

**CONGEN**

**SureFood® GMO SCREEN 4plex  
BAR/NPTII/PAT/CTP2:CP4-  
EPSPS**

Art. No. S2127  
100 rxn

**User Manual**



**March 2023**



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## **1 General Information**

### **1.1 Description**

The SureFood® GMO SCREEN 4plex BAR/NPTII/PAT/CTP2:CP4-EPSPS is a real-time PCR for the direct, qualitative detection and differentiation of following specific DNA sequences:

- Phosphinothricin-Acetyltransferase gene (BAR) from *Streptomyces hygroscopicus*
- Antibiotics-resistance gene Neomycin-Phosphotransferase (nptII)
- Phosphinothricin-Acetyltransferase gene (PAT) from *Streptomyces viridochromogenes*
- the transition from CTP2 (Chloroplast-Transpeptide-signal sequence from *Arabidopsis thaliana*) to herbicide tolerance-gene CP4 EPSPS (5-Enolpyruvylshikimat-3-Phosphat Syntheses gene from *Agrobacterium tumefaciens* strain CP4)

This kit can be used for screening of genetically modified organisms (GMOs) in food, feed and seeds.

The detections are according to the official collection of detection methods of §64 German food law, especially according to technical specification BVL L-00.00-154.

The real-time PCR assay can be performed with commonly used real-time PCR instruments, equipped for detection of four fluorescence emissions at the channels FAM, VIC/HEX, ROX and Cy5 at the same time. The technical verification of instruments was performed on Roche LightCycler® 480 II, Applied Biosystems 7500, Bio-Rad CFX96, R-Biopharm RIDA®CYCLER and Agilent Mx3005P.

### **1.2 Limit of Detection**

The SureFood® GMO SCREEN 4plex BAR/NPTII/PAT/CTP2:CP4-EPSPS real-time PCR has a limit of detection of  $\leq 5$  DNA copies.

The assay limit of detection depends on sample matrix, processing grade, DNA preparation and DNA content.

The SureFood® PCR systems are very sensitive and therefore even a small amount of target DNA is sufficient for a successful analysis. The concentration of total DNA in the sample does not allow a conclusion on the quantity and quality of the target DNA.

**Note:** Inconsistent mixing ratios\* may cause a loss of sensitivity in the low concentration channel in mixed samples especially with high amplicon concentrations (Cp value < 20).

\* e.g. 99.9 MON88017 Corn and 0.1 % Bt176 Corn

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## 1.3 DNA-preparation

For DNA-preparation of raw material the use of SureFood® PREP Basic (Art. No. S1052), SureFast® Mag PREP Food (Art. No. F1060) and for highly processed food and feed the use of SureFood® PREP Advanced (Art. No. S1053) is recommended. SureFood® PREP Add On (Art. No. S1055) is intended to be used for the extraction of DNA from raw materials as well as processed food and feed with sample weight of 2 g. It is used in conjunction with the SureFood® PREP Basic.

## 1.4 Kit components and storage

Kit Code	Reagent	Amount	Lid Color
1	Reaction Mix	2 x 1050 µl	Yellow
2	Taq Polymerase	1 x 80 µl	Dark Red
3	Positive Control	1 x 190 µl	Light Blue

**Store all reagents at –20°C and protected from light. The Taq Polymerase can be stored at +2 to +8°C for multiple uses on the same day.**

**Note: The Taq Polymerase may be in a frozen or unfrozen state. This does not affect the quality of the Taq Polymerase or the performance of the real-time PCR.**

## 1.5 Additionally required equipment and materials

- DNA-Extraction kit  
(e.g. SureFood® PREP Basic Art. No. S1052 / SureFood® PREP Advanced Art. No. S1053 / SureFood® PREP Add On Art. No. S1055 / SureFast® Mag PREP Food Art. No. F1060)
- real-time PCR instrument with four detection channels (510 nm, 580 nm, 610 nm and 660 nm)
- real-time PCR consumable (plates, tubes, foils, caps)
- pipettes with filter tips
- powder-free disposable gloves
- Vortex mixer
- micro centrifuge with a rotor for the reaction tubes

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**1.6 Setup**

	<b>Blockcycler &amp; R-Biopharm RIDA®CYCLER</b>	<b>Rotorcycler</b>
Initial Denaturation (HOLD)	5 min, 95°C	1 min, 95°C
Cycles	45	45
Denaturation	15 sec, 95°C	10 sec, 95°C
Annealing/Extension (CYCLE)	30 sec, 60°C	15 sec, 60°C
Temperature Transition Rate/ Ramp Rate	Maximum	Maximum

**1.7 Detection channel Set-up**

<b>Real-time PCR device</b>	<b>Detection</b>	<b>Detection channel</b>	<b>Quencher</b>	<b>Note</b>
<b>Agilent Mx3005P</b>	nptII	FAM	+	
	PAT	HEX	+	
	CTP2:CP4-EPSPS	ROX	+	
	BAR	Cy5	+	
<b>Applied Biosystems 7500</b>	nptII	FAM	None	Check the passive reference option ROX is none.
	PAT	VIC	None	
	CTP2:CP4-EPSPS	ROX	None	
	BAR	Cy5	None	
<b>Bio-Rad CFX96/Dx</b>	nptII	FAM	+	
	PAT	VIC/HEX	+	
	CTP2:CP4-EPSPS	ROX	+	
	BAR	Cy5	+	
<b>R-Biopharm RIDA®CYCLER</b>	nptII	green	+	
	PAT	yellow	+	
	CTP2:CP4-EPSPS	orange	+	
	BAR	red	+	
<b>Roche LightCycler® 480 II</b>	nptII	465-510	+	The SureCC Color Compensation Kit I (Art. No. F4009) is required.
	PAT	533-580	+	
	CTP2:CP4-EPSPS	533-610	+	
	BAR	618-660	+	

## 2 Qualitative Analysis

### 2.1 Protocol

#### 2.1.1 Preparation of the master-mix

Calculate the total number of reactions needed (samples and control reactions) for the specific PCR assay as well as for the inhibition control.

Recommended control reactions for the specific PCR assay: negative control, extraction control, positive control and an inhibition control per sample.

For the preparation of the inhibition control the use of the SureFood® GMO Plant PLUS (Art. No. S2049) is recommended.

#### Reactions needed for the qualitative nptII, PAT, CTP2:CP4-EPSPS and BAR detection:

3 reactions for controls (1x no-template control, 1x extraction control, 1x positive control)

For each sample: at least 1 reaction for each sample DNA

It is also recommended to prepare the master-mix with 10 % additional volume in order to compensate reagent loss. Allow the reagents to thaw, mix and centrifuge before opening and use.

#### Example for the calculation and preparation of 10 reactions:

Components of the master-mix	Amount per reaction	10 reactions (with 10% excess)
Reaction Mix	19.3 µl	212.3 µl
Taq Polymerase	0.7 µl	7.7 µl
<b>Total volume</b>	<b>20 µl</b>	<b>220 µl</b>

**Mix each master-mix well and centrifuge shortly before use.**

#### 2.1.2 Preparation of the real-time PCR-mix

- Pipette 20 µl of the master-mix into appropriate tubes/wells.
- Close the negative control (the negative control is ready for PCR without any addition).
- Pipette 5 µl of sample DNA into the designated tubes/wells and close them.
- Pipette 5 µl of Positive Control into the designated tubes/wells and close them.
- Centrifuge all tubes/plates shortly at low speed.
- Place tubes/plates into the real-time PCR instrument and start the run according to the setup.

**2.2 Interpretation of results**

The evaluation has to be made according to the usual analysis program recommended by the real-time PCR instrument manufacturer.

The control reactions have to show the correct results.

NptII DNA is detected in the FAM-channel, PAT DNA is detected in the VIC/HEX -channel, CTP2:CP4-EPSPS DNA is detected in the ROX-channel and BAR DNA is detected in the Cy5-channel (see table).

A sample is stated **positive** for the respective parameter, if the sample DNA shows amplification in the respective channel.

A sample is stated **negative** for the respective parameter, if the sample DNA shows no amplification in the respective channel and if the external inhibition control of the sample is **positive** with a shift in Cp-value ≤ 2 compared to the negative control.

If the sample DNA in the external inhibition control shows **no amplification** or a shift in Cp-value > 2 compared to the negative control, it contains PCR inhibiting substances. A significant decrease in the fluorescence signal can also show the presence of PCR inhibiting substances. Under these circumstances' DNA isolation and purification of the sample need to be improved. Alternatively, the DNA can be diluted (recommendation 1:2 in PCR-water) and analysed again for inhibition. Please note that the dilution factor also affects the detection limit of the specific nptII, PAT, CTP2:CP4-EPSPS or BAR PCR assay.

It may appear in some cases that only one of the two DNA duplicates prepared from the test sample is nptII and/or PAT and/or CTP2:CP4-EPSPS and/or BAR **positive**. This indicates that the amount of genetically modified DNA is very low and at the limit of detection. If such results are obtained in the analysis (see DIN EN ISO 24276:2013-10), the sample is stated negative.

Result in the respective channel				Result	Interpretation
FAM channel nptII	VIC/HEX channel PAT	ROX channel CTP2:CP4- EPSPS	Cy5 channel BAR	external inhibition- control	
<b>positive</b>	negative	negative	negative	<b>positive</b>	<b>nptII</b> DNA detected
negativ	<b>positive</b>	negative	negative	<b>positive</b>	<b>PAT</b> DNA detected
negativ	negative	<b>positive</b>	negative	<b>positive</b>	<b>CTP2:CP4-EPSPS</b> DNA detected
negativ	negative	negative	<b>positive</b>	<b>positive</b>	<b>BAR</b> DNA detected
negativ	negative	negative	negative	<b>positive</b>	Negative, target DNA is not detected
negativ	negative	negative	negative	negative	invalid

### **3 Further Information**

#### **3.1 Product Information**

- Detailed information about setup of several real-time PCR devices  
(Download: [www.congen.de/en/company/downloads](http://www.congen.de/en/company/downloads))
- Verification Report upon request

#### **3.2 Technical Support**

For further questions please contact your distributor or send an e-mail to [sales@r-biopharm.de](mailto:sales@r-biopharm.de).

#### **3.3 Distribution and Ordering**

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