

# Product information

Information about other products is available at: [www.demeditec.com](http://www.demeditec.com)



User's Manual

## Borrelia burgdorferi IgG ELISA



**DEBORG0040**



**96 wells**



Demeditec Diagnostics GmbH  
Lise-Meitner-Strasse 2  
24145 Kiel – Germany  
[www.demeditec.com](http://www.demeditec.com)

## CONTENTS

1. INTRODUCTION	3
2. INTENDED USE	3
3. PRINCIPLE OF THE ASSAY	3
4. MATERIALS	4
5. STABILITY AND STORAGE	4
6. REAGENT PREPARATION	4
7. SAMPLE COLLECTION AND PREPARATION	5
8. ASSAY PROCEDURE	5
9. RESULTS	6
10. SPECIFIC PERFORMANCE CHARACTERISTICS	7
11. LIMITATIONS OF THE PROCEDURE	7
12. PRECAUTIONS AND WARNINGS	8
BIBLIOGRAPHY	9
ABBREVIATIONS	9
SUMMARY OF TEST PROCEDURE	10
SYMBOLS USED WITH DEMEDITEC ASSAYS	12

## 1. INTRODUCTION

Spirochetes are motile bacteria with a periplasmic axial filament. All pathogenic species belong to the family Treponemataceae, which includes the three genera: Treponema, Borrelia, and Leptospira. The Treponemae are extremely long, flexible, filamentous cells that are usually held in a characteristic spiral, or coiled-spring shape. Borreliae are the largest Treponemataceae with very coarse and irregular spirals. Borrelia burgdorferi, the causative agent of Lyme disease, is transmitted mainly by ticks but probably also by other blood-sucking insects. Habitats are the wooded, humid and temperate regions of North America, Europe, North Africa, Australia and Japan; foresters, farmers, and anybody entering infested forests may be affected.

The degree of contamination of ticks amounts to 3-60% dependent on seasonal and regional differences; up to 30% of the population may be infected (about 1500 cases annually in USA, several hundred in Europe).

Species	Disease	Symptoms (e.g.)	Transmission route
Borrelia burgdorferi	Lyme diseases	<b>Multisystemic illness</b> <b>Early manifestations:</b> Erythema migrans; flu-like symptoms like fever, myalgias and headaches; carditis; meningitis <b>Late manifestations:</b> Chronic neurological, dermatological and / or cardiological symptoms; Acrodermatitis chronica atrophicans; Herxheimer reaction; Lyme-Arthritis	Transmission by tick bite Europe: Ixodes ricinus USA: Ixodes dammini

Infection or presence of pathogen may be identified by:

- Microscopy
- PCR
- Serology: e.g. ELISA

## 2. INTENDED USE

The Borrelia burgdorferi IgG ELISA is intended for the qualitative determination of IgG class antibodies against Borrelia burgdorferi in human serum, plasma (citrate, heparin) and CSF (cerebrospinal fluid). For the determination of intrathecally produced IgG antibodies a separate instruction for use for CSF can be obtained at Demeditec.

## 3. PRINCIPLE OF THE ASSAY

The qualitative immunoenzymatic determination of specific antibodies is based on the ELISA (Enzyme-linked Immunosorbent Assay) technique.

Microtiterplates are coated with specific antigens to bind corresponding antibodies of the sample. After washing the wells to remove all unbound sample material a horseradish peroxidase (HRP) labelled conjugate is added. This conjugate binds to the captured antibodies. In a second washing step unbound conjugate is removed. The immune complex formed by the bound conjugate is visualized by adding Tetramethylbenzidine (TMB) substrate which gives a blue reaction product.

The intensity of this product is proportional to the amount of specific antibodies in the sample.

Sulphuric acid is added to stop the reaction. This produces a yellow endpoint colour. Absorbance at 450/620 nm is read using an ELISA Microtiterplate reader

## 4. MATERIALS

### 4.1. Reagents supplied

1. **SORB MIT Microtiterplate:** 12 break-apart 8- well snap-off strips coated with recombinant *Borrelia burgdorferi* antigens; in resealable aluminium foil.
2. **SAM DIL IgG Sample Dilution Buffer:** 1 bottle containing 100 mL of phosphate buffer (10 mM) for sample dilution; pH 7.2 ± 0.2; coloured yellow; ready to use; white cap; ≤ 0.0015% (v/v) CMIT/ MIT (3:1).
3. **STOP SOLN Stop Solution:** 1 bottle containing 15 mL sulphuric acid, 0.2 mol/L; ready to use; red cap.
4. **WASH SOLN 20x Washing Buffer (20x conc.):** 1 bottle containing 50 mL of a 20-fold concentrated phosphate buffer (0.2 M), pH 7.2 ± 0.2, for washing the wells; white cap.
5. **ENZ CONJ Conjugate:** 1 bottle containing 20 mL of peroxidase labelled antibody to human IgG in phosphate buffer (10 mM); coloured blue; ready to use; black cap.
6. **SUB TMB TMB Substrate Solution:** 1 bottle containing 15 mL 3,3',5,5'-tetramethylbenzidine (TMB), < 0.1 %; ready to use; yellow cap.
7. **CAL C Positive Control:** 1 vial containing 2 mL control; coloured yellow; ready to use; red cap; ≤ 0.02% (v/v) MIT.
8. **CAL B Cut-off Control:** 1 vial containing 3 mL control; coloured yellow; ready to use; green cap; ≤ 0.02% (v/v) MIT.
9. **CAL A Negative Control:** 1 vial containing 2 mL control; coloured yellow; ready to use; blue cap; ≤ 0.0015% (v/v) CMIT/ MIT (3:1).

Controls are calibrated in arbitrary units against internal quality control specimens, since no international standard reference is available for this assay.

For hazard and precautionary statements see 12.1

For potential hazardous substances please check the safety data sheet.

### 4.2. Materials supplied

- 1 Cover foil
- 1 Instruction for use (IFU)
- 1 Plate layout

### 4.3. Materials and Equipment needed

- ELISA Microtiterplate reader, equipped for the measurement of absorbance at 450/620 nm
- Incubator 37 °C
- Manual or automatic equipment for rinsing Microtiterplates
- Pipettes to deliver volumes between 10 and 1000 µL
- Vortex tube mixer
- Distilled water
- Disposable tubes

## 5. STABILITY AND STORAGE

Store the kit at 2...8 °C. The opened reagents are stable up to the expiry date stated on the label when stored at 2...8 °C.

## 6. REAGENT PREPARATION

It is very important to bring all reagents and samples to room temperature (20...25 °C) and mix them before starting the test run!

### 6.1. Microtiterplate

The break-apart snap-off strips are coated with recombinant *Borrelia burgdorferi* antigens. Immediately after removal of the strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied and stored at 2...8 °C.

### 6.2. Washing Buffer (20x conc.)

Dilute Washing Buffer 1 + 19; e. g. 10 mL Washing Buffer + 190 mL distilled water. The diluted buffer is stable for 5 days at room temperature (20...25 °C). In case crystals appear in the concentrate, warm up the solution to 37 °C e.g. in a water bath. Mix well before dilution.

### 6.3. TMB Substrate Solution

The reagent is ready to use and has to be stored at 2...8 °C, away from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have become contaminated and should be thrown away.

## 7. SAMPLE COLLECTION AND PREPARATION

Use human serum, plasma (citrate, heparin) and CSF (cerebrospinal fluid) samples with this assay. If the assay is performed within 5 days after sample collection, the samples should be kept at 2...8 °C; otherwise they should be aliquoted and stored deep-frozen (-70...-20 °C). If samples are stored frozen, mix thawed samples well before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.

### 7.1. Sample Dilution

Before assaying, all samples should be diluted 1+100 with IgG Sample Dilution Buffer. Dispense 10 µL sample and 1 mL IgG Sample Dilution Buffer into tubes to obtain a 1+100 dilution and thoroughly mix with a Vortex.

## 8. ASSAY PROCEDURE

Please read the instruction for use carefully before performing the assay. Result reliability depends on strict adherence to the instruction for use as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend increasing the washing steps from three up to five and the volume of Washing Buffer from 300 µL to 350 µL to avoid washing effects. Pay attention to chapter 12. Prior to commencing the assay, the distribution and identification plan for all samples and standards/controls (duplicates recommended) should be carefully established on the plate layout supplied in the kit. Select the required number of microtiter strips or wells and insert them into the holder.

Perform all assay steps in the order given and without any delays.

A clean, disposable tip should be used for dispensing each standard/control and sample.

Adjust the incubator to 37 ± 1 °C.

1. Dispense 100 µL standards/controls and diluted samples into their respective wells. Leave well A1 for the Substrate Blank.
2. Cover wells with the foil supplied in the kit.
3. **Incubate for 1 hour ± 5 min at 37 ± 1 °C.**
4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 300 µL of Washing Buffer. Avoid overflows from the reaction wells. The interval between washing and aspiration should be > 5 sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!  
Note: Washing is important! Insufficient washing results in poor precision and false results.
5. Dispense 100 µL Conjugate into all wells except for the Substrate Blank well A1.
6. **Incubate for 30 min at room temperature (20...25 °C).** Do not expose to direct sunlight.
7. Repeat step 4.
8. Dispense 100 µL TMB Substrate Solution into all wells.
9. **Incubate for exactly 15 min at room temperature (20...25 °C) in the dark.** A blue colour occurs due to an enzymatic reaction.
10. Dispense 100 µL Stop Solution into all wells in the same order and at the same rate as for the TMB Substrate Solution, thereby a colour change from blue to yellow occurs.
11. Measure the absorbance at 450/620 nm within 30 min after addition of the Stop Solution.

### 8.1. Measurement

Adjust the ELISA Microtiterplate reader **to zero** using the **Substrate Blank**.

If - due to technical reasons - the ELISA Microtiterplate reader cannot be adjusted to zero using the Substrate Blank, subtract its absorbance value from all other absorbance values measured in order to obtain reliable results!

**Measure the absorbance** of all wells at **450 nm** and record the absorbance values for each standard/control and sample in the plate layout.

Bichromatic measurement using a reference wavelength of 620 nm is recommended.

**Where applicable calculate the mean absorbance values of all duplicates.**

## 9. RESULTS

### 9.1. Run Validation Criteria

In order for an assay run to be considered valid, these Instructions for Use have to be strictly followed and the following criteria must be met:

- **Substrate Blank:** Absorbance value < **0.100**
- **Negative Control:** Absorbance value < **0.200** and < **Cut-off**
- **Cut-off Control:** Absorbance value **0.150 – 1.300**
- **Positive Control:** Absorbance value > **Cut-off**

If these criteria are not met, the test is not valid and must be repeated.

### 9.2. Calculation of Results

The Cut-off is the mean absorbance value of the Cut-off Control determinations.

Example: Absorbance value Cut-off Control 0.44 + absorbance value Cut-off control 0.42 = 0.86 / 2 = 0.43  
Cut-off = 0.43

#### 9.2.1. Results in Units [U]

$$\frac{\text{Sample (mean) absorbance value} \times 10}{\text{Cut-off}} = [\text{Units} = \text{U}]$$

Example: 
$$\frac{1.591 \times 10}{0.43} = 37 \text{ U (Units)}$$

### 9.3. Interpretation of Results

Cut-off	10 U	-
Positive	> 11 U	Antibodies against the pathogen are present. There has been a contact with the antigen (pathogen resp. vaccine).
Equivocal	9 – 11 U	Antibodies against the pathogen could not be detected clearly. It is recommended to repeat the test with a fresh sample in 2 to 4 weeks. If the result is equivocal again the sample is judged as <b>negative</b> .
Negative	< 9 U	The sample contains no antibodies against the pathogen. A previous contact with the antigen (pathogen resp. vaccine) is unlikely.
Diagnosis of an infectious disease should not be established on the basis of a single test result. A precise diagnosis should take into consideration clinical history, symptomatology as well as serological data. In immunocompromised patients and newborns serological data only have restricted value.		

#### 9.3.1. Antibody Isotypes and State of Infection

Serology	Significance
IgM	Characteristic of the primary antibody response High IgM titer with low IgG titer: → suggests a current or very recent infection Rare: → persisting IgM
IgG	Characteristic of the secondary antibody response May persist for several years High IgG titer with low IgM titer: → may indicate a past infection

## 10. SPECIFIC PERFORMANCE CHARACTERISTICS

The results refer to the groups of samples investigated; these are not guaranteed specifications. For further information about the specific performance characteristics please contact Demeditec Diagnostics GmbH.

### 10.1. Precision

<b>Intraassay</b>	<b>n</b>	<b>Mean (E)</b>	<b>CV (%)</b>
#1	24	0.517	4.00
#2	24	1.790	4.23
#3	24	2.339	9.04
<b>Interassay</b>	<b>n</b>	<b>Mean (U)</b>	<b>CV (%)</b>
#1	12	37.2	4.17
#2	12	45.3	9.28
#3	12	1.9	9.94

### 10.2. Diagnostic Specificity

The diagnostic specificity is defined as the probability of the assay of scoring negative in the absence of the specific analyte.

It is 95.48% (95% confidence interval: 90.92% - 98.17%).

### 10.3. Diagnostic Sensitivity

The diagnostic sensitivity is defined as the probability of the assay of scoring positive in the presence of the specific analyte.

It is 98.39% (95% confidence interval: 91.34% - 99.96%).

### 10.4. Interferences

Interferences with hemolytic, lipemic or icteric samples are not observed up to a concentration of 10 mg/mL hemoglobin, 5 mg/mL triglycerides and 0.5 mg/mL bilirubin.

### 10.5. Cross Reactivity

By the use of recombinant antigens, cross reactions with antibodies against the following pathogens can be excluded, as best as possible:

- Treponema pallidum
- Leptospira
- Borrelia recurrentis

A lues-infection should be excluded, because antibodies against p41i could appear.

## 11. LIMITATIONS OF THE PROCEDURE

Bacterial contamination or repeated freeze-thaw cycles of the sample may affect the absorbance values.

## 12. PRECAUTIONS AND WARNINGS

- The test procedure, the information, the precautions and warnings in the instructions for use have to be strictly followed. The use of the testkits with analyzers and similar equipment has to be validated. Any change in design, composition and test procedure as well as for any use in combination with other products not approved by the manufacturer is not authorized; the user himself is responsible for such changes. The manufacturer is not liable for false results and incidents for these reasons. The manufacturer is not liable for any results by visual analysis of the patient samples.
- Only for in-vitro diagnostic use.
- All materials of human or animal origin should be regarded and handled as potentially infectious.
- All components of human origin used for the production of these reagents have been tested for anti-HIV antibodies, anti-HCV antibodies and HBsAg and have been found to be non-reactive.
- Do not interchange reagents or Microtiterplates of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry date stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and standard/control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette patient samples and dispense reagents without splashing accurately into the wells.
- The ELISA is only designed for qualified personnel following the standards of good laboratory practice (GLP).
- For further internal quality control each laboratory should additionally use known samples.

### 12.1. Safety note for reagents containing hazardous substances

Reagents may contain CMIT/MIT (3:1) or MIT (see 4.1 oder refers to 4.1)

Therefore, the following hazard and precautionary statements apply.

#### Warning



H317	May cause an allergic skin reaction.
P261	Avoid breathing spray
P280	Wear protective gloves/ protective clothing.
P302+P352	IF ON SKIN: Wash with plenty of soap and water.
P333+P313	If skin irritation or rash occurs: Get medical advice/ attention.
P362+P364	Take off contaminated and Wash it before reuse.

Further information can be found in the safety data sheet

### 12.2. Disposal Considerations

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.



## BIBLIOGRAPHY

- Asbrink, Eva; Hovmark, Anders (1988): Early and late cutaneous manifestations in Ixodes-borne borreliosis (erythema migrans borreliosis, Lyme borreliosis). In *Annals of the New York Academy of Sciences* 539, pp. 4–15.
- Barbour, Alan G. (2006): Relapsing Fever and Other Borrelia Diseases. In Richard L. Guerrant, David H. Walker, Peter F. Weller (Eds.): Tropical infectious diseases. Principles, pathogens & practice. 2nd ed. Philadelphia: Churchill Livingstone, pp. 499–510.
- Deutsche Borreliose-Gesellschaft e.V. (2011): Diagnostik und Therapie der Lyme-Borreliose. Leitlinien der Deutschen Borreliose-Gesellschaft.
- Eicken, Christoph; Sharma, Vivek; Klabunde, Thomas; Lawrenz, Matthew B.; Hardham, John M.; Norris, Steven J.; Sacchetti, James C. (2002): Crystal structure of Lyme disease variable surface antigen VlsE of Borrelia burgdorferi. In *The Journal of biological chemistry* 277 (24), pp. 21691–21696. DOI: 10.1074/jbc.M201547200.
- Jauris-Heipke, Sigrid; Rößle, Birgit; Wanner, Gerhard; Habermann, Christiane; Rössler, Dieter; Fingerle, Volker et al. (1999): Osp17, a novel immunodominant outer surface protein of Borrelia afzelii: recombinant expression in Escherichia coli and its use as a diagnostic antigen for serodiagnosis of Lyme borreliosis. In *Medical microbiology and immunology* 187 (4), pp. 213–219.
- Johnson, Russell C.; Schmid, George P.; Hyde, Fred W.; Steigerwalt, A. G.; Brenner, Don J. (1984): Borrelia burgdorferi sp. nov. Etiologic Agent of Lyme Disease. In *International journal of systematic bacteriology* 34 (4), pp. 496–497. DOI: 10.1099/00207713-34-4-496.
- Karlsson, Mats; Hovind-Hougen, Kari; Svenungsson, Bo; Stiernstedt, Göran (1990): Cultivation and characterization of spirochetes from cerebrospinal fluid of patients with Lyme borreliosis. In *Journal of Clinical Microbiology* 28 (3), pp. 473–479.
- Luther, Birgit; Moskopid, Matthäus (1990): Antigenic cross-reactivity between Borrelia burgdorferi, Borrelia recurrentis, Treponema pallidum, and Treponema phagedenis. In *Zentralblatt für Bakteriologie : international journal of medical microbiology* 274 (2), pp. 214–226.
- Magnarelli, Louis A.; Fikrig, Erol; Padula, Steven J.; Anderson, John F.; Flavell, Richard A. (1996): Use of recombinant antigens of Borrelia burgdorferi in serologic tests for diagnosis of lyme borreliosis. In *Journal of Clinical Microbiology* 34 (2), pp. 237–240.
- Ohnishi, Jun; Schneider, Brad; Messer, William B.; Piesman, Joseph; Silva, Aravinda M. de (2003): Genetic variation at the vlsE locus of Borrelia burgdorferi within ticks and mice over the course of a single transmission cycle. In *Journal of Bacteriology* 185 (15), pp. 4432–4441. DOI: 10.1128/JB.185.15.4432-4441.2003.
- Preac-Mursic, V.; Wilske, Bettina; Schierz, G. (1986): European Borrelia burgdorferi isolated from humans and ticks culture conditions and antibiotic susceptibility. In *Zentralblatt für Bakteriologie, Mikrobiologie, und Hygiene. Series A, Medical microbiology, infectious diseases, virology, parasitology* 263 (1-2), pp. 112–118.
- Schulte-Spechtel, Ulrike; Lehnert, Gisela; Liegl, Gaby; Fingerle, Volker; Heimerl, Christiane; Johnson, Barbara J. B.; Wilske, Bettina (2003): Significant improvement of the recombinant Borrelia-specific immunoglobulin G immunoblot test by addition of VlsE and a DbpA homologue derived from Borrelia garinii for diagnosis of early neuroborreliosis. In *Journal of Clinical Microbiology* 41 (3), pp. 1299–1303. DOI: 10.1128/JCM.41.3.1299-1303.2003.
- Wang, Dachun; Botkin, Douglas J.; Norris, Steven J. (2003): Characterization of the vls antigenic variation loci of the Lyme disease spirochaetes Borrelia garinii lp90 and Borrelia afzelii ACAI. In *Molecular Microbiology* 47 (5), pp. 1407–1417. DOI: 10.1046/j.1365-2958.2003.03386.x.
- Wilske, Bettina (2003): Diagnosis of lyme borreliosis in europe. In *Vector borne and zoonotic diseases (Larchmont, N.Y.)* 3 (4), pp. 215–227. DOI: 10.1089/153036603322662200.
- Wilske, Bettina; Preac-Mursic, V.; Fuchs, R.; Schierz, G. (1990): Diagnostik der Lyme Borreliose. Diagnose und Labor. In *Laboratoriumsblätter* 40, pp. 24–36.

## ABBREVIATIONS

CMIT	5-chloro-2-methyl-4-isothiazolin-3-one
MIT	2-methyl-2H-isothiazol-3-one

**SUMMARY OF TEST PROCEDURE****SCHEME OF THE ASSAY**

Borrelia burgdorferi IgG ELISA

**Test Preparation**











Prepare reagents and samples as described.  
 Establish the distribution and identification plan for all samples and standards/controls on the plate layout supplied in the kit.  
 Select the required number of microtiter strips or wells and insert them into the holder.

**Assay Procedure**

	Substrate Blank (A1)	Negative Control	Cut-off Control	Positive Control	Sample (diluted 1+100)
Negative Control	-	100 µL	-	-	-
Cut-off Control	-	-	100 µL	-	-
Positive Control	-	-	-	100 µL	-
Sample (diluted 1+100)	-	-	-	-	100 µL
Cover wells with foil supplied in the kit <b>Incubate for 1 h at 37 ± 1 °C</b> Wash each well three times with 300 µL of Washing Buffer					
Conjugate	-	100 µL	100 µL	100 µL	100 µL
<b>Incubate for 30 min at room temperature (20...25 °C)</b> Do not expose to direct sunlight Wash each well three times with 300 µL of Washing Buffer					
TMB Substrate Solution	100 µL	100 µL	100 µL	100 µL	100 µL
<b>Incubate for exactly 15 min at room temperature (20...25 °C) in the dark</b>					
Stop Solution	100 µL	100 µL	100 µL	100 µL	100 µL
Photometric measurement at 450 nm (reference wavelength: 620 nm)					



## SYMBOLS USED WITH DEMEDITEC ASSAYS

Symbol	English	Deutsch	Francais	Espanol	Italiano
	European Conformity	CE-Konformitäts-kennzeichnung	Conforme aux normes européennes	Conformidad europea	Conformità europea
	Consult instructions for use	Gebrauchsanweisung beachten	Consulter les instructions d'utilisation	Consulte las Instrucciones	Consultare le istruzioni per l'uso
	In vitro diagnostic device	In-vitro-Diagnostikum	Usage Diagnostic in vitro	Diagnóstico in vitro	Per uso Diagnostica in vitro
	For research use only	Nur für Forschungszwecke	Seulement dans le cadre de recherches	Sólo para uso en investigación	Solo a scopo di ricerca
	Catalogue number	Katalog-Nr.	Référence	Número de catálogo	No. di Cat.
	Lot. No. / Batch code	Chargen-Nr.	No. de lot	Número de lote	Lotto no
	Contains sufficient for <n> tests/	Ausreichend für "n" Ansätze	Contenu suffisant pour "n" tests	Contenido suficiente para <n> ensayos	Contenuto sufficiente per "n" saggi
	Note warnings and precautions	Warnhinweise und Vorsichtsmaßnahmen beachten	Avertissements et mesures de précaution font attention	Tiene en cuenta y advertencias precauciones	Annoti avvisi e le precauzioni
	Storage Temperature	Lagerungstemperatur	Temperature de conservation	Temperatura de conservacion	Temperatura di conservazione
	Expiration Date	Mindesthaltbarkeitsdatum	Date limite d'utilisation	Fecha de caducidad	Data di scadenza
	Legal Manufacturer	Hersteller	Fabricant	Fabricante	Fabbricante
<i>Distributed by</i>	Distributor	Vertreiber	Distributeur	Distribuidor	Distributore