.
- El **lavador ELISA** es extremadamente importante para el rendimiento global del ensayo. El lavador debe ser validado de forma minuciosa previamente, revisado para comprobar que suministra el volumen de dispensación correcto y enviado regularmente a mantenimiento de acuerdo con las instrucciones de uso del fabricante. En particular, deben lavarse minuciosamente las sales con agua desionizada del lavador al final de la carga de trabajo diaria. Antes del uso, debe suministrarse extensivamente solución de lavado diluida al lavador. Debe enviarse el instrumento semanalmente a descontaminación según se indica en su manual (se recomienda descontaminación con NaOH 0.1 M). Para asegurar que el ensayo se realiza conforme a los rendimientos declarados, basta con 5 ciclos de lavado (aspiración + dispensado de 350 µl/pocillo de solución de lavado + 20 segundos de remojo = 1 ciclo). Si no es posible

remojar, añadir un ciclo de lavado adicional. Un ciclo de lavado incorrecto o agujas obstruidas con sal son las principales causas de falsas reacciones positivas.

- Los tiempos de incubación deben tener un margen de $\pm 5\%$.
- El lector de microplaca ELISA debe estar provisto de un filtro de lectura de 450nm y de un segundo filtro de 620-630 nm, obligatorio para reducir interferencias en la lectura. El procedimiento estándar debe contemplar: a) Ancho de banda ≤ 10 nm b) Rango de absorbancia de 0 a 4, c) Linealidad a 4, reproducibilidad $\geq 1\%$. El blanco se prueba en el pocillo indicado en la sección "Procedimiento del ensayo". El sistema óptico del lector debe ser calibrado periódicamente para garantizar que se mide la densidad óptica correcta. Periódicamente debe procederse al mantenimiento según las instrucciones del fabricante.
- En caso de usar un sistema automatizado de ELISA, los pasos críticos (dispensado, incubación, lavado, lectura, agitación y procesamiento de datos) deben ser cuidadosamente fijados, calibrados, controlados y periódicamente ajustados, para garantizar los valores indicados en las secciones "Control interno de calidad" y "Procedimiento del ensayo". El protocolo del ensayo debe ser instalado en el sistema operativo de la unidad y validado tanto para el lavador como para el lector. Por otro lado, la parte del sistema que maneja los líquidos (dispensado y lavado) debe ser validada y fijada correctamente. Debe prestarse particular atención a evitar el arrastre por las agujas de dispensación y las de lavado, a fin de minimizar la posibilidad de ocurrencia de falsos positivos por contaminación de los pocillos adyacentes por muestras fuertemente reactivas. Se recomienda el uso de sistemas automatizados para el pesquaje en unidades de sangre y cuando la cantidad de muestras supera las 20-30 unidades por ensayo.
- El servicio de atención al cliente en Dia.Pro, ofrece apoyo al usuario para calibrar, ajustar e instalar los equipos a usar en combinación con el equipo, con el propósito de asegurar el cumplimiento de los requerimientos descritos.

L. OPERACIONES Y CONTROLES PREVIOS AL ENSAYO.

- Compruebe la fecha de caducidad indicada en la parte externa del equipo (envase primario). No usar si ha caducado.
- Compruebe que los componentes líquidos no están contaminados con partículas o agregados visibles. Asegúrese de que el cromógeno (TMB) es incoloro o azul pálido, aspirando un pequeño volumen de este con una pipeta estéril de plástico. Compruebe que no han ocurrido rupturas ni derrames de líquido dentro de la caja (envase primario) durante el transporte. Asegurarse de que la bolsa de aluminio que contiene la microplaca no esté rota o dañada.
- Diluir totalmente la solución de lavado 20x concentrada, como se ha descrito anteriormente.
- Disolver el Calibrador como se ha descrito anteriormente y mezclar suavemente usando un vórtex.
- Dejar los componentes restantes alcanzar la temperatura ambiente (aprox. 1 hora), mezclar luego suavemente en el vórtex todos los reactivos líquidos.
- Ajustar la incubadora de ELISA a 37°C y cebar el lavador de ELISA utilizando la solución de lavado, según las instrucciones del fabricante. Fijar el número de ciclos de lavado según se indica en la sección específica.
- Comprobar que el lector de ELISA esté conectado al menos 20 minutos antes de realizar la lectura.
- En caso de trabajar automáticamente, conectar el equipo y comprobar que los protocolos estén correctamente programados.
- Comprobar que las micropipetas estén fijadas en el volumen requerido.
- Asegurarse de que el equipamiento a usar esté en perfecto estado, disponible y listo para el uso.

11. En caso de surgir algún problema, se debe detener el ensayo y avisar al responsable.

M. PROCEDIMIENTO DEL ENSAYO.

El ensayo debe realizarse según las instrucciones que siguen a continuación, es importante mantener en todas las muestras el mismo tiempo de incubación.

1. Poner el número necesario de tiras en el soporte plástico. Dejar el pocillo A1 vacío para el blanco. Almacenar las tiras restantes en la bolsa con el desecante a temperaturas entre 2 y 8°C.

2. Dispensar 100µl del Control Negativo, por triplicado, 100µl del Control Positivo una vez y, posteriormente, añadir 100µl de muestras. Comprobar que los controles y muestras se han añadido correctamente.

Después incubar la microplaca durante **60 minutos a +37°C**.

3. Lavar la microplaca según lo descrito previamente (sección I.3).

4. Dispensar 100µl de Conjugado en todos los pocillos, excepto A1; comprobar que los reactivos se han añadido correctamente. Incubar la microplaca durante **60 minutos a +37°C**.

Nota importante: Tener cuidado de no tocar la pared interna del pocillo con la punta de la pipeta al dispensar el conjugado. Podría producirse contaminación.

5. Lavar la microplaca según lo descrito previamente (sección I.3).

6. Dispensar 100µl del Cromógeno/Substrato en todos los pocillos, incluido el A1.

Incubar la microplaca protegida de la luz a **temperatura ambiente (18-24°C) durante 20 minutos**.

Nota importante: No exponer directamente a fuerte iluminación, de lo contrario se generan interferencias.

7. Dispensar 100µl de ácido sulfúrico en todos los pocillos para detener la reacción enzimática, usar la misma secuencia que en el paso 6. La adición de la solución de parada cambia el color del Control Negativo y las muestras negativas de azul a amarillo.

8. Medir la intensidad del color de la solución en cada pocillo, según se indica en la sección I.5, con un filtro de 450 nm (lectura) y otro de 620-630 nm (substracción del fondo, obligatorio), calibrando el instrumento con el pocillo A1 (blanco).

Notas importantes:

- Asegurarse de que no hay impresiones digitales en el fondo de los pocillos antes de leer. Podrían generarse falsos positivos en la lectura.
- La lectura debe hacerse inmediatamente después de añadir la solución de parada y, en cualquier caso, nunca transcurridos 20 minutos después de su adición. Se podría producir auto oxidación del cromógeno causando un elevado fondo.
- El uso del calibrador (CAL), un control negativo bajo, no es obligatorio para el ensayo ya que el calibrador (CAL) no afecta al cálculo del valor de corte. El calibrador (CAL) puede usarse como un control negativo bajo si la gestión requiere un control interno de calidad del laboratorio. Dispensar 100µl del calibrador (CAL), posiblemente por duplicado, cuando se utilice para este propósito.

N. ESQUEMA DEL ENSAYO.

Controles/Calibrador	100 µl
Muestras	100 µl
1ª incubación	60 min
Temperatura	+37°C
Lavado	5 ciclos con 20" de remojo o 6 ciclos sin remojo
Conjugado	100 µl
2ª incubación	60 min
Temperatura	+37°C
Lavado	5 ciclos con 20" de remojo o 6 ciclos sin remojo
Mezcla TMB/H2O2	100 µl
3ª incubación	20 min
Temperatura	t.a.*
Ácido Sulfúrico	100 µl
Lectura D.O.	450nm / 620-630nm

t.a.* temperatura ambiente

A continuación se describe un ejemplo del esquema de dispensado (incluido el calibrador (CAL):

		Microplaca											
		1	2	3	4	5	6	7	8	9	10	11	12
A	BL	M2											
B	CN	M3											
C	CN	M4											
D	CN	M5											
E	CAL	M6											
F	CAL	M7											
G	CP	M8											
H	M1	M9											

Leyenda: BL = Blanco CN = Control Negativo
CAL = Calibrador CP = Control Positivo M = Muestra

O. CONTROL DE CALIDAD INTERNO.

Se realiza un grupo de pruebas con los controles negativo y positivo cada vez que se usa el equipo, y con el calibrador la primera vez que se usa el equipo, para verificar si los valores DO450nm o Co/M son los esperados.

Asegurar el cumplimiento de los siguientes parámetros:

Parámetro	Exigencia
Pocillo Blanco	valor < 0.100 DO450nm
Control Negativo (CN)	> 1.000 DO450nm después de leer el blanco Si es menor, controle cuidadosamente el proceso de lavado y disminuya los ciclos o el tiempo entre los mismos. Coeficiente de variación < 30%
Control Positivo (CP)	DO450 nm < CN/10
Calibrador (CAL)	CP ≤ DO450nm < (CN+CP)/5

Si los resultados del ensayo coinciden con lo establecido anteriormente, pase a la siguiente sección.

En caso contrario, no siga adelante y compruebe:

Problema	Compruebe que
Pocillo blanco > 0.100DO450nm	la solución cromógeno/substrato no se ha contaminado durante el ensayo.
Control	1. el proceso de lavado y los parámetros

<p>Negativo (CN) < 1.000 DO450nm después de leer el blanco</p> <p>Coefficiente de variación > 20%</p>	<p>del lavador estén validados según los estudios previos de calificación.</p> <p>2. se ha usado la solución de lavado apropiada y que el lavador ha sido cebado con la misma antes del uso.</p> <p>3. no se han cometido errores en el procedimiento (dispensar el control positivo en lugar del negativo).</p> <p>4. no ha existido contaminación del control negativo o de sus pocillos debido a muestras positivas derramadas, o al conjugado.</p> <p>5. las micropipetas no se han contaminado con muestras positivas o con el conjugado.</p> <p>6. las agujas del lavador no estén parcial o totalmente obstruidas.</p>
<p>Calibrador DO450nm Fuera de rango</p>	<p>1. el procedimiento ha sido realizado correctamente.</p> <p>2. no ha habido errores durante su distribución (dispensar el control negativo en lugar del calibrador).</p> <p>3. el proceso de lavado y los parámetros del lavador estén validados según los estudios previos de calificación.</p> <p>4. no ha ocurrido contaminación externa del calibrador.</p>
<p>Control Positivo DO450nm > CN/10</p>	<p>1. el procedimiento ha sido realizado correctamente.</p> <p>2. no se han cometido errores en el procedimiento (dispensar el control negativo en lugar del positivo).</p> <p>3. el proceso de lavado y los parámetros del lavador estén validados según los estudios previos de calificación.</p> <p>4. no ha ocurrido contaminación externa del control positivo.</p>

Si ocurre alguno de los problemas anteriores, informe al responsable para tomar las medidas pertinentes.

Nota importante:

El análisis debe seguir el paso de lectura descrito en la sección M, punto 8.

P. RESULTADOS.

Los resultados se calculan por medio de un valor de corte (cut-off) hallado con la siguiente fórmula:

$$\text{Valor de corte} = (\text{CN} + \text{CP}) / 5$$

Nota importante: Cuando el cálculo de los resultados se halla mediante el sistema operativo de un equipo de ELISA automático, asegurarse de que la formulación usada para el cálculo del valor de corte, y para la interpretación de los resultados sea correcta.

Q. INTERPRETACIÓN DE LOS RESULTADOS.

La interpretación de los resultados se realiza mediante la razón entre las DO a 450nm / 620-630nm de las muestras y el Valor de corte Co/M.

Los resultados se interpretan según la siguiente tabla:

Co/M	Interpretación
< 0.9	Negativo
0.9 - 1.1	Equívoco
> 1.1	Positivo

Un resultado negativo indica que el paciente no está infectado por HDV.

Cualquier paciente, cuya muestra resulte equívoca debe someterse a una nueva prueba con una segunda muestra de sangre colectada 1 ó 2 semanas después de la inicial.

Un resultado positivo es indicativo de infección por HDV y por consiguiente el paciente debe ser tratado adecuadamente.

Notas importantes:

1. La interpretación de los resultados debe hacerse bajo la vigilancia del responsable del laboratorio para reducir el riesgo de errores de juicio y de interpretación.
2. Cuando se transmiten los resultados de la prueba, del laboratorio a otras instalaciones, debe ponerse mucha atención para evitar el traslado de datos erróneos.
3. El diagnóstico de infección con un virus de la hepatitis debe ser evaluado y comunicado al paciente por un médico calificado.

A continuación se incluye un ejemplo de los cálculos (datos obtenidos siguiendo el paso de lectura descrito en la sección M, punto 8).

Los siguientes datos no deben usarse en lugar de los valores reales obtenidos en el laboratorio.

Control Negativo: 2.100 – 2.200 – 2.000 DO450nm

Valor medio: 2.100 DO450nm

Mayor de 1.000 – Válido

Control Positivo: 0.100 DO450nm

Menor de CN/10 – Válido

$$\text{Valor de corte} = (2.100 + 0.100) / 5 = 0.440$$

Calibrador: 0.300-0.260 DO450nm

Valor medio: 0.280 DO450nm

Dentro del rango $CP \leq DO450nm < (CN+CP)/5$ – Válido

Muestra 1: 0.020 DO450nm

Muestra 2: 1.900 DO450nm

Muestra 1 Co/M > 1.1 positiva

Muestra 2 Co/M < 0.9 negativa

R. FUNCIONAMIENTO.

La evaluación del funcionamiento ha sido realizada según lo reportado en las Especificaciones Técnicas Comunes (ETC) (art. 5, Capítulo 3 de las Directivas IVD 98/79/EC).

1. LÍMITE DE DETECCIÓN.

En ausencia de un estándar internacional, la sensibilidad del ensayo ha sido calculada por medio de un producto denominado Accurun n° 127 suministrado por Boston Biomedical Inc., Estados Unidos.

La siguiente tabla muestra los valores de DO450nm para esta preparación, diluido en suero bovino fetal (SFB), para construir la curva de dilución límite en tres lotes diferentes:

	Valores Co/M					
	DAB.CE	Lote # 1102	DAB.CE	Lote # 0103	DAB.CE	Lote # 0403
Accurun # 127	DO450 nm	Co/M valor	DO450 nm	Co/M valor	DO450 nm	Co/M valor
1x	0.171	3.0	0.163	2.9	0.156	2.8
2x	0.187	2.7	0.176	2.6	0.179	2.5
4x	0.230	2.2	0.220	2.1	0.202	2.2
8x	0.298	1.7	0.285	1.6	0.271	1.6
16x	0.417	1.2	0.405	1.1	0.402	1.1
32x	0.514	1.0	0.490	0.9	0.482	0.9
64x	0.717	0.7	0.700	0.7	0.705	0.6
128x	1.063	0.5	1.006	0.5	1.015	0.4
CTRL (-)	2.484	////////	2.261	////////	2.114	////////

2. ESPECIFICIDAD Y SENSIBILIDAD DIAGNÓSTICA.

La evaluación del procedimiento diagnóstica se realizó mediante un ensayo con más de 400 muestras frente a un equipo de referencia. Este ensayo clínico fue conducido por el Prof. M. Rizzetto, Departamento de Gastro-Hepatología del hospital S. Giovanni Battista de Turín, Italia.

Se examinaron muestras negativas, positivas y otras que pudieran provocar interferencia.

Se emplearon además plasma sometido a métodos de tratamiento estándar (citrato, EDTA y heparina) y suero humanos. No se ha observado falsa reactividad debida a los métodos de tratamiento de muestras.

A continuación se muestran brevemente los resultados obtenidos:

Sensibilidad	> 98 %
Especificidad	> 98 %

3. PRECISIÓN.

Se realizó un estudio con 3 lotes y dos muestras de diferente reactividad anti-HDV, examinadas en 16 réplicas, en tres tandas separadas. Los valores medios obtenidos se reportan a continuación:

DAB.CE: lote #1102

Control Negativo (N = 16)

Valores medios	1ª tanda	2ª tanda	3ª tanda	Valor Promedio
DO 450nm	2.342	2.428	2.433	2.401
Desviación estándar	0.113	0.106	0.122	0.114
CV %	4.8	4.4	5.0	4.7

Calibrador (N = 16)

Valores medios	1ª tanda	2ª tanda	3ª tanda	Valor Promedio
DO 450nm	0.298	0.289	0.286	0.291
Desviación estándar	0.023	0.027	0.026	0.025
CV %	7.7	9.3	9.1	8.7
Co/M	1.6	1.7	1.7	1.7

DAB.CE: lote #0103

Control Negativo (N = 16)

Valores medios	1ª tanda	2ª tanda	3ª tanda	Valor Promedio
DO 450nm	2.208	2.237	2.246	2.230
Desviación estándar	0.105	0.108	0.108	0.107
CV %	4.7	4.8	4.8	4.8

Calibrador (N = 16)

Valores medios	1ª tanda	2ª tanda	3ª tanda	Valor Promedio
DO 450nm	0.269	0.277	0.266	0.271
Desviación estándar	0.026	0.024	0.025	0.025
CV %	9.8	8.5	9.5	9.3
Co/M	1.7	1.7	1.7	1.7

DAB.CE: lote # 0403

Control Negativo (N = 16)

Valores medios	1ª tanda	2ª tanda	3ª tanda	Valor Promedio
DO 450nm	2.246	2.221	2.182	2.216
Desviación estándar	0.097	0.103	0.118	0.106
CV %	4.3	4.6	5.4	4.8

Calibrador (N = 16)

Valores medios	1ª tanda	2ª tanda	3ª tanda	Valor Promedio
DO 450nm	0.286	0.273	0.280	0.280
Desviación estándar	0.027	0.023	0.026	0.025
CV %	9.3	8.5	9.1	9.0
Co/M	1.6	1.7	1.6	1.6

La variabilidad mostrada en las tablas no dió como resultado una clasificación errónea de las muestras.

Nota importante:

Los datos de rendimiento se obtuvieron siguiendo el paso de lectura descrito en la sección M, punto 8.

S. LIMITACIONES.

La contaminación bacteriana de las muestras o la inactivación por calor pueden modificar los valores de absorbancia con la consiguiente alteración de los niveles del analito. Este ensayo es adecuado solo para el análisis de muestras individuales y no para mezclas.

El diagnóstico de una enfermedad infecciosa no se debe formular en base al resultado de un solo ensayo, sino que es necesario tomar en consideración la historia clínica y la sintomatología del paciente así como otros datos diagnósticos.

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Todos los productos de diagnóstico in vitro fabricados por la empresa son controlados por un sistema certificado de control de calidad aprobado por un organismo notificado para el mercado CE. Cada lote se somete a un control de calidad y se libera al mercado únicamente si se ajusta a las especificaciones técnicas y criterios de aceptación de la CE.

Fabricante:
Dia.Pro Diagnostic Bioprobes S.r.l.
Via G. Carducci n° 27 – Sesto San Giovanni (Mi) – Italia



0318

HBcAb

**Competitive Enzyme Immunoassay for
the determination of antibodies
to Hepatitis B core Antigen
in human serum and plasma**

- for "in vitro" diagnostic use only -



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HBcAb

A. INTENDED USE

Competitive Enzyme ImmunoAssay (ELISA) for the determination of antibodies to Hepatitis B core Antigen in human plasma and sera.

The kit is intended for the screening of blood units and the follow-up of HBV-infected patients.

For "in vitro" diagnostic use only.

B. INTRODUCTION

The World Health Organization (WHO) defines Hepatitis B as follows:

"Hepatitis B is one of the major diseases of mankind and is a serious global public health problem. Hepatitis means inflammation of the liver, and the most common cause is infection with one of 5 viruses, called hepatitis A,B,C,D, and E. All of these viruses can cause an acute disease with symptoms lasting several weeks including yellowing of the skin and eyes (jaundice); dark urine; extreme fatigue; nausea; vomiting and abdominal pain. It can take several months to a year to feel fit again. Hepatitis B virus can cause chronic infection in which the patient never gets rid of the virus and many years later develops cirrhosis of the liver or liver cancer.

HBV is the most serious type of viral hepatitis and the only type causing chronic hepatitis for which a vaccine is available. Hepatitis B virus is transmitted by contact with blood or body fluids of an infected person in the same way as human immunodeficiency virus (HIV), the virus that causes AIDS. However, HBV is 50 to 100 times more infectious than HIV. The main ways of getting infected with HBV are: (a) perinatal (from mother to baby at the birth); (b) child-to-child transmission; (c) unsafe injections and transfusions; (d) sexual contact.

Worldwide, most infections occur from infected mother to child, from child to child contact in household settings, and from reuse of un-sterilized needles and syringes. In many developing countries, almost all children become infected with the virus. In many industrialized countries (e.g. Western Europe and North America), the pattern of transmission is different. In these countries, mother-to-infant and child-to-child transmission accounted for up to one third of chronic infections before childhood hepatitis B vaccination programmes were implemented. However, the majority of infections in these countries are acquired during young adulthood by sexual activity, and injecting drug use. In addition, hepatitis B virus is the major infectious occupational hazard of health workers, and most health care workers have received hepatitis B vaccine.

Hepatitis B virus is not spread by contaminated food or water, and cannot be spread casually in the workplace. High rates of chronic HBV infection are also found in the southern parts of Eastern and Central Europe. In the Middle East and Indian sub-continent, about 5% are chronically infected. Infection is less common in Western Europe and North America, where less than 1% are chronically infected.

Young children who become infected with HBV are the most likely to develop chronic infection. About 90% of infants infected during the first year of life and 30% to 50% of children infected between 1 to 4 years of age develop chronic

infection. The risk of death from HBV-related liver cancer or cirrhosis is approximately 25% for persons who become chronically infected during childhood.

Chronic hepatitis B in some patients is treated with drugs called *interferon or lamivudine*, which can help some patients. Patients with cirrhosis are sometimes given liver transplants, with varying success. It is preferable to prevent this disease with vaccine than to try and cure it.

Hepatitis B vaccine has an outstanding record of safety and effectiveness. Since 1982, over one billion doses of hepatitis B vaccine have been used worldwide. The vaccine is given as a series of three intramuscular doses. Studies have shown that the vaccine is 95% effective in preventing children and adults from developing chronic infection if they have not yet been infected. In many countries where 8% to 15% of children used to become chronically infected with HBV, the rate of chronic infection has been reduced to less than 1% in immunized groups of children. Since 1991, WHO has called for all countries to add hepatitis B vaccine into their national immunization programmes."

Hepatitis B core Antigen (or HBcAg) is the major component of the core particles of HBV.

HBcAg is composed of a single polypeptide of about 17 kD that is released upon disaggregating the core particles; the antigen contains at least one immunological determinant.

Upon primary infection, anti HBcAg antibodies are one of the first markers of HBV hepatitis appearing in the serum of the patient, slightly later than HBsAg, the viral surface antigen.

Anti HBcAg antibodies are produced usually at high titers and their presence is detectable even years after infection. Isolated HBcAb, in absence of other HBV markers, have been observed in infected blood units, suggesting the use of this test for screening HBV, in addition of HBsAg.

The determination of HBcAb has become important for the classification of the viral agent, together with the detection of the other markers of HBV infection, in sera and plasma.

C. PRINCIPLE OF THE TEST

The assay is based on the principle of competition where the antibodies in the sample compete with a monoclonal antibody for a fixed amount of antigen on the solid phase.

A purified recombinant HBcAg is coated to the microwells.

The patient's serum/plasma is added to the microwell together with an additive able to block interferences present in the sample.

In the second incubation after washing, a monoclonal antibody, conjugated with Horseradish Peroxidase (HRP) and specific for HBcAg is added and binds to the free rec-HBcAg coated on the plastic.

After incubation, microwells are washed to remove any unbound conjugate and then the chromogen/substrate is added. In the presence of peroxidase enzyme the colorless substrate is hydrolyzed to a colored end-product.

The color intensity is inversely proportional to the amount of antibodies to HBcAg present in the sample.

D. COMPONENTS

Each kit contains sufficient reagents to perform 96 tests.

1. Microplate **MICROPLATE**

8x12 microwell strips coated with recombinant HBcAg and sealed into a bag with desiccant. Allow the microplate to reach room temperature before opening; reseal unused strips in the bag with desiccant and store at 2..8°C.

2. Negative Control CONTROL -

1x1.0ml/vial. Ready to use. Contains 5% bovine serum albumin, 10 mM phosphate buffer pH 7.4 +/-0.1, 0.09% sodium azide and 0.1% Kathon GC as preservatives. The negative control is pale yellow color coded.

3. Positive Control CONTROL +

1x1.0ml/vial. Ready to use. Contains 5% bovine serum albumin, anti HBcAg antibodies at a concentration of about 10 PEI U/ml, (calibrated on PEI HBc Reference Material 82), 10 mM phosphate buffer pH 7.4 +/-0.1, 0.09% sodium azide and 0.1% Kathon GC as preservatives. The positive control is green color coded.

4. Calibrator CAL ...

n° 1 vial. Lyophilised. To be dissolved with EIA grade water as reported in the label. Contains fetal bovine serum, human antibodies to HBcAg at a concentration of 2 PEI U/ml +/-10% (calibrated on PEI HBc Reference Material 82) and 0.1% Kathon GC as preservative.

Note: The volume necessary to dissolve the content of the vial may vary from lot to lot. Please use the right volume reported on the label .

5. Wash buffer concentrate WASHBUF 20X

1x60ml/bottle. 20x concentrated solution. Once diluted, the wash solution contains 10 mM phosphate buffer pH 7.0+/-0.2, 0.05% Tween 20 and 0.1% Kathon GC.

6. Enzyme Conjugate CONJ

1x16ml/vial. Ready-to-use solution. Contains 5% bovine serum albumine, 10 mM tris buffer pH 6.8 +/-0.1, Horseradish peroxidase conjugated mouse monoclonal antibody to HBcAg in presence of 0.3 mg/ml gentamicine sulphate and 0.1% Kathon GC as preservatives. The component is red colour coded .

7. Chromogen/Substrate SUBS TMB

1x16ml/vial. Contains a 50 mM citrate-phosphate buffered solution at pH 3.6 +/-0.1, 0.03% tetra-methyl-benzidine (TMB), 0.02% hydrogen peroxide (H₂O₂) and 4% dimethylsulphoxide

Note: To be stored protected from light as sensitive to strong illumination.

8. Specimen Diluent DILSPE

4x3ml/vial. 10 mM tris buffered solution pH 8.0 +/-0.1 containing 0.1% Kathon GC for the pre-treatment of samples and controls in the plate, blocking interference.

Note: Use all the content of one vial before opening a second one. The reagent is sensitive to oxidation.

9. Sulphuric Acid H₂SO₄ 0.3 M

1x15ml/vial. Contains 0.3 M H₂SO₄ solution. Attention: Irritant (Xi R36/38; S2/26/30)

10. Plate sealing foil n°2

11. Instruction manual n°1

E. MATERIALS REQUIRED BUT NOT PROVIDED

1. Calibrated Micropipettes (100ul and 50ul) and disposable plastic tips.
2. EIA grade water (double distilled or deionised, charcoal treated to remove oxidizing chemicals used as disinfectants).
3. Timer with 60 minute range or higher.
4. Absorbent paper tissues.
5. Calibrated ELISA microplate thermostatic incubator (dry or wet) set at +37°C.
6. Calibrated ELISA microwell reader with 450nm (reading) and with 620-630nm (blanking) filters.
7. Calibrated ELISA microplate washer.
8. Vortex or similar mixing tools.

F. WARNINGS AND PRECAUTIONS

1. The kit has to be used by skilled and properly trained technical personnel only, under the supervision of a medical doctor responsible of the laboratory.
2. When the kit is used for the screening of blood units and blood components, it has to be used in a laboratory certified and qualified by the national authority in that field (Ministry of Health or similar entity) to carry out this type of analysis.
3. All the personnel involved in performing the assay have to wear protective laboratory clothes, talc-free gloves and glasses. The use of any sharp (needles) or cutting (blades) devices should be avoided. All the personnel involved should be trained in biosafety procedures, as recommended by the Center for Disease Control, Atlanta, U.S. and reported in the National Institute of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
4. All the personnel involved in sample handling should be vaccinated for HBV and HAV, for which vaccines are available, safe and effective.
5. The laboratory environment should be controlled so as to avoid contaminants such as dust or air-born microbial agents, when opening kit vials and microplates and when performing the test. Protect the Chromogen (TMB) from strong light and avoid vibration of the bench surface where the test is undertaken.
6. Upon receipt, store the kit at 2-8°C into a temperature controlled refrigerator or cold room.
7. Do not interchange components between different lots of the kits. It is recommended that components between two kits of the same lot should not be interchanged.
8. Check that the reagents are clear and do not contain visible heavy particles or aggregates. If not, advise the laboratory supervisor to initiate the necessary procedures.
9. Avoid cross-contamination between serum/plasma samples by using disposable tips and changing them after each sample.
10. Avoid cross-contamination between kit reagents by using disposable tips and changing them between the use of each one.
11. Do not use the kit after the expiration date stated on external (primary container) and internal (vials) labels.
12. Treat all specimens as potentially infective. All human serum specimens should be handled at Biosafety Level 2, as recommended by the Center for Disease Control, Atlanta, U.S. in compliance with what reported in the Institutes of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
13. The use of disposable plastic-ware is recommended in the preparation of the washing solution or in transferring components into other containers of automated workstations, in order to avoid contamination.
14. Waste produced during the use of the kit has to be discarded in compliance with national directives and laws concerning laboratory waste of chemical and biological substances. In particular, liquid waste generated from the washing procedure, from residuals of controls and from samples has to be treated as potentially infective material and inactivated. Suggested procedures of inactivation are treatment with a 10% final concentration of household bleach for 16-18 hrs or heat inactivation by autoclave at 121°C for 20 min..
15. Accidental spills have to be adsorbed with paper tissues soaked with household bleach and then with water. Tissues should then be discarded in proper containers designated for laboratory/hospital waste.
16. The Sulphuric Acid is an irritant. In case of spills, wash the surface with plenty of water.
17. Other waste materials generated from the use of the kit (example: tips used for samples and controls, used microplates) should be handled as potentially infective and

disposed according to national directives and laws concerning laboratory wastes.

G. SPECIMEN: PREPARATION AND RECOMMANDATIONS

1. Blood is drawn aseptically by venepuncture and plasma or serum is prepared using standard techniques of preparation of samples for clinical laboratory analysis. No influence has been observed in the preparation of the sample with citrate, EDTA and heparin.
2. Avoid any addition of preservatives to samples; especially sodium azide as this chemical would affect the enzymatic activity of the conjugate.
3. Samples have to be clearly identified with codes or names in order to avoid misinterpretation of results. When the kit is used for the screening of blood units, bar code labeling and electronic reading is strongly recommended.
4. Haemolysed (red) and visibly hyperlipemic ("milky") samples have to be discarded as they could generate false results. Samples containing residues of fibrin or heavy particles or microbial filaments and bodies should be discarded as they could give rise to false results.
5. Sera and plasma can be stored at +2°.8°C for up to five days after collection. For longer storage periods, samples can be stored frozen at -20°C for several months. Any frozen samples should not be frozen/thawed more than once as this may generate particles that could affect the test result.
6. If particles are present, centrifuge at 2.000 rpm for 20 min or filter using 0.2-0.8µ filters to clean up the sample for testing.

H. PREPARATION OF COMPONENTS AND WARNINGS

A study conducted on an opened kit has not pointed out any relevant loss of activity up to 6 re-uses of the device and up to 6 months.

1. Microplates:

Allow the microplate to reach room temperature (about 1 hr) before opening the container. Check that the desiccant has not turned dark green, indicating a defect in storage. In this case, call Dia.Pro's customer service.

Unused strips have to be placed back inside the aluminum pouch, with the desiccant supplied, firmly zipped and stored at +2°.8°C. After first opening, remaining strips are stable until the humidity indicator inside the desiccant bag turns from yellow to green.

2. Negative Control:

Ready to use. Mix well on vortex before use.

3. Positive Control:

Ready to use. Mix well on vortex before use.

4. Calibrator:

Add the volume of ELISA grade water, reported on the label, to the lyophilised powder; let fully dissolve and then gently mix on vortex.

Note: *The dissolved calibrator is not stable. Store it frozen in aliquots at -20°C.*

5. Wash buffer concentrate:

The whole content of the concentrated solution has to be diluted 20x with bidistilled water and mixed gently end-over-end before use. During preparation avoid foaming as the presence of bubbles could impact on the efficiency of the washing cycles.

Note: *Once diluted, the wash solution is stable for 1 week at +2°.8°C.*

6. Enzyme conjugate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, dust or microbes. If this component has to be transferred, use only plastic, and if possible, sterile disposable containers.

7. Chromogen/Substrate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, air-driven dust or microbes. Do not expose to strong light, oxidizing agents and metallic surfaces.

If this component has to be transferred use only plastic, and if possible, sterile disposable container.

8. Specimen Diluent

Ready to use solution. Mix gently on vortex before use. Use all the content of one vial before opening a second one. The reagent is sensitive to oxidation.

9. Sulphuric Acid:

Ready to use. Mix well on vortex before use.

Attention: Irritant (Xi R36/38; S2/26/30)

Legenda: R 36/38 = Irritating to eyes and skin.

S 2/26/30 = In case of contact with eyes, rinse immediately with plenty of water and seek medical advice.

I. INSTRUMENTS AND TOOLS USED IN COMBINATION WITH THE KIT

1. Micropipettes have to be calibrated to deliver the correct volume required by the assay and must be submitted to regular decontamination (70% ethanol, 10% solution of bleach, hospital grade disinfectants) of those parts that could accidentally come in contact with the sample or the components of the kit. They should also be regularly maintained in order to show a precision of 1% and a trueness of ±2%.
2. The ELISA incubator has to be set at +37°C (tolerance of ±0.5°C) and regularly checked to ensure the correct temperature is maintained. Both dry incubators and water baths are suitable for the incubations, provided that the instrument is validated for the incubation of ELISA tests.
3. The ELISA washer is extremely important to the overall performances of the assay. The washer must be carefully validated and correctly optimized using the kit controls/calibrator and reference panels, before using the kit for routine laboratory tests. Usually 4-5 washing cycles (aspiration + dispensation of 350 µl/well of washing solution = 1 cycle) are sufficient to ensure that the assay performs as expected. A soaking time of 20-30 seconds between cycles is suggested. In order to set correctly their number, it is recommended to run an assay with the kit controls/calibrator and well characterized negative and positive reference samples, and check to match the values reported below in the sections "Validation of Test" and "Assay Performances". Regular calibration of the volumes delivered and maintenance (decontamination and cleaning of needles) of the washer has to be carried out according to the instructions of the manufacturer.
4. Incubation times have a tolerance of ±5%.
5. The ELISA microplate reader has to be equipped with a reading filter of 450nm and with a second filter (620-630nm, strongly recommended) for blanking purposes. Its standard performances should be (a) bandwidth ≤ 10 nm; (b) absorbance range from 0 to ≥ 2.0; (c) linearity to ≥ 2.0; repeatability ≥ 1%. Blanking is carried out on the well identified in the section "Assay Procedure". The optical system of the reader has to be calibrated regularly to ensure that the correct optical density is measured. It should be regularly maintained according to the manufacturer 's instructions.

- When using an ELISA automated work station, all critical steps (dispensation, incubation, washing, reading, shaking, data handling) have to be carefully set, calibrated, controlled and regularly serviced in order to match the values reported in the sections "Validation of Test" and "Assay Performances". The assay protocol has to be installed in the operating system of the unit and validated as for the washer and the reader. In addition, the liquid handling part of the station (dispensation and washing) has to be validated and correctly set. Particular attention must be paid to avoid carry over by the needles used for dispensing samples and for washing. This must be studied and controlled to minimize the possibility of contamination of adjacent wells due to strongly reactive samples, leading to false positive results. The use of ELISA automated work stations is recommended for blood screening and when the number of samples to be tested exceed 20-30 units per run.
- Dia.Pro's customer service offers support to the user in the setting and checking of instruments used in combination with the kit, in order to assure full compliance with the requirements described. Support is also provided for the installation of new instruments to be used with the kit.

L. PRE ASSAY CONTROLS AND OPERATIONS

- Check the expiration date of the kit printed on the external label (primary container). Do not use if expired.
- Check that the liquid components are not contaminated by visible particles or aggregates. Check that the Chromogen (TMB) is colourless or pale blue by aspirating a small volume of it with a sterile plastic pipette. Check that no breakage occurred in transportation and no spillage of liquid is present inside the box (primary container). Check that the aluminium pouch, containing the microplate, is not punctured or damaged.
- Dilute all the content of the 20x concentrated Wash Solution as described above.
- Dissolve the Calibrator as described above and gently mix.
- Allow all the other components to reach room temperature (about 1 hr) and then mix gently on vortex all liquid reagents.
- Set the ELISA incubator at +37°C and prepare the ELISA washer by priming with the diluted washing solution, according to the manufacturers instructions. Set the right number of washing cycles as found in the validation of the instrument for its use with the kit.
- Check that the ELISA reader is turned on or ensure it will be turned on at least 20 minutes before reading.
- If using an automated work station, turn on, check settings and be sure to use the right assay protocol.
- Check that the micropipettes are set to the required volume.
- Check that all the other equipment is available and ready to use.
- In case of problems, do not proceed further with the test and advise the supervisor.

M. ASSAY PROCEDURE

The assay has to be performed according to the procedure given below, taking care to maintain the same incubation time for all the samples being tested.

- Place the required number of strips in the plastic holder and carefully identify the wells for controls, calibrator and samples.
- Leave the A1 well empty for blanking purposes.
- Dispense 50 µl Specimen Diluent into all the control and sample wells.
- Pipette 50 µl of the Negative Control in triplicate, 50 µl of the Calibrator in duplicate and then 50 µl of the Positive Control in single. Then dispense 50 µl of each of the samples.
- Incubate the microplate for **60 min at +37°C**.

Important note: Strips have to be sealed with the adhesive sealing foil, only when the test is performed manually. Do not cover strips when using ELISA automatic instruments.

- When the first incubation is finished, wash the microwells as previously described (section I.3)
- Pipette 100 µl Enzyme Conjugate in all the wells, except A1; incubate the microplate for **60 min at +37°C**.

Important note: Be careful not to touch the plastic inner surface of the well with the tip filled with the Enzyme Conjugate. Contamination might occur.

- When the second incubation is finished, wash the microwells as previously described (section I.3)
- Pipette 100 µl Chromogen/Substrate into all the wells, A1 included.

Important note: Do not expose to strong direct light. as a high background might be generated.

- Incubate the microplate protected from light at **room temperature (18-24°C) for 20 minutes**. Wells dispensed with negative control and negative samples will turn from clear to blue (competitive method).
- Pipette 100 µl Sulphuric Acid into all the wells using the same pipetting sequence as in step 9 to stop the enzymatic reaction. Addition of the stop solution will turn the negative control and negative samples from blue to yellow.
- Measure the colour intensity of the solution in each well, as described in section I.5 using a 450nm filter (reading) and a 620-630nm filter (background subtraction, strongly recommended), blanking the instrument on A1.

Important notes:

- If the second filter is not available, ensure that no finger prints are present on the bottom of the microwell before reading at 450nm. Finger prints could generate false positive results on reading.
- Reading has should ideally be performed immediately after the addition of the Stop Solution but definitely no longer than 20 minutes afterwards. Some self oxidation of the chromogen can occur leading to a higher background.

N. ASSAY SCHEME

Specimen Diluent	50 µl
Controls&calibrator and samples	50 µl
1st incubation	60 min
Temperature	+37°C
Wash	n° 4-5
Enzyme Conjugate	100 µl
2nd incubation	60 min
Temperature	+37°C
Wash	n° 4-5
TMB/H ₂ O ₂ mix	100 µl
3rd incubation	20 min
Temperature	r.t.
Sulphuric Acid	100 µl
Reading OD	450nm

An example of dispensation scheme is reported below:

Microplate

	1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	S2										
B	NC	S3										
C	NC	S4										
D	NC	S5										
E	CAL	S6										
F	CAL	S7										
G	PC	S8										
H	S1	S9										

Legenda: BLK = Blank NC = Negative Control
CAL = Calibrator PC = Positive Control S = Sample

O. INTERNAL QUALITY CONTROL

A check is performed on the controls/calibrator any time the kit is used in order to verify whether the expected OD450nm or Co/S values have been matched in the analysis.

Ensure that the following parameters are met:

Parameter	Requirements
Blank well	< 0.050 OD450nm value
Negative Control (NC)	> 1.000 OD450nm after blanking coefficient of variation < 20%
Calibrator (about 2 PEI U/ml)	Co/S > 1
Positive Control	< 0.200 OD450nm

If the results of the test match the requirements stated above, proceed to the next section.

If they do not, do not proceed any further and perform the following checks:

Problem	Check
Blank well > 0.050 OD450nm	that the Chromogen/Substrate solution has not become contaminated during the assay
Negative Control (NC) < 1.000 OD450nm after blanking coefficient of variation > 20%	1. that the washing procedure and the washer settings are as validated in the pre qualification study; 2. that the proper washing solution has been used and the washer has been primed with it before use; 3. that no mistake has been done in the assay procedure (dispensation of positive control instead of negative control); 4. that no contamination of the negative control or of the wells where the control was dispensed has occurred due to positive samples, to spills or to the enzyme conjugate; 5. that micropipettes have not become contaminated with positive samples or with the enzyme conjugate 6. that the washer needles are not blocked or partially obstructed.
Calibrator Co/S < 1	1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution (ex.: dispensation of negative control instead 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the calibrator has occurred.
Positive Control > 0.200 OD450nm	1. that the procedure has been correctly performed; 2. that no mistake has occurred during the distribution of the control (dispensation of negative control instead of positive control). 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the positive control has occurred.

If any of the above problems have occurred, report the problem to the supervisor for further actions.

P. RESULTS

The results are calculated by means of a cut-off value determined with the following formula:

$$\text{Cut-Off} = (\text{NC} + \text{PC}) / 5$$

Important note: When the calculation of results is performed by the operating system of an ELISA automated work station, ensure that the proper formulation is used to calculate the cut-off value and generate the correct interpretation of results.

Q. INTERPRETATION OF RESULTS

Results are interpreted as ratio between the cut-off value and the sample OD450nm or Co/S.

Results are interpreted according to the following table:

Co/S	Interpretation
< 0.9	Negative
0.9 - 1.1	Equivocal
> 1.1	Positive

A negative result indicates that the patient has not been infected by HBV.

Any patient showing an equivocal result should be re-tested on a second sample taken 1-2 weeks after the initial sample. The blood unit should not be transfused.

A positive result is indicative of HBV infection and therefore the patient should be treated accordingly or the blood unit should be discarded.

Important notes:

1. Interpretation of results should be done under the supervision of the laboratory supervisor to reduce the risk of judgement errors and misinterpretations.
2. When test results are transmitted from the laboratory to another facility, attention must be paid to avoid erroneous data transfer.
3. Diagnosis of viral hepatitis infection has to be taken by and released to the patient by a suitably qualified medical doctor.

An example of calculation is reported below.

The following data must not be used instead of real figures obtained by the user.

Negative Control: 2.000 – 2.200 – 2.000 OD450nm
Mean Value: 2.100 OD450nm
Higher than 1.000 – Accepted

Positive Control: 0.100 OD450nm
Lower than 0.200 – Accepted

$$\text{Cut-Off} = (2.100 + 0.100) / 5 = 0.440$$

Calibrator: 0.400-0.360 OD450nm
Mean value: 0.380 OD450nm
Co/S > 1 – Accepted

Sample 1: 0.028 OD450nm
Sample 2: 1.890 OD450nm
Sample 1 Co/S > 1.1 positive
Sample 2 Co/S < 0.9 negative

R. PERFORMANCES

Evaluation of Performances has been conducted in accordance to what reported in the Common Technical Specifications or CTS (art. 5, Chapter 3 of IVD Directive 98/79/EC).

1. LIMIT OF DETECTION:

The sensitivity of the assay has been calculated by means of the reference preparation for HBcAb supplied by Paul Erlich Institute (PEI HBc Reference Material 82). The assay shows a sensitivity of about 1.25 PEI U/ml.

The table below reports the Co/S values shown by the PEI standard diluted as suggested by the manufacturer to prepare a limiting dilution curve in Fetal Calf Serum (FCS).

PEI U/ml	Lot 1001	Lot 0702	Lot 0702/2	Lot 1202
5	22.6	18.0	19.0	17.7
2.5	8.0	5.5	5.4	5.0
1.25	1.1	1.3	1.0	1.0
0.625	0.4	0.4	0.4	0.4

In addition Accurun 1 – series 3000 – supplied by Boston Biomedica Inc., USA, was tested to determine its Co/S value. Results are reported in the table below:

Accurun 1 – series 3000

Value	Lot 1001	Lot 0702	Lot 1202
Co/S	2.9	2.3	2.2

2. DIAGNOSTIC SPECIFICITY AND SENSITIVITY

The Performance Evaluation of the device was carried out in a trial conducted on more than total 6000 samples.

2.1 Diagnostic Specificity

It is defined as the probability of the assay of scoring negative in the absence of specific analyte. A total of more 5000 unselected donors, including 1st time donors, were examined.

In a first study 2023 samples were tested against a US company as reference. A specificity of 99.5% was found. In a second study 1588 samples were examined against a European company. A specificity of 99.7% was found. In the last study 1565 samples were assayed against the same US company; a value of 99.8% was found.

In addition to the above population, 206 samples from hospitalized patients were tested against the European company. A value of 99.3% specificity was found.

Moreover, diagnostic specificity was assessed by testing 164 potentially interfering specimens (other infectious diseases, patients affected by non viral hepatic diseases, dialysis patients, pregnant women, hemolyzed, lipemic, etc.) against the European company. A value of specificity of 100% was assessed.

Finally, both human plasma, derived with different standard techniques of preparation (citrate, EDTA and heparin), and human sera have been used to determine the specificity.

No false reactivity due to the method of specimen preparation has been observed.

2.2 Diagnostic Sensitivity

It defined as the probability of the assay of scoring positive in the presence of specific analyte.

373 positive specimens were tested against the European company; a diagnostic sensitivity of 99.7 was found.

3. PRECISION

The mean values obtained from a study conducted on three lots and on two samples of different anti-HBcAg reactivity, examined in 16 replicates in three separate runs is reported below:

BCAB.CE: lot # 1202

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	1.943	1.939	1.924	1.935
Std.Deviation	0.081	0.078	0.103	0.087
CV %	4.2	4.0	5.3	4.5

Calibrator (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.143	0.147	0.148	0.146
Std.Deviation	0.014	0.017	0.018	0.016
CV %	9.8	11.4	12.1	11.1
Co/S	2.8	2.7	2.6	2.7

BCAB.CE: lot # 0702

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.163	2.110	2.106	2.126
Std.Deviation	0.105	0.088	0.139	0.111
CV %	4.9	4.2	6.6	5.2

Calibrator (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.182	0.193	0.195	0.190
Std.Deviation	0.018	0.023	0.019	0.020
CV %	10.0	12.0	9.9	10.6
Co/S	2.5	2.2	2.3	2.3

BCAB.CE: lot # 0702/2

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.278	2.098	2.130	2.169
Std.Deviation	0.135	0.126	0.159	0.140
CV %	5.9	6.0	7.5	6.5

Calibrator (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.193	0.190	0.199	0.134
Std.Deviation	0.023	0.023	0.027	0.025
CV %	12.1	12.3	13.5	12.6
Co/S	2.4	2.2	2.2	2.3

The variability shown in the tables did not result in sample misclassification.

S. LIMITATIONS OF THE PROCEDURE

Bacterial contamination or heat inactivation of the specimen may affect the absorbance values of the samples with consequent alteration of the level of the analyte. This test is suitable only for testing single samples and not pooled ones.

Diagnosis of an infectious disease should not be established on the basis of a single test result. The patient's clinical history, symptomatology, as well as other diagnostic data should be considered.

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All the IVD Products manufactured by the company are under the control of a certified Quality Management System approved by an EC Notified Body. Each lot is submitted to a quality control and released into the market only if conforming with the EC technical specifications and acceptance criteria.

Produced by
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HBe Ag&Ab

**Enzyme Immunoassay (ELISA) for the
determination of Hepatitis B Virus
"e" Antigen and Antibody
in human plasma and sera.**

- for "in vitro" diagnostic use only -



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HBe Ag&Ab

A. INTENDED USE

Enzyme ImmunoAssay (ELISA) for the determination of Hepatitis B Virus "e" Antigen and Antibody in human plasma and sera.

The kit is intended for the follow-up of acute infection and of chronic patients under therapy.

For "in vitro" diagnostic use only.

B. INTRODUCTION

Hepatitis B "e" Antigen or HBeAg is known to be intimately associated with Hepatitis B Virus or HBV replication and the presence of infectious Dane particles in the blood.

Recently, it has been found that HBeAg is a product of proteolytic degradation of Hepatitis B core Antigen or HBcAg, occurring in hepatocytes, whose expression is under the control of the precore region of HBV genome.

If HBeAg is considered a specific marker of infectivity, the presence of anti HBeAg antibodies in blood is recognised to be a clinical sign of recovery from infection to convalescence.

The determination of these two analytes in samples from HBV patients has become important for the classification of the phase of illness and as a prognostic value in the follow up of infected patients.

C. PRINCIPLE OF THE TEST

HBeAg:

HBeAg, if present in the sample, is captured by a specific monoclonal antibody, in the 1st incubation.

In the 2nd incubation, after washing, a tracer, composed of a mix of two specific anti HBeAg monoclonal antibodies, labeled with peroxidase (HRP), is added to the microplate and binds to the captured HBeAg.

The concentration of the bound enzyme on the solid phase is proportional to the amount of HBeAg in the sample and its activity is detected by adding the chromogen/substrate in the 3rd incubation.

The presence of HBeAg in the sample is determined by means of a cut-off value that allows for the semiquantitative detection of the antigen.

HBeAb

Anti HBeAg antibodies, if present in the sample, compete with a recombinant HBeAg preparation for a fixed amount of an anti HBeAg antibody, coated on the microplate wells.

The competitive assay is carried out in two incubations, the first with the sample and rechBeAg, and the second with a tracer, composed of two anti HBeAg monoclonal antibodies, labeled with peroxidase (HRP).

The concentration of the bound enzyme on the solid phase becomes inversely proportional to the amount of anti HBeAg antibodies in the sample and its activity is detected by adding the chromogen/substrate in the third incubation.

The concentration of HBeAg specific antibodies in the sample is determined by means of a cut-off value that allows for the semi quantitative detection of anti HBeAg antibodies.

D. COMPONENTS

The kit contains reagents for total 96 tests.

1. Microplate: MICROPLATE

n° 1 coated microplate

12 strips of 8 breakable wells coated with anti HBeAg specific monoclonal antibody, postcoated with bovine serum proteins and sealed into a bag with desiccant. Allow the microplate to reach room temperature before opening; reseal unused strips in the bag with desiccant and store at 2..8°C.

2. Negative Control: CONTROL

1x2.0ml/vial. Ready to use control. It contains bovine serum, 0.09% sodium azide and 0.045% ProClin 300 as preservatives. The negative control is colorless.

3. Antigen Positive Control: CONTROL + Ag

1x1.0ml/vial. Ready to use control. It contains 2% bovine serum albumin, non infectious recombinant HBeAg, 100 mM tris buffer pH 7.4+/-0.1, 0.09% sodium azide and 0.045% ProClin 300 as preservatives.

The positive control is green color coded.

4. Antibody Positive Control: CONTROL + Ab

1x1.0ml/vial. Ready to use control. It contains 2% bovine serum albumin, human anti HbeAg positive plasma at about 10 PEI U/ml, 100 mM tris buffer pH 7.4+/-0.1, 0.09% sodium azide and 0.045% ProClin 300 as preservatives. The label is red colored.

The positive control is yellow color coded.

5. Antigen Calibrator: CALAG ...ml

n° 1 vial. Lyophilized calibrator for HBeAg. To be dissolved with EIA grade water as reported in the label. It contains fetal bovine serum, non infectious recombinant HBeAg at 1 PEI U/ml +/-10%, 0.02% gentamicine sulphate and 0.045% ProClin 300 as preservatives.

Important Note: The volume necessary to dissolve the content of the vial may vary from lot to lot. Please use the right volume reported on the label.

6. Antibody Calibrator: CALAB ...ml

n° 1 vial. Lyophilized calibrator for anti HBeAg antibody. To be dissolved with EIA grade water as reported in the label. It contains fetal bovine serum, positive plasma at 0.25 PEI U/ml +/-10%, 0.02% gentamicine sulphate and 0.045% ProClin 300 as preservatives. The label is red colored.

Important Note: The volume necessary to dissolve the content of the vial may vary from lot to lot. Please use the right volume reported on the label.

7. Wash buffer concentrate: WASHBUF 20X

1x60ml/bottle. 20x concentrated solution.

Once diluted, the wash solution contains 10 mM phosphate buffer pH 7.0+/-0.2, 0.05% Tween 20 and 0.045% ProClin 300.

8. Enzyme conjugate: CONJ

1x16ml/vial. Ready to use conjugate. It contains Horseradish peroxidase conjugated with a mix of monoclonal antibodies to HBeAg, 10 mM Tris buffer pH 6.8+/-0.1, 2% BSA, 0.045% ProClin 300 and 0.02% gentamicine sulphate as preservatives. The reagent is red color coded.

9. HBe Antigen: Ag-HBe

1x10ml/vial. Ready to use reagent. It contains recombinant HBeAg, fetal bovine serum, buffered solution pH 8.0+/-0.1, 0.045% ProClin 300 and 0.09% sodium azide as preservatives. The reagent is blue color coded.

10. Chromogen/Substrate: SUBS TMB

1x16ml/vial. Ready-to-use component. It contains a 50 mM citrate-phosphate buffered solution at pH 3.5-3.8, 4% dimethylsulphoxide, 0.03% tetra-methyl-benzidine or TMB and 0.02% hydrogen peroxide or H₂O₂.

Note: To be stored protected from light as sensitive to strong illumination.

11. Sulphuric Acid: H₂SO₄ 0.3 M

1x15ml/vial. It contains 0.3 M H₂SO₄ solution.

Attention: Irritant (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

12. Plate sealing foils n°2

13. Package insert n°1

E. MATERIALS REQUIRED BUT NOT PROVIDED

1. Calibrated Micropipettes (150ul, 100ul and 50ul) and disposable plastic tips.
2. EIA grade water (double distilled or deionised, charcoal treated to remove oxidizing chemicals used as disinfectants).
3. Timer with 60 minute range or higher.
4. Absorbent paper tissues.
5. Calibrated ELISA microplate thermostatic incubator (dry or wet) set at +37°C.
6. Calibrated ELISA microwell reader with 450nm (reading) and with 620-630nm (blinking) filters.
7. Calibrated ELISA microplate washer.
8. Vortex or similar mixing tools.

F. WARNINGS AND PRECAUTIONS

1. The kit has to be used by skilled and properly trained technical personnel only, under the supervision of a medical doctor responsible of the laboratory.
2. All the personnel involved in performing the assay have to wear protective laboratory clothes, talc-free gloves and glasses. The use of any sharp (needles) or cutting (blades) devices should be avoided. All the personnel involved should be trained in biosafety procedures, as recommended by the Center for Disease Control, Atlanta, U.S. and reported in the National Institute of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
3. All the personnel involved in sample handling should be vaccinated for HBV and HAV, for which vaccines are available, safe and effective.
4. The laboratory environment should be controlled so as to avoid contaminants such as dust or air-born microbial agents, when opening kit vials and microplates and when performing the test. Protect the Chromogen/Substrate (TMB) from strong light and avoid vibration of the bench surface where the test is undertaken.
5. Upon receipt, store the kit at 2-8°C into a temperature controlled refrigerator or cold room.
6. Do not interchange components between different lots of the kits. It is recommended that components between two kits of the same lot should not be interchanged.
7. Check that the reagents are clear and do not contain visible heavy particles or aggregates. If not, advise the laboratory supervisor to initiate the necessary procedures.
8. Avoid cross-contamination between serum/plasma samples by using disposable tips and changing them after each sample. Do not reuse disposable tips.
9. Avoid cross-contamination between kit reagents by using disposable tips and changing them between the use of each one. Do not reuse disposable tips.
10. Do not use the kit after the expiration date stated on external (primary container) and internal (vials) labels.
11. Treat all specimens as potentially infective. All human serum specimens should be handled at Biosafety Level 2, as recommended by the Center for Disease Control, Atlanta, U.S. in compliance with what reported in the Institutes of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
12. The use of disposable plastic-ware is recommended in the preparation of the washing solution or in transferring components into other containers of automated workstations, in order to avoid contamination.
13. Waste produced during the use of the kit has to be discarded in compliance with national directives and laws concerning laboratory waste of chemical and biological substances. In particular, liquid waste generated from the washing procedure, from residuals of controls and from samples has to be treated as potentially infective material and inactivated. Suggested procedures of inactivation are

treatment with a 10% final concentration of household bleach for 16-18 hrs or heat inactivation by autoclave at 121°C for 20 min..

14. Accidental spills have to be adsorbed with paper tissues soaked with household bleach and then with water. Tissues should then be discarded in proper containers designated for laboratory/hospital waste.

15. The Stop Solution is an irritant. In case of spills, wash the surface with plenty of water

16. Other waste materials generated from the use of the kit (example: tips used for samples and controls, used microplates) should be handled as potentially infective and disposed according to national directives and laws concerning laboratory wastes.

G. SPECIMEN: PREPARATION AND RECOMMENDATIONS

1. Blood is drawn aseptically by venepuncture and plasma or serum is prepared using standard techniques of preparation of samples for clinical laboratory analysis. No influence has been observed in the preparation of the sample with citrate, EDTA and heparin.
2. Avoid any addition of preservatives; especially sodium azide as this chemical would affect the enzymatic activity of the conjugate, generating false negative results.
3. Samples have to be clearly identified with codes or names in order to avoid misinterpretation of results.
4. Haemolysed and visibly hyperlipemic ("milky") samples have to be discarded as they could generate false results. Samples containing residues of fibrin or heavy particles or microbial filaments and bodies should be discarded as they could give rise to false results.
5. Sera and plasma can be stored at +2°...+8°C in primary collection tubes for up to five days after collection. Do not freeze primary tubes of collection. For longer storage periods, sera and plasma samples, carefully removed from the primary collection tube, can be stored frozen at -20°C for at least 12 months. Any frozen samples should not be frozen/thawed more than once as this may generate particles that could affect the test result.
6. If particles are present, centrifuge at 2.000 rpm for 20 min or filter using 0.2-0.8u filters to clean up the sample for testing.

H. PREPARATION OF COMPONENTS AND WARNINGS

A study conducted on an opened kit has not pointed out any relevant loss of activity up to 6 re-uses of the device and up to 3 months.

1. Microplate:

Allow the microplate to reach room temperature (about 1 hr) before opening the container. Check that the desiccant has not turned dark green, indicating a defect in manufacturing. In this case, call Dia.Pro's customer service.

Unused strips have to be placed back into the aluminum pouch, with the desiccant supplied, firmly zipped and stored at +2°-8°C. When opened the first time, unused strips are stable until the humidity indicator inside the desiccant bag turns from yellow to green.

2. Negative Control:

Ready to use. Mix well on vortex before use.

3. Antigen Positive Control:

Ready to use. Mix well on vortex before use.

4. Antibody Positive Control:

Ready to use. Mix well on vortex before use.

5. Antigen Calibrator:

Add the volume of ELISA grade water, reported on the label, to the lyophilized powder; let fully dissolve and then gently mix on vortex.

Note: The dissolved calibrator is not stable. Store it frozen in aliquots at -20°C.

6. Antibody Calibrator:

Add the volume of ELISA grade water, reported on the label, to the lyophilized powder; let fully dissolve and then gently mix on vortex.

Note: The dissolved calibrator is not stable. Store it frozen in aliquots at -20°C.

7. Wash buffer concentrate:

The whole content of the 20x concentrated solution has to be diluted with bidistilled water up to 1200 ml and mixed gently end-over-end before use.

During preparation avoid foaming as the presence of bubbles could impact on the efficiency of the washing cycles.

Note: Once diluted, the wash solution is stable for 1 week at +2..8° C.

8. Enzyme conjugate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, air-driven dust or microbes. If this component has to be transferred, use only plastic, and if possible, sterile disposable containers.

9. HBe Antigen:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, air-driven dust or microbes. If this component has to be transferred, use only plastic, and if possible, sterile disposable containers.

10. Chromogen/Substrate:

Ready to use. Mix well on vortex before use.

Avoid contamination of the liquid with oxidizing chemicals, air-driven dust or microbes. Do not expose to strong light, oxidizing agents and metallic surfaces.

If this component has to be transferred use only plastic, and if possible, sterile disposable container.

11. Sulphuric Acid:

Ready to use. Mix well on vortex before use.

Attention: Irritant (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Legenda:

Warning H statements:

H315 – Causes skin irritation.

H319 – Causes serious eye irritation.

Precautionary P statements:

P280 – Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352 – IF ON SKIN: Wash with plenty of soap and water.

P332 + P313 – If skin irritation occurs: Get medical advice/attention.

P305 + P351 + P338 – IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P337 + P313 – If eye irritation persists: Get medical advice/attention.

P362 + P363 – Take off contaminated clothing and wash it before reuse.

I. INSTRUMENTS AND TOOLS USED IN COMBINATION WITH THE KIT

- Micropipettes have to be calibrated to deliver the correct volume required by the assay and must be submitted to regular decontamination (household alcohol, 10% solution of bleach, hospital grade disinfectants) of those parts that could accidentally come in contact with the sample. Decontamination of spills or residues of kit components should also be carried out regularly. They should also be regularly maintained in order to show a precision of 1% and a trueness of $\pm 2\%$.
- The ELISA incubator has to be set at +37°C (tolerance of $\pm 0.5^\circ\text{C}$) and regularly checked to ensure the correct temperature is maintained. Both dry incubators and water baths are suitable for the incubations, provided that the instrument is validated for the incubation of ELISA tests.
- The **ELISA washer** is extremely important to the overall performances of the assay. The washer must be carefully validated in advance, checked for the delivery of the right dispensation volume and regularly submitted to maintenance according to the manufacturer's instructions for use. In particular the washer, at the end of the daily workload, has to be extensively cleaned out of salts with deionized water. Before use, the washer has to be extensively primed with the diluted Washing Solution. The instrument weekly has to be submitted to decontamination according to its manual (NaOH 0.1 M decontamination suggested).
5 washing cycles (aspiration + dispensation of 350ul/well of washing solution + 20 sec soaking = 1 cycle) are sufficient to ensure the assay with the declared performances. If soaking is not possible add one more cycle of washing. An incorrect washing cycle or salt-blocked needles are the major cause of false positive reactions.
- Incubation times have a tolerance of $\pm 5\%$.
- The ELISA reader has to be equipped with a reading filter of 450nm and with a second filter of 620-630nm, mandatory for blanking purposes. Blanking is carried out on the well identified in the section "Assay Procedure". The optical system of the reader has to be calibrated regularly to ensure the correct optical density is measured. It should be regularly maintained according to the manufacturer's instructions.
- When using an ELISA automated work station, all critical steps (dispensation, incubation, washing, reading, data handling) have to be carefully set, calibrated, controlled and regularly serviced in order to match the values reported in the section "Internal Quality Control". The assay protocol has to be installed in the operating system of the unit and validated as for the washer and the reader. In addition, the liquid handling part of the station (dispensation and washing) has to be validated and correctly set. Particular attention must be paid to avoid carry over by the needles used for dispensing and for washing. This must be studied and controlled to minimize the possibility of contamination of adjacent wells. The use of ELISA automated work stations is recommended when the number of samples to be tested exceed 20-30 units per run.
- Dia.Pro's customer service offers support to the user in the setting and checking of instruments used in combination with the kit, in order to assure compliance with the requirements described. Support is also provided for the installation of new instruments to be used with the kit.

L. PRE ASSAY CONTROLS AND OPERATIONS

- Check the expiration date of the kit printed on the external label (primary container). Do not use if expired.
- Check that the liquid components are not contaminated by visible particles or aggregates. Check that the Chromogen/Substrate (TMB+H₂O₂) is colourless or pale blue by aspirating a small volume of it with a sterile plastic pipette. Check that no breakage occurred in transportation

and no spillage of liquid is present inside the box (primary container). Check that the aluminium pouch, containing the microplate, is not punctured or damaged.

- Dilute all the content of the 20x concentrated Wash Solution as described above.
- Dissolve the Calibrator as described above and gently mix.
- Allow all the other components to reach room temperature (about 1 hr) and then mix gently on vortex all liquid reagents.
- Set the ELISA incubator at +37°C and prepare the ELISA washer by priming with the diluted washing solution, according to the manufacturers instructions. Set the right number of washing cycles as reported in the specific section.
- Check that the ELISA reader is turned on or ensure it will be turned on at least 20 minutes before reading.
- If using an automated work station, turn on, check settings and be sure to use the right assay protocol.
- Check that the micropipettes are set to the required volume.
- Check that all the other equipment is available and ready to use.

In case of problems, do not proceed further with the test and advise the supervisor.

M. ASSAY PROCEDURE

The assay has to be performed according to the procedure given below, taking care to maintain the same incubation time for all the samples being tested.

A) HBe Antigen:

- Place the required number of strips in the plastic holder and carefully identify the wells for controls, calibrator and samples.
- Leave the A1 well empty for blanking purposes.
- Pipette 100 µl of the Negative Control in triplicate, 100 µl of the Antigen Calibrator in duplicate and then 100 µl of the Antigen Positive Control in single.
- Then dispense 100 µl of samples in the proper wells.
- Check for the presence of samples in wells by naked eye (there is a marked colour difference between empty and full wells) or by reading at 450/620nm (samples show OD values higher than 0.100).
- Incubate the microplate for **60 min at +37°C**.

Important note: Strips have to be sealed with the adhesive sealing foil, only when the test is performed manually. Do not cover strips when using ELISA automatic instruments.

- When the first incubation is finished, wash the microwells as previously described (section I.3)
- Dispense 100 µl Enzyme Conjugate in all wells, except for A1, used for blanking operations.

Important note: Be careful not to touch the inner surface of the well with the pipette tip and not to immerse the top of it into samples or controls. Contamination might occur.

- Check that the reagent has been dispensed properly and then incubate the microplate for **60 min at +37°C**.
- When the second incubation is finished, wash the microwells as previously described (section I.3)
- Pipette 100 µl Chromogen/Substrate into all the wells, A1 included.

Important note: Do not expose to strong direct light as a high background might be generated.

- Incubate the microplate protected from light at **room temperature (18-24°C) for 20 minutes**. Wells dispensed with positive control and positive samples will turn from clear to blue.

- Pipette 100 µl Sulphuric Acid into all the wells using the same pipetting sequence as in step 11. Addition of the stop solution will turn the positive control and positive samples from blue to yellow.

- Measure the color intensity of the solution in each well, as described in section I.5 using a 450nm filter (reading) and a 620-630nm filter (background subtraction, mandatory), blanking the instrument on A1.

B) HBe Antibody:

- Place the required number of strips in the plastic holder and carefully identify the wells for controls, calibrator and samples.
- Leave the A1 well empty for blanking purposes.
- Pipette 50 µl of the Negative Control in triplicate, 50 µl of the Antibody Calibrator in duplicate and then 50 µl of the Antibody Positive Control in single.
- Then dispense 50 µl of samples in the proper wells.
- Check for the presence of samples in wells by naked eye (there is a marked color difference between empty and full wells) or by reading at 450/620nm (samples show OD values higher than 0.100).
- Dispense then 50 µl of HBe Antigen in all the wells, except for A1.
- Incubate the microplate for **60 min at +37°C**.

Important note: Strips have to be sealed with the adhesive sealing foil, only when the test is performed manually. Do not cover strips when using ELISA automatic instruments.

- When the first incubation is finished, wash the microwells as previously described (section I.3)
- Finally proceed as described for the HBeAg assay from point 8 to the last one.

Important notes:

- Ensure that no finger prints are present on the bottom of the microwell before reading. Finger prints could generate false positive results on reading.
- Reading should ideally be performed immediately after the addition of the Stop Solution but definitely no longer than 20 minutes afterwards. Some self oxidation of the chromogen can occur leading to a higher background.
- The Calibrator (CAL) does not affect the cut-off calculation and therefore the test results calculation. The Calibrator may be used only when a laboratory internal quality control is required by the management.

N. ASSAY SCHEME

HBe antigen test

Controls and calibrator	100 ul
Samples	100 ul
1st incubation	60 min
Temperature	+37°C
Wash step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
Enzyme Conjugate	100 ul
2nd incubation	60 min
Temperature	+37°C
Wash step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
TMB/H2O2 mix	100 ul
3rd incubation	20 min
Temperature	r.t.
Sulphuric Acid	100 ul
Reading OD	450nm/620-630nm

HBe antibody test

Controls and calibrator	50 ul
Samples	50 ul
Neutralising antigen	50 ul
1st incubation	60 min
Temperature	+37°C
Wash step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
Enzymatic conjugate	100 ul
2nd incubation	60 min
Temperature	+37°C
Wash step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
TMB/H2O2 mixture	100 ul
3rd incubation	20 min
Temperature	r.t.
Sulphuric Acid	100 ul
Reading OD	450nm/620-630nm

An example of dispensation scheme is reported below:

Microplate

	1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	S2										
B	NC	S3										
C	NC	S4										
D	NC	S5										
E	CAL	S6										
F	CAL	S7										
G	PC	S8										
H	S1	S9										

Legenda: BLK = Blank // NC = Negative Control
PC = Positive Control // CAL = Calibrators // S = Sample

O. INTERNAL QUALITY CONTROL

A validation check is carried out on the controls any time the kit is used in order to verify whether the performances of the assay are as qualified.

Control that the following data are matched:

HBe Antigen

Check	OD450nm
Blank well	< 0.100 OD450nm
Negative Control (NC)	< 0.150 OD450nm after blanking coefficient of variation < 30%
Antigen Calibrator	S/Co > 2.0
Positive Control (PC)	> 1.500 OD450nm

HBe Antibody

Check	OD450nm
Blank well	< 0.100 OD450nm
Negative Control (NC)	> 1.000 OD450nm after blanking coefficient of variation < 10%
Antibody Calibrator	OD450nm < NC/1.5
Positive Control (PC)	OD450nm < NC/10

If the results of the test match the requirements stated above, proceed to the next section.

If they do not, don't proceed any further and perform the following checks:

HBeAg

Problem	Check
Blank well > 0.100 OD450nm	1. that the Chromogen/Substrate solution has not become contaminated during the assay
Negative Control (NC) > 0.150 OD450nm after blanking coefficient of variation > 30%	1. that the washing procedure and the washer settings are as validated in the pre qualification study; 2. that the proper washing solution has been used and the washer has been primed with it before use; 3. that no mistake has been done in the assay procedure (dispensation of positive control instead of negative control); 4. that no contamination of the negative control or of the wells where the control was dispensed has occurred due to positive samples, to spills or to the enzyme conjugate; 5. that micropipettes have not become contaminated with positive samples or with the enzyme conjugate 6. that the washer needles are not blocked or partially obstructed.
Calibrator S/Co < 2	1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution (ex.: dispensation of negative control instead); 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the calibrator has occurred.
Positive Control < 1.500 OD450nm	1. that the procedure has been correctly performed; 2. that no mistake has occurred during the distribution of the control (dispensation of negative control instead of positive control); 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the positive control has occurred.

HBe antibody

Problem	Check
Blank well > 0.100 OD450nm	1. that the Chromogen/Substrate solution has not become contaminated during the assay
Negative Control (NC) < 1.000 OD450nm after blanking coefficient of variation > 10%	1. that the washing procedure and the washer settings are as validated in the pre qualification study; 2. that the proper washing solution has been used and the washer has been primed with it before use; 3. that no mistake has been done in the assay procedure (e.g.: dispensation of positive control instead of negative control; no dispensation of the Neutralizing Antigen; no dispensation of the Enzyme Conjugate); 4. that no contamination of the negative control or of the wells where the control was dispensed has occurred; 5. that micropipettes have not become contaminated with positive samples; 6. that the washer needles are not blocked or partially obstructed.

Calibrator OD450nm > NC/1.5	<ol style="list-style-type: none"> 1. that the procedure has been correctly performed; 2. that no mistake has occurred during its distribution (ex.: dispensation of negative control instead; no dispensation of the Neutralizing Antigen; no dispensation of the Enzyme Conjugate); 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the calibrator has occurred.
Positive Control OD450nm > NC/10	<ol style="list-style-type: none"> 1. that the procedure has been correctly performed; 2. that no mistake has occurred during the distribution of the control; 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the positive control has occurred.

If any of the above problems have occurred, report the problem to the supervisor for further actions.

Important note:

The analysis must be done proceeding as the reading step described in the section M, point 14.

P. CALCULATION OF THE CUT-OFF

The results are calculated by means of a cut-off value determined with the following formula:

HBeAg:

$$NC + 0.100 = \text{Cut-Off (Co)}$$

The value found for the test is used for the interpretation of results as described in the next paragraph.

HBeAb:

$$(NC + PC) / 3 = \text{Cut-Off (Co)}$$

Important note: *When the calculation of results is performed by the operating system of an ELISA automated work station, ensure that the proper formulation is used to calculate the cut-off value and generate the correct interpretation of results.*

Q. INTERPRETATION OF RESULTS

Results are interpreted as follows:

HBeAg:

S/Co	Interpretation
< 0.9	Negative
0.9 - 1.1	Equivocal
> 1.1	Positive

HBeAb:

Co/S	Interpretation
< 0.9	Negative
0.9 - 1.1	Equivocal
> 1.1	Positive

Note:

$S = OD450nm/620-630nm$ of the sample

$Co = \text{cut-off value}$

An example of calculation for HBeAg assay is reported below (data obtained proceeding as the the reading step described in the section M, point 14):

The following data must not be used instead of real figures obtained by the user.

Negative Control: 0.020 – 0.030 – 0.025 OD450nm

Mean Value: 0.025 OD450nm

Lower than 0.150 – Accepted

Positive Control: 2.489 OD450nm

Higher than 1.500 – Accepted

Cut-Off = 0.025+0.100 = 0.125

Calibrator: 0.520 - 0.540 OD450nm

Mean value: 0.530 OD450nm

S/Co = 4.2

S/Co higher than 2.0 – Accepted

Sample 1: 0.030 OD450nm

Sample 2: 1.800 OD450nm

Sample 1 S/Co < 0.9 = negative

Sample 2 S/Co > 1.1 = positive

An example of calculation for HBeAb is reported below (data obtained proceeding as the the reading step described in the section M, point 14):

The following data must not be used instead of real figures obtained by the user.

Negative Control: 2.100 – 2.200 – 2.000 OD450nm

Mean Value: 2.100 OD450nm

Higher than 1.000 – Accepted

Positive Control: 0.100 OD450nm

Lower than NC/10 – Accepted

Cut-Off = (2.100 + 0.100) / 3 = 0.733

Calibrator: 0.720-0.760 OD450nm

Mean value: 0.740 OD450nm

OD450nm < NC/1.5 – Accepted

Sample 1: 0.020 OD450nm

Sample 2: 1.900 OD450nm

Sample 1 Co/S > 1.1 positive

Sample 2 Co/S < 0.9 negative

Important notes:

1. Interpretation of results should be done under the supervision of the laboratory director to reduce the risk of judgment errors and misinterpretations.
2. The Identification of the clinical status of a HBV patient (acute, chronic, asymptomatic hepatitis) has to be done on the basis also of the other markers of HBV infection (HBsAg, HBsAb, HBcAb, HBcIgM);
3. When test results are transmitted from the laboratory to another facility, attention must be paid to avoid erroneous data transfer.
4. Diagnosis of viral hepatitis infection has to be taken by and released to the patient by a suitably qualified medical doctor.

R. PERFORMANCE CHARACTERISTICS

A) HBeAg

1. Limit of detection

The limit of detection of the assay has been calculated by means of the International Standard for HBeAg, supplied by Paul Erlich Institute (PEI).

The data obtained by examining the limit of detection on three lots is reported in the table below.

HBE.CE Lot ID	PEI U/ml HBeAg
0103	0.25
0103/2	0.25
0303	0.25

In addition the preparation Accurun # 51, produced by Boston Biomedica Inc., USA, has been tested, upon dilution in FCS. Results are reported for three lots of products.

BBI's Accurun 51 (S/Co)

HBE.CE Lot ID	1 x	2 x	4 x	8 x	16x
0103	4.1	1.6	0.9	0.6	0.4
0103/2	4.1	1.7	0.9	0.6	0.4
0303	4.0	1.6	0.9	0.5	0.4

2. Diagnostic Sensitivity:

The diagnostic sensitivity has been tested on panels of samples classified positive by a US FDA approved kit.

Positive samples were collected from different HBV pathologies (acute, chronic) bearing HBeAg reactivity.

An overall value > 98% has been found in the study conducted on a total number of more than 200 samples.

Moreover the Panel of Seroconversion code PHM 935B, produced by BBI, was examined.

Data are reported below and compared with those reported by BBI for two other commercial products.

Sample ID	HBE.CE S/Co	Abbott EIA S/Co	Sorin EIA S/Co
21	5.4	4.5	6.3
22	3.7	4.3	5.4
23	1.9	3.2	3.1
24	1.1	2.4	1.5
25	1.0	2.1	1.2
26	0.6	1.7	0.7
27	0.2	0.8	0.3
28	0.2	0.6	0.2
29	0.2	0.4	0.2
30	0.2	0.3	0.2
31	0.1	0.3	0.2
32	0.1	0.3	0.2

Finally the Performance Panel code PHJ 201, produced by BBI, was tested. Data are reported below and compared with those reported by BBI for an other commercial product.

Member	PEI U/ml	HBE.CE	Sorin EIA
1	3	3.3	7.0
2	6	17.5	21.9
3	26	30.1	37.1
4	31	29.4	23.5
5	1	1.1	2.2
6	2	2.3	6.9
7	35	30.1	24.6
8	38	29.2	31.9
9	4	16.6	10.8
10	-	0.3	0.2

11	1	3.4	3.6
12	< 1	0.2	1.2
13	< 1	0.9	1.4
14	-	0.2	0.2
15	-	0.4	0.1
16	-	0.5	0.1
17	-	0.3	0.2
18	-	0.2	0.2
19	-	0.2	0.1
20	-	0.2	0.1
21	-	0.3	1.0
22	-	0.3	0.1
23	-	0.4	0.1
24	-	0.2	0.2
25	-	0.3	0.2

3. Diagnostic Specificity:

The diagnostic specificity has been determined on panels of negative samples from normal individuals and blood donors, classified negative with a FDA approved kit.

Both plasma, derived with different standard techniques of preparation (citrate, EDTA and heparin), and sera have been used to determine the specificity.

No false reactivity due to the method of specimen preparation has been observed.

Frozen specimens have also been tested to check whether this interferes with the performance of the test. No interference was observed on clean and particle free samples.

Samples derived from patients with different viral (HCV and HAV) and non viral pathologies of the liver that may interfere with the test were examined.

No cross reaction were observed.

The Performance Evaluation study conducted in a qualified external reference center on more than 500 samples has provided a value > 98% .

4. Precision

It has been calculated on two samples examined in 16 replicate in three different runs on three lots.

The values found were as follows:

HBE.CE: lot # 0103

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.030	0.027	0.032	0.029
Std.Deviation	0.002	0.002	0.003	0.002
CV %	7.4	8.2	7.9	7.8

PEI 1 U/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.569	0.559	0.575	0.568
Std.Deviation	0.027	0.029	0.028	0.028
CV %	4.7	5.3	4.9	4.9
S/Co	4.4	4.4	4.4	4.4

HBE.CE: lot # 0103/2

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.033	0.031	0.030	0.032
Std.Deviation	0.003	0.003	0.002	0.003
CV %	7.9	8.5	7.4	8.0

PEI 1 U/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.565	0.573	0.568	0.569
Std.Deviation	0.026	0.025	0.024	0.025
CV %	4.7	4.3	4.2	4.4
S/Co	4.2	4.4	4.4	4.3

HBE.CE: lot # 0303

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.029	0.034	0.038	0.034
Std.Deviation	0.003	0.003	0.004	0.003
CV %	9.7	9.8	9.2	9.6

PEI 1 U/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.579	0.573	0.564	0.572
Std.Deviation	0.023	0.028	0.025	0.025
CV %	4.1	4.8	4.5	4.5
S/Co	4.5	4.3	4.1	4.3

B) HBe Antibody

1. Limit of detection

The limit of detection of the assay has been calculated by means of the International Standard for HBeAb, supplied by Paul Erlich Institute (PEI).

The data obtained by examining the limit of detection on three lots is reported in the table below.

HBE.CE Lot ID	PEI U/ml HBeAb
0103	0.25
0103/2	0.25
0303	0.25

In addition the preparation Accurun # 52, produced by Boston Biomedica Inc., USA, has been tested, upon dilution in FCS. Results are reported for three lots of products.

Accurun 52 (Co/S)

HBE.CE Lot ID	1 x	2 x	4 x	8 x	16x
0103	1.0	0.8	0.6	0.4	0.4
0103/2	1.0	0.8	0.6	0.5	0.4
0303	1.0	0.8	0.6	0.4	0.4

2. Diagnostic sensitivity:

The diagnostic sensitivity has been tested on panels of samples classified positive for HBeAb by a US FDA approved kit. Positive samples were collected from different HBV pathologies bearing anti HBeAg antibody reactivity.

An overall value > 98% has been found in the study conducted on a total number of more than 200 samples.

Moreover the Panel of Seroconversion code PHM 935B, produced by BBI, was examined.

Data are reported below and compared with those reported by BBI for two other commercial products.

Sample ID	HBE.CE Co/S	Abbott EIA Co/S	Sorin EIA Co/S
21	0.4	0.4	0.5
22	0.4	0.5	0.6
23	0.4	0.6	0.5
24	0.4	0.5	0.6
25	0.4	0.6	0.5
26	0.5	0.6	0.6
27	0.6	0.8	0.7
28	0.7	0.9	0.7
29	0.6	0.9	0.7
30	0.8	1.0	0.9
31	1.0	1.3	1.1
32	1.0	1.2	1.0

Finally the Performance Panel code PHJ 201, produced by BBI, was tested. Data are reported below and compared with those reported by BBI for another commercial product.

Member	PEI U/ml	HBE.CE	Sorin EIA
1	-	0.3	0.5
2	-	0.2	0.5
3	-	0.2	0.5
4	-	0.2	0.5
5	-	0.3	0.6
6	-	0.3	0.6
7	-	0.2	0.4
8	-	0.2	0.4
9	-	0.2	0.5
10	-	1.9	0.6
11	-	0.3	0.5
12	-	0.4	0.9
13	2	4.4	9.1
14	1	3.8	2.9
15	< 1	1.0	1.5
16	> 50	4.3	120.9
17	< 1	1.0	1.0
18	5	5.6	21.8
19	1	2.7	6.4
20	11	5.0	47.3
21	2	1.9	10.0
22	26	28.1	90.7
23	-	0.3	0.5
24	< 1	0.8	1.3
25	50	28.1	167.4

3. Diagnostic specificity:

The clinical specificity has been determined as described before for HBeAg.

The Performance Evaluation study conducted in a qualified external reference center on more than 500 samples has provided a value > 98%.

4. Precision:

It has been calculated on two samples examined in 16 replicate in three different runs on three lots.

The values found were as follows:

HBE.CE: lot # 0103

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.484	2.420	2.471	2.458
Std.Deviation	0.129	0.160	0.142	0.144
CV %	5.2	6.6	5.7	5.9

PEI 0.25 U/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.867	0.800	0.878	0.848
Std.Deviation	0.043	0.060	0.050	0.051
CV %	5.0	7.5	5.7	6.1
Co/S	1.0	1.0	1.0	1.0

HBE.CE: lot # 0103/2

Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.316	2.361	2.413	2.363
Std.Deviation	0.127	0.144	0.146	0.139
CV %	5.5	6.1	6.0	5.9

PEI 0.25 U/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.767	0.793	0.785	0.781
Std.Deviation	0.041	0.050	0.046	0.046
CV %	5.4	6.3	5.8	5.8
Co/S	1.0	1.0	1.0	1.0

All the IVD Products manufactured by the company are under the control of a certified Quality Management System approved by an EC Notified Body. Each lot is submitted to a quality control and released into the market only if conforming with the EC technical specifications and acceptance criteria.

HBE.CE: lot #0303
Negative Control (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	2.334	2.415	2.437	2.395
Std.Deviation	0.146	0.155	0.158	0.153
CV %	6.3	6.4	6.5	6.4

Manufacturer:
Dia.Pro Diagnostic Bioprobes S.r.l.
Via G. Carducci n° 27 – Sesto San Giovanni (MI) – Italy



0318

PEI 0.25 U/ml (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.850	0.867	0.876	0.864
Std.Deviation	0.052	0.051	0.048	0.050
CV %	6.1	5.9	5.5	5.8
Co/S	0.9	1.0	1.0	1.0

Important note:

The performance data have been obtained proceeding as the reading step described in the section M, point 14.

S. LIMITATIONS

Frozen samples containing fibrin particles or aggregates may generate false positive results.

Bacterial contamination or heat inactivation of the specimen may affect the absorbance values of the samples with consequent alteration of the level of the analyte.

This test is suitable only for testing single samples and not pooled ones.

Diagnosis of an infectious disease should not be established on the basis of a single test result. The patient's clinical history, symptomatology, as well as other diagnostic data should be considered.

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HCV Ab

**Version 4.0 Enzyme Immunoassay
for the determination of
anti Hepatitis C Virus antibody
in human serum and plasma**

- for "in vitro" diagnostic use only -



DIA.PRO

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REF CVAB.CE
96,192,480,960 Tests

HCV Ab

A. INTENDED USE

Version 4.0 Enzyme ImmunoAssay (ELISA) for the determination of antibodies to Hepatitis C Virus in human plasma and sera. The kit is intended for the screening of blood units and the follow-up of HCV-infected patients.

For "in vitro" diagnostic use only.

B. INTRODUCTION

The World Health Organization (WHO) define Hepatitis C infection as follows:

"Hepatitis C is a viral infection of the liver which had been referred to as parenterally transmitted "non A, non B hepatitis" until identification of the causative agent in 1989. The discovery and characterization of the hepatitis C virus (HCV) led to the understanding of its primary role in post-transfusion hepatitis and its tendency to induce persistent infection.

HCV is a major cause of acute hepatitis and chronic liver disease, including cirrhosis and liver cancer. Globally, an estimated 170 million persons are chronically infected with HCV and 3 to 4 million persons are newly infected each year. HCV is spread primarily by direct contact with human blood. The major causes of HCV infection worldwide are use of unsterilized blood transfusions, and re-use of needles and syringes that have not been adequately sterilized. No vaccine is currently available to prevent hepatitis C and treatment for chronic hepatitis C is too costly for most persons in developing countries to afford. Thus, from a global perspective, the greatest impact on hepatitis C disease burden will likely be achieved by focusing efforts on reducing the risk of HCV transmission from nosocomial exposures (e.g. blood transfusions, unsafe injection practices) and high-risk behaviours (e.g. injection drug use).

Hepatitis C virus (HCV) is one of the viruses (A, B, C, D, and E), which together account for the vast majority of cases of viral hepatitis. It is an enveloped RNA virus in the *flaviviridae* family which appears to have a narrow host range. Humans and chimpanzees are the only known species susceptible to infection, with both species developing similar disease.

An important feature of the virus is the relative mutability of its genome, which in turn is probably related to the high propensity (80%) of inducing chronic infection. HCV is clustered into several distinct genotypes which may be important in determining the severity of the disease and the response to treatment.

The incubation period of HCV infection before the onset of clinical symptoms ranges from 15 to 150 days. In acute infections, the most common symptoms are fatigue and jaundice; however, the majority of cases (between 60% and 70%), even those that develop chronic infection, are asymptomatic. About 80% of newly infected patients progress to develop chronic infection. Cirrhosis develops in about 10% to 20% of persons with chronic infection, and liver cancer develops in 1% to 5% of persons with chronic infection over a period of 20 to 30 years. Most patients suffering from liver cancer who do not have hepatitis B virus infection have evidence of HCV infection. The mechanisms by which HCV infection leads to liver cancer are still unclear. Hepatitis C also exacerbates the severity of underlying liver disease when it coexists with other hepatic conditions. In particular, liver disease progresses more rapidly among persons with

alcoholic liver disease and HCV infection. HCV is spread primarily by direct contact with human blood. Transmission through blood transfusions that are not screened for HCV infection, through the reuse of inadequately sterilized needles, syringes or other medical equipment, or through needle-sharing among drug-users, is well documented. Sexual and perinatal transmission may also occur, although less frequently. Other modes of transmission such as social, cultural, and behavioural practices using percutaneous procedures (e.g. ear and body piercing, circumcision, tattooing) can occur if inadequately sterilized equipment is used. HCV is not spread by sneezing, hugging, coughing, food or water, sharing eating utensils, or casual contact.

In both developed and developing countries, high risk groups include injecting drug users, recipients of unsterilized blood, haemophiliacs, dialysis patients and persons with multiple sex partners who engage in unprotected sex. In developed countries, it is estimated that 90% of persons with chronic HCV infection are current and former injecting drug users and those with a history of transfusion of unsterilized blood or blood products. In many developing countries, where unsterilized blood and blood products are still being used, the major means of transmission are unsterilized injection equipment and unsterilized blood transfusions. In addition, people who use traditional scarification and circumcision practices are at risk if they use or re-use unsterilized tools.

WHO estimates that about 170 million people, 3% of the world's population, are infected with HCV and are at risk of developing liver cirrhosis and/or liver cancer. The prevalence of HCV infection in some countries in Africa, the Eastern Mediterranean, South-East Asia and the Western Pacific (when prevalence data are available) is high compared to some countries in North America and Europe.

Diagnostic tests for HCV are used to prevent infection through screening of donor blood and plasma, to establish the clinical diagnosis and to make better decisions regarding medical management of a patient. Diagnostic tests commercially available today are based on Enzyme immunoassays (EIA) for the detection of HCV specific antibodies. EIAs can detect more than 95% of chronically infected patients but can detect only 50% to 70% of acute infections. A recombinant immunoblot assay (RIBA) that identifies antibodies which react with individual HCV antigens is often used as a supplemental test for confirmation of a positive EIA result. Testing for HCV circulating by amplification tests RNA (e.g. polymerase chain reaction or PCR, branched DNA assay) is also being utilized for confirmation of serological results as well as for assessing the effectiveness of antiviral therapy. A positive result indicates the presence of active infection and a potential for spread of the infection and or/the development of chronic liver disease.

Antiviral drugs such as interferon taken alone or in combination with ribavirin, can be used for the treatment of persons with chronic hepatitis C, but the cost of treatment is very high. Treatment with interferon alone is effective in about 10% to 20% of patients. Interferon combined with ribavirin is effective in about 30% to 50% of patients. Ribavirin does not appear to be effective when used alone.

There is no vaccine against HCV. Research is in progress but the high mutability of the HCV genome complicates vaccine development. Lack of knowledge of any protective immune response following HCV infection also impedes vaccine research. It is not known whether the immune system is able to eliminate the virus.

Some studies, however, have shown the presence of virus neutralizing antibodies in patients with HCV infection. In the absence of a vaccine, all precautions to prevent infection must be taken including (a) screening and testing of blood and organ donors; (b) Virus inactivation of plasma derived products; (c) implementation and maintenance of infection control practices in health care settings, including appropriate sterilization of medical and dental equipment; (d) promotion of behaviour change among the general public and health care workers to reduce overuse of injections and to use safe injection practices; and (e) Risk reduction counselling for persons with high-risk drug and sexual practices. “

The genome encodes for structural components, a nucleocapsid protein and two envelope glycoproteins, and functional constituents involved in the virus replication and protein processing. The nucleocapsid-encoding region seems to be the most conservative among the isolates obtained all over the world.

C. PRINCIPLE OF THE TEST

Microplates are coated with HCV-specific antigens derived from “core” and “ns” regions encoding for conservative and immunodominant antigenic determinants (Core peptide, recombinant NS3, NS4 and NS5 peptides).

The solid phase is first treated with the diluted sample and HCV Ab are captured, if present, by the antigens.

After washing out all the other components of the sample, in the 2nd incubation bound HCV antibodies, IgG and IgM as well, are detected by the addition of polyclonal specific anti hlgG&M antibodies, labelled with peroxidase (HRP).

The enzyme captured on the solid phase, acting on the substrate/chromogen mixture, generates an optical signal that is proportional to the amount of anti HCV antibodies present in the sample. A cut-off value let optical densities be interpreted into HCV antibody negative and positive results.

D. COMPONENTS

Code CVAB.CE contains reagents for 192 tests.

1. Microplate **MICROPLATE**

n° 2 microplates

12 strips of 8 microwells coated with Core peptide, recombinant NS3, NS4 and NS5 peptides. Plates are sealed into a bag with desiccant.

2. Negative Control **CONTROL -**

1x4.0ml/vial. Ready to use control. It contains 1% goat serum proteins, 10 mM Na-citrate buffer pH 6.0 +/-0.1, 0.5% Tween 20, 0.09% Na-azide and 0.045% ProClin 300 as preservatives. The negative control is olive green colour coded.

3. Positive Control **CONTROL +**

1x4.0ml/vial. Ready to use control. It contains 1% goat serum proteins, human antibodies positive to HCV, 10 mM Na-citrate buffer pH 6.0 +/-0.1, 0.5% Tween 20, 0.09% Na-azide and 0.045% ProClin 300 as preservatives. The Positive Control is blue colour coded.

4. Calibrator **CAL ...**

n° 2 vials. Lyophilized calibrator. To be dissolved with the volume of EIA grade water reported on the label. It contains foetal bovine serum proteins, human antibodies to HCV whose content is calibrated on the NIBSC Working Standard code 99/588-003-WI, 10 mM Na-citrate buffer pH 6.0 +/-0.1, 0.3 mg/ml gentamicine sulphate and 0.045% ProClin 300 as preservatives.

Note: The volume necessary to dissolve the content of the vial may vary from lot to lot. Please use the right volume reported on the label .

5. Wash buffer concentrate **WASHBUF 20X**

2x60ml/bottle. 20x concentrated solution. Once diluted, the wash solution contains 10 mM phosphate buffer pH 7.0+/-0.2, 0.05% Tween 20 and 0.045% ProClin 300.

6. Enzyme Conjugate **CONJ**

2x16ml/vial. Ready to use and pink/red colour coded reagent. It contains Horseradish Peroxidase conjugated goat polyclonal antibodies to human IgG and IgM, 5% BSA, 10 mM Tris buffer pH 6.8+/-0.1, 0.045% ProClin 300 and 0.02% gentamicine sulphate as preservatives.

7. Chromogen/Substrate **SUBS TMB**

2x16ml/vial. Ready-to-use component. It contains 50 mM citrate-phosphate buffer pH 3.5-3.8, 4% dimethylsulphoxide, 0.03% tetra-methyl-benzidine or TMB and 0.02% hydrogen peroxide or H₂O₂.

Note: To be stored protected from light as sensitive to strong illumination.

8. Assay Diluent **DILAS**

1x15ml/vial. 10 mM tris buffered solution pH 8.0 +/-0.1 containing 0.045% ProClin 300 for the pre-treatment of samples and controls in the plate, blocking interference.

9. Sulphuric Acid **H₂SO₄ 0.3 M**

1x32ml/bottle. It contains 0.3 M H₂SO₄ solution. Attention: Irritant (H315; H319; P280; P302+P352; P332+P313; P305+P351+P338; P337+P313; P362+P363)

10. Sample Diluent: **DILSPE**

2x50ml/bottle. It contains 1% goat serum proteins, 10 mM Na-citrate buffer pH 6.0 +/-0.1, 0.5% Tween 20, 0.09% Na-azide and 0.045% ProClin 300 as preservatives. To be used to dilute the sample.

Note: The diluent changes colour from olive green to dark bluish green in the presence of sample.

11. Plate sealing foils n° 4

12. Package insert n° 1

Important note: Only upon specific request , Dia.Pro can supply reagents for 96, 480, 960 tests , as reported below:

1. Microplate	n°1	n°5	n°10
2.NegativeControl	1x2.0ml/vial	1x10ml/vial	1x20.ml/vial
3.PositiveControl	1x2.0ml/vial	1x10ml/vial	1x20.ml/vial
4.Calibrator	n° 1 vial	n° 5 vials	n° 10 vials
5.Wash buff conc	1x60ml/bottle	5x60ml/bottles	4x150ml/bottles
6.Enz. Conjugate	1x16ml/vial	2x40ml/bottles	4x40ml/bottles
7.Chromog/Subs	1x16ml/vial	2x40ml/bottles	4x40ml/bottles
8.Assay Diluent	1x8ml/vial	1x40ml/bottle	1x80ml/bottle
9.Sulphuric Acid	1x15ml/vial	2x40ml/bottle	2x80ml/bottles
10.SampleDiluent	1x50ml/vial	5x50ml/bottles	4x125ml/bottles
11.Plate seal foils	n° 2	n° 10	n° 20
12. Pack. insert	n° 1	n° 1	n° 1
Number of tests	96	480	960
Code	CVAB.CE.96	CVAB.CE.480	CVAB.CE.960

E. MATERIALS REQUIRED BUT NOT PROVIDED

1. Calibrated Micropipettes (200ul and 10ul) and disposable plastic tips.
2. EIA grade water (bidistilled or deionised, charcoal treated to remove oxidizing chemicals used as disinfectants).
3. Timer with 60 minute range or higher.
4. Absorbent paper tissues.
5. Calibrated ELISA microplate thermostatic incubator capable to provide a temperature of +37°C.
6. Calibrated ELISA microwell reader with 450nm (reading) and with 620-630nm (blinking) filters.
7. Calibrated ELISA microplate washer.
8. Vortex or similar mixing tools.

F. WARNINGS AND PRECAUTIONS

1. The kit has to be used by skilled and properly trained technical personnel only, under the supervision of a medical doctor responsible of the laboratory.
2. When the kit is used for the screening of blood units and blood components, it has to be used in a laboratory certified and qualified by the national authority in that field (Ministry of Health or similar entity) to carry out this type of analysis.
3. All the personnel involved in performing the assay have to wear protective laboratory clothes, talc-free gloves and glasses. The use of any sharp (needles) or cutting (blades) devices should be avoided. All the personnel involved should be trained in biosafety procedures, as recommended by the Center for Disease Control, Atlanta, U.S. and reported in the National Institute of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
4. All the personnel involved in sample handling should be vaccinated for HBV and HAV, for which vaccines are available, safe and effective.
5. The laboratory environment should be controlled so as to avoid contaminants such as dust or air-borne microbial agents, when opening kit vials and microplates and when performing the test. Protect the Chromogen/Substrate from strong light and avoid vibration of the bench surface where the test is undertaken.
6. Upon receipt, store the kit at 2.8°C into a temperature controlled refrigerator or cold room.
7. Do not interchange components between different lots of the kits. It is recommended that components between two kits of the same lot should not be interchanged.
8. Check that the reagents are clear and do not contain visible heavy particles or aggregates. If not, advise the laboratory supervisor to initiate the necessary procedures for kit replacement.
9. Avoid cross-contamination between serum/plasma samples by using disposable tips and changing them after each sample. Do not reuse disposable tips.
10. Avoid cross-contamination between kit reagents by using disposable tips and changing them between the use of each one. Do not reuse disposable tips.
11. Do not use the kit after the expiration date stated on the external container and internal (vials) labels.
12. Treat all specimens as potentially infective. All human serum specimens should be handled at Biosafety Level 2, as recommended by the Center for Disease Control, Atlanta, U.S. in compliance with what reported in the Institutes of Health's publication: "Biosafety in Microbiological and Biomedical Laboratories", ed. 1984.
13. The use of disposable plastic-ware is recommended in the preparation of the liquid components or in transferring components into automated workstations, in order to avoid cross contamination.
14. Waste produced during the use of the kit has to be discarded in compliance with national directives and laws concerning laboratory waste of chemical and biological substances. In particular, liquid waste generated from the washing procedure, from residuals of controls and from samples has to be treated as potentially infective material and inactivated

before waste. Suggested procedures of inactivation are treatment with a 10% final concentration of household bleach for 16-18 hrs or heat inactivation by autoclave at 121°C for 20 min..

15. Accidental spills from samples and operations have to be adsorbed with paper tissues soaked with household bleach and then with water. Tissues should then be discarded in proper containers designated for laboratory/hospital waste.
16. The Sulphuric Acid is an irritant. In case of spills, wash the surface with plenty of water
17. Other waste materials generated from the use of the kit (example: tips used for samples and controls, used microplates) should be handled as potentially infective and disposed according to national directives and laws concerning laboratory wastes.

G. SPECIMEN: PREPARATION AND RECOMMANDATIONS

1. Blood is drawn aseptically by venipuncture and plasma or serum is prepared using standard techniques of preparation of samples for clinical laboratory analysis. No influence has been observed in the preparation of the sample with citrate, EDTA and heparin.
2. Avoid any addition of preservatives to samples; especially sodium azide as this chemical would affect the enzymatic activity of the conjugate, generating false negative results.
3. Samples have to be clearly identified with codes or names in order to avoid misinterpretation of results. When the kit is used for the screening of blood units, bar code labeling and electronic reading is strongly recommended.
4. Haemolysed (red) and visibly hyperlipemic ("milky") samples have to be discarded as they could generate false results. Samples containing residues of fibrin or heavy particles or microbial filaments and bodies should be discarded as they could give rise to false results.
5. Sera and plasma can be stored at +2°...+8°C in primary collection tubes for up to five days after collection. Do not freeze primary tubes of collection. For longer storage periods, sera and plasma samples, carefully removed from the primary collection tube, can be stored frozen at -20°C for several months. Any frozen samples should not be frozen/thawed more than once as this may generate particles that could affect the test result.
6. If particles are present, centrifuge at 2.000 rpm for 20 min or filter using 0.2-0.8u filters to clean up the sample for testing.

H. PREPARATION OF COMPONENTS AND WARNINGS

A study conducted on an opened kit has not pointed out any relevant loss of activity up to 6 re-use of the device and up to 6 months.

1. Microplates:

Allow the microplate to reach room temperature (about 1 hr) before opening the container. Check that the desiccant is not turned to dark green, indicating a defect of manufacturing. In this case call Dia.Pro's customer service. Unused strips have to be placed back into the aluminium pouch, in presence of desiccant supplied, firmly zipped and stored at +2°..8°C. When opened the first time, residual strips are stable till the indicator of humidity inside the desiccant bag turns from yellow to green.

2. Negative Control:

Ready to use. Mix well on vortex before use.

3. Positive Control:

Ready to use. Mix well on vortex before use. Handle this component as potentially infective, even if HCV, eventually present in the control, has been chemically inactivated.

4. Calibrator:

Dissolve carefully the content of the lyophilised vial with the volume of EIA grade water reported on its label. Mix well on vortex before use.

Handle this component as potentially infective, even if HCV, eventually present in the control, has been chemically inactivated.

Note: *When dissolved the Calibrator is not stable. Store in aliquots at -20°C.*

5. Wash buffer concentrate:

The 20x concentrated solution has to be diluted with EIA grade water up to 1200 ml and mixed gently end-over-end before use. As some salt crystals may be present into the vial, take care to dissolve all the content when preparing the solution.

In the preparation avoid foaming as the presence of bubbles could give origin to a bad washing efficiency.

Note: *Once diluted, the wash solution is stable for 1 week at +2..8° C.*

6. Enzyme conjugate:

Ready to use. Mix well on vortex before use.

Be careful not to contaminate the liquid with oxidizing chemicals, air-driven dust or microbes.

If this component has to be transferred use only plastic, possibly sterile disposable containers.

7. Chromogen/Substrate:

Ready to use. Mix well on vortex before use.

Be careful not to contaminate the liquid with oxidizing chemicals, air-driven dust or microbes.

Do not expose to strong illumination, oxidizing agents and metallic surfaces.

If this component has to be transferred use only plastic, possible sterile disposable container.

8. Assay Diluent:

Ready to use. Mix well on vortex before use.

9. Sulphuric Acid:

Ready to use. Mix well on vortex before use.

Attention: Irritant (H315; H319; P280; P302+P352; P332+P313; P305+P351+P338; P337+P313; P362+P363).

Precautionary P statements:

P280 – Wear protective gloves/protective clothing/eye protection/face protection.

P302 + P352 – IF ON SKIN: Wash with plenty of soap and water.

P332 + P313 – If skin irritation occurs: Get medical advice/attention.

P305 + P351 + P338 – IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

P337 + P313 – If eye irritation persists: Get medical advice/attention.

P362 + P363 - Take off contaminated clothing and wash it before reuse.

10. Sample Diluent:

Ready to use. Mix well on vortex before use.

I. INSTRUMENTS AND TOOLS USED IN COMBINATION WITH THE KIT

1. Micropipettes have to be calibrated to deliver the correct volume required by the assay and must be submitted to regular decontamination (household alcohol, 10% solution of bleach, hospital grade disinfectants) of those parts that could accidentally come in contact with the sample. They should also be regularly maintained in order to show a precision of 1% and a trueness of +/-2%. Decontamination of spills or residues of kit components should also be carried out regularly.
2. The ELISA incubator has to be set at +37°C (tolerance of +/-0.5°C) and regularly checked to ensure the correct temperature is maintained. Both dry incubators and water

baths are suitable for the incubations, provided that the instrument is validated for the incubation of ELISA tests.

3. The **ELISA washer** is extremely important to the overall performances of the assay. The washer must be carefully validated in advance, checked for the delivery of the right dispensation volume and regularly submitted to maintenance according to the manufacturer's instructions for use. In particular the washer, at the end of the daily workload, has to be extensively cleaned out of salts with deionized water. Before use, the washer has to be extensively primed with the diluted Washing Solution. The instrument weekly has to be submitted to decontamination according to its manual (NaOH 0.1 M decontamination suggested). 5 washing cycles (aspiration + dispensation of 350ul/well of washing solution + 20 sec soaking = 1 cycle) are sufficient to ensure the assay with the declared performances. If soaking is not possible add one more cycle of washing. An incorrect washing cycle or salt-blocked needles are the major cause of false positive reactions.
4. Incubation times have a tolerance of ±5%.
5. The ELISA microplate reader has to be equipped with a reading filter of 450nm and with a second filter of 620-630nm, mandatory for blanking purposes. Its standard performances should be (a) bandwidth ≤ 10 nm; (b) absorbance range from 0 to ≥ 2.0; (c) linearity to ≥ 2.0; (d) repeatability ≥ 1%. Blanking is carried out on the well identified in the section "Assay Procedure". The optical system of the reader has to be calibrated regularly to ensure that the correct optical density is measured. It should be regularly maintained according to the manufacturer 's instructions.
6. When using an ELISA automated work station, all critical steps (dispensation, incubation, washing, reading, data handling) have to be carefully set, calibrated, controlled and regularly serviced in order to match the values reported in the section O "Internal Quality Control". The assay protocol has to be installed in the operating system of the unit and validated as for the washer and the reader. In addition, the liquid handling part of the station (dispensation and washing) has to be validated and correctly set. Particular attention must be paid to avoid carry over by the needles used for dispensing and for washing. This must be studied and controlled to minimize the possibility of contamination of adjacent wells. The use of ELISA automated work stations is recommended for blood screening when the number of samples to be tested exceed 20-30 units per run.
7. When using automatic devices, in case the vial holder of the instrument does not fit with the vials supplied in the kit, transfer the solution into appropriate containers and label them with the same label peeled out from the original vial. This operation is important in order to avoid mismatching contents of vials, when transferring them. When the test is over, return the secondary labeled containers to 2..8°C, firmly capped.
8. Dia.Pro's customer service offers support to the user in the setting and checking of instruments used in combination with the kit, in order to assure compliance with the requirements described. Support is also provided for the installation of new instruments to be used with the kit.

L. PRE ASSAY CONTROLS AND OPERATIONS

1. Check the expiration date of the kit printed on the external label of the kit box. Do not use if expired.
2. Check that the liquid components are not contaminated by naked-eye visible particles or aggregates. Check that the Chromogen/Substrate is colorless or pale blue by aspirating a small volume of it with a sterile transparent plastic pipette. Check that no breakage occurred in transportation and no spillage of liquid is present inside the box. Check that the

- aluminum pouch, containing the microplate, is not punctured or damaged.
- Dilute all the content of the 20x concentrated Wash Solution as described above.
 - Dissolve the Calibrator as described above.
 - Allow all the other components to reach room temperature (about 1 hr) and then mix as described.
 - Set the ELISA incubator at +37°C and prepare the ELISA washer by priming with the diluted washing solution, according to the manufacturers instructions. Set the right number of washing cycles as reported in the specific section.
 - Check that the ELISA reader has been turned on at least 20 minutes before reading.
 - If using an automated workstation, turn it on, check settings and be sure to use the right assay protocol.
 - Check that the micropipettes are set to the required volume.
 - Check that all the other equipment is available and ready to use.
 - In case of problems, do not proceed further with the test and advise the supervisor.

M. ASSAY PROCEDURE

The assay has to be carried out according to what reported below, taking care to maintain the same incubation time for all the samples in testing.

Automated assay:

In case the test is carried out automatically with an ELISA system, we suggest to make the instrument aspirate 200 ul Sample Diluent and then 10 ul sample.

All the mixture is then carefully dispensed directly into the appropriate sample well of the microplate. Before the next sample is aspirated, needles have to be duly washed to avoid any cross-contamination among samples.

Do not dilute controls/calibrator as they are ready to use.

Dispense 200 ul controls/calibrator in the appropriate control/calibration wells.

Important Note: *Visually monitor that samples have been diluted and dispensed into appropriate wells. This is simply achieved by checking that the colour of dispensed samples has turned to dark bluish-green while the colour of the negative control has remained olive green.*

For the next operations follow the operative instructions reported below for the Manual Assay.

It is strongly recommended to check that the time lap between the dispensation of the first and the last sample will be calculated by the instrument and taken into consideration by delaying the first washing operation accordingly.

Manual assay:

- Place the required number of Microwells in the microwell holder. Leave the 1st well empty for the operation of blanking.
- Dispense 200 ul of Negative Control in triplicate, 200 ul Calibrator in duplicate and 200 ul Positive Control in single in proper wells. Do not dilute Controls and Calibrator as they are pre-diluted, ready to use !
- Add 200 ul of Sample Diluent (DILSPE) to all the sample wells; then dispense 10 ul sample in each properly identified well. Mix gently the plate, avoiding overflowing and contaminating adjacent wells, in order to fully disperse the sample into its diluent.

Important note: *Check that the colour of the Sample Diluent, upon addition of the sample, changes from light green to dark bluish green, monitoring that the sample has been really added.*

- Dispense 50 ul Assay Diluent (DILAS) into all the controls/calibrator and sample wells. Check that the color of samples has turned to dark blue.
- Incubate the microplate for **45 min at +37°C**.

Important note: *Strips have to be sealed with the adhesive sealing foil, supplied, only when the test is carried out manually. Do not cover strips when using ELISA automatic instruments.*

- Wash the microplate with an automatic washer by delivering and aspirating 350ul/well of diluted washing solution as reported previously (section I.3).
- Pipette 100ul Enzyme Conjugate into each well, except the 1st blanking well, and cover with the sealer. Check that this pink/red coloured component has been dispensed in all the wells, except A1.

Important note: *Be careful not to touch the plastic inner surface of the well with the tip filled with the Enzyme Conjugate. Contamination might occur.*

- Incubate the microplate for **45 min at +37°C**.
- Wash microwells as in step 6.
- Pipette 100ul Chromogen/Substrate mixture into each well, the blank well included. Then incubate the microplate at **room temperature (18-24°C) for 15 minutes**.

Important note: *Do not expose to strong direct illumination. High background might be generated.*

- Pipette 100ul Sulphuric Acid into all the wells using the same pipetting sequence as in step 10 to stop the enzymatic reaction. Addition of acid will turn the positive control and positive samples from blue to yellow/brown.
- Measure the colour intensity of the solution in each well, as described in section I.5, at 450nm filter (reading) and at 620-630nm (background subtraction), blanking the instrument on A1 (mandatory).

Important notes:

- Ensure that no finger prints are present on the bottom of the microwell before reading. Finger prints could generate false positive results on reading.
- Reading has to be carried out just after the addition of the Stop Solution and anyway not any longer than 20 minutes after its addition. Some self oxidation of the chromogen can occur leading to high background.
- Shaking at 350 ±150 rpm during incubation has been proved to increase the sensitivity of the assay of about 20%.
- The Calibrator (CAL) does not affect the cut-off calculation and therefore the test results calculation. The Calibrator may be used only when a laboratory internal quality control is required by the management.

N. ASSAY SCHEME

Method	Operations
Controls & Calibrator Samples	200 ul 200ul dil.+10ul
Assay Diluent (DILAS)	50 ul
1st incubation	45 min
Temperature	+37°C
Wash step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
Enzyme conjugate	100 ul
2nd incubation	45 min
Temperature	+37°C
Wash step	n° 5 cycles with 20" of soaking OR n° 6 cycles without soaking
TMB/H ₂ O ₂	100 ul
3rd incubation	15 min
Temperature	r.t.
Sulphuric Acid	100 ul
Reading OD	450nm / 620-630nm

An example of dispensation scheme is reported below:

		Microplate											
		1	2	3	4	5	6	7	8	9	10	11	12
A	BLK	S2											
B	NC	S3											
C	NC	S4											
D	NC	S5											
E	CAL	S6											
F	CAL	S7											
G	PC	S8											
H	S1	S9											

Legenda: BLK = Blank NC = Negative Control
CAL = Calibrator PC = Positive Control S = Sample

O. INTERNAL QUALITY CONTROL

A check is carried out on the controls and the calibrator any time the kit is used in order to verify whether their OD450nm values are as expected and reported in the table below.

Check	Requirements
Blank well	< 0.100 OD450nm value
Negative Control (NC)	< 0.050 mean OD450nm value after blanking
Calibrator	S/Co > 1.1
Positive Control	> 1.000 OD450nm value

If the results of the test match the requirements stated above, proceed to the next section.

If they do not, do not proceed any further and operate as follows:

Problem	Check
Blank well > 0.100 OD450nm	1. that the Chromogen/Sustrate solution has not got contaminated during the assay
Negative Control (NC) > 0.050 OD450nm after blanking	1. that the washing procedure and the washer settings are as validated in the pre qualification study; 2. that the proper washing solution has been used and the washer has been primed with it before use; 3. that no mistake has been done in the assay procedure (dispensation of positive control instead of negative control); 4. that no contamination of the negative control or of their wells has occurred due to positive

	samples, to spills or to the enzyme conjugate; 5. that micropipettes haven't got contaminated with positive samples or with the enzyme conjugate 6. that the washer needles are not blocked or partially obstructed.
Calibrator S/Co < 1.1	1. that the procedure has been correctly executed; 2. that no mistake has been done in its distribution (ex.: dispensation of negative control instead of control serum) 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the calibrator has occurred.
Positive Control < 1.000 OD450nm	1. that the procedure has been correctly executed; 2. that no mistake has been done in the distribution of controls (dispensation of negative control instead of positive control. In this case, the negative control will have an OD450nm value > 0.150, too. 3. that the washing procedure and the washer settings are as validated in the pre qualification study; 4. that no external contamination of the positive control has occurred.

Should these problems happen, after checking, report any residual problem to the supervisor for further actions.

P. CALCULATION OF THE CUT-OFF

The tests results are calculated by means of a cut-off value determined with the following formula on the mean OD450nm value of the Negative Control (NC):

$$NC + 0.350 = \text{Cut-Off (Co)}$$

The value found for the test is used for the interpretation of results as described in the next paragraph.

Important note: When the calculation of results is done by the operative system of an ELISA automated work station be sure that the proper formulation is used to calculate the cut-off value and generate the right interpretations of results.

Q. INTERPRETATION OF RESULTS

Test results are interpreted as ratio of the sample OD450nm and the Cut-Off value (or S/Co) according to the following table:

S/Co	Interpretation
< 0.9	Negative
0.9 - 1.1	Equivocal
> 1.1	Positive

A negative result indicates that the patient has not been infected by HCV or that the blood unit may be transfused.

Any patient showing an equivocal result should be tested again on a second sample taken 1-2 weeks later from the patient and examined. The blood unit should not be transfused.

A positive result is indicative of HCV infection and therefore the patient should be treated accordingly or the blood unit should be discarded.

Important notes:

1. Interpretation of results should be done under the supervision of the responsible of the laboratory to reduce the risk of judgment errors and misinterpretations.
2. Any positive result should be confirmed by an alternative method capable to detect IgG and IgM antibodies (confirmation test) before a diagnosis of viral hepatitis is formulated.
3. As proved in the Performance Evaluation of the product, the assay is able to detect seroconversion to anti HCV core antibodies **earlier** than some other commercial kits. Therefore a positive result, not confirmed with these commercial kits, does not have to be ruled out as a false positive result ! The sample has to be anyway submitted to a confirmation test (supplied upon request by DiaPro srl, code CCONF).
4. As long as the assay is able to detect also IgM antibodies some discrepant results with other commercial products for the detection of anti HCV antibodies - lacking anti hIgM conjugate in the formulation of the enzyme tracer and therefore missing IgM reactivity - may be present. The real positivity of the sample for antibodies to HCV should be then confirmed by examining also IgM reactivity, important for the diagnosis of HCV infection.
5. When test results are transmitted from the laboratory to an informatics centre, attention has to be done to avoid erroneous data transfer.
6. Diagnosis of viral hepatitis infection has to be done and released to the patient only by a qualified medical doctor.

An example of calculation is reported below:

The following data must not be used instead of real figures obtained by the user.

Negative Control: 0.019 – 0.020 – 0.021 OD450nm
 Mean Value: 0.020 OD450nm
 Lower than 0.050 – Accepted
 Positive Control: 2.189 OD450nm
 Higher than 1.000 – Accepted
 Cut-Off = 0.020+0.350 = 0.370
 Calibrator: 0.550 - 0.530 OD450nm
 Mean value: 0.540 OD450nm S/Co = 1.4
 S/Co higher than 1.1 – Accepted
 Sample 1: 0.070 OD450nm
 Sample 2: 1.690 OD450nm
 Sample 1 S/Co < 0.9 = negative
 Sample 2 S/Co > 1.1 = positive

R. PERFORMANCES

Evaluation of Performances has been conducted in accordance to what reported in the Common Technical Specifications or CTS (art. 5, Chapter 3 of IVD Directive 98/79/EC).

1. LIMIT OF DETECTION

The limit of detection of the assay has been calculated by means of the British Working Standard for anti-HCV, NIBSC code 99/588-003-WI. The table below reports the mean OD450nm values of this standard when diluted in negative plasma and then examined.

Dilution	Lot # 1	Lot # 2
Factor	S/Co	S/Co
1 X	2.0	2.0
2 X	1.1	1.2
4 X	0.7	0.8
8 X	0.5	0.5
Negative plasma	0.3	0.3

In addition the sample coded Accurun 1 – series 3000 - supplied by Boston Biomedica Inc., USA, has been evaluated "in toto" showing the results below:

CVAB.CE Lot ID	Accurun 1 Series	S/Co
1201	3000	1.5
0602	3000	1.5
1202	3000	1.9

In addition, n° 7 samples, tested positive for HCV Ab with Ortho HCV 3.0 SAve, code 930820, lot. # EXE065-1, were diluted in HCV Ab negative plasma in order to generate limiting dilutions and then tested again on CVAB.CE, lot. # 1202, and Ortho. The following table reports the data obtained.

Sample n°	Limit Dilution	CVAB.CE S/Co	Ortho 3.0 S/Co
1	256 X	1.9	1.3
2	256 X	1.9	0.7
3	256 X	2.4	1.0
4	128 X	2.5	3.2
5	85 X	3.3	1.4
6	128 X	2.2	0.8
7	135 X	3.2	2.2

2. DIAGNOSTIC SPECIFICITY AND SENSITIVITY

The Performance Evaluation of the device was carried out in a trial conducted on more than total 5000 samples.

2.1 Diagnostic specificity:

It is defined as the probability of the assay of scoring negative in the absence of specific analyte. In addition to the first study, where a total of 5043 unselected blood donors, (including 1st time donors), 210 hospitalized patients and 162 potentially interfering specimens (other infectious diseases, E.coli antibody positive, patients affected by non viral hepatic diseases, dialysis patients, pregnant women, hemolized, lipemic, etc.) were examined, the diagnostic specificity was recently assessed by testing a total of 2876 negative blood donors on six different lots. A value of specificity of 100% was found. No false reactivity due to the method of specimen preparation has been observed. Both plasma, derived with different standard techniques of preparation (citrate, EDTA and heparin), and sera have been used to determine the value of specificity. Frozen specimens have been tested, as well, to check for interferences due to collection and storage. No interference was observed.

2.2 Diagnostic Sensitivity

It defined as the probability of the assay of scoring positive in the presence of specific analyte. The diagnostic sensitivity has been assessed externally on a total number of 359 specimens; a diagnostic sensitivity of 100% was found. Internally more than other 50 positive samples were tested, providing a value of diagnostic sensitivity of again 100%. Positive samples from infections carried out by different genotypes of HCV were tested as well. Furthermore, most of seroconversion panels available from Boston Biomedica Inc., USA, (PHV) and Zeptometrix, USA, (HCV) have been studied. Results are reported below for some of them.

Panel	N° samples	DiaPro*	Ortho* **
PHV 901	11	9	9
PHV 904	7	2	4
PHV 905	9	3	4
PHV 906	7	7	7
PHV 907	7	3	2
PHV 908	13	10	8
PHV 909	3	2	2
PHV 910	5	3	3
PHV 911	5	3	3
PHV 912	3	1	1
PHV 913	4	2	2
PHV 914	9	5	5
PHV 915	4	3	0
PHV 916	8	4	3
PHV 917	10	6	6
PHV 918	8	2	0
PHV 919	7	3	3
PHV 920	10	6	6
HCV 10039	5	2	0
HCV 6212	9	6	7
HCV 10165	9	5	4

Note: * Positive samples detected

** HCV v.3.0

Finally the Product has been tested on the panel EFS Ac HCV, lot n° 01/08.03.22C/01/A, supplied by the Etablissement Francais Du Sang (EFS), France, with the following results:

EFS Panel Ac HCV

Sample	Lot # 1	Lot # 2	Lot # 2	Results expected
	S/Co	S/Co	S/Co	
HCV 1	2.2	2.4	2.6	positive
HCV 2	1.6	2.0	2.1	positive
HCV 3	1.5	1.7	1.6	positive
HCV 4	5.2	6.5	5.5	positive
HCV 5	1.6	1.8	1.6	positive
HCV 6	0.4	0.4	0.4	negative

3. PRECISION:

It has been calculated on two samples, one negative and one low positive, examined in 16 replicates in three separate runs. Results are reported as follows:

Lot # 1202

Negative Sample (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.094	0.099	0.096	0.096
Std.Deviation	0.008	0.007	0.008	0.007
CV %	8.7	6.6	7.9	7.7

Cal # 2 – 7K (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.396	0.403	0.418	0.406
Std.Deviation	0.023	0.029	0.027	0.026
CV %	5.9	7.1	6.4	6.5
S/Co	1.1	1.1	1.2	1.1

Lot # 0602

Negative Sample (N = 16)

Mean values	1st run	2nd run	3 rd run	Average
OD 450nm	0.097	0.096	0.094	0.096
Std.Deviation	0.009	0.010	0.008	0.009
CV %	8.9	10.1	8.4	9.1

Cal # 2 – 7K (N = 16)

Mean values	1st run	2nd run	3 rd run	Average value
OD 450nm	0.400	0.395	0.393	0.396
Std.Deviation	0.021	0.025	0.026	0.024
CV %	5.4	6.2	6.6	6.1
S/Co	1.2	1.2	1.1	1.2

Lot # 0602/2

Negative Sample (N = 16)

Mean values	1st run	2nd run	3 rd run	Average
OD 450nm	0.087	0.091	0.088	0.089
Std.Deviation	0.009	0.007	0.008	0.008
CV %	10.0	8.2	8.6	8.9

Cal # 2 – 7K (N = 16)

Mean values	1st run	2nd run	3 rd run	Average
OD 450nm	0.386	0.390	0.391	0.389
Std.Deviation	0.023	0.021	0.023	0.022
CV %	6.0	5.3	5.8	5.7
S/Co	1.1	1.2	1.2	1.2

The variability shown in the tables above did not result in sample misclassification.

S. LIMITATIONS

Repeatable false positive results, not confirmed by RIBA or similar confirmation techniques, were assessed as less than 0.1% of the normal population. Frozen samples containing fibrin particles or aggregates after thawing have been observed to generate some false results.

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Manufacturer:
Dia.Pro Diagnostic Bioprobes Srl.
Via G. Carducci n° 27 – Sesto San Giovanni (MI) - Italy



HCV Ab

**Versión 4.0 del Ensayo
Inmunoenzimático para la determinación
de anticuerpos frente Virus de la
Hepatitis C
en plasma y suero humanos.**

Uso exclusivo para diagnóstico "in vitro"



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HCV Ab

A. OBJETIVO DEL EQUIPO.

Versión 4.0 del Ensayo Inmunoenzimático (ELISA) para la determinación de anticuerpos al virus de la Hepatitis C en plasma y suero humanos.

El equipo está diseñado para el cribado en unidades de sangre así como para el seguimiento de pacientes infectados con HCV. Uso exclusivo para diagnóstico "in vitro".

B. INTRODUCCIÓN.

La Organización Mundial de la Salud (OMS) define la infección por el virus de la Hepatitis C como:

"La Hepatitis C es una infección viral del hígado, definida como hepatitis de transmisión parenteral "no A no B" hasta el descubrimiento del agente causal en 1989. El descubrimiento y la caracterización del virus de la hepatitis C (HCV) ha permitido comprender su papel primario en la hepatitis post-transfusional y su tendencia a inducir la infección persistente. El virus de la hepatitis C es la causa principal de hepatitis aguda y enfermedad hepática crónica, incluyendo cirrosis y cáncer de hígado. A nivel mundial se estima que 170 millones de personas estén infectadas de forma crónica con HCV y que de 3 a 4 millones se infecten cada año.

El virus se transmite por contacto directo con sangre humana. Las causas principales de infección por HCV en el mundo son las transfusiones sanguíneas no controladas y la reutilización de jeringuillas y agujas sin una correcta esterilización previa. En la actualidad aún no existe una vacuna eficaz contra el virus y el tratamiento para la hepatitis C crónica es demasiado costoso para la mayoría de las personas en países en vías de desarrollo. Desde una perspectiva global, el mayor impacto contra la hepatitis C puede lograrse a través de esfuerzos orientados hacia la prevención y el control de la transmisión por exposiciones nosocomiales (como las transfusiones sanguíneas y las prácticas invasoras inseguras) y los comportamientos que conllevan alto riesgo (como el consumo de drogas inyectables).

El virus de la hepatitis C aparece en la mayoría de los casos de hepatitis viral. Es un virus RNA envuelto, perteneciente a la familia Flaviviridae y que parece tener un estrecho margen de huéspedes. Humanos y chimpancés son las únicas especies susceptibles conocidas y ambas desarrollan una enfermedad similar. Una característica importante del virus es su variabilidad genómica, la cual pudiera estar relacionada a su elevada capacidad (80%) de inducir infección crónica. El HCV ha sido agrupado por genotipos, lo cual puede ser útil para determinar la gravedad de la enfermedad y la respuesta al tratamiento.

El periodo de incubación varía desde 15 hasta 150 días. En la infección aguda los síntomas más comunes son fatiga e ictericia, sin embargo la mayoría de los casos (entre el 60% y el 70%), incluso aquellos que desarrollan la infección crónica, son asintomáticos. Cerca del 80% de los nuevos pacientes infectados progresan a la infección crónica. Del 10 al 20% de las personas con infección crónica desarrollan cirrosis, mientras que el cáncer de hígado lo presentan entre el 1 y el 5% de las personas con este tipo de infección, en un periodo de 20 a 30 años. Muchos pacientes que padecen cáncer de hígado y no están infectados por el virus de la hepatitis B, presentan evidencias de infección por el virus de la hepatitis C. Los mecanismos que relacionan la infección por HCV y el desarrollo de cáncer hepático no han sido aún esclarecidos. La hepatitis C puede exacerbar la gravedad de una enfermedad subyacente

del hígado cuando coexiste con otras disfunciones hepáticas; particularmente la enfermedad progresa más rápidamente en personas alcohólicas e infectadas por HCV. Las formas de transmisión más frecuentes son a través de transfusiones sanguíneas sin controlar y por la reutilización de agujas, jeringuillas y material médico contaminados. La transmisión sexual y perinatal puede suceder aunque es menos frecuente. Determinadas prácticas y comportamientos sociales y culturales (perforaciones en orejas y otras partes del cuerpo (piercing), circuncisiones y tatuajes) pueden constituir modos de transmisión si existe una inadecuada esterilización de los instrumentos usados. El HCV no se transmite por estornudos, tos, abrazos, agua o alimentos, estrechar la mano, compartir cubiertos o en general por contactos casuales. Tanto en países desarrollados como en aquellos en vías de desarrollo, los grupos de alto riesgo incluyen drogadictos, receptores de transfusiones sin analizar, hemofílicos, pacientes sometidos a diálisis y personas con actividad sexual promiscua y sin la debida protección. En los países desarrollados, se ha estimado que el 90% de las personas con infección crónica por HCV son o han sido drogadictos o han recibido donaciones de sangre o hemoderivados contaminados. En muchos países en vías de desarrollo, donde aún se utilizan transfusiones o hemoderivados sin analizar, los principales medios de transmisión son los instrumentos para inyecciones y las transfusiones sin analizar.

La OMS estima que cerca de 170 millones de personas, es decir el 3% de la población mundial, están infectadas por el HCV y bajo riesgo de desarrollar cirrosis y/o cáncer hepático. La prevalencia de la infección por HCV en países de África, el Mediterráneo oriental, Sudeste Asiático y el Pacífico Occidental es alta, comparada con países de Norteamérica y Europa.

Las pruebas de diagnóstico para el HCV contribuyen a prevenir la infección mediante el cribado de la sangre y plasma del donante, son útiles para establecer un diagnóstico clínico y en el seguimiento de los pacientes. Las pruebas de diagnóstico comerciales disponibles en la actualidad, se basan en ensayos enzimáticos de inmunoabsorción (EIA) para la detección de anticuerpos específicos contra HCV. Estos métodos pueden detectar más del 95% de los pacientes con infección crónica, pero solo entre el 50 y el 70% de las infecciones agudas. Para confirmar los resultados positivos por EIA se usa frecuentemente el sistema inmunoblot recombinante (RIBA), el cual identifica anticuerpos contra los antígenos individuales del HCV. Por otra parte, algunas técnicas de biología molecular (amplificación de ácidos nucleicos: Reacción en Cadena de la Polimerasa (PCR) y DNA ramificado) han sido utilizadas para confirmar los resultados serológicos así como para determinar la efectividad de la terapia antiviral. Un resultado positivo indica la presencia de una infección activa, de una fuente potencial de transmisión y/o del desarrollo de una enfermedad hepática crónica.

Para el tratamiento de personas con hepatitis C crónica se emplean fármacos antivirales como el interferón (administrado solo o en combinación con la ribavirina), pero el costo del tratamiento es elevado. Si se emplea solo el tratamiento con interferón, la eficacia en los pacientes es de 10 a 20%, mientras que en combinación con la ribavirina es eficaz en cerca del 30-50% de los casos. El tratamiento solo con ribavirina no parece ser efectivo.

No existe en la actualidad una vacuna contra HCV, debido en parte, a la alta frecuencia de mutaciones del virus. El escaso conocimiento de la respuesta inmune protectora que sigue a la infección por HCV ha dificultado el desarrollo de la vacuna. No se conoce tampoco acerca de los mecanismos del sistema inmune para la eliminación del virus. Algunos estudios, sin embargo, han demostrado la aparición de anticuerpos neutralizantes en pacientes con infección HCV. En ausencia de la vacuna, es conveniente tomar todas las medidas posibles para prevenir la infección (a) cribado y análisis de sangre y órganos de donantes; (b) inactivación del virus en productos derivados del plasma; (c) implementación y mantenimiento de las prácticas para el control de la infección incluyendo la

esterilización del material médico y dental; (d) promover cambios en la conducta entre el público en general y el personal sanitario para evitar las prácticas incorrectas y (e) vigilancia de los grupos de riesgo (personas con promiscuidad sexual y drogadictos).”

El genoma codifica para componentes estructurales: una proteína de la nucleocápside y dos glicoproteínas de la envoltura, así como para proteínas funcionales involucradas en la replicación viral y la síntesis de proteínas. La región que codifica para la nucleocápside parece estar altamente conservada entre los aislamientos obtenidos en todo el mundo.

C. PRINCIPIOS DEL ENSAYO.

Las microplacas están recubiertas con antígenos específicos del HCV correspondientes a las regiones del “core” y “ns” que codifican para determinantes antigénicos inmunodominantes y conservados (péptido del core y péptidos recombinantes NS3, NS4 y NS5).

Se añade la muestra diluida y los anticuerpos contra HCV, presentes en la muestra, son capturados por los antígenos de la fase sólida.

Después del lavado, en la 2ª incubación, los anticuerpos IgG e IgM son detectados mediante anticuerpos policlonales específicos anti-IgG/IgM humanos, conjugados con Peroxidasa (HPR).

La enzima capturada en la fase sólida, combinada con la mezcla sustrato/cromógeno, genera una señal óptica proporcional a la cantidad de anticuerpos anti-HCV presentes en la muestra. Posteriormente, mediante un valor de corte calculado, las densidades ópticas pueden interpretarse como resultados negativos o positivos a la presencia de anticuerpos al HCV.

D. COMPONENTES.

Cada equipo (Código CVAB.CE) contiene reactivos suficientes para realizar 192 pruebas.

1. Microplaca: MICROPLATE

n° 2 microplacas

12 tiras de 8 pocillos recubiertos con péptidos recombinantes para el “core” y para NS3, NS4 y NS5. Las placas están empaquetadas en bolsas selladas con desecante.

2. Control Negativo: CONTROL -

1x4.0ml/vial

Listo para el uso. Contiene 1% de proteínas del suero de cabra, tampón Citrato sódico 10mM pH 6.0 +/-0.1, 0.5% de Tween 20, además de azida sódica 0.09% y ProClin 300 al 0,045% como conservantes. El control negativo está codificado con el color verde olivo.

3. Control Positivo: CONTROL +

1x4.0ml/vial

Listo para el uso. Contiene 1% de proteínas del suero de cabra, anticuerpos humanos anti-HCV, tampón Citrato sódico 10mM pH 6.0 +/-0.1, 0.5% de Tween 20, así como azida sódica 0.09% y ProClin 300 al 0,045% como conservantes. El control positivo está codificado con el color azul.

4. Calibrador CAL

n° 2 viales

Liofilizado. Para disolver en agua calidad EIA como se indica en la etiqueta. Contiene suero fetal bovino, anticuerpos humanos al HCV, calibrados según el código Estándar de Trabajo de NIBSC 99/588-003-W1, tampón Citrato sódico 10mM pH 6.0 +/-0.1, además de sulfato de gentamicina 0.3 mg/ml y ProClin 300 al 0,045% como conservantes.

Nota: El volumen necesario para disolver el contenido del frasco varía en cada lote. Se recomienda usar el volumen indicado en la etiqueta.

5. Tampón de Lavado Concentrado: WASHBUF 20X

2x60ml/botella. Solución concentrada 20x.

Una vez diluida, la solución de lavado contiene tampón fosfato 10 mM a pH 7.0 +/- 0.2, Tween 20 al 0.05% y ProClin 300 al 0,045%.

6. Conjugado CONJ

2x16ml/vial. Solución lista para el uso. Contiene 5% de albúmina de suero bovino, tampón Tris 10mM a pH 6.8 +/- 0.1, anticuerpo policlonal de cabra anti-IgM/IgG humanos conjugado con peroxidasa (HPR) en presencia de 0.2 % de sulfato de gentamicina y ProClin 300 al 0,045% como conservantes. El conjugado está codificado con el color rosa/rojo.

7. Cromógeno/Substrato SUBS TMB

2x16ml/vial. Contiene una solución tamponada citrato-fosfato 50mM pH 3.5-3.8, tetra-metil-benzidina (TMB) 0.03% y peróxido de hidrógeno (H₂O₂) 0.02% así como dimetilsulfóxido 4%.

Nota: Evitar la exposición a la luz, la sustancia es fotosensible.

8. Diluyente de ensayo: DILAS

1x15ml/vial. Contiene una solución tamponada Tris 10 mM pH 8.0 +/- 0.1 y 0.1% de ProClin 300 al 0,045% para el pretratamiento de muestras y controles, bloquea posibles interferencias.

Nota: Usar todo el contenido del vial antes de abrir un segundo. El reactivo es sensible a oxidación.

9. Ácido Sulfúrico: H₂SO₄ 0.3 M

1x32ml/vial. Contiene solución de H₂SO₄ 0.3M

Atención: Irritante (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

10. Diluyente de muestras DILSPE

2x50ml. Contiene una solución tamponada citrato sódico 10 mM pH 6.0 +/- 0.1, 1% de proteínas del suero de cabra, 0.5% de Tween 20, azida sódica 0.09% y ProClin 300 al 0,045% como conservantes. Se usa para diluir las muestras.

11. Sellador adhesivo, n° 4

12. Manual de instrucciones, n° 1

Nota importante: A solicitud del cliente, Dia.Pro puede suministrar reactivos para realizar 96, 480 ó 960 pruebas, según se reporta a continuación:

1.Microplaca	n°1	n°5	n°10
2.ControlNegativo	1x2.0ml/vial	1x10ml/vial	1x20.ml/vial
3.ControlPositivo	1x2.0ml/vial	1x10ml/vial	1x20.ml/vial
4.Calibrador	n° 1 vial	n° 5 vials	n° 10 vials
5.Soluc. Lav. conc	1x60ml/bot.	5x60ml/frasc.	4x150ml/frasc.
6.Conjugado	1x16ml/vial	2x40ml/frasc.	4x40ml/frasc.
7.Cromóg/Subs	1x16ml/vial	2x40ml/frasc.	4x40ml/frasc.
8.Diluent. ensayo	1x8ml/vial	1x40ml/ frasc.	1x80ml/frasc.
9.Acido Sulfúrico	1x15ml/vial	2x40ml/ frasc.	2x80ml/frasc.
10.Diluent.muestr.	1x50ml/vial	5x50ml/frasc.	4x125ml/frasc.
11.Sellador adhes.	n° 2	n° 10	n° 20
12.Manual de instrucciones	n° 1	n° 1	n° 1
Número de pruebas	96	480	960
Código	CVAB.CE.96	CVAB.CE.480	CVAB.CE.960

E. MATERIALES NECESARIOS NO SUMINISTRADOS.

1. Micropipetas calibradas (200µl y 10µl) y puntas plásticas desechables.
2. Agua de calidad EIA (bidestilada o desionizada, tratada con carbón para remover químicos oxidantes usados como desinfectantes).
3. *Timer* con un rango de 60 minutos como mínimo.
4. Papel absorbente.
5. Incubador termostático de microplacas ELISA, calibrado (en seco o húmedo) fijo a 37°C.
6. Lector calibrado de microplacas de ELISA con filtros de 450nm (lectura) y de filtros de 620-630 nm.
7. Lavador calibrado de microplacas ELISA.
8. Vórtex o similar.

F. ADVERTENCIAS Y PRECAUCIONES.

1. El equipo debe ser usado por personal técnico adecuadamente entrenado, bajo la supervisión de un doctor responsable del laboratorio.
2. Cuando el equipo es usado para cribado en unidades de sangre, el laboratorio debe estar certificado y calificado para realizar este tipo de análisis (Ministerio de Salud o entidad similar).
3. Todas las personas encargadas de la realización de las pruebas deben llevar las ropas protectoras adecuadas de laboratorio, guantes y gafas. Evitar el uso de objetos cortantes (cuchillas) o punzantes (agujas). El personal debe ser adiestrado en procedimientos de bioseguridad, según ha sido recomendado por el Centro de Control de Enfermedades de Atlanta, Estados Unidos, y publicado por el Instituto Nacional de Salud: "Biosafety in Microbiological and Biomedical Laboratories", ed.1984.
4. Todo el personal involucrado en el manejo de muestras debe estar vacunado contra HBV y HAV, para lo cual existen vacunas disponibles, seguras y eficaces.
5. Se debe controlar el ambiente del laboratorio para evitar la contaminación de los componentes con polvo o agentes microbianos cuando se abran los equipos, así como durante la realización del ensayo. Evitar la exposición del sustrato a la luz y las vibraciones de la mesa de trabajo durante el ensayo.
6. Conservar el equipo a temperaturas entre 2-8 °C, en un refrigerador con temperatura regulada o en cámara fría.
7. No intercambiar reactivos de diferentes lotes ni tampoco de diferentes equipos.
8. Comprobar que los reactivos no contienen precipitados ni agregados en el momento del uso. De darse el caso, informar al responsable para realizar el procedimiento pertinente y reemplazar el equipo.
9. Evitar contaminación cruzada entre muestras de suero/plasma usando puntas desechables y cambiándolas después de cada uso. No reutilizar puntas desechables
10. Evitar contaminación cruzada entre los reactivos del equipo usando puntas desechables y cambiándolas después de cada uso. No reutilizar puntas desechables
11. No usar el producto después de la fecha de caducidad indicada en el equipo e internamente en los reactivos. Según estudios realizados, no se ha detectado pérdida relevante de actividad en equipos abiertos, en uso por un período de hasta 6 meses.
12. Tratar todas las muestras como potencialmente infecciosas. Las muestras de suero humano deben ser manipuladas al nivel 2 de bioseguridad, según ha sido recomendado por el Centro de Control de Enfermedades de Atlanta, Estados Unidos y publicado por el Instituto Nacional de Salud: "Biosafety in Microbiological and Biomedical Laboratories", ed.1984.
13. Se recomienda el uso de material plástico desechable para la preparación de las soluciones de lavado y para la transferencia de los reactivos a los diferentes equipos automatizados a fin de evitar contaminaciones.

14. Los desechos producidos durante el uso del equipo deben ser eliminados según lo establecido por las directivas nacionales y las leyes relacionadas con el tratamiento de los residuos químicos y biológicos de laboratorio. En particular, los desechos líquidos provenientes del proceso de lavado deben ser tratados como potencialmente infecciosos y deben ser inactivados. Se recomienda la inactivación con lejía al 10% de 16 a 18 horas o el uso de la autoclave a 121°C por 20 minutos.
15. En caso de derrame accidental de algún producto, se debe utilizar papel absorbente embebido en lejía y posteriormente en agua. El papel debe eliminarse en contenedores designados para este fin en hospitales y laboratorios.
16. El ácido sulfúrico es irritante. En caso de derrame, se debe lavar la superficie con abundante agua.
17. Otros materiales de desecho generados durante la utilización del equipo (por ejemplo: puntas usadas en la manipulación de las muestras y controles, microplacas usadas) deben ser manipuladas como fuentes potenciales de infección de acuerdo a las directivas nacionales y leyes para el tratamiento de residuos de laboratorio.

G. MUESTRA: PREPARACIÓN Y RECOMENDACIONES.

1. Extraer la sangre asépticamente por punción venosa y preparar el suero o plasma según las técnicas estándar de los laboratorios de análisis clínico. No se ha detectado que el tratamiento con citrato, EDTA o heparina afecte las muestras.
2. Evitar el uso de conservantes, en particular azida sódica, ya que pudiera afectar la actividad enzimática del conjugado, generando resultados falsos negativos.
3. Las muestras deben estar identificadas claramente mediante código de barras o nombres, a fin de evitar errores en los resultados. Cuando el equipo se emplea para el cribado en unidades de sangre, se recomienda el uso del código de barras.
4. Las muestras hemolizadas (color rojo) o hiperlipémicas (aspecto lechoso) deben ser descartadas para evitar falsos resultados, al igual que aquellas donde se observe la presencia de precipitados, restos de fibrina o filamentos microbianos.
5. El suero y el plasma pueden conservarse a una temperatura entre +2° y +8°C en tubos de recolección principales hasta cinco días después de la extracción. No congelar tubos de recolección principales. Para periodos de almacenamiento más prolongados, las muestras de plasma o suero, retiradas cuidadosamente del tubo de extracción principal, pueden almacenarse congeladas a -20°C durante varios meses, evitando luego descongelar cada muestra más de una vez, ya que se pueden generar partículas que podrían afectar al resultado de la prueba.
6. Si hay presencia de agregados, la muestra se puede aclarar mediante centrifugación a 2000 rpm durante 20 minutos o por filtración con un filtro de 0,2-0,8 micras.

H. PREPARACIÓN DE LOS COMPONENTES Y PRECAUCIONES.

Según estudios realizados, no se ha detectado pérdida relevante de actividad en equipos abiertos, utilizados hasta 6 veces, en un período de hasta 6 meses.

1. Microplacas:

Dejar la microplaca a temperatura ambiente (aprox. 1 hora) antes de abrir el envase. Compruebe que el desecante no esté de un color verde oscuro, lo que indicaría un defecto de fabricación. De ser así, debe solicitar el servicio de Dia.Pro: atención al cliente.

Las tiras de pocillos no utilizadas, deben guardarse herméticamente cerradas en la bolsa de aluminio con el

deseicante a 2-8°C. Una vez abierto el envase, las tiras sobrantes, se mantienen estables hasta que el indicador de humedad dentro de la bolsa del desecante cambie de amarillo a verde.

2. Control Negativo:

Listo para el uso. Mezclar bien con la ayuda de un vórtex, antes de usar.

3. Control Positivo:

Listo para el uso. Mezclar bien con la ayuda de un vórtex, antes de usar. Manipule este reactivo como potencialmente infeccioso, aunque las partículas virales presentes en el control han sido inactivadas químicamente.

4. Calibrador:

Disolver cuidadosamente el contenido del vial en el volumen de agua de calidad EIA indicado en la etiqueta. Mezclar bien con el vórtex antes de usar.

Manipule este reactivo como potencialmente infeccioso, aunque las partículas virales presentes en el control han sido inactivadas químicamente.

Nota: Una vez reconstituida, la solución no es estable. Se recomienda mantenerla congelada en alícuotas a -20°C.

5. Solución de Lavado Concentrada:

Todo el contenido de la solución concentrada 20x debe diluirse con agua bidestilada fino a 1200 ml y mezclarse suavemente antes de usarse.

Por que en los frascos pueden estar presente los cristales, cuando se prepara la solución prestar mucha atención en diluir todo el contenido. Durante la preparación evitar la formación de espuma y burbujas, lo que podría influir en la eficiencia de los ciclos de lavado.

Nota: Una vez diluida, la solución es estable por una semana a temperaturas entre +2 y 8°C.

6. Conjugado:

Listo para el uso. Mezclar bien con un vórtex antes de usar. Evitar posible contaminación del líquido con oxidantes químicos, polvo o microbios. En caso de que deba transferirse el reactivo, usar contenedores de plástico, estériles y desechables, siempre que sea posible.

7. Cromógeno/ Substrato:

Listo para el uso. Mezclar bien con un vórtex antes de usar. Evitar posible contaminación del líquido con oxidantes químicos, polvo o microbios. Evitar la exposición a la luz, agentes oxidantes y superficies metálicas. En caso de que deba transferirse el reactivo, usar contenedores de plástico, estériles y desechables, siempre que sea posible.

8. Diluyente de ensayo:

Listo para el uso. Mezclar bien con un vórtex antes de usar.

9. Ácido Sulfúrico:

Listo para el uso. Mezclar bien con un vórtex antes de usar. Atención: Irritante (H315, H319; P280, P302+P352, P332+P313, P305+P351+P338, P337+P313, P362+P363).

Leyenda:

Indicación de peligro, **Frases H**

H315 – Provoca irritación cutánea.

H319 – Provoca irritación ocular grave.

Consejo de prudencia, **Frases P**

P280 – Llevar guantes/prendas/gafas/máscara de protección.

P302 + P352 – EN CASO DE CONTACTO CON LA PIEL: Lavar con agua y jabón abundantes.

P332 + P313 – En caso de irritación cutánea: Consultar a un médico.

P305 + P351 + P338 – EN CASO DE CONTACTO CON LOS OJOS: Aclarar cuidadosamente con agua durante varios minutos. Quitar las lentes de contacto, si lleva y resulta fácil. Seguir aclarando.

P337 + P313 – Si persiste la irritación ocular: Consultar a un médico.

P362 + P363 – Quitarse las prendas contaminadas y lavarlas antes de volver a usarlas.

10. Diluyente de muestras :

Listo para el uso. Mezclar bien con un vórtex antes de usar.

I. INSTRUMENTOS Y EQUIPAMIENTO UTILIZADOS EN COMBINACIÓN CON EL EQUIPO.

1. Las micropipetas deben ser calibradas para dispensar correctamente el volumen requerido en el ensayo y sometidas a una descontaminación periódica de las partes que pudieran entrar accidentalmente en contacto con la muestra o los reactivos (lejía 10%, de calidad de los desinfectantes hospitalarios). Deben además, ser regularmente revisadas para mantener una precisión del 1% y una confiabilidad de +/- 2%. Deben descontaminarse periódicamente los residuos de los componentes del equipo.
2. La incubadora de ELISA debe ser ajustada a 37°C (+/- 0.5°C) y controlada periódicamente para mantener la temperatura correcta. Pueden emplearse incubadoras secas o baños de agua siempre que estén validados para la incubación de pruebas de ELISA.
3. El **lavador ELISA** es extremadamente importante para el rendimiento global del ensayo. El lavador debe ser validado de forma minuciosa previamente, revisado para comprobar que suministra el volumen de dispensación correcto y enviado regularmente a mantenimiento de acuerdo con las instrucciones de uso del fabricante. En particular, deben lavarse minuciosamente las sales con agua desionizada del lavador al final de la carga de trabajo diaria. Antes del uso, debe suministrarse extensivamente solución de lavado diluida al lavador. Debe enviarse el instrumento semanalmente a descontaminación según se indica en su manual (se recomienda descontaminación con NaOH 0.1 M). Para asegurar que el ensayo se realiza conforme a los rendimientos declarados, basta con 5 ciclos de lavado (aspiración + dispensado de 350 µl/pocillo de solución de lavado + 20 segundos de remojo = 1 ciclo). Si no es posible remojar, añadir un ciclo de lavado adicional. Un ciclo de lavado incorrecto o agujas obstruidas con sal son las principales causas de falsas reacciones positivas.
4. Los tiempos de incubación deben tener un margen de ±5%.
5. El lector de microplacas ELISA debe estar provisto de un filtro de lectura de 450 nm y de un segundo filtro de 620-630 nm, obligatorio para el blanco. El procedimiento estándar debe contemplar: a) Ancho de banda ≤ 10 nm; b) Rango de absorbancia de 0 a ≥ 2,0; c) Linealidad ≥ 2,0; d) Reproducibilidad ≥ 1%. El blanco se prueba en el pocillo indicado en la sección "Procedimiento del ensayo". El sistema óptico del lector debe calibrarse periódicamente para garantizar que se mide la densidad óptica correcta. Periódicamente se debe proceder al mantenimiento según las instrucciones del fabricante.
6. En caso de usar un sistema automatizado de ELISA, los pasos críticos (dispensado, incubación, lavado, lectura, agitación y procesamiento de datos) deben ser cuidadosamente fijados, calibrados, controlados y periódicamente ajustados, para garantizar los valores indicados en la sección "Control interno de calidad". El protocolo del ensayo debe ser instalado en el sistema operativo de la unidad y validado tanto para el lavador como para el lector. Por otro lado, la parte del sistema que maneja los líquidos (dispensado y lavado) debe ser validada y fijada correctamente. Debe prestarse particular

atención a evitar el arrastre por las agujas de dispensación y de lavado, a fin de minimizar la posibilidad de ocurrencia de falsos positivos por contaminación de los pocillos adyacentes por muestras fuertemente reactivas. Se recomienda el uso de sistemas automatizados para el cribado en unidades de sangre y cuando la cantidad de muestras supera las 20-30 unidades por ensayo.

7. Cuando se utilizan instrumentos automáticos, en el caso en que los contenedores para los frascos del instrumento no sean adecuados a los frascos del kit, transferir la solución en ellos contenida en frascos idóneos al instrumento y etiquetarlos con la misma etiqueta utilizada en el frasco original. Esta operación es importante para evitar el cambio del contenido de los frascos durante el transferimiento. Cuando el test a terminado colocar los contenedores secundarios etiquetados y tapados a 2.8°C.
8. El servicio de atención al cliente en Dia.Pro, ofrece apoyo al usuario para calibrar, ajustar e instalar los equipos a usar en combinación con el equipo, con el propósito de asegurar el cumplimiento de los requerimientos descritos.

L. OPERACIONES Y CONTROLES PREVIOS AL ENSAYO.

1. Compruebe la fecha de caducidad indicada en la parte externa del equipo (envase primario). No usar si ha caducado.
2. Compruebe que los componentes líquidos no están contaminados con partículas o agregados visibles. Asegúrese de que el cromógeno (TMB) es incoloro o azul pálido, aspirando un pequeño volumen de este con una pipeta estéril de plástico. Compruebe que no han ocurrido rupturas ni derrames de líquido dentro de la caja (envase primario) durante el transporte. Asegurarse de que la bolsa de aluminio que contiene la microplaca no esté rota o dañada.
3. Diluir totalmente la solución de lavado concentrada 20X, como se ha descrito anteriormente.
4. Disolver el Calibrador como se ha descrito anteriormente y mezclar suavemente.
5. Dejar los componentes restantes alcanzar la temperatura ambiente (aprox. 1 hora), mezclar luego suavemente en el vórtex todos los reactivos líquidos.
6. Ajustar la incubadora de ELISA a 37°C y cebar el lavador de ELISA utilizando la solución de lavado, según las instrucciones del fabricante. Fijar el número de ciclos de lavado según se indica en la sección específica.
7. Comprobar que el lector de ELISA esté conectado al menos 20 minutos antes de realizar la lectura.
8. En caso de trabajar automáticamente, conectar el equipo y comprobar que los protocolos estén correctamente programados.
9. Comprobar que las micropipetas estén fijadas en el volumen requerido.
10. Asegurarse de que el equipamiento a usar esté en perfecto estado, disponible y listo para el uso.
11. En caso de surgir algún problema, se debe detener el ensayo y avisar al responsable.

M. PROCEDIMIENTO DEL ENSAYO.

El ensayo debe realizarse según las instrucciones que siguen a continuación, es importante mantener en todas las muestras el mismo tiempo de incubación.

Ensayos Automatizados.

En el caso de que el ensayo se realice de manera automatizada con un sistema ELISA, se recomienda programar al equipo para aspirar 200µl de Diluyente de Muestras, y posteriormente 10µl de muestra.

La mezcla debe ser dispensada cuidadosamente en los pocillos correspondientes a cada muestra. Antes de aspirar la muestra siguiente, las agujas deben lavarse debidamente para evitar cualquier contaminación cruzada entre las muestras.

No diluir el Calibrador ni los controles ya que están listos para el uso.

Dispensar 200µl de controles/Calibrador en los pocillos correspondientes.

Nota importante: Controle a simple vista que las muestras han sido diluidas y dispensadas en los pocillos adecuados, para lo cual el color de las muestras dispensadas debe ser verde azul oscuro, mientras que el del control negativo debe permanecer verde olivo.

Para las operaciones siguientes, consulte las instrucciones que aparecen debajo para el Ensayo Manual.

Es muy importante comprobar que el tiempo entre el dispensado de la primera y la última muestra sea calculado por el instrumento y considerado para los lavados.

Ensayo Manual.

1. Poner el número de tiras necesarias en el soporte de plástico. Dejar el primer pocillo vacío para el blanco.
2. Dispensar 200µl del Control Negativo, por triplicado, 200µl de Calibrador por duplicado y 200µl del Control Positivo. No diluir el Calibrador ni los controles ya que están listos para el uso!
3. Dispensar 200µl del Diluyente de muestras (DILSPE) a todos los pocillos de muestras, después dispensar 10 µl de cada muestra en su pocillo correspondiente. Resuspender suavemente evitando la formación de espuma y la contaminación de los pocillos adyacentes.

Nota importante: Comprobar que el color del Diluyente de muestras, después de adicionada la misma, cambia de verde a verde azul oscuro.

4. Dispensar 50 µl de Diluyente de ensayo (DILAS) en los pocillos de los controles/Calibrador y muestras. Compruebe que el color de las muestras sea azul oscuro.
5. Incubar la microplaca **45 min a +37°C**.

Nota importante: Las tiras se deben sellar con el adhesivo suministrado solo cuando se hace el test manualmente. No sellar cuando se emplean equipos automatizados de ELISA.

6. Lavar la microplaca con el lavador automático dispensando y aspirando 350 µl/pocillo de solución de lavado diluida, según según se indica (sección 1.3).
7. Dispensar 100µl del Conjugado en todos los pocillos, excepto en el A1 y cubrir con el sellador. Compruebe que este reactivo de color rosa/rojo ha sido añadido en todos los pocillos excepto el A1.

Nota importante: Tener cuidado de no tocar la pared interna del pocillo con la punta de la pipeta al dispensar el conjugado. Podría producirse contaminación.

8. Incubar la microplaca **45 min a +37°C**.
9. Lavar la microplaca, de igual forma que en el paso 6.
10. Dispensar 100µl del Cromógeno/Substrato en todos los pocillos, incluido el A1. Incubar la microplaca a **temperatura ambiente (18-24°C) durante 15 minutos**.

Nota importante: No exponer directamente a fuerte iluminación, de lo contrario se generan interferencias.

11. Dispensar 100µl de ácido sulfúrico en todos los pocillos para detener la reacción enzimática, usar la misma secuencia que en el paso 10. La adición de la solución de parada cambia el color del Control Positivo y las muestras positivas de azul a amarillo/marrón.

12. Medir la intensidad del color de la solución en cada pocillo, según se indica en la sección I.5, con un filtro de 450 nm (lectura) y, otro de 620-630 nm (substracción del fondo), calibrando el instrumento con el pocillo A1 (blanco, obligatorio).

Notas importantes:

1. Asegurarse de que no hay impresiones digitales ni polvo en el fondo de los pocillos antes de leer. Podrían generarse falsos positivos en la lectura.
2. La lectura debe hacerse inmediatamente después de añadir la solución de parada y, en cualquier caso, nunca transcurridos 20 minutos después de su adición. Se podría producir auto oxidación del cromógeno causando un elevado fondo.
3. Se ha probado que la agitación a 350 +/- 150 rpm, durante la incubación, aumenta en un 20% la sensibilidad del ensayo.
4. El calibrador (CAL) no afecta al cálculo del valor de corte y, por lo tanto, no afecta al cálculo de los resultados de la prueba. El calibrador (CAL) se usa solo si la gestión requiere un control interno de calidad del laboratorio.

N. ESQUEMA DEL ENSAYO.

Método	Operaciones
Controles & Calibrador	200 µl
Muestras	200µl dil.+10µl
Diluyente de ensayo (DILAS)	50 µl
1ª incubación	45 min
Temperatura	+37°C
Lavado	5 ciclos con 20" de remojo o 6 ciclos sin remojo
Conjugado	100 µl
2ª incubación	45 min
Temperatura	+37°C
Lavado	5 ciclos con 20" de remojo o 6 ciclos sin remojo
TMB/H2O2	100 µl
3ª incubación	15 min
Temperatura	18-24°C
Acido Sulfúrico	100 µl
Lectura D.O.	450nm / 620-630nm

A continuación se describe un ejemplo del esquema de dispensado.

		Microplaca											
		1	2	3	4	5	6	7	8	9	10	11	12
A	BL	M2											
B	CN	M3											
C	CN	M4											
D	CN	M5											
E	CAL	M6											
F	CAL	M7											
G	CP	M8											
H	M 1	M9											

Leyenda: BL = Blanco CN = Control Negativo CAL = Calibrador CP = Control Positivo M = Muestra

O. CONTROL DE CALIDAD INTERNO.

Se realiza un grupo de pruebas con los controles/calibrador cada vez que se usa el equipo para verificar si los valores DO450nm son los esperados.

Asegurar el cumplimiento de los siguientes parámetros:

Parámetro	Exigencia
Pocillo Blanco	Valor < 0.100 DO450nm
Control Negativo (CN)	Valor medio < 0.050 DO450nm después de leer el blanco
Calibrador	M/Co > 1.1
Control Positivo	Valor > 1.000 DO450nm

Si los resultados del ensayo coinciden con lo establecido anteriormente, pase a la siguiente sección.

En caso contrario, detenga el ensayo y compruebe:

Problema	Compruebe que
Pocillo blanco > 0.100DO450nm	la solución cromógeno/substrato no se ha contaminado durante el ensayo.
Control Negativo (CN) > 0.050 DO450nm después de leer el blanco	1. el proceso de lavado y los parámetros del lavador estén validados según los estudios previos de calificación. 2. se ha usado la solución de lavado apropiada y que el lavador ha sido cebado con la misma antes del uso. 3. no se han cometido errores en el procedimiento (dispensar el control positivo en lugar del negativo). 4. no ha existido contaminación del control negativo o de sus pocillos debido a muestras positivas derramadas, o al conjugado. 5. las micropipetas no se han contaminado con muestras positivas o con el conjugado. 6. las agujas del lavador no estén parcial o totalmente obstruidas.
Calibrador M/Co < 1.1	1. el procedimiento ha sido realizado correctamente. 2. no ha habido errores durante su distribución (dispensar el control negativo en lugar del calibrador). 3. el proceso de lavado y los parámetros del lavador estén validados según los estudios previos de calificación. 4. no ha ocurrido contaminación externa del calibrador.
Control Positivo < 1.000 DO450nm	1. el procedimiento ha sido realizado correctamente. 2. no se han cometido errores en el procedimiento (dispensar el control negativo en lugar del positivo). En este caso el control negativo debe tener un valor de DO450nm > 0.150. 3. el proceso de lavado y los parámetros del lavador estén validados según los estudios previos de calificación. 4. no ha ocurrido contaminación externa del control positivo.

Si ocurre alguno de los problemas anteriores, después de comprobar, informe al responsable para tomar las medidas pertinentes.

P. CÁLCULO DEL VALOR DE CORTE.

Los resultados se calculan por medio de un valor de corte (cut-off) hallado con la siguiente fórmula:

$$\text{Valor de corte} = \text{CN medio DO450nm} + 0.350$$

El valor encontrado para el ensayo se usa para la interpretación de los resultados, según se describe a continuación:

Nota Importante: Cuando el cálculo de los resultados se halla mediante el sistema operativo de un equipo de ELISA automático, asegurarse de que la formulación usada para el cálculo del valor de corte, y para la interpretación de los resultados sea correcta.

Control Positivo: 2.189 DO450nm
 Mayor de 1.000 – Válido
 Valor de corte = $0.020 + 0.350 = 0.370$

Calibrador: 0.550 - 0.530 DO450nm
 Valor medio: 0.540 DO450nm M/Co = 1.4
 M/Co Mayor de 1.1 – Válido

Q. INTERPRETACIÓN DE LOS RESULTADOS.

La interpretación de los resultados se realiza mediante la razón entre las DO a 450nm de las muestras y el Valor de corte (M/Co).

Los resultados se interpretan según la siguiente tabla:

(M/Co)	Interpretación
< 0.9	Negativo
0.9 – 1.1	Equívoco
> 1.1	Positivo

Un resultado negativo indica que el paciente no está infectado por HCV y la unidad de sangre se puede transfundir.

Cualquier paciente, cuya muestra resulte equívoca debe someterse a una nueva prueba con una segunda muestra de sangre colectada 1 ó 2 semanas después de la inicial. En este caso la unidad de sangre no debe ser transfundida.

Un resultado positivo es indicativo de infección por HCV y por consiguiente el paciente debe ser tratado adecuadamente. La unidad de sangre debe ser descartada.

Notas importantes:

1. La interpretación de los resultados debe hacerse bajo la vigilancia del responsable del laboratorio para reducir el riesgo de errores de juicio y de interpretación.
2. Antes de formular un diagnóstico de hepatitis viral, los resultados positivos deben comprobarse a través de un método alternativo, capaz de detectar anticuerpos IgG e IgM (prueba confirmatoria).
3. Según se demuestra en la Evaluación del Performance del producto, el ensayo es capaz de detectar los anticuerpos anti HCV core, en etapas más tempranas en comparación con otros equipos comerciales. Sin embargo, un resultado positivo, no confirmado con estos equipos comerciales, no debe necesariamente considerarse como falso positivo! Es necesario realizar una prueba de confirmación (suministrada, bajo solicitud del cliente, por Dia.pro srl. Codificada CCONF).
4. Como el ensayo es capaz de detectar además anticuerpos IgM, pueden presentarse resultados discrepantes (pérdida de reactividad IgM) con respecto a otros productos comerciales para la detección de anticuerpos anti-HCV. La positividad real de una muestra debe confirmarse probando la reactividad IgM, lo cual resulta muy importante para el diagnóstico de infección por HCV.
5. Cuando se transmiten los resultados de la prueba, del laboratorio a otras instalaciones, debe ponerse mucha atención para evitar el traslado de datos erróneos.
6. El diagnóstico de infección con un virus de la hepatitis debe ser evaluado y comunicado al paciente por un médico calificado.

A continuación, un ejemplo de los cálculos a realizar:

Los siguientes datos no deben usarse en lugar de los valores reales obtenidos en el laboratorio.

Control Negativo: 0.019 – 0.020 – 0.021 DO450nm
 Valor medio: 0.020 DO450nm
 Menor de 0.050 – Válido

R. FUNCIONAMIENTO.

La evaluación del funcionamiento ha sido realizada según lo reportado en las Especificaciones Técnicas Comunes (ETC) (art. 5, Capítulo 3 de las Directivas IVD 98/79/EC).

1. LÍMITE DE DETECCIÓN.

El límite de detección ha sido calculado por medio del estándar de trabajo británico anti-HCV NIBSC, código 99/558-003-WI). La siguiente tabla muestra los valores medios de DO450nm de este estándar diluido en plasma negativo y examinado:

Dilución	Lote # 1	Lote # 2
Factor	M/Co	M/Co
1 X	2.0	2.0
2 X	1.1	1.2
4 X	0.7	0.8
8 X	0.5	0.5
Plasma Negativo	0.3	0.3

Se evaluó además la muestra Accurun 1 –serie 3000– suministrado por Boston Biomedica Inc., Estados Unidos.

Los resultados son los siguientes:

CVAB.CE Lote ID	Accurun 1 Serie	M/Co
1201	3000	1.5
0602	3000	1.5
1202	3000	1.9

Por otra parte, un total de 7 muestras, positivas para HCVAb según Ortho HCV 3.0 SAVe, código 930820, lote # EXE065-1, fueron diluidas en plasma negativo a HCVAb con el fin de obtener diluciones limitantes y luego fueron probadas nuevamente en CVAB.CE, lote # 1202, y Ortho.

Las tablas siguientes reflejan los resultados obtenidos:

Muestra n°	Dilución Límite	CVAB.CE M/Co	Ortho 3.0 M/Co
1	256 X	1.9	1.3
2	256 X	1.9	0.7
3	256 X	2.4	1.0
4	128 X	2.5	3.2
5	85 X	3.3	1.4
6	128 X	2.2	0.8
7	135 X	3.2	2.2

2. ESPECIFICIDAD Y SENSIBILIDAD DIAGNÓSTICAS.

La evaluación del procedimiento diagnóstico se realizó mediante un ensayo con más de 5000 muestras.

2.1 Especificidad Diagnóstica:

Se define como la probabilidad del ensayo de detectar negativos en ausencia del analito específico.

Además del primer estudio, donde se examinaron en total 5043 muestras de donantes de sangre no seleccionados, (incluyendo donantes por 1ª vez), 210 muestras de pacientes hospitalizados y 162 muestras que pudieran provocar interferencia (otras enfermedades infecciosas, positivas para anticuerpos de E. coli, pacientes con enfermedades hepáticas no virales, pacientes en diálisis, mujeres embarazadas, hemolizadas, lipémicas, etc.), la especificidad diagnóstica se evaluó recientemente examinando un total de 2876 muestras de donantes de sangre negativas en seis lotes distintos. Se observó un valor de especificidad de 100%.

Se emplearon además, plasma sometido a métodos de tratamiento estándar (citrato, EDTA y heparina) y suero humanos. No se ha observado falsa reactividad debida a los métodos de tratamiento de muestras.

Por último se analizaron muestras congeladas, para determinar posibles interferencias debidas a la toma de muestra y al almacenamiento. No se observaron interferencias.

2.2 Sensibilidad Diagnóstica.

Se define como la probabilidad del ensayo de detectar positivos en presencia del analito específico.

La sensibilidad diagnóstica ha sido estimada de forma externa en un total de 359 muestras, el valor obtenido fue de 100%. Más de 50 muestras positivas fueron probadas de forma interna, en este caso el resultado fue también de 100%.

Se evaluaron además, muestras positivas producto de infecciones por diferentes genotipos de HCV, así como también se estudió gran parte de los paneles de seroconversión de Boston Biomedica Inc (PHV) y Zeptometrix, USA (HCV), disponibles.

Los resultados para algunos de ellos se describen a continuación:

Panel	N° samples	DiaPro*	Ortho**
PHV 901	11	9	9
PHV 904	7	2	4
PHV 905	9	3	4
PHV 906	7	7	7
PHV 907	7	3	2
PHV 908	13	10	8
PHV 909	3	2	2
PHV 910	5	3	3
PHV 911	5	3	3
PHV 912	3	1	1
PHV 913	4	2	2
PHV 914	9	5	5
PHV 915	4	3	0
PHV 916	8	4	3
PHV 917	10	6	6
PHV 918	8	2	0
PHV 919	7	3	3
PHV 920	10	6	6
HCV 10039	5	2	0
HCV 6212	9	6	7
HCV 10165	9	5	4

Note: * Positive samples detected

** HCV v.3.0

Por último, el producto ha sido probado contra el panel EFS Ac HCV, lote n° 01/08.03.22C/01/A, suministrado por Etablissement Francais Du Sang (EFS), Francia, obteniéndose los siguientes resultados:

EFS Panel Ac HCV

Muestra	Lote # 1 M/Co	Lote # 2 M/Co	Lote# 3 M/Co	Resultados esperados
HCV 1	2.2	2.4	2.6	positivo
HCV 2	1.6	2.0	2.1	positivo
HCV 3	1.5	1.7	1.6	positivo
HCV 4	5.2	6.5	5.5	positivo
HCV 5	1.6	1.8	1.6	positivo
HCV 6	0.4	0.4	0.4	negativo

3. PRECISIÓN.

Ha sido calculada utilizando dos muestras, una negativa y una débil positiva, examinadas en 16 réplicas en tres corridas separadas.

Los resultados se muestran a continuación:

Lote # 1202

Muestra Negativa (N = 16)

Valores medios	1ª corrida	2ª corrida	3ª corrida	Valor Promedio
DO 450nm	0.094	0.099	0.096	0.096
Desviación estándar	0.008	0.007	0.008	0.007
CV %	8.7	6.6	7.9	7.7

Cal # 2 – 7K (N = 16)

Valores medios	1ª corrida	2ª corrida	3ª corrida	Valor Promedio
DO 450nm	0.396	0.403	0.418	0.406
Desviación estándar	0.023	0.029	0.027	0.026
CV %	5.9	7.1	6.4	6.5
M/Co	1.1	1.1	1.2	1.1

Lote # 0602

Muestra Negativa (N = 16)

Valores medios	1ª corrida	2ª corrida	3ª corrida	Valor Promedio
DO 450nm	0.097	0.096	0.094	0.096
Desviación estándar	0.009	0.010	0.008	0.009
CV %	8.9	10.1	8.4	9.1

Cal # 2 – 7K (N = 16)

Valores medios	1ª corrida	2ª corrida	3ª corrida	Valor Promedio
DO 450nm	0.400	0.395	0.393	0.396
Desviación estándar	0.021	0.025	0.026	0.024
CV %	5.4	6.2	6.6	6.1
M/Co	1.2	1.2	1.1	1.2

Lote # 0602/2

Muestra Negativa (N = 16)

Valores medios	1 ^{ra} corrida	2 ^{da} corrida	3 ^{ra} corrida	Valor Promedio
DO 450nm	0.087	0.091	0.088	0.089
Desviación estándar	0.009	0.007	0.008	0.008
CV %	10.0	8.2	8.6	8.9

Cal # 2 - 7K (N = 16)

Valores medios	1 ^{ra} corrida	2 ^{da} corrida	3 ^{ra} corrida	Valor Promedio
DO 450nm	0.386	0.390	0.391	0.389
Desviación estándar	0.023	0.021	0.023	0.022
CV %	6.0	5.3	5.8	5.7
M/Co	1.1	1.2	1.2	1.2

La variabilidad mostrada en las tablas no dió como resultado una clasificación errónea de las muestras.

S. LIMITACIONES.

Los falsos positivos repetibles, no confirmados por RIBA o similares técnicas de confirmación, fueron estimados como menos del 0.1% de la población normal.

Las muestras que después de ser descongeladas presentan partículas de fibrina o partículas agregadas, generan algunos resultados falsos positivos.

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Todos los productos de diagnóstico in vitro fabricados por la empresa son controlados por un sistema certificado de control de calidad aprobado por un organismo notificado para el mercado CE. Cada lote se somete a un control de calidad y se libera al mercado únicamente si se ajusta a las especificaciones técnicas y criterios de aceptación de la CE.

Fabricante:
Dia.Pro Diagnostic Bioprobes S.r.l.
Via G. Carducci n° 27 – Sesto San Giovanni
(Milán) – Italia



0318



Dia.Pro
Diagnostic
Bio**Probes**

Letter of Authorization

We, "Dia.Pro Diagnostic Bioprobes S.r.l." located at Via G. Carducci, Nr. 27 – Sesto San Giovanni (Milan) 20099, Italy, authorize

GLOBAL BIOMARKETING GROUP – MOLDOVA SRL
Str. Tighina 65, Oficiu 607
MD-2001 CHISINAU
REP. MOLDOVA

as our exclusive distributor for the territory of the Republic of Moldova, to participate in various tenders with **Dia.Pro** ELISA products.

We, Dia.Pro Diagnostic Bioprobes S.r.l shall supply our distributor GLOBAL BIOMARKETING GROUP – MOLDOVA SRL with all products in strict compliance with the existing "Distribution Agreement" rev.0121 valid until 31-Dec-2023, with possibility of renewal upon agreement between both parties for an additional period.

Dia.Pro Diagnostic Bioprobes S.r.l will grant the supply of all awarded tenders until their natural expiry, of which a documental proof has to be provided to Dia.Pro by the distributor GLOBAL BIOMARKETING GROUP – MOLDOVA SRL.

Sincerely yours,

Date: **Milan, 04-February-2021**

Dia.Pro Diagnostic Bioprobes S.r.l.

DIA. PRO.
DIAGNOSTIC BIOPROBES S.r.l.

Dr.ssa Fiorenza Scozzesi

Legal Representative

DIA.PRO Diagnostic Bioprobes S.r.l.

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Capitale sociale €50.000,00 I.V. – P.IVA: 11924660159 – Reg. Imp. 11924660159 – REA 1509959

DECLARATION OF CONFORMITY

1) Manufacturer (Name, department): **Monobind Inc.**

Address: **100 North Pointe, LAKE FOREST, CA 92630. UNITED STATES**

and

2) European authorized representative: **CEpartner4U BV,**

Address: **ESDOORNLAAN 13, 3951DB MAARN, THE NETHERLANDS;**

(on product labels printed as:

CEpartner4U , ESDOORNLAAN 13, 3951DB MAARN, THE NETHERLANDS Tel.: +31 (0)6 516 536 26;

or as: CEpartner4U, 3951DB; 13. NL tel: +31 (0)6 – 516.536.26)

3) Product(s) (name, type or model/batch number, etc.):

Immunoassay products;

ELISA,

CLIA,

Control,

Instruments

(see appendix)

4) The product(s) described above is in conformity with:

<u>Document No.</u>	<u>Title</u>	<u>Edition / Date of issue</u>
L 331; 98/79/EC	In-Vitro-Diagnostic Directive	1998-10-27

5) Additional information (conformity procedure, Notified Body, CE certificate, etc.):

Conformity assessment procedure for CE marking: IVD Directive, Annex III

Lake Forest, USA;2011-09-27

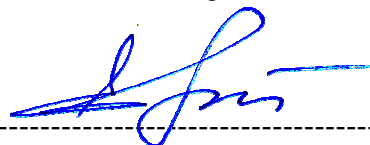


Tony Shatola; QA Director, Monobind Inc.

(Place & date of issue (yyyy-mm-dd))

(name, function and signature of manufacturer)

Maarn, NL; 2011-09-27



Olga Teirlinck; Consultant, CEpartner4U BV

(Place & date of issue (yyyy-mm-dd))

(name; function and signature of authorized representative)

Appendix

Date: 2011-09-26

<i>Device types</i>	<i>Item# ELISA</i>	<i>Item# CLIA</i>	<i>Item# Control</i>	<i>Item# Instrument</i>	<i>EDMS code</i>	<i>Risk Class</i>	<i>Certificate #</i>	<i>First date of CE-marking</i>
Thyroid								
T3 – Triiodothyronine	125-300	175-300			12.04.01.05.00	Low		2005-11-11
fT3 – Free Triiodothyronine	1325-300	1375-300			12.04.01.01.00	Low		2005-11-11
T4 – Thyroxine	225-300	275-300			12.04.01.07.00	Low		2005-11-11
fT4 – Free Thyroxine	1225-300	1275-300			12.04.01.02.00	Low		2005-11-11
TSH – Thyrotropin	325-300	375-300			12.04.01.11.00	Low		2005-11-11
Rapid TSH – Rapid Thyrotropin	6025-300	6075-300			12.04.01.11.00	Low		2010-06-29
T3U – Triiodothyronine Uptake	525-300	575-300			12.04.01.06.00	Low		2005-11-11
TBG – Thyroxine-Binding Globulin	3525-300	3575-300			12.04.01.09.00	Low		2005-11-11
Tg – Thyroglobulin	2225-300	2275-300			12.04.01.08.00	Low		2005-11-11
T3, T4 & TSH – Triiodothyronine, Thyroxine & Thyrotropin Combo (VAST)	8025-300	8075-300			12.04.01.01.00	Low		2005-11-11
T3 – Triiodothyronine (SBS)	8125-300	8175-300			12.04.01.01.00	Low		2010-06-29
T4- Thyroxine (SBS)	8225-300	8275-300			12.04.01.01.00	Low		2010-06-29
fT3, fT4 & TSH – Free Triiodothyronine, Free Thyroxine & Thyrotropin Combo (VAST)	7025-300	7075-300			12.04.01.01.00	Low		2010-06-29
Neonatal Thyroid & Genetics								
NTSH – Neonatal Thyrotropin	3425-300	3475-300			12.04.01.90.00	Low		2005-11-11
NT4 – Neonatal Thyroxine	2625-300	2675-300			12.04.01.12.00	Low		2005-11-11
N 17OHP – Neonatal 17 OH Progesterone	5525-300				12.05.01.07	Low		2008-02-01
Biotinidase	8825-300				12 07 02 90 00	Low		2011-09-26
Autoimmune Thyroid								
Anti-Tg – Anti-Thyroglobulin Antigen	1025-300	1075-300			12.10.03.04.00	Low		2005-11-11
Anti-TPO – Anti-Thyropoxidase Antigen	1125-300	1175-300			12.10.03.01.00	Low		2005-11-11
Fertility & Prenatal								
LH – Lutropin	625-300	675-300			12.05.01.05.00	Low		2005-11-11
FSH – Follitropin	425-300	475-300			12.05.01.04.00	Low		2005-11-11
PRL – Prolactin	725-300	775-300			12.05.01.08.00	Low		2005-11-11
PRL – Prolactin Sequential	6025-300	6075-300			12.05.01.08.00	Low		2005-11-11
hCG – Human Chorionic Gonadotropin	825-300	875-300			12.05.02.05.00	Low		2005-11-11
Rapid hCG – Rapid Human Chorionic Gonadotropin	3325-300				12.05.02.05.00	Low		2005-11-11
FSH, LH, hCG, sPRL Combo (VAST)	8325-300	8375-300			12.05.01.90.00	Low		2006-08-24
AFP, hCG, uE3 Combo (VAST)	8525-300	8575-300			12.05.01.90.00	Low		2010-06-29
Steroid								
Cortisol	3625-300	3675-300			12.06.02.04.00	Low		2005-11-11
DHEA-S – Dehydroepiandrosterone sulfate	5125-300	5175-300			12.05.01.02.00	Low		2010-06-29
DHEA - Dehydroepiandrosterone	7425-300	7475-300			12.05.01.02.00	Low		2011-09-26

<i>Device types</i>	<i>Item# ELISA</i>	<i>Item# CLIA</i>	<i>Item# Control</i>	<i>Item# Instrument</i>	<i>EDMS code</i>	<i>Risk Class</i>	<i>Certificate #</i>	<i>First date of CE-marking</i>
E2 – Estradiol	4925-300	4975-300			12.05.01.03.00	Low		2010-06-29
uE3 – Estriol, Unconjugated	5025-300	5075-300			12.05.02.02.00	Low		2010-06-29
Progesterone	4825-300	4875-300			12.05.01.06.00	Low		2010-06-29
Testosterone	3725-300	3775-300			12.05.01.10.00	Low		2007-11-01
Free Testosterone	5325-300	5375-300			12.05.01.10.00	Low		2010-06-29
17OHP - 17-Hydroxyprogesterone	5225-300	5275-300			12.05.01.07.00	Low		2010-06-29
17OHP - 17-Hydroxyprogesterone Ext. Range	9925-300	9975-300			12.05.01.07.00	Low		2010-10-18
Vitamin D3 – 25-Hydroxyvitamin D3	7725-300	7775-300			12.06.03.10.00	Low		2011-09-26
Growth & Bone Metabolism								
hGH - Human Growth Hormone	1725-300	1775-300			12.06.04.02.00	Low		2005-11-11
PTH - Parathyroid Hormone	7825-300	7875-300			12.06.03.13.00	Low		2011-09-26
Diabetes								
Insulin	2425-300	2475-300			12.06.01.03.00	Low		2005-11-11
Insulin Rapid	5825-300				12.06.01.03.00	Low		2010-06-29
C-peptide	2725-300	2775-300			12.06.01.01.00	Low		2005-11-11
Insulin & C-peptide Combo (VAST)	7325-300	7375-300			12.06.01.03.00	Low		2005-11-11
Cardiac Markers								
CKMB – Circulating Creatine Kinase (MB)	2925-300	2975-300			12.13.01.02.00	Low		2005-11-11
CTnl – Troponin I	3825-300	3875-300			12.13.01.07.00	Low		2005-11-11
DIG – Digoxin	925-300	975-300			12.08.01.01.00	Low		2005-11-11
HS-CRP – High Sensitivity C- Reactive Protein	3125-300	3175-300			12.13.01.90.00	Low		2005-11-11
Myoglobin	3225-300	3275-300			12.13.01.05.00	Low		2005-11-11
Infectious Diseases								
IgG – Anti/H. Pylori	1425-300	1475-300			15.01.04.03.00	Low		2005-11-11
IgM – Anti/H. Pylori	1525-300	1575-300			15.01.04.03.00	Low		2005-11-11
IgA – Anti/H. Pylori	1625-300	1675-300			15.01.04.03.00	Low		2005-11-11
Cancer Markers								
AFP – Alpha-Fetoprotein	1925-300	1975-300			12.03.90.01.00	Low		2005-11-11
CA 125 Ovarian Cancer Antigen	3025-300	3075-300			12.03.01.06.00	Low		2005-11-11
CA 15-3 Breast Cancer Antigen	5625-300	5675-300			12.03.01.02.00	Low		2010-06-29
CA 19-9 - Pancreatic Cancer Antigen	3925-300	3975-300			12.03.01.03.00	Low		2005-11-11
CEA – Carcinoembryonic Antigen	1825-300	1875-300			12.03.01.31.00	Low		2005-11-11
CEA - Carcinoembryonic Antigen Next Generation	4625-300	4675-300			12.03.01.31.00	Low		2010-06-29
fβhCG – Free Beta Human Chorionic Gonadotropin	2025-300	2075-300			12.03.01.90.00	Low		2005-11-11
Allergy & Anemia								
Ferritin	2825-300	2875-300			12.07.01.02.00	Low		2005-11-11
Folate	7525-300	7575-300			12.07.01.03.00	Low		2010-06-29
IgE – Immunoglobulin E	2525-300	2575-300			12.02.01.02.00	Low		2005-11-11
sTfR - Transferrin Soluble Receptor	8625-300	8675-300			12.07.01.06.00	Low		2010-06-29
Vitamin B12	7625-300	7675-300			12.07.02.04.00	Low		2011-09-26

Miscellaneous Controls							
Anti-Tg & Anti-TPO – Positive & Negative - Anti-Thyroglobulin, Anti-Thyroperoxidase			AIT-101		12.50.01.16.00	Low	2010-06-29
High Level Fertility Control – Single Level – Progesterone, Estradiol, Human Chorionic Gonadotropin			FC-300		12.50.01.16.00	Low	2010-06-29
Maternal Control – Tri Level - Human Chorionic Gonadotropin, Free Beta Human Chorionic Gonadotropin Subunit, Alpha Feta Protein, Estriol			MC-300		12.50.01.16.00	Low	2010-06-29
Thyroglobulin Control – Tri Level			TG-300		12.50.01.16.00	Low	2010-06-29
H. Pylori IgG Control – Positive & Negative			HPy-IgG-300		12.50.01.16.00	Low	2010-06-29
Miscellaneous Instruments							
IC hardware + dedicated accessories + software – Autoplex ELISA Analyzer & CLIA Processor				IN006	21.02.10.01	Low	2010-06-29
IC hardware + dedicated accessories + software – Lumax Chemiluminescence Strip Reader				IN001	21.02.10.01	Low	2006-08-24
IC hardware + dedicated accessories + software – Neo-Lumax Chemiluminescence Strip Reader				IN010	21.02.10.01	Low	2011-09-26
IC hardware + dedicated accessories + software – Impulse 2 Chemiluminescence Strip Reader				IN005	21.02.10.01	Low	2006-08-24
IC hardware + dedicated accessories + software – Impulse 3 Chemiluminescence Strip Reader				IN007	21.02.10.01	Low	2010-06-29
IC hardware + dedicated accessories + software – Lumax96 Chemiluminescence Plate Reader				IN004	21.02.10.01	Low	2007-03-01
IC hardware + dedicated accessories + software – LuMatic Chemiluminescence Plate Reader				IN008	21.02.10.01	Low	2011-09-26
IC hardware + dedicated accessories + software – Eldex 3.8 ELISA Strip Reader				IN003	21.02.10.01	Low	2007-09-10
IC hardware + dedicated accessories + software – Neo-Eldex ELISA Strip Reader				IN009	21.02.10.01	Low	2011-09-26
IC hardware + dedicated accessories + software – Microplate Washer				IN002	21.02.10.01	Low	2010-06-29



Total Prostate Specific Antigen (tPSA) Test System

Product Code: 2125-300

1.0 INTRODUCTION

Intended Use: The Quantitative Determination of Total Prostate Specific Antigen (tPSA) Concentration in Human Serum by a Microplate Immunoassay

2.0 SUMMARY AND EXPLANATION OF THE TEST

Prostate Specific antigen (PSA) is a serine protease with chymotrypsin-like activity (1,2). The protein is a single chain glycoprotein with a molecular weight of 28.4 kDa (3). PSA derives its name from the observation that it is a normal antigen of the prostate but is not found in any other normal or malignant tissue.

PSA is found in benign, malignant and metastatic prostate cancer. Since prostate cancer is the second most prevalent form of male malignancy, the detection of elevated PSA levels plays an important role in the early diagnosis. Serum PSA levels have been found to be more useful than prostatic acid phosphatase (PAP) in the diagnosis and management of patients due to increased sensitivity (4).

In this method, PSA calibrator, patient specimen or control is first added to a streptavidin coated well. Biotinylated monoclonal and enzyme labeled antibodies (directed against distinct and different epitopes of PSA) are added and the reactants mixed. Reaction between the various PSA antibodies and native PSA forms a sandwich complex that binds with the streptavidin coated to the well.

After the completion of the required incubation period, the enzyme-PSA antibody bound conjugate is separated from the unbound enzyme-PSA conjugate by aspiration or decantation. The activity of the enzyme present on the surface of the well is quantitated by reaction with a suitable substrate to produce color.

The employment of several serum references of known prostate specific antigen (PSA) levels permits the construction of a dose response curve of activity and concentration. From comparison to the dose response curve, an unknown specimen's activity can be correlated with PSA concentration.

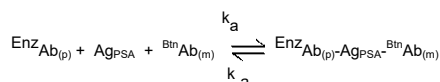
3.0 PRINCIPLE

Immunoassay (TYPE 3):

The essential reagents required for an immunoassay include high affinity and specificity antibodies (enzyme and immobilized), with different and distinct epitope recognition, in excess, and native antigen. In this procedure, the immobilization takes place during the assay at the surface of a microplate well through the interaction of streptavidin coated on

the well and exogenously added biotinylated monoclonal anti-PSA antibody.

Upon mixing monoclonal biotinylated antibody, the enzyme-labeled antibody and a serum containing the native antigen, reaction results between the native antigen and the antibodies, without competition or steric hindrance, to form a soluble sandwich complex. The interaction is illustrated by the following equation:



$\text{B}^{\text{tn}}\text{Ab}_{(m)}$ = Biotinylated Antibody (Excess Quantity)

Ag_{PSA} = Native Antigen (Variable Quantity)

$\text{EnzAb}_{(p)}$ = Enzyme labeled Antibody (Excess Quantity)

$\text{EnzAb}_{(p)}\text{-Ag}_{\text{PSA}}\text{-B}^{\text{tn}}\text{Ab}_{(m)}$ = Antigen-Antibodies Complex

k_a = Rate Constant of Association

k_{-a} = Rate Constant of Dissociation

Simultaneously, the complex is deposited to the well through the high affinity reaction of streptavidin and biotinylated antibody. This interaction is illustrated below:



$\text{Streptavidin}_{\text{C.W.}}$ = Streptavidin immobilized on well

Immobilized complex = complex bound to the solid surface

After equilibrium is attained, the antibody-bound fraction is separated from unbound antigen by decantation or aspiration. The enzyme activity in the antibody-bound fraction is directly proportional to the native antigen concentration. By utilizing several different serum references of known antigen values, a dose response curve can be generated from which the antigen concentration of an unknown can be ascertained.

4.0 REAGENTS

Materials Provided:

A. Prostate Specific antigen (PSA) 1ml/vial – Icons A-F
Six (6) vials of references PSA Antigen at levels of 0(A), 5(B), 10(C), 25(D), 50(E) and 100(F) ng/ml. Store at 2-8°C. A preservative has been added.

Note: The calibrators, human serum based, were calibrated using a reference preparation, which was assayed against the 1st IS 96/670.

B. PSA Enzyme Reagent – 13ml/vial - Icon
One (1) vial containing enzyme labeled antibody, biotinylated monoclonal mouse IgG in buffer, dye, and preservative. Store at 2-8°C.

C. Streptavidin Coated Plate – 96 wells - Icon
One 96-well microplate coated with streptavidin and packaged in an aluminum bag with a drying agent. Store at 2-8°C.

D. Wash Solution Concentrate – 20 ml - Icon
One (1) vial containing a surfactant in buffered saline. A preservative has been added. Store at 2-30°C.

E. Substrate A – 7ml/vial - Icon S^A
One (1) bottle containing tetramethylbenzidine (TMB) in buffer. Store at 2-8°C.

F. Substrate B – 7ml/vial - Icon S^B
One (1) bottle containing hydrogen peroxide (H₂O₂) in buffer. Store at 2-8°C.

G. Stop Solution – 8ml/vial - Icon
One (1) bottle containing a strong acid (1N HCl). Store at 2-30°C.

H. Product Instructions.

Note 1: Do not use reagents beyond the kit expiration date.

Note 2: Avoid extended exposure to heat and light. **Opened reagents are stable for sixty (60) days when stored at 2-8°C. Kit and component stability are identified on the label.**

Note 3: Above reagents are for a single 96-well microplate

4.1 Required But Not Provided:

- Pipette(s) capable of delivering 25 & 50µl volumes with a precision of better than 1.5%.
- Dispenser(s) for repetitive deliveries of 0.100ml and 0.350ml volumes with a precision of better than 1.5%.
- Microplate washers or a squeeze bottle (optional).
- Microplate Reader with 450nm and 620nm wavelength absorbance capability.
- Absorbent Paper for blotting the microplate wells.
- Plastic wrap or microplate cover for incubation steps.
- Vacuum aspirator (optional) for wash steps.
- Timer.
- Quality control materials

5.0 PRECAUTIONS

**For In Vitro Diagnostic Use
Not for Internal or External Use in Humans or Animals**

All products that contain human serum have been found to be non-reactive for Hepatitis B Surface Antigen, HIV 1&2 and HCV Antibodies by FDA licensed reagents. Since no known test can offer complete assurance that infectious agents are absent, all human serum products should be handled as potentially hazardous and capable of transmitting disease. Good laboratory procedures for handling blood products can be found in the Center for Disease Control / National Institute of Health, "Biosafety in Microbiological and Biomedical Laboratories," 2nd Edition, 1988, HHS Publication No. (CDC) 88-8395.

Safe Disposal of kit components must be according to local regulatory and statutory requirement.

6.0 SPECIMEN COLLECTION AND PREPARATION

The specimens shall be blood, serum in type and the usual precautions in the collection of venipuncture samples should be observed. For accurate comparison to established normal values, a fasting morning serum sample should be obtained. The blood should be collected in a plain redtop venipuncture tube without additives or anti-coagulants. Allow the blood to clot. Centrifuge the specimen to separate the serum from the cells. Samples may be refrigerated at 2-8°C for a maximum period of five (5) days. If the specimen(s) cannot be assayed within this time, the sample(s) may be stored at temperatures of -20°C for up to 30 days. Avoid use of contaminated devices. Avoid repetitive freezing and thawing. When assayed in duplicate, 0.050ml of the specimen is required.

7.0 QUALITY CONTROL

Each laboratory should assay controls at levels in the low, normal and elevated range for monitoring assay performance. These controls should be treated as unknowns and values determined in every test procedure performed. Quality control charts should be maintained to follow the performance of the supplied reagents. Pertinent statistical methods should be employed to ascertain trends. Significant deviation from established performance can indicate unnoticed change in experimental conditions or degradation of kit reagents. Fresh reagents should be used to determine the reason for the variations.

8.0 REAGENT PREPARATION:

- Wash Buffer**
Dilute contents of wash concentrate to 1000ml with distilled or deionized water in a suitable storage container. Store at room temperature 20-27°C for up to 60 days.
- Working Substrate Solution**
Pour the contents of the amber vial labeled Solution 'A' into the clear vial labeled Solution 'B'. Place the yellow cap on the clear vial for easy identification. Mix and label accordingly. Store at 2 - 8°C.

Note1 : Do not use the working substrate if it looks blue.

Note 2: Do not use reagents that are contaminated or have bacteria growth.

9.0 TEST PROCEDURE

Before proceeding with the assay, bring all reagents, serum references and controls to room temperature (20 - 27°C).

****Test Procedure should be performed by a skilled individual or trained professional****

- Format the microplates' wells for each serum reference, control and patient specimen to be assayed in duplicate. **Replace any unused microwell strips back into the aluminum bag, seal and store at 2-8°C.**
- Pipette 0.025 ml (25µl) of the appropriate serum reference, control or specimen into the assigned well.
- Add 0.100 ml (100µl) of the PSA Enzyme Reagent to each well. **It is very important to dispense all reagents close to the bottom of the coated well.**
- Swirl the microplate gently for 20-30 seconds to mix and cover.
- Incubate 30 minutes at room temperature.
- Discard the contents of the microplate by decantation or aspiration. If decanting, tap and blot the plate dry with absorbent paper.
- Add 350µl of wash buffer (see Reagent Preparation Section), decant (tap and blot) or aspirate. Repeat two (2) additional times for a total of three (3) washes. **An automatic or manual plate washer can be used. Follow the manufacturer's instruction for proper usage. If a squeeze bottle is employed, fill each well by depressing the container (avoiding air bubbles) to dispense the wash. Decant the wash and repeat two (2) additional times.**
- Add 0.100 ml (100µl) of working substrate solution to all wells (see Reagent Preparation Section). **Always add reagents in the same order to minimize reaction time differences between wells.**
- Incubate at room temperature for fifteen (15) minutes.
- Add 0.050ml (50µl) of stop solution to each well and mix gently for 15-20 seconds. **Always add reagents in the same order to minimize reaction time differences between wells.**
- Read the absorbance in each well at 450nm (using a reference wavelength of 620-630nm to minimize well imperfections) in a microplate reader. **The results should be read within thirty (30) minutes of adding the stop solution.**

10.0 CALCULATION OF RESULTS

A dose response curve is used to ascertain the concentration of PSA in unknown specimens.

- Record the absorbance obtained from the printout of the microplate reader as outlined in Example 1.
- Plot the absorbance for each duplicate serum reference versus the corresponding PSA concentration in ng/ml on linear graph paper (do not average the duplicates of the serum references before plotting).
- Draw the best-fit curve through the plotted points.
- To determine the concentration of PSA for an unknown, locate the average absorbance of the duplicates for each unknown on the vertical axis of the graph, find the intersecting point on the curve, and read the concentration (in ng/ml) from the horizontal axis of the graph (the duplicates of the unknown may be averaged as indicated). In the following example, the average absorbance (1.142) intersects the dose response curve at (23.6 ng/ml) PSA concentration (See Figure 1).

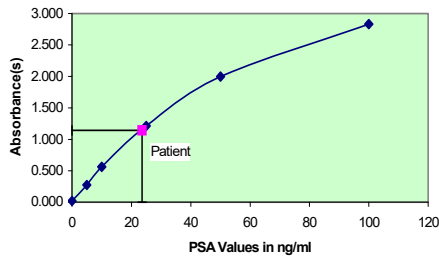
Note: Computer data reduction software designed for ELISA assays may also be used for the data reduction. **If such software is utilized, the validation of the software should be ascertained.**

EXAMPLE 1

Sample I.D.	Well Number	Abs (A)	Mean Abs (B)	Value (ng/ml)
Cal A	A1	0.019	0.019	0
	B1	0.019		
Cal B	C1	0.279	0.276	5
	D1	0.273		
Cal C	E1	0.567	0.563	10
	F1	0.559		
Cal D	G1	1.248	1.213	25
	H1	1.179		
Cal E	A2	2.051	1.999	50
	B2	1.947		
Cal F	C2	2.892	2.833	100
	D2	2.775		
Patient	E3	1.186	1.142	23.6
	F3	1.099		

*The data presented in Example 1 and Figure 1 is for illustration only and **should not** be used in lieu of a dose response curve prepared with each assay.

Figure 1



11.0 Q.C. PARAMETERS

In order for the assay results to be considered valid the following criteria should be met:

1. The absorbance (OD) of calibrator F should be ≥ 1.3 .
2. Four out of six quality control pools should be within the established ranges.

12.0 RISK ANALYSIS

The MSDS and Risk Analysis Form for this product is available on request from Monobind Inc.

12.1 Assay Performance

1. It is important that the time of reaction in each well is held constant to achieve reproducible results.
2. Pipetting of samples should not extend beyond ten (10) minutes to avoid assay drift.

3. Highly lipemic, hemolyzed or grossly contaminated specimen(s) should not be used.
4. If more than one (1) plate is used, it is recommended to repeat the dose response curve.
5. The addition of substrate solution initiates a kinetic reaction, which is terminated by the addition of the stop solution. Therefore, the substrate and stop solution should be added in the same sequence to eliminate any time-deviation during reaction.
6. Plate readers measure vertically. Do not touch the bottom of the wells.
7. Failure to remove adhering solution adequately in the aspiration or decantation wash step(s) may result in poor replication and spurious results.
8. Use components from the same lot. No intermixing of reagents from different batches.
9. Patient specimens with PSA concentrations above 100 ng/ml may be diluted (for example 1/10 or higher) with normal female serum (PSA = 0 ng/ml) and re-assayed. The sample's concentration is obtained by multiplying the result by the dilution factor (10).
10. Accurate and precise pipetting, as well as following the exact time and temperature requirements prescribed are essential. Any deviation from Monobind's IFU may yield inaccurate results.
11. All applicable national standards, regulations and laws, including, but not limited to, good laboratory procedures, must be strictly followed to ensure compliance and proper device usage.
12. It is important to calibrate all the equipment e.g. Pipettes, Readers, Washers and/or the automated instruments used with this device, and to perform routine preventative maintenance.
13. Risk Analysis- as required by CE Mark IVD Directive ISO 14971:2009 - for this and other devices, made by Monobind, can be requested via email from Monobind@monobind.com.

12.2 Interpretation

1. **Measurements and interpretation of results must be performed by a skilled individual or trained professional.**
2. Laboratory results alone are only one aspect for determining patient care and should not be the sole basis for therapy, particularly if the results conflict with other determinants.
3. For valid test results, adequate controls and other parameters must be within the listed ranges and assay requirements.
4. If test kits are altered, such as by mixing parts of different kits, which could produce false test results, or if results are incorrectly interpreted, **Monobind shall have no liability.**
5. If computer controlled data reduction is used to interpret the results of the test, it is imperative that the predicted values for the calibrators fall within 10% of the assigned concentrations.
6. PSA is elevated in benign prostrate hypertrophy (BPH). Clinically an elevated **PSA value alone is not of diagnostic value as a specific test for cancer** and should only be used in conjunction with other clinical manifestations (observations) and diagnostic procedures (prostate biopsy). Free PSA determinations may be helpful in regard to the discrimination of BPH and prostrate cancer conditions (5).

13.0 EXPECTED RANGES OF VALUES

Healthy males are expected to have values below 4 ng/ml (4).

TABLE I
Expected Values for the PSA Elisa Test System

Healthy Males	<4 ng/ml
---------------	----------

It is important to keep in mind that establishment of a range of values which can be expected to be found by a given method for a population of "normal"-persons is dependent upon a multiplicity of factors: the specificity of the method, the population tested and the precision of the method in the hands of the analyst. For these reasons each laboratory should depend upon the range of expected values established by the Manufacturer only until an in-house range can be determined by

the analysts using the method with a population indigenous to the area in which the laboratory is located.

14.0 PERFORMANCE CHARACTERISTICS

14.1 Precision

The within and between assay precisions of the tPSA AccuBind™ ELISA test system were determined by analyses on three different levels of control sera. The number, mean value, standard deviation and coefficient of variation for each of these control sera are presented in Table 2 and Table 3.

TABLE 2
Within Assay Precision (Values in ng/ml)

Sample	N	X	S.D.	C.V.
Level 1	20	0.7	0.05	7.1%
Level 2	20	4.5	0.20	4.4%
Level 3	20	28.3	1.07	3.7%

TABLE 3
Between Assay Precision* (Values in ng/ml)

Sample	N	X	S.D.	C.V.
Level 1	10	0.8	0.09	11.3%
Level 2	10	4.3	0.25	5.8%
Level 3	10	27.5	1.42	5.2%

*As measured in ten experiments in duplicate.

14.2 Sensitivity

The tPSA AccuBind™ ELISA test system has a sensitivity of 0.012 ng. This is equivalent to a sample containing 0.5 ng/ml tPSA concentration.

14.3 Accuracy

The tPSA AccuBind™ ELISA method was compared with a reference Elisa method. Biological specimens from low, normal, and elevated concentrations were assayed. The total number of such specimens was 241. The least square regression equation and the correlation coefficient were computed for the tPSA AccuBind™ ELISA test method in comparison with the reference method. The data obtained is displayed in Table 4.

TABLE 4
Least Square Regression Analysis

Method	Mean	Regression Analysis	Correlation Coefficient
This Method (X)	5.62	$y = -0.0598 + 0.98(X)$	0.987
Reference (Y)	5.57		

Only slight amounts of bias between the tPSA AccuBind™ ELISA method and the reference method are indicated by the closeness of the mean values. The least square regression equation and correlation coefficient indicates excellent method agreement.

No interference was detected with the performance of tPSA AccuBind™ ELISA method and the reference method are indicated by the closeness of the mean values. The least square regression equation and correlation coefficient indicates excellent method agreement.

14.4 Specificity:

No interference was detected with the performance of tPSA AccuBind™ ELISA test system upon addition of massive amounts of the following substances to a human serum pool.

Acetylsalicylic Acid	100 µg/ml
Ascorbic Acid	100 µg/ml
Caffeine	100 µg/ml
CEA	10 µg/ml
AFP	10 µg/ml
CA-125	10,000 U/ml
hCG	1000 IU/ml
hLH	10 IU/ml
hTSH	100 mIU/ml
hPRL	100 µg/ml

15.0 REFERENCES

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6. Prestigiacomo AF, Stamey TA, "Physiological variations of serum prostate antigen in the (4-10 ng/ml) range in male volunteers' *J Urol*, **155**, 1977-80 (1996).
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9. Horton GL, Bahnson RR, Datt M, Cfan KM, Catalona WJ and Landenson JH, "Differences in values obtained with two assays of Prostate Specific Antigen", *J Urol*, **139**, 762-72 (1988).
10. Stenman UH, Leinonen J, Alfthan H, Rannikko S, Tuhkanen K and Alfthan O, "A complex between prostate specific antigen and α_1 -anticymotrypsin is the major form of prostate specific antigen in serum of patients with prostate cancer: assay of complex improves clinical sensitivity for cancer", *Cancer Res*, **51**, 222-26 (1991).

Revision: 3 Date: 081511 DCO: 0524

Cat #: 2125-300

Size	96(A)	192(B)
Reagent (fill)	A)	1ml set
	B)	1 (13ml) 2 (13ml)
	C)	1 plate 2 plates
	D)	1 (20ml) 1 (20ml)
	E)	1 (7ml) 2 (7ml)
	F)	1 (7ml) 2 (7ml)
	G)	1 (8ml) 2 (8ml)

For Orders and Inquiries, please contact



Tel: 949-951-2665 Email: info@monobind.com
 Fax: 949-951-3539 Web: www.monobind.com

Please visit our website to learn more about our other interesting products and services.



Avantor Performance Materials Poland Spółka Akcyjna
Sowińskiego 11
44-101 Gliwice
Tel. 48 32 2392 000

Declaration of conformity

Avantor Performance Materials Poland S.A. who is an established manufacturer of reagents and products for diagnostic in vitro located at:

Sowińskiego 11 Street
44-101, Gliwice
Poland

Herewith declares the following:

Reagents mentioned in attached list are labeled with J.T.Baker label, comply with the In Vitro Diagnostic Medical Devices Directive 98/79/EC and the requirements of ISO 13485 Standard. This declaration is the basic for CE marking of the In Vitro Diagnostic Medical Devices.

The products are not part of List A and List B of Annex II of the IVD Directive 98/79/EC but are subject to self registration.

This declaration is valid for all the IVD medical devices described above and which are placed on the market by ourselves on or after the date hereof and which bear the CE marking

Gliwice, Poland

January 25, 2019

A handwritten signature in blue ink that reads 'Anna Szuba'.

Anna Szuba
Quality Director

J.T.Baker product list for CE marked products

Product	Product number	Pack size
Diluents		
Diluid™ 100 Plus	3961	20 L
Diluid™ 22	2990.9010PC	10 L
Diluid™ 610	3969	20 L
	3969-00	20 L
Diluid™ Abacus	3430.9020	20 L
	3430.9010	10 L
	3430-00	20 L
Diluid™ AC 900	3996	20 L
Diluid™ APR	3476.9020PC	20 L
Diluid™ Azide free	3957	20 L
Diluid™ III Diff	3963	20 L
	3963.9010	10 L
	3963-00	20 L
Diluid™ Erma	3459.9020	20 L
	3459-00	20 L
Diluid™ Mindray	3439.9020PC	20 L
	3439-00	20 L
Diluid™ NR	3483.9020PC	20 L
	3483-00	20 L
Diluid™ Ruby	2987.9020PC	20 L
Diluid™/Sheath 3200-4000	3832.9020	20 L
Diluid™ ST1600/2000	3976	20 L
Sheath D	3495.9010PC	10 L
Sheath Fluid 3000/3500	3471.9020PC	20 L
Lyses		
CN-free Lyse Diff AC 900	3998	5 L
CyMet™ 22 CN Free	2986.0500PE	500 ml
CyMet™ 3000	3469.9010PC	10 L
CyMet™ 3200 CN free	3823.1000	1 L
CyMet™ 3500	3839.5000PC	5 L
CyMet™ 3500 CN free	3825	5 L
CyMet™ 610 CN free	3970	10 L
	3970-00	10 L
	3977	5 L
CyMet™ Abacus CN free	3431.1000	1 L
	3431-00	1 L
CyMet™ APR Baso II	3479.1000PE	1 L
CyMet™ APR CN free	3417.0500PE	500 ml
CyMet™ APR EO	3478.1000PE	1 L
CyMet™ ASA	2950.2500PE	2.5 L
CyMet™ ASB	2951.0500PE	500 ml
CyMet™ AS CN free	2952.9010PC	10 L
CyMet™ BS3 CN free	2982.0500PE	500 ml
CyMet™ III Diff	3968	1 L
	3968-00	500 ml
CyMet™ III Diff CN free	3511.1000	1 L
	3511-00	5 L
CyMet™ Erma	3416-00	500 ml
	3416.0500	500 ml
CyMet™ H20	3853.1000	1 L
CyMet™ KX CN Free	3425-00	500 ml
	3425.0500	500 ml
CyMet™ Micro	3852.1000	1 L
CyMet™ Micro CN free	3863.1000	1 L micros
	3863-00	1 L micros
CyMet™ Mindray	3441-00	500 ml
CyMet™ Mindray CN Free	3440.0500PE	500 ml

J.T.Baker product list for CE marked products

Product	Product number	Pack size
CyMet™ NR III	3484.1000PE	1 L
CyMet™ NR III CN Free	3486-00	1 L
	3486.1000PE	1 L
CyMet™ NR V	3485.1000PE	1 L
CyMet™ Ruby CN Free	2988.5000PC	5 L
CyMet™ ST 1600/2000 CN free	3759.5000	5 L
LeucoLyse	3475.5000PC	5 L
LeucoLyse Ruby	2989.5000PC	5 L
Cleaners		
Blanking Solution 1600/2000	3947	20 L
DetectoTerge™	3763	5 L
	3766	1 L
DetectoTerge™ BS	2970.0900PE	900 ml
ProClean™	3900	5 L
	3900-00	5 L
	3768,1000	1 L micros
ProClean™ Abacus	3432,5000	5 L
	3432.1000PE	1 L
ProClean™ CD	3902.0100PE	100 ml
ProClean™ Extra	3862,5000	5 L
	3862.9020PC	20 L
	3862-00	5 L
	3867-00	1 L micros
	3867.1000PE	1 L micros
ProClean™ Plus	3901	100 ml
Rinse Mindray	3442.5000PE	5 L
Hematology Controls		
8-Parameter Control L/N/H	3427/3428/3429	2.5 ml
	3463/3464/3465	2.5 ml
8-Parameter Control 4xN	3747	4 x 2.5 ml
8-Parameter Control 1xL+4xN+1xH	3751	6 x 2.5 ml
8-Parameter Control extended L/N/H	3633/3634/3635	2.5 ml
3-Diff Control L/N/H	3433/3434/3435	2.5 ml
	3502/3503/3504	4.5 ml
3-Diff Control extended L/N/H	3421/3422/3423	2.5 ml
CD-Diff Control L/N/H	3452/3453/3454	3.0 ml
CD-Diff Control 2xL+2xN+2xH	3838	6 x 3.0 ml
K-Diff Control L/N/H	3455/3456/3457	2.5 ml
Platelet Control- Extended value	3424	5 x 3.0 ml
WBC Reduced RBC L/H	3698/3699	3.0 ml
XE-Diff Control L/N/H	3731/3732/3733	4.5 ml
Fixatives		
Cervix Spray Fixative	3869,1200	12 x 125 ml
10% v/v Buffered Formaldehyde (4% w/v)	3933,1000	1 L
	3933.5000PC	5 L
	3933,9010	10 L
	3933,9020	20 L
	3933.1000MB	1000 L
	3933.9020PE	20 L
	3933.9010JL	10 L
	3933.9020JL	20 L
Clearing agents		
UltraClear™	3905.2500PE	2.5 L
	3905.5000PE	5 L
	3905.9010PE	10 L

J.T.Baker product list for CE marked products

Product	Product number	Pack size
Stains and Dyes		
Eosin-Y Alcoholic	3800.1000PE	1 L
	3800.2500PE	2.5 L
Giemsa	3856,1000	1 L
	3856,2500	2.5 L
	3856.9180ST	180 L
Hematoxylin er (Mayer)	3870,1000	1 L
	3870,2500	2.5 L
Hematoxylin Modified (Harris, Gill II)	3873,1000	1 L
	3873,2500	2.5 L
May-Grünwald	3855,1000	1 L
	3855,2500	2.5 L
Papanicolaou 2A	3554.1000PE	1 L
	3554.2500PE	2.5 L
Papanicolaou 2B	3555.1000PE	1 L
	3555,2500PE	2,5 L
Papanicolaou 3B	3556,1000PE	1 L
	3556.2500PE	2.5 L
Mounting media		
UltraKitt™	3921,0500	500 ml
	3921,0600	6 x 100 ml
	3921,9025ST	25 L
Mounting medium High	3882,0500	500 ml
Mounting medium Low	3883,0500	500 ml
PBS		
PBS	3059	20 L
	3059.9010PC	10 L

Declaration of CE conformity

Avantor Performance Materials B.V. reg. No. 38013066 who is an established manufacturer of Hematology- Reagents, Stains, Controls and Calibrators and products for Histopathology located at:

Teugseweg 20
7418 AM Deventer
the Netherlands

herewith declares the following:

The reagents (see attached list) are labeled with the J.T. Baker label and have the CE mark on the label where applicable. The devices comply with the In Vitro Diagnostic Medical Devices Directive 98/79/EC and the conformity assessment procedure according to Annex III.

The products are not part of List A and List B of Annex II of the IVD Directive 98/79/EC but are subject to self registration.

This declaration is valid for all the IVD medical devices described above and which are placed on the market by ourselves on or after the date hereof and which bear the CE marking.

Deventer, the Netherlands.

22 November 2011



Dr. J. Mittendorf
QA & RA Manager

J.T.Baker product list for CE marked products

Prod.no.	Product	Pack size
Reagents for diluting and lysing		
3961	Diluid™ 100 Plus	20 liter
3954	Diluid 590	20 liter
3969	Diluid 610	20 liter
3430.9010	Diluid Abacus	10 liter
3430.9020	Diluid Abacus	20 liter
3996	Diluid AC 900	20 liter
3996.9010PC	Diluid AC 900	10 liter
3476.9020PC	Diluid APR	20 liter
3957	Diluid Azide free	20 liter
3958	Diluid Azide free	10 liter
3963.9010	Diluid III Diff	10 liter
3963	Diluid III Diff	20 liter
3974	Diluid III Diff Seaccontainer	20 liter
3459.9020	Diluid Erma	20 liter
3483.9020PC	Diluid NR	20 liter
3439.9020PC	Diluid Mindray	20 liter
3832.9020	Diluid Sheath 3200-4000	20 liter
3976	Diluid ST 1600/2000	20 liter
3496.9020PC	Diluid M5	20 liter
3495.9010PC	Sheath D	10 liter
3826	Sheath Fluid 3000/3500	20 liter
3826.5000	Sheath Fluid 3000/3500	5 liter
3827.5000PC	LeucoLyse	5 liter
3998	CN-free Lyse Diff AC 900	5 liter
3744	CyMet™ 1000 CN free	5 liter
3773.5000PC	CyMet 4500 CN free	5 liter
3824	CyMet 3000	10 liter
3823.1000	CyMet 3200 CN free	1 liter
3825	CyMet 3500 CN free	5 liter
3839.5000PC	CyMet 3500	5 liter
3975	CyMet 530+ CN free	10 liter
3971	CyMet 590 CN free	5 liter
3970	CyMet 610 CN free	10 liter
3977	CyMet 610 CN free	5 liter
3918.5000	CyMet 9000 CN free	5 liter
3431.1000	CyMet Abacus CN free	1 liter
3444.1000PE	CyMet Abacus EO	1 liter
3445.1000PE	CyMet Abacus Baso	1 liter
3477.0500PE	CyMet APR CN free	500 ml
3478.1000PE	CyMet APR EO	1 liter
3479.1000PE	CyMet APR Baso II	1 liter
3755	CyMet Automated	5 liter
3757	CyMet Automated	500 ml
3780	CyMet Automated CN Free	1 liter
3460.0500	CyMet Erma	500 ml
3841.1000PE	CyMet H12 CN Free	1 liter
3842.1000	EO Reagent Autocounter	1 liter
3853.1000	CyMet H20	1 liter
3968	CyMet III Diff	1 liter
3964	CyMet III Diff	5 liter
3972.1000	CyMet III Diff CN free	1 liter
3972.5000	CyMet III Diff CN free	5 liter
3740.0500	CyMet KX CN Free	500 ml
3852.1000	CyMet Micro	1 liter
3852.0500	CyMet Micro	500 ml
3857.1000	CyMet Micro CN free	1 liter
3857.0500	CyMet Micro CN free	500 ml

3863.1000	CyMet Micro CN free	1L micros
3440.0500PE	CyMet Mindray CN Free	500 ml
3441.0500PE	CyMet Mindray	500 ml
3480.5000PC	CyMet SF Baso	5L
3481.5000PC	CyMet SF Diff 1	5L
3482.0500PE	CyMet SF Diff 2	500 ml
3775.1000	CyMet ST 1600/2000	1 liter
3759.1000	CyMet ST 1600/2000 CN free	1 liter
3759.5000	CyMet ST 1600/2000 CN free	5 liter
3788	CyMet STX/STL	1 liter
3919	CyMet STX/STL	5 liter
3484.1000PE	CyMet NR III	1 liter
3486.1000PE	CyMet NR III, CN Free	1 liter
3485.1000PE	CyMet NR V	1 liter
3497.0500PE	CyMet MH CN Free	500 ml
3489.1000PE	CyMet MBA	1 liter
3487.1000PE	CyMet MD(I)	1 liter
3488.0500PE	CyMet MD(II)	500 ml
3077	LyzerGlobin™	500 ml
3769	LyzerGlobin	6 x 15 ml
3771	LyzerGlobin PCE	6 x 15 ml
3770	LyzerGlobin II	10 x 10 ml
3850	LyzerGlobin CN free	6 x 15 ml
Cleaners		
3766.0500	DetectoTerge	500 ml
3763	DetectoTerge	5 liter
3766	DetectoTerge	1 liter
3900	ProClean™	5 liter
3768.1000	ProClean	1L micros
3867.1000PE	ProClean Extra	1L micros
3862.1000	ProClean Extra	1 liter
3862.5000	ProClean Extra	5 liter
3901	ProClean Plus	100 ml
3902.0100PE	ProClean CD	100 ml
3432.5000	ProClean Abacus	5 liter
3946	Blanking Solution Hgb	20 liter
3947	Blanking Solution 1600/2000	20 liter
3917	Hypochlorite 0.5%	1liter
3917.5000	Hypochlorite 0.5%	5 liter
3936.1000	Hypochlorite 5%	1liter
3442.5000PE	Rinse Mindray	5 liter
3915	Rinsing Solution Serono 9000	20 liter
3941.1000PE	HypoChlorite NR	1 liter
3941.5000PC	HypoChlorite NR	5 liter
3498.1000PE	ProClean MX5	1 liter
Reagents for 5-part WBC diff. on STKS and MaxM.		
3938	RBCLyse™	1 liter
3938G.1000PE	RBCLyse G	1 liter
3939	WBCStabilise™	500 ml
3492.0090	RetiCount MH	6 x 15 ml
3493.0500PE	RetiClear MHG	500 ml
3493.1000PE	RetiClear MHG	1 liter
3494.0200PE	RetiCount G	200 ml
3774	Reticount™	30 ml
3777	Reticount CD	15 x 3.5 ml

Hematology Controls		
3721/3722/3723	8 PMC Low/Normal/High	8 ml
3724/3725/3726	8 PMC Low/Normal/High	2.5 ml
3633/3634/3635	8 PMC Low/Normal/High ext	2.5 ml
3701/3702/3703	8 PMC Low/Normal/High	4.5 ml
3922/3923/3924	8 PMC L/N/H Swelab	4.5 ml
3746	8 PMC 1 x L, 1 x N, 1 x H	3 x 2.5 ml
3747	8 PMC 4 x Normal	4 x 2.5 ml
3748	8 PMC 4 x Normal	4 x 8 ml
3749	8 PMC 4 x Low	4 x 2.5 ml
3751	8 PMC 1 x L, 4 x N, 1 x H	6 x 2.5 ml
3734/3735/3736	3-Diff Control L/N/H	2.5 ml
3630/3631/3632	3-Diff Control L/N/H ext	2.5 ml
3820/3821/3822	3-Diff Control L/N/H	4.5 ml
3752	3-Diff Control 4 x Low	4 x 2.5 ml
3753	3-Diff Control 4 x Norm	4 x 2.5 ml
3754	3-Diff Control 4 x High	4 x 2.5 ml
3782/3783/3784	CA-Diff Control L/N/H	4.5 ml
3607/3608/3609	CA-Diff Control L/N/H	2.5 ml
3610/3611/3612	DIA Diff 5 Control L/N/H	4.5 ml
3731/3732/3733	XE-Diff Control L/N/H	4.5 ml
3693/3694/3695	SF-Diff Control L/N/H	4.5 ml
3613/3614/3615	BC Diff 5 Control L/N/H	4.5 ml

3684/3685/3686	ADV-Diff Control L/N/H	3.5 ml
3690/3691/3692	ADV Retic 1/2/3	4.0 ml
3828/3829/3830	CD-Diff Control	3.0 ml
3838	CD-Diff Control 2x L _N ,H	6 x 3.0 ml
3687/3688	CD 4K Retic 1/2	3.0 ml
3892/3893/3894	AC-Diff Control	2.5 ml
3896/3897/3898	K-Diff Control	2.5 ml
3696/3697	WBC reduced Plt Control L/H	3.0 ml
3698/3699	WBC reduced RBC Control L/H	3.0 ml
Laser controls for Coulter MaxM, GenS and STKS		
3681/3682/3683	5D Control Low /N /H	5.0 ml
Calibration Set for Cell Analysers.		
3940	Cal Set 1	2 x 2.5 ml
3720	Platelet Control Ext. value	5 x 3 ml
Phosphate Buffered Saline.		
3059	PBS, diluting fluid for bloodgrouping	20 liter
3059.9010PC	PBS, diluting fluid for bloodgrouping	10 liter

Number	Product	Content
Stains and Dyes		
3554.1000PE	Papanicolaou Solution 2A	1 liter
3554.2500PE	Papanicolaou Solution 2A	2.5 liter
3554.9200PE	Papanicolaou Solution 2A	200 liter
3555.1000PE	Papanicolaou Solution 2B	1 liter
3555.2500PE	Papanicolaou Solution 2B	2.5 liter
3556.1000PE	Papanicolaou Solution 3B	1 liter
3556.2500PE	Papanicolaou Solution 3B	2.5 liter
3556.9200PE	Papanicolaou Solution 3B	200 liter
3800.1000PE	Eosine-Y Alcoholic	1 liter
3800.2500PE	Eosine-Y Alcoholic	2.5 liter
3801.1000PE	Eosin Y 0.5% Aqueous	1 liter
3801.2500PE	Eosin Y 0.5% Aqueous	2.5 liter
3871.1000	Eosine Solution 0.2% ready to use	1 liter
3871.2500	Eosine Solution 0.2% ready to use	2.5 liter
3856.0100	Giemsa	0.1 liter
3856.0500	Giemsa	0.5 liter
3856.1000	Giemsa	1 liter
3856.2500	Giemsa	2.5 liter
3870.1000	Hematoxyline er (Mayer)	1 liter
3870.2500	Hematoxyline er (Mayer)	2.5 liter
3873.1000	Hematoxyline (Harris, Gill II)	1 liter
3873.2500	Hematoxyline (Harris, Gill II)	2.5 liter
3879.1000	Leishman	1 liter
3855.0500	May Grünwald	0.5 liter
3855.1000	May Grünwald	1 liter
3855.2500	May Grünwald	2.5 liter

3864.1000	Papanicolaou 2A OG6	1 liter
3864.2500	Papanicolaou 2A OG6	2.5 liter
3865.1000	Papanicolaou 2B Orange II	1 liter
3865.2500	Papanicolaou 2B Orange II	2.5 liter
3866.1000	Papanicolaou 3B EA 50	1 liter
3866.2500	Papanicolaou 3B EA 50	2.5 liter
3876.1000	Shorr	1 liter
3878.1000	Wright	1 liter
Clearing agent		
3905.2500PE	UltraClear	2.5 liter
3905.5000PE	UltraClear	5 liter
3905.9010PE	UltraClear	10 liter
3905.9200	UltraClear	200 liter
Mounting media		
3921.0500	UltraKitt	500 ml
3921.0600	UltraKitt	6 x 100 ml
Fixatives		
3933.1000	10% v/v Buffered Formaldehyde	1 liter
3933.5000PC	10% v/v Buffered Formaldehyde	5 liter
3933.9010 (PE)	10% v/v Buffered Formaldehyde	10 liter (PE)
3933.9020 (PE)	10% v/v Buffered Formaldehyde	20 liter (PE)
3869.1200	Cervix Fixative	12 x 125 ml
3880.1000	Bouin's Fixative	1 liter
3058.9010	Immuno PBS 20x concentrated	10 liter

22 November 2011

To whom this may concern

Date: March 01, 2021

Letter of Authorization

Avantor Performance Materials Poland S.A., reg. No. 0000010108 who is an established manufacturer of Hematology- Reagents, Stains, Controls and Calibrators and products for Histology located at:

Sowińskiego 11
44-101 Gliwice
Poland

herewith confirms that:

I.M Global Biomarketing Group Moldova S.R.L
Republic of Moldova
MD-2001, Chisinau
Tighina str. 65, 607 office
Tel (373 22) 549 120, 549 121
Fax (373 22) 547 373

is authorized to act as our distributor for our hematology/histology reagents and controls (Products) in Moldova

We declare that we will supply the Products for the needs of tenders.

We declare that we will supply the Products for tenders with warranty as per the Avantor General Conditions of Sale.

Furthermore I.M Global Biomarketing Group is duly entitled to:

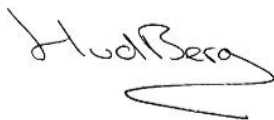
- Register, promote, offer, negotiate prices and sell our Products in Moldova;
- carry out the required product training of the medical and technical personnel who will use these products.

The product specialists of I.M Global Biomarketing Group have been duly trained and are qualified for providing all services in regards to consulting, sales, maintenance and training.

In all the above activities I.M Global Biomarketing Group is acting in its own name and on its own account.

This authorization letter is valid until about 1 year after date.

Avantor Performance Materials S.A.
Poland



H van den Berg,
Marketing Product Manager Diagnostics



Certificate of Completion

This is to certify

Mr. Alexei Legun

Has successfully completed

The technical maintenance training course

On

Fully Automatic Blood Cell Counter

PCE-210

Particle(Blood Cell)Counter

PCE-170/PCE-170N

Hemoglobin meter

Hb-20N

March 24, 2005

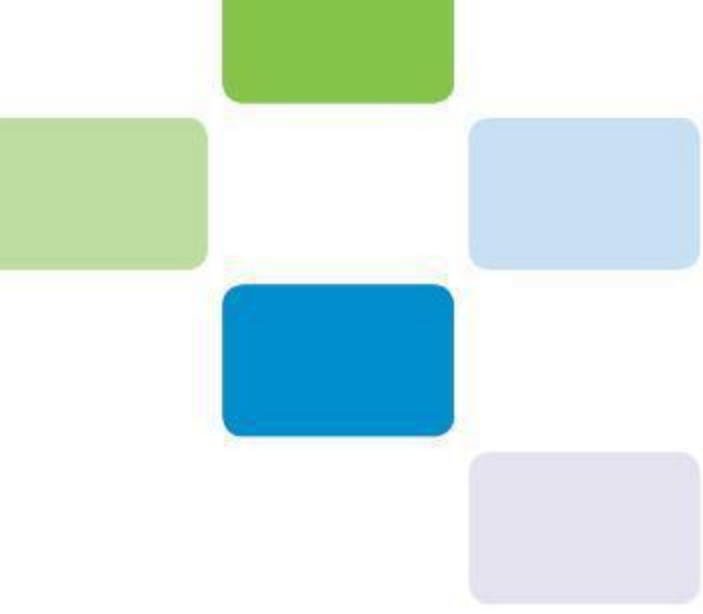
H. Shimosaka

Hiroshi Shimosaka

President

ERMA INC.





BeneSphera™
3 PART
DIFFERENTIAL
Hematology Analyzer

 **BeneSphera™ TRAINING**

Mr /-Ms Sergiu Sorocovici
Global Biomarketing Group
str. Tighina 65, of. 607
2001 Chisinau, Moldau

has attended a 2-days training on goods manufactured or distributed by us.
April 12th – April 13th, 2012

Deventer, The Netherlands
Place, Date 13.04.2012

H. J. Daas




CERTIFICATO N° 505DM07

CERTIFICATE N° 505DM07

Si certifica che il
this is to certify that

Sistema di Gestione per la Qualità

Quality Management System

messo in atto da
implemented by

APTACA S.p.A.

Via Monte Bianco, 4 – IT 20900 MONZA (MB)

nella Sede Operativa di
Operative Unit

Regione Monforte, 30 – IT 14053 CANELLI (AT)

è conforme alla norma
is in compliance with the standard

UNI CEI EN ISO 13485-2016 (ISO 13485-2016)

per i seguenti Processi
concerning the following kinds of Processes

Gestione della fabbricazione e immissione in commercio di tamponi sterili per il prelievo di campioni biologici in orifizi naturali e in ambito chirurgico.

Progettazione e fabbricazione di dispositivi medico diagnostici per laboratori di analisi.


Gestione della fabbricazione ed immissione in commercio di dispositivi medici invasivi in relazione agli orifizi del corpo in Classe I Sterile. Fabbricazione di dispositivi medici invasivi in relazione agli orifizi del corpo in Classe I Sterile. Commercializzazione di dispositivi medici e diagnostici in vitro.

Management of the manufacturing and placing on the market of sterile tampons for sampling of biological specimens in natural orifice and in surgical field. Design and manufacturing of diagnostic medical devices for laboratories of analysis. Management of the manufacturing and placing on the market of invasive medical devices with respect to body orifices (class I sterile). Manufacturing of invasive medical devices with respect to body orifices (class I sterile). Marketing of medical and diagnostic devices in vitro.

Il presente Certificato è soggetto al rispetto delle condizioni stabilite dai Regolamenti per la certificazione in vigore applicabili.
This Certificate shall satisfy the requirements established in the Rules for the certification in force applicable.

In caso di discordanza tra le lingue utilizzate nella traduzione del contenuto del presente certificato, fare riferimento alla lingua italiana
In cases of discrepancy between the languages used in the translation of the content of this certificate, please refer to the Italian language

L'AMMINISTRATORE DELEGATO
MANAGING DIRECTOR


Dr. Ing. Roberto Cusolito

Data di Prima Emissione
First Issue Date
2007-10-30

Data di Prima Emissione ITALCERT
First Issue Date ITALCERT
2011-10-30

Data di Rinnovo
Renewal Date
2020-10-30

Data di Scadenza
Expiration Date
2023-10-29



SGQ N° 023A

Membro degli Accordi di Mutuo Riconoscimento EA, IAF e ILAC
Signatory of EA, IAF and ILAC Mutual Recognition Agreements

CERTIFICATO N° 505SGQ05

CERTIFICATE N° 505SGQ05

Si certifica che il
this is to certify that

Sistema di Gestione per la Qualità

Quality Management System

messo in atto da
implemented by

APTACA S.p.A.

Via Monte Bianco, 4 – IT 20900 MONZA (MB)

nella Sede Operativa di
Operative Unit

Regione Monforte, 30 – IT 14053 CANELLI (AT)

è conforme alla norma
is in compliance with the standard

UNI EN ISO 9001-2015 (ISO 9001-2015)

per i seguenti Processi
concerning the following kinds of Processes

Gestione della fabbricazione ed immissione in commercio di tamponi sterili per il prelievo di campioni biologici in orifizi naturali e in ambito chirurgico. Progettazione e fabbricazione di dispositivi medico diagnostici per laboratori di analisi. Gestione della fabbricazione ed immissione in commercio di dispositivi medici invasivi in relazione agli orifizi del corpo in Classe I Sterile. Fabbricazione di dispositivi medici invasivi in relazione agli orifizi del corpo in Classe I Sterile. Commercializzazione di dispositivi medici e diagnostici in vitro.


Commercializzazione di articoli da laboratorio

Management of the manufacturing and placing on the market of sterile tampons for sampling of biological specimens in natural orifice and in surgical field. Design and manufacturing of diagnostic medical devices for laboratories of analysis. Management of the manufacturing and placing on the market of invasive medical devices with respect to body orifices (class I sterile). Manufacturing of invasive medical devices with respect to body orifices (class I sterile). Marketing of medical and diagnostic devices in vitro. Marketing of laboratory articles.

Il presente Certificato è soggetto al rispetto delle condizioni stabilite dai Regolamenti per la certificazione in vigore applicabili.
This Certificate shall satisfy the requirements established in the Rules for the certification in force applicable.

In caso di discordanza tra le lingue utilizzate nella traduzione del contenuto del presente certificato, fare riferimento alla lingua italiana
In cases of discrepancy between the languages used in the translation of the content of this certificate, please refer to the Italian language.

L'AMMINISTRATORE DELEGATO
MANAGING DIRECTOR



Dr. Ing. Roberto Cusolito

Data di Prima Emissione
First Issue Date

1998-07-23

Data di Prima Emissione ITALCERT
First Issue Date ITALCERT

2011-10-30

Data di Rinnovo
Renewal Date

2020-10-30

Data di Scadenza
Expiration Date

2023-10-29

Settore IAF 14 - 29



SGQ N° 023A

Membro degli Accordi di Mutuo Riconoscimento EA, IAF e ILAC
Signatory of EA, IAF and ILAC Mutual Recognition Agreements

DICHIARAZIONE DI CONFORMITÀ CE
EC DECLARATION OF CONFORMITY

conforme all'Allegato III della Direttiva 98/79/CE "Dispositivi Medico-Diagnostici In Vitro" e s.m.i.
according to Annex III of the Directive 98/79/EC on "In Vitro Diagnostic Medical Devices" as amended

fabbricante **VACUTEST KIMA S.r.l. - articoli per laboratori analisi**
manufacturer **disposable labware**

indirizzo **Via dell'Industria, 12**
address **35020 Arzergrande (PD) - Italia**

telefono **+39-049-9720624** fax **+39-049-9720182** posta elettronica **info@vacutestkima.it**
phone e-mail

identificazione dei prodotti **Sistema di prelievo di sangue e altri liquidi biologici**
product identification **mediante provette con vuoto predeterminato in plastica**
"VACUTEST KIMA".

"VACUTEST KIMA" vacuum blood and biological liquids
collection tubes in plastic.

nome commerciale **"VACUTEST KIMA"**
brand name

classificazione dei prodotti **dispositivi diversi da quelli elencati nell'Allegato II della Direttiva 98/79/CE e s.m.i.**
product classification **devices other than those mentioned in Annex II of the Directive 98/79/EC as amended**

Si dichiara

sotto la propria responsabilità che tutti i dispositivi sopraelencati rispettano le disposizioni applicabili della Direttiva 98/79/CE e s.m.i. "Dispositivi Medico-Diagnostici In Vitro".

Tutta la documentazione tecnica richiesta dall'Allegato III della succitata Direttiva e comprovante il rispetto dei Requisiti Essenziali di cui all'Allegato I della Direttiva, è conservata a cura del Fabbricante

Hereby we declare

under our sole responsibility that the above mentioned devices meet the applicable provisions of the Directive 98/79/EC as amended on "In Vitro Diagnostic Medical Devices".


All the supporting documents, as required by Annex III, in order to prove conformity to the Essential Requirements as listed in Annex I, are retained under the premises of the Manufacturer

luogo e data
place and date

Arzergrande, 01/01/2015

firma
signature

Assicuratore Qualità / Quality Manager
Giovanni Chiarin





IQNet, the association of the world's first class certification bodies, is the largest provider of management System Certification in the world.
IQNet is composed of more than 30 bodies and counts over 150 subsidiaries all over the globe.

CERTIFICATO n. **4265/5**
CERTIFICATE No. _____

SI CERTIFICA CHE IL SISTEMA DI GESTIONE PER LA QUALITÀ DI
WE HEREBY CERTIFY THAT THE QUALITY MANAGEMENT SYSTEM OPERATED BY

GRUPPO VACUTEST KIMA

Sede / Head Office

Via dell'Industria, 12 – 35020 Arzergrande (PD) - Italia

Unità Operative / Operative Units

MEUS S.r.l. - Via Leonardo da Vinci, 24B – 26 – 28 – Zona Industriale Tognana – 35028 Piove di Sacco (PD) - Italia

MEUS S.r.l. - Via dell'Industria, 2 - 16 – 35020 Arzergrande (PD) - Italia

ROLL S.R.L. - Via Leonardo Da Vinci, 24A – Zona Industriale Tognana – 35028 Piove di Sacco (PD) - Italia

KIMA S.R.L. - Via Leonardo da Vinci, 22 – Zona Industriale Tognana – 35028 Piove di Sacco (PD) - Italia

VACUTEST KIMA S.r.l. - Via dell'Industria, 12 – 35020 Arzergrande (PD) – Italia

VACUTEST KIMA S.r.l. via L. Da Vinci, 22 Zona Industriale Tognana – 35028 Piove di Sacco (PD) - Italia

È CONFORME ALLA NORMA / IS IN COMPLIANCE WITH THE STANDARD

UNI CEI EN ISO 13485:2016

Sistema di Gestione per la Qualità / Quality Management System

PER LE SEGUENTI ATTIVITÀ / FOR THE FOLLOWING ACTIVITIES

Progettazione e produzione di provette con vuoto predeterminato ad uso prelievo ematico, liquidi biologici e urine.
Produzione di provette per microprelievi di sangue. Progettazione e produzione di Holders (camicie) per prelievo sottovuoto. Progettazione e produzione di kit diagnostici per l'analisi del sangue e dei liquidi biologici. Progettazione e produzione di terreni di coltura per microbiologia. Progettazione e produzione di aghi e dispositivi sterili per il prelievo ematico. Commercializzazione di prodotti del Gruppo: kit diagnostici, terreni di coltura per microbiologia, articoli in plastica per laboratorio analisi, provette con vuoto predeterminato e aghi sterili. Progettazione e produzione di stampi per articoli in plastica per laboratorio analisi. Stampaggio di materie termoplastiche ad iniezione per articoli medicali.

Design and production of test tubes with predetermined vacuum for collection of haematological samples, biological liquids and urine samples. Production of test tubes for micro-collection of haematological samples. Design and production of Holders for vacuum sampling. Design and production of diagnostic kits for blood and biological liquids analysis. Design and production of culture media for microbiology. Design and production of sterile needles and devices for collection of haematological samples. Trading of the products of the Group: diagnostic kits, culture media for microbiology, plastic disposable labware, test tubes with predetermined vacuum and sterile needles. Design and production of moulds for plastic labware. Injection moulding of thermoplastic materials for medical devices.

Riferirsi alla documentazione del Sistema di Gestione per la Qualità aziendale per l'applicabilità dei requisiti della norma di riferimento.

Refer to the documentation of the Quality Management System for details of application to reference standard requirements.

Il presente certificato è soggetto al rispetto del documento ICIM "Regolamento per la certificazione dei sistemi di gestione" e al relativo Schema specifico.

The use and the validity of this certificate shall satisfy the requirements of the ICIM document "Rules for the certification of company management systems" and Specific Scheme.

Per informazioni puntuali e aggiornate circa eventuali variazioni intervenute nello stato della certificazione di cui al presente certificato,

si prega di contattare il n° telefonico +39 02 725341 o indirizzo e-mail info@icim.it.

For timely and updated information about any changes in the certification status referred to in this certificate,

please contact the number +39 02 725341 or email address info@icim.it.

DATA EMISSIONE
FIRST ISSUE
18/01/2007

EMISSIONE CORRENTE
CURRENT ISSUE
18/01/2022

DATA DI SCADENZA
EXPIRING DATE
17/01/2025

Vincenzo Delacqua
Rappresentante Direzione / Management Representative
ICIM S.p.A.

Piazza Don Enrico Mapelli, 75 – 20099 Sesto San Giovanni (MI)
www.icim.it



SGQ N° 004 A



www.cisq.com

CISQ è la Federazione Italiana di Organismi di Certificazione dei sistemi di gestione aziendali.
CISQ is the Italian Federation of management system Certification Bodies.

Zertifikat

**Qualitätsmanagementsystem
EN ISO 13485:2016**

Registrier-Nr.: SX 1614112-1

Organisation: KABE-Labortechnik GmbH
Jägerhofstr. 17
51588 Nümbrecht
Deutschland

Geltungsbereich: Entwicklung, Herstellung und Vertrieb von In-vitro-Diagnostik-Produkten und Verbrauchsmaterialien für die Probengewinnung, -vorbereitung und -aufbewahrung sowie von Medizinprodukten zur einmaligen Anwendung

Die Zertifizierungsstelle der TÜV Rheinland LGA Products GmbH bescheinigt, dass die Organisation ein Qualitätsmanagementsystem für Medizinprodukte eingeführt hat und anwendet. Der Nachweis wurde erbracht, dass die Forderungen der oben genannten Norm erfüllt sind. Das Qualitätsmanagementsystem unterliegt einer jährlichen Überwachung.

Bericht Nr.: 1092786-40
Gültig ab: 25.10.2021
Gültig bis: 15.10.2024
Datum: 25.10.2021



Dipl.-Ing. F. Schwingen

TÜV Rheinland LGA Products GmbH
Tillystraße 2 · 90431 Nürnberg · Deutschland



Deutsche
Akkreditierungsstelle
D-ZM-14169-01-02

Zertifikat



**Qualitätsmanagementsystem
EN ISO 13485:2016**

Registrier-Nr.: SX 1614112-1
Organisation: KABE-Labortechnik GmbH
Jägerhofstr. 17
51588 Nümbrecht
Deutschland

Der Geltungsbereich beinhaltet folgende zusätzlichen Standorte:

Nr.	Standorte	Geltungsbereich
/01	c/o KABE-Labortechnik GmbH Jägerhofstr. 17 51588 Nümbrecht Deutschland	Entwicklung, Herstellung und Vertrieb von In-vitro-Diagnostik-Produkten und Verbrauchsmaterialien für die Probengewinnung, -vorbereitung und -aufbewahrung sowie von Medizinprodukten zur einmaligen Anwendung
/02	c/o KABE-Labortechnik GmbH Werner-von-Siemens-Str. 1 51674 Wiehl Deutschland	Lager

Bericht Nr.: 1092786-40
Gültig ab: 25.10.2021
Gültig bis: 15.10.2024
Datum: 25.10.2021



Dipl.-Ing. F. Schwingen
TÜVRheinland LGA Products GmbH
Tillystraße 2 | 90431 Nürnberg · Deutschland

EC Certificate
Directive 93/42/EEC Annex II, excluding Section 4
Full Quality Assurance System
Medical Devices

Registration No.: HD 60150763 0001

Report No.: 21234760 013

Manufacturer: KABE LABORTECHNIK GmbH
Jägerhofstr. 17
51588 Nümbrecht
Deutschland

Products:

- Cannulas for blood collection
- MBU Capillaries

(see attachment for details)

Replaces certificate, Registration No.: HD 60105393 0001

Expiry Date: 2024-05-26

The Notified Body hereby declares that the requirements of Annex II, excluding section 4 of the directive 93/42/EEC have been met for the listed products. The above named manufacturer has established and applies a quality assurance system, which is subject to periodic surveillance, defined by Annex II, section 5 of the aforementioned directive. For placing on the market of class III devices covered by this certificate an EC design-examination certificate according to Annex II, section 4 is required.

Effective Date: 2020-10-07

Date: 2020-10-07

Notified Body


Dr. K. Kluge



TÜV Rheinland LGA Products GmbH - Tillystraße 2 - 90431 Nürnberg
TÜV Rheinland LGA Products GmbH is a Notified Body according to Directive 93/42/EEC concerning medical devices with the identification number 0197.

**TÜV Rheinland
LGA Products GmbH
Tillystraße 2, 90431 Nürnberg**

**Attachment to
Certificate**

Registration No.: HD 60150763 0001
Report No.: 21234760 013

Manufacturer: KABE LABORTECHNIK GmbH
Jägerhofstr. 17
51588 Nümbrecht
Deutschland

Products included:

- Cannulas for blood collection

For the following devices the scope covers only
the aspects of the manufacture concerned with
the securing and maintaining sterile conditions:

- MBU Capillaries

Date: 2020-10-07

Notified Body

Dr. K. Kluge
Dr. K. Kluge



EC Certificate

**Full Quality Assurance System
Directive 98/79/EC on In Vitro Diagnostic Medical Devices,
Annex IV excluding (4, 6)**

Registration No.: HL 1038121-1

Manufacturer: MACHEREY-NAGEL GmbH & Co. KG
Valenciener Str. 11
52355 Düren
Germany

Products: Products for self-testing
- Single and multi-parameter disposable test strips for urine analysis
- Indicator test strips and papers for measurement of pH in urine

Replaces Certificate, Registration No.: HL 60119814 0001

The Notified Body hereby declares that the requirements of Annex IV, excluding sections 4 and 6 of the directive 98/79/EC have been met for the listed products. The above named manufacturer has established and applies a quality assurance system, which is subject to periodic surveillance, defined by Annex IV, section 5 of the aforementioned directive. For placing on the market of List A devices covered by this certificate an EC design-examination certificate according to Annex IV, section 4 and a verification of manufactured products according to section 6 is required.

Report No.: 1106581-20

Effective date: 2022-02-16

Expiry date: 2025-05-26

Issue date: 2022-02-16



Dipl.-Ing. Sven Hoffmann
TÜV Rheinland LGA Products GmbH
Tillystraße 2 · 90431 Nürnberg · Germany

TÜV Rheinland LGA Products GmbH is a Notified Body according to Directive 98/79/EC concerning in vitro diagnostic medical devices with the identification number 0197.

EC Certificate

**Full Quality Assurance System
Directive 98/79/EC on In Vitro Diagnostic Medical Devices,
Annex IV excluding (4, 6)**

Registration No.: HL 1038121-1

Manufacturer: MACHEREY-NAGEL GmbH & Co. KG
Valenciener Str. 11
52355 Düren
Germany

The scope of certification includes the following manufacturing sites:

No.	Location	Product groups manufactured
/01	MACHEREY-NAGEL GmbH & Co. KG Valenciener Str. 11 52355 Düren Germany	Design and development, manufacture and quality control
/02	MACHEREY-NAGEL GmbH & Co. KG Bahnstr. 120 52355 Düren Germany	Warehousing and logistics

Report No.: 1106581-20

Effective date: 2022-02-16

Expiry date: 2025-05-26

Issue date: 2022-02-16



TÜV Rheinland LGA Products GmbH
TÜVRheinland®
Zertifizierungsstelle

Dipl.-Ing. Sven Hoffmann
TÜV Rheinland LGA Products GmbH
Tillystraße 2 · 90431 Nürnberg · Germany

TÜV Rheinland LGA Products GmbH is a Notified Body according to Directive 98/79/EC concerning in vitro diagnostic medical devices with the identification number 0197.

Certificate ES10/81671

SGS

The management system of

DELTALAB, S.L.

Polígono Industrial La Llana, Plaza de la Verneda 1, 08191 Rubi, Barcelona, Spain

has been assessed and certified as meeting the requirements of

ISO 13485:2016

EN ISO 13485:2016

For the following activities

Design, manufacture and sale of sterile and non-sterile medical devices for the collection, transport and conservation of biological samples for clinical and IVD analysis.

Distribution of non-active medical devices and in vitro diagnostic medical devices.

Diseño, fabricación y comercialización de productos sanitarios estériles y no estériles para la toma, transporte y conservación de muestras biológicas para análisis clínicos y de IVD.

Distribución de productos sanitarios no activos y productos sanitarios para diagnóstico in vitro.

Disseny, fabricació i comercialització de productes sanitaris estèrils i no estèrils per a la presa, transport i conservació de mostres biològiques per a anàlisis clíniques i de IVD.

Distribució de productes sanitaris no actius i productes sanitaris per a diagnòstic in vitro.

This certificate is valid from 12 October 2022 until 11 October 2025 and remains valid subject to satisfactory surveillance audits.

Issue 10. Certified since 12 October 2010.

Jonathan M. Hall

Jonathan Hall
Global Head - Certification Services

SGS United Kingdom Ltd
Rossmore Business Park, Ellesmere Port, Cheshire, CH65 3EN, UK
t +44 (0)151 350-6666 - www.sgs.com



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Система Сертификации

Продукции, Работ и Услуг, Систем Менеджмента

ИнтерСертТест

**ОРГАН ПО СЕРТИФИКАЦИИ
ОБЩЕСТВО С ОГРАНИЧЕННОЙ ОТВЕТСТВЕННОСТЬЮ
«ИСО КОНСАЛТИНГ»**

121352, г. Москва, ул. Давыдовская, дом 3, этаж 2, блок 2, пом. 126, 127, 128, 129

СЕРТИФИКАТ СООТВЕТСТВИЯ

Выпуск 1. СМК сертифицирована с января 2021 года

№ РОСС RU.С.04ША.СК.1558

Выдан: Обществу с ограниченной ответственностью

Научно-производственная фирма «ВИНАР»

(ООО «НПФ «ВИНАР»)

ИНН 5023001024

105094, г. Москва, Госпитальный вал, д.5, стр.7А, пом. VIII

НАСТОЯЩИЙ СЕРТИФИКАТ УДОСТОВЕРЯЕТ:

*СИСТЕМА МЕНЕДЖМЕНТА КАЧЕСТВА ПРИМЕНИТЕЛЬНО К РАЗРАБОТКЕ, ПРОИЗВОДСТВУ И РЕАЛИЗАЦИИ ПРОДУКЦИИ;
ХИМИЧЕСКИХ И БИОЛОГИЧЕСКИХ ИНДИКАТОРОВ КОНТРОЛЯ СТЕРИЛИЗАЦИИ, ДЕЗИНФЕКЦИИ И ОБЕЗЗАРАЖИВАНИЯ; УСТРОЙСТВ
КОНТРОЛЯ ПРОЦЕССА СТЕРИЛИЗАЦИИ; ИНДИКАТОРОВ ЭКСПРЕСС-КОНТРОЛЯ КОНЦЕНТРАЦИЙ РАБОЧИХ РАСТВОРОВ
ДЕЗИНФИЦИРУЮЩИХ И СТЕРИЛИЗУЮЩИХ СРЕДСТВ; ИНДИКАТОРОВ КОНТРОЛЯ ЭФФЕКТИВНОСТИ ПРЕДСТЕРИЛИЗАЦИОННОЙ
ОЧИСТКИ МЕДИЦИНСКИХ ИНСТРУМЕНТОВ; УПАКОВОЧНЫХ МАТЕРИАЛОВ ДЛЯ ФИПИШНОЙ СТЕРИЛИЗАЦИИ И СТИРКИ;
ИНДИКАТОРОВ КОНТРОЛЯ ХОЛОДОВОЙ ЦЕПИ; РАСХОДНЫХ МАТЕРИАЛОВ ДЛЯ СТЕРИЛИЗАЦИОННЫХ, ОПЕРАЦИОННЫХ, ЧИСТЫХ
ПОМЕЩЕНИЙ; АНТИСЕПТИЧЕСКИХ И ДЕЗИНФИЦИРУЮЩИХ СРЕДСТВ*

СООТВЕТСТВУЕТ ТРЕБОВАНИЯМ ГОСТ ISO 13485-2017 (ISO 13485:2016)

Основание: Решение Органа по сертификации № 1558 от 22 января 2021 года

НАСТОЯЩИЙ СЕРТИФИКАТ ОБЯЗЫВАЕТ ОРГАНИЗАЦИЮ ПОДДЕРЖИВАТЬ СОСТОЯНИЕ СИСТЕМЫ МЕНЕДЖМЕНТА КАЧЕСТВА В РАБОТОСПОСОБНОМ СОСТОЯНИИ В СООТВЕТСТВИИ С ТРЕБОВАНИЯМИ ВЫПУСКАЕМОГО СТАНДАРТА, ПОДТВЕРЖДАТЬ ЭТО СООТВЕТСТВИЕ РЕЗУЛЬТАТАМИ ПРОХОЖДЕНИЯ ЕЖЕГОДНОГО ИНСПЕКЦИОННОГО КОНТРОЛЯ В ОС ООО «ИСО КОНСАЛТИНГ» ВО ВРЕМЯ ВСЕГО СРОКА ДЕЙСТВИЯ СЕРТИФИКАТА.

Дата выдачи: 25.01.2021

Срок действия до: 25.01.2024

(при прохождении инспекционного контроля)

Срок прохождения первого инспекционного контроля: не позднее 25.01.2022

Срок прохождения второго инспекционного контроля: не позднее 25.01.2023



Т.В. ГРИЧАНАЯ

Заместитель Руководителя Органа

С.Т. БУТКИНА

Эксперт

№ 005153