



130201035M:100 tests/kit 130601035M: 50 tests/kit

MAGLUMI® Free PSA (CLIA)

INTENDED USE

The kit is an *in vitro* chemiluminescence immunoassay for the quantitative determination of Free PSA (fPSA) in human serum and plasma using the MAGLUMI series Fully-auto chemiluminescence immunoassay analyzer and Biolumi series Integrated System, and the assay is used for an aid in distinguishing prostate cancer from benion prostatic conditions with MAGLUMI Total PSA value in the range 4.00 no/mL to 10.0 no/mL to

SUMMARY

Prostate specific antigen (PSA) is an androgen-regulated serine protease and member of the tissue kallikrein family of proteases¹⁻³. It is produced primarily by prostate ductal and acinar epithelium and is secreted into the lumen, where its function is to cleave semenogelin I and II in the seminal coagulum, resulting in increased sperm motility¹⁻³. PSA can be detected in normal prostate epithelium, benign prostatic hypertrophy (BPH), malignant prostate epithelium, prostatic fluid and serum⁴. It is not detectable in significant quantities in other human tissues⁴.

There are 3 major molecular forms of serum PSA. PSA-ACT, PSA complex with α-1-antichymotrypsin (ACT); PSA-AMG, PSA complex with α-2-macroglobulin (AMG); and free PSA uncomplexed to serum protease binding proteins⁵. ACT and AMG, occur at large molar excess to PSA in the extracellular fluids and possess the major capacity to regulate the PSA activity in blood in vitro by the formation of covalent complexes with the active single-chain form of PSA⁶. In the formation of sodium dodecyl sulphate-stable complexes with AMG, no PSA epitopes remain exposed on the PSA-AMG complexes unless denaturing conditions are used to disrupt their conformation⁶. Therefore, presently available immunoassays for PSA fail to recognize and correctly measure the concentration of PSA-AMG complexes in serum at nondenaturing conditions⁶.

Prostate cancer is the most common visceral cancer and the second-leading cause of death from cancer in men in the United States*. The high mortality rate may be due to its frequent detection in advanced stages since the cancer often remains asymptomatic until it has spread beyond the organ*. The findings of investigation indicate that the serum PSA concentration is directly correlated with patient age, and this phenomenon is primarily due to the increase in prostatic volume with advancing age8.9 BPH tissue contributes to the serum PSA concentration and has a high prevalence in men more than 50 years old*. Because approximately 25% of the patients with BPH only will have an elevated serum PSA concentration and BPH tissue contributes to this PSA value in a variable manner from patient to patient, it is unlikely that PSA by itself will become an effective screening tool for the early diagnosis of prostate cancer. However, if combined with digital rectal examination and/or transrectal ultrasound it may become a vital part of any early defection program*. Men with prostate cancer had a significantly lower percentage of free PSA than men with BPH only*. Measurement of the percentage of free PSA gives predictive information about the presence or absence of prostate cancer above that provided by other clinical indexes such as age, total PSA level, suspicious results on rectal examination*. Use of the percentage of free PSA can reduce unnecessary biopsies in patients undergoing evaluation for prostate cancer, with a minimal loss in sensitivity in detecting cancer*. The proportion of free to total PSA is recommended for patients with PSA values between 4.0 and 10.0 ng/mL and a paplably benign gland, regardless of patients age or prostate size*. Men with total PSA values between 4.0 and 10.0 ng/mL, the proportion of free to total PSA significantly differentiated between patients with benign and malignant histologic conditions, whereas the total PSA did not*. The proportion of free to total PSA at the same level of sensit

■ TEST PRINCIPLE

Sandwich chemiluminescence immunoassay.

The sample, buffer, magnetic microbeads coated with anti-PSA monoclonal antibody, ABEI labeled with anti-fPSA monoclonal antibody are mixed thoroughly, reacting to form sandwich complexes and incubating. After precipitation in a magnetic field, the supernatant is decanted and then a wash cycle is performed. Subsequently, the Starter 1+2 are added to initiate a chemiluminescent reaction. The light signal is measured by a photomultiplier as relative light units (RLUs), which is proportional to the concentration of fPSA present in the sample.

REAGENTS

Component	Description	100 tests/kit	50 tests/kit	30 tests/kit
Magnetic Microbeads	Magnetic microbeads coated with anti-PSA monoclonal antibody (~4.00 μg/mL) in PBS buffer, NaN ₃ (<0.1%).	2.5 mL	1.5 mL	1.0 mL
Calibrator Low	A low concentration of PSA antigen in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Calibrator High	A high concentration of PSA antigen in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Buffer	PBS buffer, NaN ₃ (<0.1%).	10.5 mL	6.0 mL	3.9 mL
ABEI Label	ABEI labeled with anti-fPSA monoclonal antibody (~0.125 μg/mL) in PBS buffer, NaN ₃ (<0.1%).	11.5 mL	6.5 mL	4.2 mL
Control 1	A low concentration of PSA antigen (0.500 ng/mL) in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
Control 2	A high concentration of PSA antigen (4.00 ng/mL) in PBS buffer, NaN ₃ (<0.1%).	1.0 mL	1.0 mL	1.0 mL
All reagents are pr	ovided ready-to-use.			

The control barcode labels are provided.

Warnings and Precautions

- For in vitro diagnostic use
 For professional use only.
- Exercise the normal precautions required for handling all laboratory reagents.
- Personal protective measures should be taken to prevent any part of the human body from contacting samples, reagents, and controls, and should comply with local operating requirements for the assay.
- A skillful technique and strict adherence to the package insert are necessary to obtain reliable results.
- Do not use kit beyond the expiration date indicated on the label.
- Do not interchange reagent components from different reagents or lots.
- Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).
- All waste associated with biological samples, biological reagents and disposable materials used for the assay should be considered potentially infectious and should be disposed of in accordance with local guidelines.
- This product contains sodium azide. Sodium azide may react with lead or copper plumbing to form highly explosive metal azides. Immediately after disposal, flush
 with a large volume of water to prevent azide build-up. For additional information, see Safety Data Sheets available for professional user on request.

Note: If any serious incident has occurred in relation to the device, please report to Shenzhen New Industries Biomedical Engineering Co., Ltd. (Snibe) or our authorized representative and the competent authority of the Member State in which you are established.

Reagent Handling

- To avoid contamination, wear clean gloves when operating with a reagent kit and sample. When handling reagent kit, replace the gloves that have been in contact
 with samples, since introduction of samples will result in unreliable results.
- Do not use kit in malfunction conditions; e.g., the kit leaking at the sealing film or elsewhere, obviously turbid or precipitation is found in reagents (except for Magnetic Microbeads) or control value is out of the specified range repeatedly. When kit in malfunction conditions, please contact Snibe or our authorized distributor.
- To avoid evaporation of the liquid in the opened reagent kits in refrigerator, it is recommended that the opened reagent kits to be sealed with reagent seals
 contained within the packaging. The reagent seals are single use, and if more seals are needed, please contact Snibe or our authorized distributor.
- Over time, residual liquids may dry on the septum surface. These are typically dried salts and have no effect on assay efficacy.
- Use always the same analyzer for an opened reagent integral.
- · For magnetic microbeads mixing instructions, refer to the Preparation of the Reagent section of this package insert.
- · For further information about the reagent handing during system operation, please refer to Analyzer Operating Instructions.

Storage and Stability

- Do not freeze the integral reagents.
- Store the reagent kit upright to ensure complete availability of the magnetic microbeads.

· Protect from direct sunlight.

Stability of the Reagents	
Unopened at 2-8°C	until the stated expiration date
Opened at 2-8°C	6 weeks
On-board	4 weeks

Stability of Controls				
Unopened at 2-8°C	until the stated expiration date			
Opened at 2-8°C	6 weeks			
Opened at 15-25°C	6 hours			
Frozen at -20°C	3 months			
Frozen and thawed cycles	no more than 3 times			

SPECIMEN COLLECTION AND PREPARATION

Specimen Types

Only the specimens listed below were tested and found acceptable.				
Specimen Types	Collection Tubes			
Serum	Tubes without additive/accessory, or tubes containing clot activator or clot activator with gel			
Plasma	K2-EDTA			

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes
of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some
cases. Follow tube manufacturers' instructions carefully when using collection tubes.

Specimen Conditions

- Do not use heat-inactivated samples or grossly hemolyzed/hyperlipidaemia specimens and specimens with obvious microbial contamination.
- Ensure that complete clot formation in serum specimens has taken place prior to centrifugation. Some serum specimens, especially those from patients receiving
 anticoagulant or thrombolytic therapy, may exhibit increased clotting time. If the serum specimen is centrifuged before a complete clotting, the presence of fibrin
 may cause erroneous results.
- · Samples must be free of fibrin and other particulate matter.
- . To prevent cross contamination, use of disposable pipettes or pipette tips is recommended.

Preparation for Analysis

- Inspect all specimens for foam. Remove foam with an applicator stick before analysis. Use a new applicator stick for each specimen to prevent cross contamination.
- Frozen specimens must be completely thawed before mixing. Mix thawed specimens thoroughly by low speed vortexing or by gently inverting. Visually inspect the
 specimens. If layering or stratification is observed, mix until specimens are visibly homogeneous. If specimens are not mixed thoroughly, inconsistent results may
 be obtained.
- Specimens should be free of fibrin, red blood cells, or other particulate matter. Such specimens may give reliable results and must be centrifuged prior to testing.
 Transfer clarified specimen to a sample cup or secondary tube for testing. For centrifuged specimens with a lipid layer, transfer only the clarified specimen and not the lipemic material.
- The sample volume required for a single determination of this assay is 10 µL.

Specimen Storage

Specimens removed from the separator, red blood cells or clot may be stored up to 8 hours at 15-25°C, or 5 days at 2-8°C, or 3 months frozen at -20°C or colder. Frozen specimens subjected to up to 1 freeze/thaw cycle have been evaluated.

Specimen Shipping

- Package and label specimens in compliance with applicable local regulations covering the transport of clinical specimens and infectious substances.
- Do not exceed the storage limitations listed above.

Specimen Dilution

- Samples, fPSA concentrations above the analytical measuring interval, can be diluted with manual dilution procedure. The recommended dilution ratio is 1:5. The
 concentration of the diluted sample must be >10 ng/mL.
- After manual dilution, multiply the result by the dilution factor.
- Please choose applicable diluents or ask Snibe for advice before manual dilution.

PROCEDURE

Materials Provided

Free PSA (CLIA) assay, control barcode labels. Materials Required (But Not Provided)

General laboratory equipment.

- Fully-auto chemiluminescence immunoassay analyzer Maglumi 600, Maglumi 800, Maglumi 1000, Maglumi 2000, Maglumi 2000 Plus, Maglumi 4000, Maglumi 4000 Plus, MAGLUMI X8, MAGLUMI X3, MAGLUMI X6, or Integrated System Biolumi 8000 and Biolumi CX8.
- Additional accessories of test required for the above analyzers include Reaction Module, Starter 1+2, Wash Concentrate, Light Check, Tip, and Reaction Cup. Specific accessories and accessories' specification for each model refer to corresponding Analyzer Operating Instructions.
- · Please use accessories specified by Snibe to ensure the reliability of the test results.

Assay Procedure

Preparation of the Reagent

- Take the reagent kit out of the box and visually inspect the integral vials for leaking at the sealing film or elsewhere. If there is no leakage, please tear off the sealing film carefully.
- Open the reagent area door; hold the reagent handle to get the RFID label close to the RFID reader (for about 2s); the buzzer will beep; one beep sound indicates successful sensing.
- Keeping the reagent straight insert to the bottom along the blank reagent track.
- Observe whether the reagent information is displayed successfully in the software interface, otherwise repeat the above two steps.
- Resuspension of the magnetic microbeads takes place automatically when the kit is loaded successfully, ensuring the magnetic microbeads are totally resuspended homogenous prior to use.

Assav Calibration

- Select the assay to be calibrated and execute calibration operation in reagent area interface. For specific information on ordering calibrations, refer to the calibration section of Analyzer Operating Instructions.
- Execute recalibration according to the calibration interval required in this package insert.

Quality Control

- When new lot used, check or edit the quality control information.
- Scan the control barcode, choose corresponding quality control information and execute testing. For specific information on ordering quality controls, refer to the
 quality control section of the Analyzer Operating Instructions.

Sample Testing

After successfully loading the sample, select the sample in interface and edit the assay for the sample to be tested and execute testing. For specific information
on ordering patient specimens, refer to the sample ordering section of the Analyzer Operating Instructions.

To ensure proper test performance, strictly adhere to Analyzer Operating Instructions.

Calibration

Traceability: This method has been standardized against the WHO 1st International Standard 96/668.

Test of assay specific calibrators allows the detected relative light unit (RLU) values to adjust the master curve.

Recalibration is recommended as follows:

Whenever a new lot of Reagent or Starter 1+2 is used.

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- Every 28 days.
- · The analyzer has been serviced.
- · Control values lie outside the specified range.

Quality Control

Controls are recommended for the determination of quality control requirements for this assay and should be run in singlicate to monitor the assay performance. Refer to published guidelines for general quality control recommendations, for example Clinical and Laboratory Standards Institute (CLSI) Guideline C24 or other published

Quality control is recommended once per day of use, or in accordance with local regulations or accreditation requirements and your laboratory's quality control procedures, quality control could be performed by running the Free PSA assay:

- · Whenever the kit is calibrated
- Whenever a new lot of Starter 1+2 or Wash Concentrate is used.

Controls are only applicable with MAGLUMI and Biolumi system and only used matching with the same top seven LOT numbers of corresponding reagents. For each target value and range refer to the label.

The performance of other controls should be evaluated for compatibility with this assay before they are used. Appropriate value ranges should be established for all quality control materials used.

Control values must lie within the specified range, whenever one of the controls lies outside the specified range, calibration should be repeated and controls retested. If control values lie repeatedly outside the predefined ranges after successful calibration, patient results must not be reported and take the following

- · Verify that the materials are not expired
- · Verify that required maintenance was performed.
- Verify that the assay was performed according to the package insert.
- If necessary, contact Snibe or our authorized distributors for assistance.

If the controls in kit are not enough for use, please order Free PSA (CLIA) Controls (REF: 160201222MT) from Snibe or our authorized distributors for more.

RESULTS

Calculation

The analyzer automatically calculates the fPSA concentration in each sample by means of a calibration curve which is generated by a 2-point calibration master curve procedure. The results are expressed in ng/mL. For further information please refer to the Analyzer Operating Instructions

Interpretation of Results

The cut-off for the Free PSA assay was obtained by testing 394 male subjects (260 biopsy negative and 134 biopsy positive) with a total PSA range of 4 to 10 ng/mL and a DRE non-suspicious for prostate cancer in China, gave a cut-off of 25%. At the cutoff of 25%, the sensitivity and specificity of the Free PSA assay were 88,06% and 56.92%, respectively.

Results may differ between laboratories due to variations in population and test method. It is recommended that each laboratory establish its own reference interval.

LIMITATIONS

- The Free PSA assay should be used only with the MAGLUMI Total PSA (CLIA) assay to calculate the ratio fPSA/tPSA (%fPSA).
- · Results should be used in conjunction with patient's medical history, clinical examination and other findings
- . If the fPSA results are inconsistent with clinical evidence, additional testing is needed to confirm the result.
- · The assay is not suitable for screening of the general population.
- Digital rectal examination (DRE), prostatic massage, ultrasonography, and needle biopsy can lead to PSA elevations 16.17. PSA levels may also be increased following ejaculation18.
- Hormonal therapy may affect PSA expression, resulting in a decrease in PSA levels¹⁹.
- Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies (HAMA). Such specimens may show either falsely elevated or depressed values when tested with assay kits which employ mouse monoclonal antibodies^{20,21} Additional information may be required for diagnosis.
- · Heterophilic antibodies in human serum can react with reagent immunoglobulins, interfering with in vitro immunoassays. Patients routinely exposed to animals or animal serum products can be prone to this interference and anomalous values may be observed²².
- Bacterial contamination or heat inactivation of the specimens may affect the test results

■ SPECIFIC PERFORMANCE CHARACTERISTICS

Representative performance data are provided in this section. Results obtained in individual laboratories may vary.

Precision

Precision was determined using the assay, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): duplicates at two

independent runs per day for 5 days at three different sites using three lots of reagent kits (n =180). The following results were obtained:

Sample	Mean (ng/mL) Within-Run		-Run	Between-Run		Reproducibility	
Sample	(n=180)	SD (ng/mL)	%CV	SD (ng/mL)	%CV	SD (ng/mL)	%CV
Serum Pool 1	2.020	0.068	3.37	0.053	2.62	0.101	5.00
Serum Pool 2	10.119	0.336	3.32	0.166	1.64	0.496	4.90
Serum Pool 3	30.180	0.891	2.95	0.355	1.18	1.398	4.63
Plasma Pool 1	2.022	0.085	4.20	0.048	2.37	0.120	5.93
Plasma Pool 2	9.923	0.312	3.14	0.211	2.13	0.48	5.44
Plasma Pool 3	29.889	0.957	3.20	0.143	0.48	1.174	3.93
Control 1	0.506	0.022	4.35	0.002	0.40	0.029	5.73
Control 2	4.054	0.143	3.53	0.089	2.20	0.226	5.57

Linear Range

0.020-50.0 ng/mL (defined by the Limit of Quantitation and the maximum of the master curve).

Reportable Interval

0.015-250 ng/mL (defined by the Limit of Detection and the maximum of the master curve×Recommended Dilution Ratio).

Analytical Sensitivity

Limit of Blank (LoB) =0.010 na/mL.

Limit of Detection (LoD) =0.015 ng/mL. Limit of Quantitation (LoQ) =0.020 ng/mL.

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Analytical Specificity

Interference was determined using the assay, three samples containing different concentrations of analyte were spiked with potential endogenous and exogenous interferents in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Interference	No interference up to	Interference	No interference up to		
Bilirubin	66 mg/dL	Cisplatin	165 μg/mL		
Hemoglobin	1000 mg/dL	Methotrexate	30 μg/mL		
Intralipid	1500 mg/dL	5-Fluorouracil	360 μg/mL		
HAMA	40 ng/mL	Paclitaxel	67 μg/mL		
Rheumatoid factor	1500 IU/mL	Vinblastine sulfate	1.5 μg/mL		
ANA	6 (S/CO) strong positive	Doxorubicin hydrochloride	50 μg/mL		
Cyclophosphamide monohydrate	500 μg/mL	Carboplatin	500 μg/mL		
Megestrol	90 μg/mL	Carbopiatiii	эоо рулпс		
Cross Penetivity					

Cross-reactivity was determined using the assay, three samples containing different concentrations of analyte were spiked with potential cross-reactants in a protocol (EP7-A2) of the CLSI. The measurement deviation of the interference substance is within ±10%. The following results were obtained:

Cross-reactant	No interference up to	Cross-reactant	No interference up to	
α1-antichymotrypsin	100 ng/mL	CEA	3000 ng/mL	
PSA-ACT	100 ng/mL	CEA	3000 fig/file	

High-Dose Hook

No high-dose hook effect was seen for fPSA concentrations up to 15000 ng/mL.

Method Comparison

A comparison of the Free PSA assay with a commercially available immunoassay, gave the following correlations (ng/mL):

Number of samples measured: 397

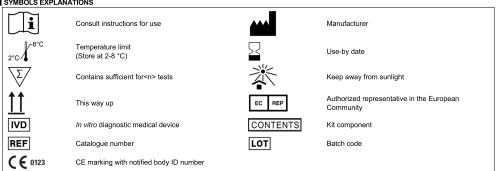
Passing-Bablok: y=1.0138x-0.0025, т=0.948.

The clinical specimen concentrations were between 0.022 and 49.26 ng/mL

■ REFERENCES

- 1. Oesterling J E. Prostate specific antigen: a critical assessment of the most useful tumor marker for adenocarcinoma of the prostate[J]. The Journal of Urology, 1991 145-907-923
- 2. Balk S P, Ko Y J, Bubley G J. Biology of Prostate-Specific Antigen[J]. Journal of Clinical Oncology, 2003, 21:383-391.
- 3. Mccormack R T, Wang T J, Rittenhouse H G, et al. Molecular forms of prostate-specific antigen and the human kallikrein gene family: a new era[J]. Urology, 1995, 45: 729-744.
- 4. Benson M C, Wang I S, Olsson C A, et al. The use of prostate specific antigen density to enhance the predictive value of intermediate levels of serum prostate specific antigen[J]. The Journal of Urology, 1992, 147:817-821.
- Becker C, Noldus J, Diamandis E, et al. The role of molecular forms of prostate-specific antigen (PSA or hK3) and of human glandular kallikrein 2 (hK2) in the diagnosis and monitoring of prostate cancer and in extra-prostatic disease[J]. Critical Reviews in Clinical Laboratory Sciences, 2001, 38(5):357-399.
- 6. Abrahamsson P A, Lilja H, Oesterling J E. Molecular forms of serum prostate-specific antigen: the clinical value of percent free prostate-specific antigen[J]. Urologic Clinics of North America, 1997, 24: 353-365.
- 7. Catalona W J, Smith D S, Ratliff T L, et al. Detection of organ-confined prostate cancer is increased through prostate-specific based Screening[J]. JAMA, 1993, 270:948-954
- 8. Oesterling J E, Jacobsen S J, Chute C G, et al. Serum prostate-specific antigen in a community-based population of healthy men establishment of age-specific reference ranges[J]. JAMA, 1993, 270:860-864.
- 9. Carter H B, Pearson J D, Metter J, et al. Longitudinal evaluation of prostate-specific antigen levels in men with and without prostate disease[J]. JAMA, 1992,
- 10. Berry S J, COFFEY D S, Walsh P C, et al. The development of human benign prostatic hyperplasia with age [J]. The Journal of Urology, 2000, 132:474-479.
- 11. Richie J E, Ratlife T L, Catalona W J, et al. Effect of patient age on early detection of prostate cancer with serum prostate-specific antigen and digital rectal examination[J]. Urology, 1993, 42(4):365-374
- 12. Catalona W J, Smith D S, Wolfert R L, et al. Evaluation of percentage of free serum prostate-specific antigen to improve specificity of prostate cancer screening[J].JAMA, 1995, 274(15):1214:1220.
- 13. Catalona W J, Partin A W, Slawin, K M, et al. Use of the percentage of free prostate-specific antigen to enhance differentiation of prostate cancer from benign prostatic disease[J].JAMA, 1998, 279(19):1542:1547.
- 14. Luderer A.A. Chen Y.T. Soriano T.F. et al. Measurement of the proportion of free to total prostate-specific antigen improves diagnostic performance of prostatespecific antigen in the diagnostic gray zone of the total prostate-specific antigen[J]. Urology. 1995, 46(2):187-194.
- 15. CLSI. Statistical Quality Control for Quantitative Measurement Procedures: Principles and Definitions. 4th ed. CLSI guideline C24. Wayne, PA: Clinical and Laboratory Standards Institute; 2016.
- 16. Collins G N, Martin P J, Wynn-Davies A, et al. The effect of digital rectal examination, flexible cystoscopy and prostatic biopsy on free and total prostate specific antigen, and the free-to-total prostate specific antigen ratio in clinical practice[J]. The Journal of Urology, 1997, 157(5):1744-1747.
- 17. Yuan J J, Coplen D E, Petros J A, et al. Effects of rectal examination, prostatic massage, ultrasonography and needle biopsy on serum prostate specific antigen levels[J]. Journal of Urology, 1992, 147(3 Part 2):810-814.
- 18. Tchetgen M B, Song J T, Strawderman M, et al. Ejaculation increases the serum prostate-specific antigen concentration[J]. Urology, 1996, 47(4):511-516.
- 19. Morgan W R, Zincke H, Rainwater L M, et al. Prostate specific antigen values after radical retropubic prostatectomy for adenocarcinoma of the prostate: impact of adjuvant treatment (hormonal and radiation)[J]. The Journal of Urology, 1991, 145(2):319-323.
- 20. Robert W. Schroff, Kenneth A. Foon, Shannon M. Beatty, et al. Human Anti-Murine Immunoglobulin Responses in Patients Receiving Monoclonal Antibody Therapy [J]. Cancer Research, 1985, 45(2):879-885.
- 21. Primus F J, Kelley E A, Hansen H J, et al. "Sandwich"-type immunoassay of carcinoembryonic antigen in patients receiving murine monoclonal antibodies for diagnosis and therapy [J]. Clinical Chemistry, 1988, 34(2):261-264.
- 22. Boscato L M, Stuart M C. Heterophilic antibodies: a problem for all immunoassays [J]. Clinical Chemistry, 1988,34(1):27-33.

SYMBOLS EXPLANATIONS



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Summary of safety and performance is available at Eudamed.

Tel: +49-40-2513175



Shenzhen New Industries Biomedical Engineering Co., Ltd. No.23, Jinxiu East Road, Pingshan District, 518122 Shenzhen, P.R. China Tel: +86-755-21536601 Fax:+86-755-28292740

Fax: +49-40-255726

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Shanghai International Holding Corp. GmbH (Europe) Eiffestrasse 80, 20537 Hamburg, Germany

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