HIMEDIA®
For Life is Precious

HiMedia Laboratories Pvt. Ltd.

Date: 07th March 2024.

**TO WHOMSOEVER IT MAY CONCERN**

We hereby certify that,

**Sanmedico SRL**
**Str. Corobceanu 7A, Apt.9,**
**MD-2012, CITY CHISINAU**
**Republic of Moldova,**
**Tel:-00-373-231 31515 / 00-373-222 60595**
**Fax:-00-373-22 62 30 32**
**E-mail: sanmedico.office@gmail.com**

have been appointed by us as our **Authorized Distributor** for selling our Products in **MOLDOVA**

***This certificate is valid upto 06th March 2026.***

This Authorization Letter shall stand effective from the date of signing and can be terminated by either party with two months advance notice.

**For HIMEDIA LABORATORIES PVT. LTD.,**

HiMedia Laboratories Pvt. Limited Thane 400604

**V.M.WARKE.**
**DIRECTOR – SALES & MARKETING**

Digitally signed by Dolgopol Iulian
Date: 2024.04.02 10:14:12 EEST
Reason: MoldSign Signature
Location: Moldova

THE INTERNATIONAL CERTIFICATION NETWORK

# CERTIFICATE

Quality Austria
has issued an IQNet recognized certificate that the organization:

## HiMedia Laboratories Pvt. Ltd.
## Plot NO. C40, ROAD - 21Y, WAGLE INDUSTRIAL ESTATE, THANE (WEST) - 400604 MAHARASHTRA, INDIA

for the following scope:

Design, Development & Testing of Microbiology, Animal Cell Culture, Plant Tissue Culture & Molecular Biology products

EAC: 34

has implemented and maintains a

## QUALITY MANAGEMENT SYSTEM

which fulfils the requirements of the following standard

## ISO 9001:2015

This attestation is directly linked to the IQNet Partner's original certificate and shall not be used as a stand-alone document

| | |
|---|---|
| Issued on: | 2022-02-28 |
| Validity date: | 2025-02-27 |
| Quality Austria certified since: | 2022-02-28 |

*Registration Number:* AT-27302/0

PARTNER OF IQNet

*Alex Stoichitoiu*
*President of IQNet*

*Mag. Friedrich Khuen-Belasi*
*Authorised Representative of Quality Austria*

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Succeed with Quality

IQNet Partners*:
AENOR *Spain* AFNOR Certification *France* APCER *Portugal* CCC *Cyprus* CISQ *Italy*
CQC *China* CQM *China* CQS *Czech Republic* Cro Cert *Croatia* DQS Holding GmbH *Germany* EAGLE Certification Group *USA*
FCAV *Brazil* FONDONORMA *Venezuela* ICONTEC *Colombia* Inspecta Sertifiointi Oy *Finland* INTECO *Costa Rica*
IRAM *Argentina* JQA *Japan* KFQ *Korea* MIRTEC *Greece* MSZT *Hungary* Nemko AS *Norway* NSAI *Ireland*
*NYCE*-SIGE *México* PCBC *Poland* Quality Austria *Austria* RR *Russia* SII *Israel* SIQ *Slovenia*
SIRIM QAS International *Malaysia* SQS *Switzerland* SRAC *Romania* TEST St Petersburg *Russia* TSE *Turkey* YUQS *Serbia*

* The list of IQNet partners is valid at the time of issue of this certificate. Updated information is available under www.iqnet-certification.com

qualityaustria
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# CERTIFICATE

Quality Austria - Trainings, Zertifizierungs und Begutachtungs GmbH awards this **quality**austria certificate to the following organisation:

This **quality**austria certificate confirms the application and further development of an effective

**HiMedia Laboratories Pvt. Ltd.**
Plot NO. C40, Road - 21Y, Wagle Industrial Estate,
Thane (West) - 400604 Maharashtra, INDIA

**QUALITY MANAGEMENT SYSTEM**
complying with the requirements of standard
**ISO 9001:2015**

Design, Development & Testing of Microbiology, Animal Cell Culture, Plant Tissue Culture & Molecular Biology products

Registration No.: 27302/0

Date of initial issue: 28 February 2022

Valid until: 27 February 2025

The validity of the **quality**austria certificate will be maintained by annual surveillance audits and one renewal audit after three years.

Vienna, 28 February 2022

Quality Austria - Trainings, Zertifizierungs und Begutachtungs GmbH,
AT-1010 Vienna, Zelinkagasse 10/3

Mag. Christoph Mondl
General Manager

Mag. Dr. Werner Paar
General Manager

Mag. Dr. Anni Koubek
Specialist representative



Dok. Nr. FO_24_028

1702280c-6c19-4683-8f36-3ea2e4167c18

The current validity of the certificate is documented exclusively on the Internet under
http://www.qualityaustria.com/en/cert EAC: 34

THE INTERNATIONAL CERTIFICATION NETWORK

# CERTIFICATE

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for the following scope:

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EAC: 34

has implemented and maintains a

## QUALITY MANAGEMENT SYSTEM

which fulfils the requirements of the following standard

## ISO 13485:2016

This attestation is directly linked to the IQNet Partner's original certificate and shall not be used as a stand-alone document

| | |
|---|---|
| Issued on: | 2022-02-28 |
| Validity date: | 2025-02-27 |
| Quality Austria certified since: | 2022-02-28 |

*Registration Number:* AT-00391/0

PARTNER OF IQNet

*Alex Stoichitoiu*
*President of IQNet*

*Mag. Friedrich Khuen-Belasi*
*Authorised Representative of Quality Austria*

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IQNet Partners*:
AENOR *Spain* AFNOR Certification *France* APCER *Portugal* CCC *Cyprus* CISQ *Italy*
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*NYCE*-SIGE *México* PCBC *Poland* Quality Austria *Austria* RR *Russia* SII *Israel* SIQ *Slovenia*
SIRIM QAS International *Malaysia* SQS *Switzerland* SRAC *Romania* TEST St Petersburg *Russia* TSE *Turkey* YUQS *Serbia*

* The list of IQNet partners is valid at the time of issue of this certificate. Updated information is available under www.iqnet-certification.com

qualityaustria
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# CERTIFICATE

Quality Austria - Trainings, Zertifizierungs und Begutachtungs GmbH awards this **quality**austria certificate to the following organisation:

This **quality**austria certificate confirms the application and further development of an effective

**QUALITY MANAGEMENT SYSTEM**
complying with the requirements of standard
**ISO 13485:2016**
Medical devices - Quality management systems - Requirements for regulatory purposes

## HiMedia Laboratories Pvt. Ltd.

Plot NO. C40, Road - 21Y, Wagle Industrial Estate,
Thane (West) - 400604 Maharashtra, INDIA

Design, Development & Testing of Biosciences Products for application in Microbiology, Animal Cell Culture & Molecular Biology products

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Mag. Dr. Anni Koubek
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Dok. Nr. FO_24_028

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The current validity of the certificate is documented exclusively on the Internet under
http://www.qualityaustria.com/en/cert EAC: 34

## DECLARATION OF CONFORMITY
## MICROBIOLOGY PRODUCTS

1) Manufacturer *(Name, department)*: **HiMedia Laboratories Pvt. Ltd.**

**Address: Plot No. C-40, Road No. 21/Y, MIDC, Wagle Industrial Area, Thane(West)-400604, Maharashtra, India**

and

2) European authorized representative: **CEpartner4U BV,**

**Address**: **ESDOORNLAAN 13, 3951DB MAARN, THE NETHERLANDS**;
(on product labels printed as:
CEpartner4U , ESDOORNLAAN 13, 3951DB MAARN, THE NETHERLANDS. www.cepartner4u.eu)

3) Product(s) *(groupnames /.)*:

| Group | Group name | NL registration no. | No. |
|---|---|---|---|
| DCM&S | Dehydrated Culture Media & Supplements | NL-CA002-2013-26442 | 1 |
| RPM | Ready Prepared Media<br>Subgroups: Ready Prepared Plates, Ready Prepared Liquid & Solid Medium, Ready Prepared Slants,Ready Prepared Dual Media, HiDip Slides, HiSafe Blood Culturing System, Transport Medium w/ swabs, Viral Transport Medium w/ swabs, L.J.Medium Slants & Kits, Biochemical Kits for Mycobacteria, UTI Diagnostic Kits, Biochemical Identification Kits | NL-CA002-2013-26448 | 2 |
| ESK | Epidemiological Screening Kit:<br>Subgroups: Hi Aureus Confirmation Kits | NL-CA002-2012-24117 | 3 |
| ASS | Antimicrobial Susceptibility Systems<br>Subgroups: Sensitivity Discs-Single & Multi Discs<br>MIC Strips: HiComb Strips, HiComb™ MIC Strip, Modified &<br>Ezy MIC Strips, HiMIC™ Plate Kit | NL-CA002-2013-26444 | 4 |
| BDA | Bacteriological Differentiation Aids<br>Subgroups: Readymade Stains, Indicators & Reagents in liquid, Differentiation Discs & Strips, HiDtect Rapid Identification Discs | NL-CA002-2013-26445 | 5 |

*type and model numbers: see appendix*

4) The product(s) described above is in conformity with:

| Title | Document No. |
|---|---|
| *In vitro* Diagnostic Medical Devices Directive | 98/79/EC |

5) Additional information *(Conformity procedure, Notified Body, CE certificate, Registration nr., etc.)*:
Conformity assessment procedure for CE marking: *In vitro* Diagnostic Medical Device Directive, Annex III

Mumbai, India; 2022-03-01

*(Place & date of issue (yyyy-mm-dd))*

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Dr. G.M.Warke , Managing Director

*(name; function and signature of manufacturer)*

# Appendix

Date: 2022-03-01

## List of devices:

| Product group | Type/ Model / Ref number | Device Name | Risk Class | Date of CE compliance |
|---|---|---|---|---|
| **Dehydrated Culture Media** | | | | |
| DCM | M1739 | A7 Agar Base (Shepard's Differential Agar Base) | Low risk | 20/12/2012 |
| DCM | MCD884 | Aeromonas Isolation HiCynth™ Medium Base | Low risk | 12/08/2015 |
| DCM | MV884 | Aeromonas Isolation HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M884 | Aeromonas Isolation Medium Base | Low risk | 20/12/2012 |
| DCM | M1284 | Aeromonas Starch DNA Agar Base | Low risk | 20/12/2012 |
| DCM | M016B | Agar Medium L (Brilliant Green, Phenol Red, Lactose Monohydrate, Sucrose Agar) | Low risk | 20/12/2012 |
| DCM | ME016 | Agar Medium L (Brilliant Green, Phenol Red, Lactose Monohydrate, Sucrose Agar) | Low risk | 20/12/2012 |
| DCM | MCD618 | Alkaline HiCynth™ Peptone Water | Low risk | 12/08/2015 |
| DCM | MV618 | Alkaline HiVeg™ Peptone Water | Low risk | 20/12/2012 |
| DCM | M618 | Alkaline Peptone Water | Low risk | 20/12/2012 |
| DCM | M1887 | Alkaline Saline Peptone Water (ASPW) | Low risk | 10/11/2020 |
| DCM | M651 | Amies Transport Medium w/ Charcoal | Low risk | 20/12/2012 |
| DCM | M684A | Amies Transport Medium, Liquid w/o charcoal | Low risk | 25/08/2016 |
| DCM | M228 | Anaerobic Agar | Low risk | 20/12/2012 |
| DCM | M491 | Anaerobic Agar (Brewer) | Low risk | 20/12/2012 |
| DCM | M230 | Anaerobic Agar w/o Dextrose | Low risk | 20/12/2012 |
| DCM | M229 | Anaerobic Agar w/o Dextrose and Eh Indicator | Low risk | 20/12/2012 |
| DCM | M1635 | Anaerobic Basal Agar | Low risk | 20/12/2012 |
| DCM | M1636 | Anaerobic Basal Broth | Low risk | 20/12/2012 |
| DCM | M1345 | Anaerobic Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M975A | Anaerobic Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M1034 | Anaerobic CNA Agar Base | Low risk | 20/12/2012 |
| DCM | MV228 | Anaerobic HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV491 | Anaerobic HiVeg™ Agar (Brewer) | Low risk | 20/12/2012 |
| DCM | MV230 | Anaerobic HiVeg™ Agar w/o Dextrose | Low risk | 20/12/2012 |
| DCM | MV229 | Anaerobic HiVeg™ Agar w/o Dextrose and Eh Indicator | Low risk | 20/12/2012 |
| DCM | MV909 | Andrade Peptone Water w/ HiVeg™ Extract No. 1 | Low risk | 20/12/2012 |
| DCM | M909 | Andrade Peptone Water w/ HM Extract | Low risk | 20/12/2012 |
| DCM | M1485 | Antibiotic Sulphonamide Sensitivity Test Agar (ASS Agar) | Low risk | 20/12/2012 |
| DCM | M1576 | Arabinose Agar Base | Low risk | 30/10/2018 |
| DCM | M1637 | Arcobacter Broth Base | Low risk | 10/11/2020 |
| DCM | M1894 | Arcobacter Selective Broth Base | Low risk | 10/11/2020 |

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|---|---|---|---|---|
| DCM | M672 | Asparagine Broth (Coccidioidin and Histoplasmin Broth) | Low risk | 20/12/2012 |
| DCM | M158 | Azide Blood Agar Base | Low risk | 20/12/2012 |
| DCM | MV158 | Azide Blood Agar Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M1271 | Azide Dextrose Broth w/ BCP | Low risk | 10/11/2020 |
| DCM | M220 | B.A.G.G. Broth Base (Buffered Azide Glucose Glycerol Broth Base) | Low risk | 20/12/2012 |
| DCM | MV220 | B.A.G.G. HiVeg™ Broth Base (Buffered Azide Glucose Glycerol HiVeg™ Broth Base) | Low risk | 20/12/2012 |
| DCM | M106 | B.C.G. - Dextrose Agar (Snyder Test Agar) | Low risk | 20/12/2012 |
| DCM | MV106 | B.C.G. - Dextrose HiVeg™ Agar (Snyder Test HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | MCD462 | B.Q.Vaccine HiCynth™ Medium (Thioglycollate HiCynth™ Broth) | Low risk | 28/04/2017 |
| DCM | MV462 | B.Q.Vaccine HiVeg™Medium (Thioglycollate Broth w/ HiVeg™ Extract No. 2) | Low risk | 20/12/2012 |
| DCM | M462 | B.Q.Vaccine Medium (Thioglycollate Broth w/ HL Extract) | Low risk | 20/12/2012 |
| DCM | M861 | B.T.B. Lactose Agar | Low risk | 20/12/2012 |
| DCM | MCD861 | B.T.B. Lactose HiCynth™ Agar | Low risk | 28/04/2017 |
| DCM | MCD1081 | B.T.B. Lactose HiCynth™ Agar, Modified | Low risk | 28/04/2017 |
| DCM | MV861 | B.T.B. Lactose HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV833 | Bacillus Cereus HiVeg™ Agar Base | Low risk | 22/04/2019 |
| DCM | M833 | Bacillus Cereus Agar Base | Low risk | 22/04/2019 |
| DCM | M805 | Bacteroides Bile Esculin Agar Base (BBE) | Low risk | 20/12/2012 |
| DCM | MV805 | Bacteroides HiVeg™ Agar Base (BBE) | Low risk | 20/12/2012 |
| DCM | M043 | Baird Parker Agar Base | Low risk | 20/12/2012 |
| DCM | M2093 | Baird Parker Agar Base w/o Egg Yolk Emulsion | Low risk | 22/04/2019 |
| DCM | MCD043 | Baird Parker HiCynth™ Agar Base | Low risk | 12/08/2015 |
| DCM | MV043 | Baird Parker HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1091 | Baird Staphylococcus Enrichment Broth Base | Low risk | 10/11/2020 |
| DCM | M694 | Bennet's Agar | Low risk | 20/12/2012 |
| DCM | M1683 | Bennet's Broth | Low risk | 20/12/2012 |
| DCM | MV694 | Bennet's HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1888 | BETA-SSA Agar (Group A Streptococci Selective Agar) | Low risk | 20/12/2012 |
| DCM | M211 | BHI Agar (Special Infusion Agar) | Low risk | 20/12/2012 |
| DCM | M211A | BHI Agar w/ 1% Agar | Low risk | 20/12/2012 |
| DCM | MV211A | BHI Agar w/ 1% Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M1069 | BHI Agar w/ 3.0% Agar | Low risk | 20/12/2012 |
| DCM | MV211 | BHI Agar, HiVeg™ (Special Infusion Agar, HiVeg™) | Low risk | 20/12/2012 |
| DCM | M1611 | BHI Agar, Modified | Low risk | 20/12/2012 |
| DCM | M210 | BHI Broth | Low risk | 20/12/2012 |
| DCM | M210I | BHI Broth | Low risk | 20/12/2012 |
| DCM | M209 | BHI CC Agar | Low risk | 20/12/2012 |
| DCM | MV209 | BHI CC Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MCD211 | BHI HiCynth™ Agar (Special Insusion HiCynth™ Agar) | Low risk | 12/08/2015 |

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|---|---|---|---|---|
| DCM | MCD210 | BHI HiCynth™ Broth | Low risk | 12/08/2015 |
| DCM | M1036 | BHI w/ 0.1% Agar | Low risk | 20/12/2012 |
| DCM | M1037 | BHI w/ 6.5% NaCl | Low risk | 20/12/2012 |
| DCM | MV1037 | BHI w/ 6.5% NaCl, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M212 | BHI w/ PABA | Low risk | 20/12/2012 |
| DCM | M213 | BHI w/ PABA and Agar | Low risk | 20/12/2012 |
| DCM | MV213 | BHI w/ PABA and Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MV212 | BHI w/ PABA, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MV1036 | BHI with 0.1% Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MV210 | BHI, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M217 | Bi.G.G.Y. Agar (Nickerson Medium) | Low risk | 20/12/2012 |
| DCM | MCD217 | Bi.G.G.Y. HiCynth™ Agar (Nickerson HiCynth™ Agar) | Low risk | 25/08/2016 |
| DCM | M1396 | Bifidobacterium Agar | Low risk | 10/11/2020 |
| DCM | M1960R | Bifidobacterium Agar (HiCrome™) | Low risk | 25/08/2016 |
| DCM | M1396R | Bifidobacterium Agar (Modified, Selective Medium, Kit) | Low risk | 04/07/2018 |
| DCM | M1858 | Bifidobacterium Agar, Modified | Low risk | 20/12/2012 |
| DCM | M1395 | Bifidobacterium Broth | Low risk | 10/11/2020 |
| DCM | M071 | Bile Broth Base | Low risk | 20/12/2012 |
| DCM | MV071 | Bile Broth Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M972A | Bile Esculin Agar,Modified | Low risk | 22/04/2019 |
| DCM | M493 | Bile Esculin Azide Agar | Low risk | 10/11/2020 |
| DCM | MV493 | Bile Esculin Azide HiVeg™ Agar | Low risk | 10/11/2020 |
| DCM | MCD493 | Bile Esculin Azide HiCynth™ Agar | Low risk | 10/11/2020 |
| DCM | M481 | Bile Peptone Transport Medium | Low risk | 20/12/2012 |
| DCM | M739 | Bile Salt Agar | Low risk | 20/12/2012 |
| DCM | MCD027 | Bismuth Sulphite HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | M027 | Bismuth Sulphite Agar | Low risk | 20/12/2012 |
| DCM | M027L | Bismuth Sulphite Agar | Low risk | 04/07/2018 |
| DCM | MU027 | Bismuth Sulphite Agar Medium | Low risk | 20/12/2012 |
| DCM | M1004 | Bismuth Sulphite Agar, Modified | Low risk | 20/12/2012 |
| DCM | MV027 | Bismuth Sulphite HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV1004 | Bismuth Sulphite HiVeg™ Agar, Modified | Low risk | 20/12/2012 |
| DCM | M073 | Blood Agar Base (Infusion Agar) | Low risk | 20/12/2012 |
| DCM | M834 | Blood Agar Base No. 2 | Low risk | 20/12/2012 |
| DCM | M834A | Blood Agar Base No. 2 w/ 1.2% Agar | Low risk | 20/12/2012 |
| DCM | MV834A | Blood Agar Base No. 2 w/ 1.2% Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MV834 | Blood Agar Base No. 2, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M834Z | Blood Agar Base No.2 | Low risk | 28/04/2017 |
| DCM | M089 | Blood Agar Base w/ Low pH | Low risk | 20/12/2012 |
| DCM | MV089 | Blood Agar Base w/ Low pH, HiVeg™ | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| DCM | M1904 | Blood Agar Base w/ Nalidixic Acid | Low risk | 20/12/2012 |
| DCM | MV073 | Blood Agar Base, HiVeg™ (Infusion Agar, HiVeg™) | Low risk | 20/12/2012 |
| DCM | M1989 | Blood Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M1318 | Blood Free Campylobacter Broth Base | Low risk | 20/12/2012 |
| DCM | MCD073 | Blood HiCynth™ Agar Base (Infusion HiCynth™ Agar Base) | Low risk | 25/08/2016 |
| DCM | MCD834 | Blood HiCynth™ Agar Base No.2 | Low risk | 25/08/2016 |
| DCM | MCD089 | Blood HiCynth™ Agar Base w/ Low pH | Low risk | 25/08/2016 |
| DCM | M175 | Bordet Gengou Agar Base | Low risk | 20/12/2012 |
| DCM | M175A | Bordet Gengou Agar Base w/ 1.6% Agar | Low risk | 20/12/2012 |
| DCM | M175SB | Bordet Gengou Agar Base, Modified | Low risk | 16/12/2017 |
| DCM | M2012 | Bordet Gengou Broth | Low risk | 25/08/2016 |
| DCM | MV175 | Bordet Gengou HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV175A | Bordet Gengou HiVeg™ Agar Base w/ 1.6% Agar | Low risk | 20/12/2012 |
| DCM | M1020 | BPL Agar | Low risk | 20/12/2012 |
| DCM | MV1020 | BPL HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M016A | Brilliant Green Agar Base w/ 1.2% Agar | Low risk | 20/12/2012 |
| DCM | M971 | Brilliant Green Agar Base w/ Phosphates | Low risk | 20/12/2012 |
| DCM | M016 | Brilliant Green Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | MCD016 | Brilliant Green Agar HiCynth™ Base, Modified | Low risk | 12/08/2015 |
| DCM | MU016 | Brilliant Green Agar Medium | Low risk | 20/12/2012 |
| DCM | MM016 | Brilliant Green Agar Medium 16 | Low risk | 20/12/2012 |
| DCM | MV016A | Brilliant Green HiVeg™ Agar Base w/ 1.2% Agar | Low risk | 20/12/2012 |
| DCM | MV971 | Brilliant Green HiVeg™ Agar Base w/ Phosphates | Low risk | 20/12/2012 |
| DCM | MV016 | Brilliant Green HiVeg™ Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M016B | Brilliant Green, Phenol Red, Lactose Monohydrate, Sucrose Agar (Agar Medium L) | Low risk | 20/12/2012 |
| DCM | ME016 | Brilliant Green, Phenol Red, Lactose Monohydrate, Sucrose Agar (Agar Medium L) | Low risk | 20/12/2012 |
| DCM | M1822 | Bromo Thymol Lactose Blue Agar | Low risk | 16/12/2017 |
| DCM | M074 | Brucella Agar Base | Low risk | 20/12/2012 |
| DCM | M1638 | Brucella Agar Base w/ 1.0% Dextrose | Low risk | 20/12/2012 |
| DCM | M1039 | Brucella Agar Base w/ Hemin and Vitamin K | Low risk | 20/12/2012 |
| DCM | M074A | Brucella Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M5392 | Brucella Broth Base | Low risk | 30/10/2018 |
| DCM | M348 | Brucella Broth Base | Low risk | 20/12/2012 |
| DCM | MV074 | Brucella HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV074A | Brucella HiVeg™ Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | MV348 | Brucella HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M822 | Brucella Selective Medium Base | Low risk | 20/12/2012 |
| DCM | M1890 | BSIBG Agar (Aeromonas Selective Agar) | Low risk | 10/11/2020 |
| DCM | M1668 | BSK - H Medium Base | Low risk | 20/12/2012 |

| | | | | |
|---|---|---|---|---|
| DCM | M1668B | BSK - H Medium Base w/o BSA | Low risk | 28/04/2017 |
| DCM | M813 | Buffered Charcoal Yeast Extract Agar Base | Low risk | 20/12/2012 |
| DCM | M813I | Buffered Charcoal Yeast Extract Agar Medium (BCYE Medium) | Low risk | 20/12/2012 |
| DCM | MCD813 | Buffered Charcoal Yeast Extract HiCynth™ Medium | Low risk | 25/08/2016 |
| DCM | M204 | Buffered Glycerol Saline Base | Low risk | 20/12/2012 |
| DCM | MCD1275 | Buffered HiCynth™ Peptone Water w/ NaCl | Low risk | 12/08/2015 |
| DCM | MV614 | Buffered HiVeg™ Peptone Water | Low risk | 22/04/2019 |
| DCM | MV1275 | Buffered HiVeg™ Peptone Water w/NaCL | Low risk | 20/12/2012 |
| DCM | M614 | Buffered Peptone Water | Low risk | 22/04/2019 |
| DCM | M1275 | Buffered Peptone Water w/ NaCl | Low risk | 20/12/2012 |
| DCM | M1851 | Buffered Peptone Water w/ Pyruvate | Low risk | 20/12/2012 |
| DCM | MH1275 | Buffered Sodium Chloride-Peptone Solution pH 7.0 | Low risk | 22/04/2019 |
| DCM | M1640 | Burkholderia Cepacia Agar Base | Low risk | 20/12/2012 |
| DCM | MCD1640 | Burkholderia cepacia HiCynth™ Agar Base | Low risk | 25/08/2016 |
| DCM | M2089 | Burkholderia Cepacia Selectie Agar | Low risk | 10/11/2020 |
| DCM | MU2089 | Burkholderia Cepacia Selective Agar (BCSA) | Low risk | 10/11/2020 |
| DCM | M470 | BYE Agar | Low risk | 20/12/2012 |
| DCM | MV470 | BYE HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M911 | C. botulinum Isolation Agar Base | Low risk | 20/12/2012 |
| DCM | MV911 | C. botulinum Isolation HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1146 | C.L.E.D. Agar Base w/o Indicator | Low risk | 20/12/2012 |
| DCM | M792 | C.L.E.D. Agar w/ Bromo Thymol Blue | Low risk | 20/12/2012 |
| DCM | MCD792 | C.L.E.D. HiCynth™ Agar w/BTB | Low risk | 12/08/2015 |
| DCM | MCD352 | C.L.E.D. HiCynth™ Agar w/Andrade Indicator | Low risk | 12/08/2015 |
| DCM | MV1146 | C.L.E.D. HiVeg™ Agar Base w/o Indicator | Low risk | 20/12/2012 |
| DCM | MV352 | C.L.E.D. HiVeg™ Agar w/ Andrade Indicator | Low risk | 20/12/2012 |
| DCM | MV792 | C.L.E.D. HiVeg™ Agar w/ Bromo Thymol Blue | Low risk | 20/12/2012 |
| DCM | M352 | C.L.E.D.Agar w/ Andrade Indicator | Low risk | 20/12/2012 |
| DCM | M352M | C.L.E.D.Agar w/ Andrades Indicator | Low risk | 22/04/2019 |
| DCM | M352A | C.L.E.D.Agar w/o Lactose & w/ Andrades Indicator | Low risk | 22/04/2019 |
| DCM | M563 | Caffeic Acid Ferric Citrate Test Agar (CAFC Medium) | Low risk | 20/12/2012 |
| DCM | M893 | CAL Agar (Cellobiose Arginine Lysine Agar) | Low risk | 20/12/2012 |
| DCM | M894 | CAL Broth (Cellobiose Arginine Lysine Broth) | Low risk | 20/12/2012 |
| DCM | MV893 | CAL HiVeg™ Agar (Cellobiose Arginine Lysine HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | MV894 | CAL HiVeg™ Broth (Cellobiose Arginine Lysine HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | MV908 | Campylo Thioglycollate HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M908 | Campylo Thioglycollate Medium Base | Low risk | 20/12/2012 |
| DCM | M994 | Campylobacter Agar Base | Low risk | 20/12/2012 |
| DCM | M1267 | Campylobacter Cefex Agar Base | Low risk | 20/12/2012 |
| DCM | M899 | Campylobacter Enrichment Broth Base (Preston Enrichment Broth Base) | Low risk | 20/12/2012 |

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| DCM | MV899 | Campylobacter Enrichment HiVeg™ Broth Base (Preston Enrichment HiVeg™ Broth Base) | Low risk | 20/12/2012 |
| DCM | MV994 | Campylobacter HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1240 | Campylobacter Nitrate Broth | Low risk | 20/12/2012 |
| DCM | MV1240 | Campylobacter Nitrate HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1602 | Candida Agar | Low risk | 20/12/2012 |
| DCM | M355 | Candida BCG Agar Base | Low risk | 20/12/2012 |
| DCM | MV355 | Candida BCG HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV104 | Candida HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M104 | Candida Medium | Low risk | 20/12/2012 |
| DCM | M202 | Cary - Blair Medium Base (Transport Medium w/o Charcoal) | Low risk | 20/12/2012 |
| DCM | M202A | Cary Blair Medium, Liquid w/o charcoal | Low risk | 25/08/2016 |
| DCM | M794 | Casitose Agar w/ 2.5% Agar | Low risk | 20/12/2012 |
| DCM | M200 | Casitose Broth | Low risk | 20/12/2012 |
| DCM | M910 | Casitose Yeast Extract Broth (CAYE) | Low risk | 20/12/2012 |
| DCM | MV910 | Casitose Yeast Extract HiVeg™ Broth (CAYE) | Low risk | 20/12/2012 |
| DCM | M201 | Casman Agar Base | Low risk | 20/12/2012 |
| DCM | M766 | Casman Broth Base | Low risk | 20/12/2012 |
| DCM | MV201 | Casman HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV766 | Casman HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | MH024 | Cetrimide Agar | Low risk | 22/04/2019 |
| DCM | M024 | Cetrimide Agar Base | Low risk | 20/12/2012 |
| DCM | M1742 | Cetrimide Agar Base (w 1.3% Agar) | Low risk | 20/12/2012 |
| DCM | M862 | Cetrimide Broth | Low risk | 20/12/2012 |
| DCM | MCD024 | Cetrimide HiCynth™ Agar Base | Low risk | 12/08/2015 |
| DCM | MV024 | Cetrimide HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV862 | Cetrimide HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M344 | Charcoal Agar Base | Low risk | 10/11/2020 |
| DCM | MV344 | Charcoal Agar Base, HiVeg™ | Low risk | 10/11/2020 |
| DCM | M1053 | Charcoal Agar Base with Niacin | Low risk | 16/12/2017 |
| DCM | M646 | Charcoal Blood Agar Base | Low risk | 10/11/2020 |
| DCM | MV646 | Charcoal Blood Agar Base, HiVeg™ | Low risk | 10/11/2020 |
| DCM | M103 | Chocolate Agar Base | Low risk | 20/12/2012 |
| DCM | MV103 | Chocolate HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1548 | Chocolate No. 2 Agar Base | Low risk | 20/12/2012 |
| DCM | MV1548 | Chocolate No. 2 HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV558 | Cholera HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M558 | Cholera Medium Base | Low risk | 20/12/2012 |
| DCM | M143 | Christensen Citrate Agar | Low risk | 20/12/2012 |
| DCM | M1820 | Chrysoidin Agar with MUG | Low risk | 16/12/2017 |

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| DCM | M497 | Clostridial Agar | Low risk | 20/12/2012 |
| DCM | MV497 | Clostridial HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M836 | Clostridium Difficile Agar Base | Low risk | 20/12/2012 |
| DCM | MV836 | Clostridium Difficile HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1976 | Clostridium difficile Mannitol Taurocholate Broth base (CCMB -TAL Broth) | Low risk | 20/12/2012 |
| DCM | M272 | Coagulase Mannitol Agar Base | Low risk | 20/12/2012 |
| DCM | M277 | Coagulase Mannitol Broth Base | Low risk | 20/12/2012 |
| DCM | MV272 | Coagulase Mannitol HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV277 | Coagulase Mannitol HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1826 | Coliform Broth w/SLS | Low risk | 22/04/2019 |
| DCM | MV1826 | Coliform HiVeg Broth w/ SLS | Low risk | 22/04/2019 |
| DCM | MH144 | Columbia Agar | Low risk | 22/04/2019 |
| DCM | M144M | Columbia Agar | Low risk | 22/04/2019 |
| DCM | M144PM | Columbia Blood Agar Base | Low risk | 22/04/2019 |
| DCM | M144R | Columbia Blood Agar Base | Low risk | 25/08/2016 |
| DCM | M144 | Columbia Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M144A | Columbia Blood Agar Base w/ 1% Agar | Low risk | 20/12/2012 |
| DCM | MV144A | Columbia Blood Agar Base w/ 1% Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M1133 | Columbia Blood Agar Base w/ Hemin | Low risk | 20/12/2012 |
| DCM | MV144 | Columbia Blood Agar Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MCD144 | Columbia Blood HiCynth™ Agar Base | Low risk | 12/08/2015 |
| DCM | MCD144A | Columbia Blood HiCynth™ Agar Base w/1% Agar | Low risk | 12/08/2015 |
| DCM | M145 | Columbia Broth Base | Low risk | 20/12/2012 |
| DCM | MV145 | Columbia Broth Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M560 | Columbia C.N.A. Agar Base | Low risk | 20/12/2012 |
| DCM | M560A | Columbia C.N.A. Agar Base w/ 1% Agar | Low risk | 20/12/2012 |
| DCM | MV560 | Columbia C.N.A. HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV560A | Columbia C.N.A. HiVeg™ Agar Base w/ 1% Agar | Low risk | 20/12/2012 |
| DCM | MCD145 | Columbia HiCynth™ Broth | Low risk | 12/08/2015 |
| DCM | M2103 | Congo Red Magnesium Oxalate (CR-MOX) Agar | Low risk | 22/04/2019 |
| DCM | M730 | Conn's Agar | Low risk | 20/12/2012 |
| DCM | M149 | Cooked M Medium (R.C .Medium) | Low risk | 16/12/2017 |
| DCM | M1040 | Cooked M Medium w/ Glucose, Hemin & Vitamin K | Low risk | 16/12/2017 |
| DCM | MV731 | Corn Meal HiVeg™ Peptone Yeast Agar | Low risk | 20/12/2012 |
| DCM | M731 | Corn Meal Peptone Yeast Agar | Low risk | 20/12/2012 |
| DCM | M897 | Crystal Violet Lactose Agar | Low risk | 10/11/2020 |
| DCM | MV897 | Crystal Violet Lactose HiVeg™ Agar | Low risk | 10/11/2020 |
| DCM | M1892 | CTAS Agar Base (Carnobacterium Selective Agar Base) | Low risk | 20/12/2012 |
| DCM | M172 | Cystine H Agar Base | Low risk | 20/12/2012 |

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| DCM | MV172 | Cystine HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M881 | Cystine Tellurite Agar Base | Low risk | 20/12/2012 |
| DCM | M160 | D.C.L.S. Agar | Low risk | 20/12/2012 |
| DCM | M178 | D.C.L.S. Agar, Hajna | Low risk | 20/12/2012 |
| DCM | MV160 | D.C.L.S. HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV178 | D.C.L.S. HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M188 | D.T.M. Agar Base (Dermatophyte Test Agar Base) | Low risk | 20/12/2012 |
| DCM | M501 | Decarboxylase Agar Base | Low risk | 20/12/2012 |
| DCM | M393 | Decarboxylase Broth Base, Moeller (Moeller Decarboxylase Broth Base) | Low risk | 20/12/2012 |
| DCM | MV501 | Decarboxylase HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV393 | Decarboxylase HiVeg™ Broth Base, Moeller (Moeller Decarboxylase HiVeg™ Broth Base) | Low risk | 20/12/2012 |
| DCM | M030 | Deoxycholate Agar | Low risk | 20/12/2012 |
| DCM | MV030 | Deoxycholate Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M065 | Deoxycholate Citrate Agar | Low risk | 20/12/2012 |
| DCM | M1639 | Deoxycholate Citrate Agar w/1.5% Agar | Low risk | 20/12/2012 |
| DCM | M222 | Deoxycholate Citrate Agar w/o Sucrose | Low risk | 20/12/2012 |
| DCM | MV065 | Deoxycholate Citrate Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MCD065 | Deoxycholate Citrate HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | M084 | Dextrose Agar | Low risk | 20/12/2012 |
| DCM | M286 | Dextrose Agar Base, Emmons (Sabouraud Dextrose Agar Base, Modified) | Low risk | 20/12/2012 |
| DCM | MV084 | Dextrose HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV286 | Dextrose HiVeg™ Agar Base, Emmons (Sabouraud Dextrose HiVeg™ AgarBase, Modified) | Low risk | 20/12/2012 |
| DCM | M734 | Dextrose Proteose Peptone Agar Base | Low risk | 20/12/2012 |
| DCM | MV734 | Dextrose Proteose Peptone HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M502 | Diagnostic Sensitivity Test Agar (D.S.T. Agar) | Low risk | 20/12/2012 |
| DCM | M111 | Diagnostic Stuart's Urea Broth Base (Urea Broth Base) | Low risk | 20/12/2012 |
| DCM | MV191 | Diagnostic Thioglycollate HiVeg™ Medium (Thioglycollate HiVeg™ Medium w/o Indicator) | Low risk | 20/12/2012 |
| DCM | M191 | Diagnostic Thioglycollate Medium (Thioglycollate Medium w/o Indicator) | Low risk | 20/12/2012 |
| DCM | M1129 | Dichloran Glycerol Medium Base | Low risk | 22/04/2019 |
| DCM | M1049 | Differential Agar for Group D Streptococci | Low risk | 10/11/2020 |
| DCM | M814 | Differential Buffered Charcoal Yeast Extract Agar Base | Low risk | 20/12/2012 |
| DCM | M1603 | Differential Reinforced Clostridial Agar | Low risk | 10/11/2020 |
| DCM | M915 | Dihydrolase Broth Base | Low risk | 20/12/2012 |
| DCM | MV915 | Dihydrolase HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | MM1276 | Dilute Sautans Medium (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M882 | Diphtheria Virulence Agar Base | Low risk | 25/08/2016 |
| DCM | M882R | Diphtheria Virulence Agar Base Modified | Low risk | 25/08/2016 |

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| DCM | MV882 | Diphtheria Virulence HiVeg™ Agar Base | Low risk | 25/08/2016 |
| DCM | M1984 | Dixon's Agar | Low risk | 20/12/2012 |
| DCM | M1419 | DNase Test Agar w/ Methyl Green | Low risk | 10/11/2020 |
| DCM | M057 | Double Sugar Agar, Russell (Russell Double Sugar Agar) | Low risk | 20/12/2012 |
| DCM | MV057 | Double Sugar HiVeg™ Agar (Russell Double Sugar HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | M916 | Doyle's Enrichment Broth Base | Low risk | 20/12/2012 |
| DCM | MV916 | Doyle's Enrichment HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1378 | Drigalski Lactose Agar, Modified | Low risk | 20/12/2012 |
| DCM | M1761 | Drigalski Lactose Selective Agar | Low risk | 20/12/2012 |
| DCM | M659 | Drigalski Litmus Lactose Agar | Low risk | 20/12/2012 |
| DCM | MV659 | Drigalski Litmus Lactose HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M5349 | DTP Medium | Low risk | 30/10/2018 |
| DCM | M067 | Dubos Broth Base | Low risk | 20/12/2012 |
| DCM | MV067 | Dubos HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M179 | Dubos Oleic Agar Base | Low risk | 20/12/2012 |
| DCM | M839 | Dubos Oleic Broth Base | Low risk | 20/12/2012 |
| DCM | MV179 | Dubos Oleic HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV839 | Dubos Oleic HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1536 | Dulcitol Selenite Broth (Selenite-F Broth w/ Dulcitol) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M854 | E.T. Medium | Low risk | 20/12/2012 |
| DCM | M1768 | EC Blue Broth | Low risk | 20/12/2012 |
| DCM | MV1768 | EC Blue HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M127 | EC Broth | Low risk | 20/12/2012 |
| DCM | M127I | EC Broth | Low risk | 20/12/2012 |
| DCM | MV127 | EC HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M748 | Edward's Medium Base, Modified | Low risk | 20/12/2012 |
| DCM | MV748 | Edward's Medium HiVeg™ Base, Modified | Low risk | 20/12/2012 |
| DCM | M294 | Edwards and Bruner Semisolid Medium | Low risk | 20/12/2012 |
| DCM | M808 | Egg Yolk Agar Base | Low risk | 20/12/2012 |
| DCM | MV808 | Egg Yolk Agar Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M1043 | Egg Yolk Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M086 | Eijkman Lactose Broth | Low risk | 20/12/2012 |
| DCM | MV086 | Eijkman Lactose HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M368 | Elliker Broth (Lactobacilli Broth) | Low risk | 10/11/2020 |
| DCM | MV368 | Elliker HiVeg™ Broth (Lactobacilli HiVeg™ Broth) | Low risk | 10/11/2020 |
| DCM | M317 | EMB Agar | Low risk | 20/12/2012 |
| DCM | M301 | EMB Agar Base | Low risk | 20/12/2012 |
| DCM | M022 | EMB Agar, Levine | Low risk | 20/12/2012 |
| DCM | M022S | EMB Agar, Levine | Low risk | 20/12/2012 |
| DCM | M503 | EMB Broth | Low risk | 20/12/2012 |

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| DCM | MV317 | EMB HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV022 | EMB HiVeg™ Agar, Levine | Low risk | 20/12/2012 |
| DCM | MV503 | EMB HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M325 | Emerson Agar | Low risk | 20/12/2012 |
| DCM | MV325 | Emerson HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M773 | Emerson YSS Agar | Low risk | 20/12/2012 |
| DCM | M029 | Endo Agar | Low risk | 20/12/2012 |
| DCM | M1077 | Endo Agar Base | Low risk | 20/12/2012 |
| DCM | M1258 | Endo Agar w/ NaCl | Low risk | 20/12/2012 |
| DCM | M1075 | Endo Agar, Modified | Low risk | 20/12/2012 |
| DCM | M029R | Endo Agar, Special | Low risk | 25/08/2016 |
| DCM | MCD029 | Endo HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | MV029 | Endo HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV1077 | Endo HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV1258 | Endo HiVeg™ Agar w/ NaCl | Low risk | 20/12/2012 |
| DCM | MV1075 | Endo HiVeg™ Agar, Modified | Low risk | 20/12/2012 |
| DCM | M738 | Enriched Thioglycollate Broth | Low risk | 20/12/2012 |
| DCM | MV738 | Enriched Thioglycollate HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | MV077 | Entamoeba HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M077 | Entamoeba Medium | Low risk | 20/12/2012 |
| DCM | M1662 | Enteric Fermentation Base | Low risk | 20/12/2012 |
| DCM | MH287 | Enterobacteria Enrichment Broth, Mossel | Low risk | 22/04/2019 |
| DCM | M426 | Ethyl Violet Azide Broth (E.V.A. Broth) | Low risk | 20/12/2012 |
| DCM | M426S | Ethyl Violet Azide Broth (E.V.A. Broth) | Low risk | 20/12/2012 |
| DCM | M1397 | Ethyl Violet Azide Dextrose Agar | Low risk | 20/12/2012 |
| DCM | MV426 | Ethyl Violet Azide HiVeg™ Broth (E.V.A. HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M428 | Eugonic Agar | Low risk | 20/12/2012 |
| DCM | M429 | Eugonic Broth | Low risk | 20/12/2012 |
| DCM | MV428 | Eugonic HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV429 | Eugonic HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1517 | Eugonic LT 100 Broth Base w/o Tween 80 | Low risk | 20/12/2012 |
| DCM | M1517Z | Eugonic LT 100 Broth Base w/o Tween 80 | Low risk | 17/06/2021 |
| DCM | M811 | Feeley Gorman Agar (F.G. Agar) | Low risk | 20/12/2012 |
| DCM | M812 | Feeley Gorman Broth (F.G. Broth) | Low risk | 20/12/2012 |
| DCM | MV811 | Feeley Gorman HiVeg™ Agar (F.G. HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | MV812 | Feeley Gorman HiVeg™ Broth (F.G. HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M827 | Fermentation Medium for Staphylococcus and Micrococcus | Low risk | 20/12/2012 |
| DCM | MV919 | Fermentation HiVeg™ Medium Base for C. perfringens | Low risk | 20/12/2012 |
| DCM | MV825 | Fermentation HiVeg™ Medium for Neisseriae | Low risk | 20/12/2012 |
| DCM | MV827 | Fermentation HiVeg™ Medium for Staphylococcus and Micrococcus | Low risk | 20/12/2012 |

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| DCM | M919 | Fermentation Medium Base for C. perfringens | Low risk | 20/12/2012 |
| DCM | M825 | Fermentation Medium for Neisseriae | Low risk | 20/12/2012 |
| DCM | M1028 | Field's Tryptic Digest Broth (Tryptic Digest Broth) | Low risk | 20/12/2012 |
| DCM | MV1028 | Field's Tryptic digest Broth, HiVeg™ (Tryptic Digest Broth, HiVeg™) | Low risk | 20/12/2012 |
| DCM | MV239 | Fletcher Leptospira HiVeg™ Medium Base (Leptospira HiVeg™ MediumBase, Fletcher) | Low risk | 20/12/2012 |
| DCM | M239 | Fletcher Leptospira Medium Base (Leptospira Medium Base, Fletcher) | Low risk | 20/12/2012 |
| DCM | M1209 | Fluconazole Testing Medium (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV013 | Fluid Sabouraud HiVeg™ Medium (Sabouraud Medium, Fluid, HiVeg™ ) | Low risk | 20/12/2012 |
| DCM | M013 | Fluid Sabouraud Medium (Sabouraud Medium, Fluid) | Low risk | 20/12/2012 |
| DCM | M1533I | Fluid Selenite Cystine Broth (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV025 | Fluid Selenite Cystine HiVeg™ Medium (Selenite Cystine HiVeg™ Broth) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M025 | Fluid Selenite Cystine Medium (Selenite Cystine Broth) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MM025 | Fluid Selenite Cystine Medium (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MU025 | Fluid Selenite Cystine Medium (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MCD032 | Fluid Tetrathionate HiCynth™ Medium w/o Iodine and BG | Low risk | 25/08/2016 |
| DCM | MV032 | Fluid Tetrathionate HiVeg™ Medium w/o Iodine and BG (Tetrathionate HiVeg™ Broth Base w/o Iodine & BG) | Low risk | 20/12/2012 |
| DCM | M032 | Fluid Tetrathionate Medium w/o Iodine and BG (Tetrathionate Broth Base w/o Iodine and BG) | Low risk | 20/12/2012 |
| DCM | MV009 | Fluid Thioglycollate HiVeg™ Medium | Low risk | 22/04/2019 |
| DCM | M009 | Fluid Thioglycollate medium (Thioglycollate medium Fluid) | Low risk | 22/04/2019 |
| DCM | M543 | Folic Acid Casei Medium | Low risk | 20/12/2012 |
| DCM | M2014 | Folic Acid Casei Medium, Modified | Low risk | 25/08/2016 |
| DCM | M1050 | Frey Mycoplasma Broth Base | Low risk | 20/12/2012 |
| DCM | M475 | Fungobiotic Agar (Mycobio Agar) | Low risk | 10/11/2020 |
| DCM | M476 | Garrod Actinomyces Medium | Low risk | 10/11/2020 |
| DCM | M1073 | GBS Medium Base | Low risk | 28/04/2017 |
| DCM | M434 | GC Agar Base | Low risk | 25/08/2016 |
| DCM | MV434 | GC HiVeg™ Agar Base | Low risk | 04/07/2018 |
| DCM | M5397 | Gifu Anaerobic Broth w/o starch & dextrose | Low risk | 22/04/2019 |
| DCM | M2079 | Gifu Anaerobic Broth, Modified (GAM) | Low risk | 04/07/2018 |
| DCM | M1746 | Glucose Agar | Low risk | 10/11/2020 |
| DCM | M435 | Glucose Citrate Broth Base | Low risk | 20/12/2012 |
| DCM | M433 | Glucose Cysteine Agar Base w/ Thiamine | Low risk | 20/12/2012 |
| DCM | MV433 | Glucose Cysteine HiVeg™ Agar Base w/ Thiamine | Low risk | 20/12/2012 |
| DCM | M070 | Glucose Phosphate Broth (Buffered Glucose Broth) | Low risk | 20/12/2012 |
| DCM | MV070 | Glucose Phosphate HiVeg™ Broth (Buffered Glucose HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M621 | Glucose Salt Teepol Broth (Twin Pack) | Low risk | 20/12/2012 |

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| DCM | MV621 | Glucose Salt Teepol HiVeg™ Broth (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1935 | Glycerol Mannitol Acetamide Cetrimide Agar | Low risk | 20/12/2012 |
| DCM | M242 | GN Broth, Hajna | Low risk | 20/12/2012 |
| DCM | MV242 | GN HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1888 | Group A Streptococci Selective Agar (BETA-SSA Agar) | Low risk | 20/12/2012 |
| DCM | M1607 | Gum Listeria Medium | Low risk | 20/12/2012 |
| DCM | M243 | H Broth | Low risk | 20/12/2012 |
| DCM | MV116 | H.S. Vaccine HiVeg™ Medium (Standard Nutrient HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M116 | H.S. Vaccine Medium (Standard Nutrient Broth) | Low risk | 20/12/2012 |
| DCM | M1259 | Haemophilus Test Agar Base | Low risk | 20/12/2012 |
| DCM | M551 | Hartley's Digest Broth | Low risk | 20/12/2012 |
| DCM | MV551 | Hartley's Digest HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M467 | Hektoen Enteric Agar | Low risk | 20/12/2012 |
| DCM | MU467 | Hektoen Enteric Agar Medium | Low risk | 20/12/2012 |
| DCM | MCD467 | Hektoen Enteric HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | MV467 | Hektoen Enteric HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M5390 | Helicobacter Pylori Selective Agar | Low risk | 30/10/2018 |
| DCM | M1158 | Hemorrhagic Coli (HC) Agar | Low risk | 20/12/2012 |
| DCM | M169 | HI Agar | Low risk | 20/12/2012 |
| DCM | MV169 | HI Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M170 | HI Broth | Low risk | 20/12/2012 |
| DCM | MV170 | HI Broth, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M1938 | HiCrome™ Acinetobacter Agar Base | Low risk | 20/12/2012 |
| DCM | M1651 | HiCrome™ Bacillus Agar | Low risk | 25/08/2016 |
| DCM | MCD1651 | HiCrome™ Bacillus HiCynth™ Agar | Low risk | 25/08/2016 |
| DCM | M1960 | HiCrome™ Bifidobacterium Agar | Low risk | 20/12/2012 |
| DCM | M1456AR | HiCrome™ Candida Differential Agar,Modified | Low risk | 25/08/2016 |
| DCM | MCD1297A | HiCrome™ Candida Differential HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | M1832 | HiCrome Coliform Agar Modified | Low risk | 22/04/2019 |
| DCM | MV1300 | HiCrome Coliform HiVeg Agar w/ SLS | Low risk | 22/04/2019 |
| DCM | MV1295 | HiCrome E. coli HiVegTM Agar | Low risk | 22/04/2019 |
| DCM | MV1293 | HiCrome ECC HiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | MV1294 | HiCrome ECC Selective HiVeg Agar Base | Low risk | 22/04/2019 |
| DCM | M1598 | HiCrome Enrichment Broth Base for EC O157:H7 | Low risk | 22/04/2019 |
| DCM | M1577 | HiCrome™ Enterobacter sakazakii Agar | Low risk | 22/04/2019 |
| DCM | M1641 | HiCrome Enterobacter sakazakii Agar, Modified | Low risk | 22/04/2019 |
| DCM | MV1577 | HiCrome Enterobacter sakazakii HiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | MV1641 | HiCrome Enterobacter sakazakii HiVeg™ Agar, Modified | Low risk | 22/04/2019 |
| DCM | M1580 | HiCrome™ Enterococcus faecium Agar Base | Low risk | 25/08/2016 |
| DCM | MCD1466 | HiCrome™ Improved Salmonella HiCynth™ Agar | Low risk | 12/08/2015 |

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| DCM | M1569 | HiCrome M-Lauryl Sulphate Agar | Low risk | 22/04/2019 |
| DCM | M1862 | HiCrome M-Modified ECO157:H7 Selective Agar Base | Low risk | 22/04/2019 |
| DCM | M1571 | HiCrome M-TEC Agar | Low risk | 22/04/2019 |
| DCM | M1713 | HiCrome M-TEC Broth | Low risk | 22/04/2019 |
| DCM | M1985 | HiCrome™ Malassezia Agar | Low risk | 20/12/2012 |
| DCM | M1953R | HiCrome™ MeReSa Agar Base (Modified) | Low risk | 25/08/2016 |
| DCM | M1953 | HiCrome™ MeReSa Agar Base (Modified) | Low risk | 25/08/2016 |
| DCM | M2010 | HiCrome™ Mueller Hinton Agar | Low risk | 25/08/2016 |
| DCM | M1974 | HiCrome™ Rapid MRSA Agar Base | Low risk | 20/12/2012 |
| DCM | M1842 | HiCrome Selective Salmonella Agar Base | Low risk | 22/04/2019 |
| DCM | M1353R | HiCrome™ UTI Agar | Low risk | 25/08/2016 |
| DCM | MCD1353 | HiCrome™ UTI HiCynth™™ Agar | Low risk | 12/08/2015 |
| DCM | MV1353R | HiCrome™ UTI HiVeg™ Agar | Low risk | 25/08/2016 |
| DCM | MV1682 | HiCrome Vibrio HiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | M2114 | HiCrome™ C.auris (MDR) Selective Agar Base | Low risk | 10/11/2020 |
| DCM | M2020 | HiCrome™ Campylobacter Agar Base | Low risk | 16/12/2017 |
| DCM | M1297A | HiCrome™ Candida Differential Agar | Low risk | 20/12/2012 |
| DCM | M1297AR | HiCrome™ Candida Differential Agar Base | Low risk | 20/12/2012 |
| DCM | M1456A | HiCrome™ Candida Differential Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | MV1297A | HiCrome™ Candida Differential HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV1456A | HiCrome™ Candida Differential HiVeg™ Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M2099 | HiCrome™ CarbaResist Agar Base | Low risk | 22/04/2019 |
| DCM | M1991I | HiCrome™ Chromogenic Coliform Agar (CCA) | Low risk | 22/04/2019 |
| DCM | M2026 | HiCrome™ Clostridial Agar Base | Low risk | 25/08/2016 |
| DCM | M1300 | HiCrome™ Coliform Agar w/ SLS | Low risk | 22/04/2019 |
| DCM | MCD1300 | HiCrome™ Coliform HiCynth™ Agar w/ SLS | Low risk | 10/11/2020 |
| DCM | M2094 | HiCrome™ Colistin Resistant Agar Base | Low risk | 30/10/2018 |
| DCM | M2062I | HiCrome™ Cronobacter Isolation Agar (CCI Agar) | Low risk | 10/11/2020 |
| DCM | M1295 | HiCrome™ E. coli Agar | Low risk | 22/04/2019 |
| DCM | M1295I | HiCrome™ E. coli Agar | Low risk | 22/04/2019 |
| DCM | MCD1295 | HiCrome™ E.coli HiCynth™ Agar | Low risk | 22/04/2019 |
| DCM | MCD1580 | HiCrome™ E.faecium HiCynth™Agar Base | Low risk | 25/08/2016 |
| DCM | M1575A | HiCrome™ EC O157 : H7 Selective Agar Base, Modified | Low risk | 10/11/2020 |
| DCM | MV1575A | HiCrome™ EC O157 : H7 Selective HiVeg™ Agar Base, Modified | Low risk | 10/11/2020 |
| DCM | MCD1575A | HiCrome™ EC O157:H7 HiCynth™ Agar Base, Modified | Low risk | 10/11/2020 |
| DCM | M1574A | HiCrome™ EC O157:H7 Agar,Modified | Low risk | 22/04/2019 |
| DCM | M1293 | HiCrome™ ECC Agar | Low risk | 22/04/2019 |
| DCM | M1294 | HiCrome™ ECC Selective Agar Base | Low risk | 22/04/2019 |
| DCM | M2056 | HiCrome™ ECC Selective Agar Base, Modified | Low risk | 22/04/2019 |
| DCM | M1488 | HiCrome™ ECD Agar w/ MUG | Low risk | 10/11/2020 |

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| DCM | MV1488 | HiCrome™ ECD HiVeg™ Agar w/ MUG | Low risk | 10/11/2020 |
| DCM | MCD1598 | HiCrome™ Enrichment HiCynth™ Broth Base for ECO157:H7 | Low risk | 10/11/2020 |
| DCM | MCD1641 | HiCrome™ Enterobacter sakazakii HiCynth™ Agar, Modified (HiCrome™ Cronobacter sakazakii HiCynth™ Agar, Modified) | Low risk | 10/11/2020 |
| DCM | M1376 | HiCrome™ Enterococci Broth | Low risk | 10/11/2020 |
| DCM | MCD1376 | HiCrome™ Enterococci HiCynth™ Broth | Low risk | 10/11/2020 |
| DCM | MV1376 | HiCrome™ Enterococci HiVeg™ Broth | Low risk | 10/11/2020 |
| DCM | MV1580 | HiCrome™ Enterococcus faecium HiVeg™ Agar Base | Low risk | 10/11/2020 |
| DCM | M1829 | HiCrome™ ESBL Agar Base | Low risk | 20/12/2012 |
| DCM | M2128 | HiCrome™ Haemophilus Agar Base | Low risk | 17/06/2021 |
| DCM | M1466 | HiCrome™ Improved Salmonella Agar | Low risk | 20/12/2012 |
| DCM | MV1466 | HiCrome™ Improved Salmonella HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1573 | HiCrome™ Klebsiella Selective Agar Base | Low risk | 10/11/2020 |
| DCM | MV1573 | HiCrome™ Klebsiella Selective HiVeg™ Agar Base | Low risk | 10/11/2020 |
| DCM | M1831 | HiCrome™ KPC Agar Base | Low risk | 20/12/2012 |
| DCM | M2009 | HiCrome™ L mono differential Agar Base | Low risk | 10/11/2020 |
| DCM | M1924 | HiCrome™ L.mono Rapid Differential Agar Base | Low risk | 10/11/2020 |
| DCM | M2065 | HiCrome™ Lactobacillus Selective Agar Base | Low risk | 10/11/2020 |
| DCM | M1417F | HiCrome™ Listeria Agar Base | Low risk | 10/11/2020 |
| DCM | M1417 | HiCrome™ Listeria Agar Base, Modified | Low risk | 10/11/2020 |
| DCM | MCD1417 | HiCrome™ Listeria HiCynth™ Agar Base, Modified | Low risk | 10/11/2020 |
| DCM | M1340 | HiCrome™ MacConkey Sorbitol Agar Base | Low risk | 20/12/2012 |
| DCM | MCD1340 | HiCrome™ MacConkey Sorbitol HiCynth™ Agar | Low risk | 25/08/2016 |
| DCM | M2058 | HiCrome™ M-Coliconfirm Agar Base | Low risk | 10/11/2020 |
| DCM | M2064 | HiCrome™ M-Coliconfirm Broth Base | Low risk | 22/04/2019 |
| DCM | M1674 | HiCrome™ MeReSa Agar Base | Low risk | 20/12/2012 |
| DCM | MCD1674 | HiCrome™ MeReSa HiCynth™ Agar Base | Low risk | 25/08/2016 |
| DCM | MV1674 | HiCrome™ MeReSa HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1393 | HiCrome™ MM Agar | Low risk | 20/12/2012 |
| DCM | M1816 | HiCrome™ MM Agar , Modified | Low risk | 04/07/2018 |
| DCM | M1816R | HiCrome™ MM Agar , Modified | Low risk | 04/07/2018 |
| DCM | MCD1816 | HiCrome™ MM HiCynth™ Agar, Modified (Hicrome™ Miller and Mallinson HiCynth™ Agar) | Low risk | 10/11/2020 |
| DCM | MV1393 | HiCrome™ MM HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MCD1571 | HiCrome™ M-TEC HiCynth™ Agar | Low risk | 10/11/2020 |
| DCM | MCD1713 | HiCrome™ M-TEC HiCynth™ Broth | Low risk | 10/11/2020 |
| DCM | M2067 | HiCrome™ Mueller Hinton Agar (for antifungal) | Low risk | 16/12/2017 |
| DCM | M1712 | HiCrome™ Nickels and Leesment Medium | Low risk | 10/11/2020 |
| DCM | MV1712 | HiCrome™ Nickels & Leesment HiVeg™ Agar Base | Low risk | 10/11/2020 |
| DCM | MCD1633 | HiCrome™ RajHans HiCynth™ Medium (Salmonella HiCynth™ Agar) | Low risk | 25/08/2016 |

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| DCM | M1633 | HiCrome™ RajHans Medium (Salmonella Agar) | Low risk | 20/12/2012 |
| DCM | M1634 | HiCrome™ RajHans Medium, Modified (Salmonella Agar, Modified) | Low risk | 20/12/2012 |
| DCM | M2011 | HiCrome™ Rapid ECC Broth | Low risk | 22/04/2019 |
| DCM | MCD1974 | HiCrome™ Rapid MRSA HiCynth™ Agar Base | Low risk | 25/08/2016 |
| DCM | M2116 | HiCrome™ Salmoconfirm Selective Agar | Low risk | 10/11/2020 |
| DCM | M1296 | HiCrome™ Salmonella Agar | Low risk | 20/12/2012 |
| DCM | MV1296 | HiCrome™ Salmonella HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MCD1842 | HiCrome™ Selective Salmonella HiCynth™ Agar Base | Low risk | 10/11/2020 |
| DCM | M1837 | HiCrome™ Staph Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M1931 | HiCrome™ Staph Selective Agar | Low risk | 10/11/2020 |
| DCM | M2092 | HiCrome™ STEC Agar Base | Low risk | 30/10/2018 |
| DCM | M1840 | HiCrome™ Strep B Selective Agar Base | Low risk | 04/07/2018 |
| DCM | M1966 | HiCrome™ Strep B Selective Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | MCD1840 | HiCrome™ Strep B Selective HiCynth™Agar Base | Low risk | 04/07/2018 |
| DCM | M1600 | HiCrome™ Universal Differential Medium | Low risk | 20/12/2012 |
| DCM | MCD1418 | HiCrome™ UTI HiCynth™ Agar, Modified | Low risk | 25/08/2016 |
| DCM | M1353 | HiCrome™ UTI Agar | Low risk | 20/12/2012 |
| DCM | M1418 | HiCrome™ UTI Agar, Modified | Low risk | 20/12/2012 |
| DCM | MV1353 | HiCrome™ UTI HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV1418 | HiCrome™ UTI HiVeg™ Agar, Modified | Low risk | 20/12/2012 |
| DCM | M1505 | HiCrome™ UTI Selective Agar | Low risk | 20/12/2012 |
| DCM | MV1505 | HiCrome™ UTI Selective HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1682 | HiCrome™ Vibrio Agar | Low risk | 22/04/2019 |
| DCM | MCD1682 | HiCrome™ Vibrio HiCynth™ Agar | Low risk | 10/11/2020 |
| DCM | M1830 | HiCrome™ VRE Agar Base | Low risk | 20/12/2012 |
| DCM | M1925 | HiCrome™ VRE Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M2025 | HiCrome™ Yersinia Agar Base | Low risk | 25/08/2016 |
| DCM | M1951 | HiCrome™M-Coliform Differential Agar Base | Low risk | 22/04/2019 |
| DCM | M2048 | HiFast™ Listeria Enrichment Broth Base | Low risk | 10/11/2020 |
| DCM | M1469 | HiFluoro Pseudomonas Agar Base | Low risk | 20/12/2012 |
| DCM | MV1469 | HiFluoro Pseudomonas HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M2126 | HiMRSA™ Confirmation Agar Base | Low risk | 18/06/2021 |
| DCM | M1218 | High Salt Nutrient Agar | Low risk | 20/12/2012 |
| DCM | M1219 | High Salt Peptone Yeast Extract Agar | Low risk | 20/12/2012 |
| DCM | M1054 | Hippurate Hydrolysis Broth | Low risk | 20/12/2012 |
| DCM | M485 | Hi-Sensitivity Test Agar | Low risk | 20/12/2012 |
| DCM | M486 | Hi-Sensitivity Test Broth | Low risk | 20/12/2012 |
| DCM | MV485 | Hi-Sensitivity Test HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV486 | Hi-Sensitivity Test HiVeg™ Broth | Low risk | 20/12/2012 |

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| DCM | M485A | HiSitest Agar | Low risk | 20/12/2012 |
| DCM | MV806 | HiVeg™ Extract Agar | Low risk | 20/12/2012 |
| DCM | MV807 | HiVeg™ Extract Broth | Low risk | 20/12/2012 |
| DCM | MV028 | HiVeg™ Peptone Water | Low risk | 20/12/2012 |
| DCM | M806 | HM Peptone B Agar | Low risk | 20/12/2012 |
| DCM | M807 | HM Peptone B Broth | Low risk | 20/12/2012 |
| DCM | M924 | Horie Arabinose Ethyl Violet Broth | Low risk | 20/12/2012 |
| DCM | M5385 | Horse Blood agar | Low risk | 30/10/2018 |
| DCM | M1425 | Hottinger Broth | Low risk | 20/12/2012 |
| DCM | MV015 | Hoyle HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M015 | Hoyle Medium Base | Low risk | 20/12/2012 |
| DCM | MV871 | Hugh Leifson Glucose HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M871 | Hugh Leifson Glucose Medium | Low risk | 20/12/2012 |
| DCM | MV826 | Hugh Leifson HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M826 | Hugh Leifson Medium | Low risk | 20/12/2012 |
| DCM | M826S | Hugh Leifson Medium | Low risk | 20/12/2012 |
| DCM | MV364 | Indole Nitrate HiVeg™ Medium (Tryptone Nitrate HiVeg™ Medium) | Low risk | 20/12/2012 |
| DCM | M364 | Indole Nitrate Medium (Tryptone Nitrate Medium) | Low risk | 20/12/2012 |
| DCM | M574 | Inositol Brilliant Green Bile Agar (Plesiomonas Differential Agar) | Low risk | 20/12/2012 |
| DCM | MV574 | Inositol Brilliant Green HiVeg™ Agar (Plesiomonas Differential HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | M1222 | Karmali Campylobacter Agar Base | Low risk | 10/11/2020 |
| DCM | M248 | KF Streptococcal Agar Base | Low risk | 22/04/2019 |
| DCM | M249 | KF Streptococcal Broth Base | Low risk | 20/12/2012 |
| DCM | MV248 | KF Streptococcal HiVeg Agar Base | Low risk | 22/04/2019 |
| DCM | MV249 | KF Streptococcal HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1007 | KF Streptococcus Agar Base w/ BCP | Low risk | 20/12/2012 |
| DCM | M1021 | KF Streptococcus Broth Base w/ BCP | Low risk | 20/12/2012 |
| DCM | MV1021 | KF Streptococcus HiVeg™ Broth Base w/ BCP | Low risk | 20/12/2012 |
| DCM | M1232 | Kimmig Fungi Agar Base | Low risk | 20/12/2012 |
| DCM | MV1232 | Kimmig Fungi HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1543 | King's Medium A Base | Low risk | 20/12/2012 |
| DCM | M1235 | King's OF Medium Base | Low risk | 20/12/2012 |
| DCM | MV1235 | Kings OF Medium Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M2040 | Kirchner Medium Base | Low risk | 28/04/2017 |
| DCM | M161 | Kirchner Medium Base, Modified | Low risk | 20/12/2012 |
| DCM | M078 | Kligler Iron Agar | Low risk | 20/12/2012 |
| DCM | M078I | Kligler Iron Agar | Low risk | 20/12/2012 |
| DCM | M078A | Kligler Iron Agar, Modified | Low risk | 20/12/2012 |
| DCM | MCD078 | Kligler Iron HiCynth™ Agar | Low risk | 12/08/2015 |

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| DCM | MV078 | Kligler Iron HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV142 | Kohn Two Tube HiVeg™ Medium No.1 Base | Low risk | 20/12/2012 |
| DCM | MV802 | Kohn Two Tube HiVeg™ Medium No.2 | Low risk | 20/12/2012 |
| DCM | M142 | Kohn Two Tube Medium No.1 Base | Low risk | 20/12/2012 |
| DCM | M802 | Kohn Two Tube Medium No.2 | Low risk | 20/12/2012 |
| DCM | M069 | Koser Citrate Medium | Low risk | 20/12/2012 |
| DCM | MV171 | Kracke Blood Culture HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M171 | Kracke Blood Culture Medium | Low risk | 20/12/2012 |
| DCM | M305 | Kupferberg Trichomonas Broth Base (Trichomonas Broth Base, Kupferberg) | Low risk | 20/12/2012 |
| DCM | MV305 | Kupferberg Trichomonas HiVeg™ Broth Base (Trichomonas HiVeg™ Broth Base, Kupferberg) | Low risk | 20/12/2012 |
| DCM | M928 | L Broth | Low risk | 20/12/2012 |
| DCM | M1312 | L Broth, Modified | Low risk | 20/12/2012 |
| DCM | M162R | L J Medium Base, Modified (Lowenstein Jensen Medium Base, Modified) | Low risk | 25/08/2016 |
| DCM | M1552 | L. mono Confirmatory Agar Base | Low risk | 20/12/2012 |
| DCM | MV1552 | L. mono Confirmatory HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M742 | L.D. Agar | Low risk | 20/12/2012 |
| DCM | M744 | L.D. Egg Yolk Agar Base | Low risk | 20/12/2012 |
| DCM | M743 | L.D. Esculin Agar | Low risk | 20/12/2012 |
| DCM | MV743 | L.D. Esculin HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV742 | L.D. HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1540 | L.mono Differential Agar Base | Low risk | 22/04/2019 |
| DCM | M1540I | HiCrome™ Listeria Ottaviani-Agosti Agar Base | Low risk | 10/11/2020 |
| DCM | M1540IR | L.mono Differential Agar Base | Low risk | 10/11/2020 |
| DCM | MCD1540 | L.mono Differential HiCynth™ Agar Base | Low risk | 22/04/2019 |
| DCM | MV1540 | L.mono Differential HiVeg™ Agar Base | Low risk | 22/04/2019 |
| DCM | M926 | Lactic Streak Agar (Reddy's Differential Agar, Modified) | Low risk | 20/12/2012 |
| DCM | MV926 | Lactic Streak HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV368 | Lactobacilli HiVeg™Broth (Elliker HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M927 | Lactobacillus Bulgaricus Agar Base | Low risk | 20/12/2012 |
| DCM | MV927 | Lactobacillus Bulgaricus HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M641 | Lactobacillus MRS Agar (MRS Agar) | Low risk | 20/12/2012 |
| DCM | M641I | Lactobacillus MRS Agar (MRS Agar) | Low risk | 20/12/2012 |
| DCM | M369 | Lactobacillus MRS Broth (MRS Broth) | Low risk | 20/12/2012 |
| DCM | MV641 | Lactobacillus MRS HiVeg™ Agar (MRS HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | MV369 | Lactobacillus MRS HiVeg™ Broth (MRS HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M1165 | Lactobacillus Selection Bile Agar Base (LBS Bile Agar) | Low risk | 20/12/2012 |
| DCM | M1081 | Lactose Blue Agar (B.T.B. Lactose Agar, Modified) | Low risk | 20/12/2012 |
| DCM | MV1081 | Lactose Blue HiVeg™ Agar (B.T.B. Lactose HiVeg™ Agar, Modified) | Low risk | 20/12/2012 |

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| DCM | M1003 | Lactose Broth | Low risk | 22/04/2019 |
| DCM | MV1003 | Lactose HiVeg™ Broth | Low risk | 22/04/2019 |
| DCM | M1047 | Lactose Lecithin Agar | Low risk | 04/07/2018 |
| DCM | M080 | Lauryl Sulphate Broth (Lauryl Tryptose Broth) | Low risk | 22/04/2019 |
| DCM | MV080 | Lauryl SulphateHiVeg™ Broth (Lauryl Tryptose HiVeg™ Broth) | Low risk | 22/04/2019 |
| DCM | M180 | Lead Acetate Agar | Low risk | 10/11/2020 |
| DCM | M1839 | Leeds Acinetobacter Agar Base | Low risk | 20/12/2012 |
| DCM | M1938R | Leeds Acinetobacter Agar Base (HiCrome™ Acinetobacter Agar Base) | Low risk | 25/08/2016 |
| DCM | M1845 | Legionella Agar Base w/o Charcoal | Low risk | 10/11/2020 |
| DCM | M1380 | Leifson Agar | Low risk | 20/12/2012 |
| DCM | MV1380 | Leifson HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1138 | Leifson's Deoxycholate Agar, Modified | Low risk | 20/12/2012 |
| DCM | MV1138 | Leifson's Deoxycholate HiVeg™ Agar, Modified | Low risk | 20/12/2012 |
| DCM | MV239 | Leptospira HiVeg™ Medium Base, Fletcher (Fletcher Leptospira HiVeg™ Medium Base) | Low risk | 20/12/2012 |
| DCM | MV457 | Leptospira HiVeg™ Medium Base, Korthof, Modified | Low risk | 20/12/2012 |
| DCM | M1009 | Leptospira Medium Base | Low risk | 20/12/2012 |
| DCM | M239 | Leptospira Medium Base, Fletcher (Fletcher Leptospira Medium Base) | Low risk | 20/12/2012 |
| DCM | M457 | Leptospira Medium Base, Korthof, Modified | Low risk | 20/12/2012 |
| DCM | MV472 | Levinthal's HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M472 | Levinthal's Medium Base | Low risk | 20/12/2012 |
| DCM | M374 | LI Agar | Low risk | 20/12/2012 |
| DCM | MV374 | LI Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M153 | LI Broth | Low risk | 20/12/2012 |
| DCM | MV153 | LI Broth, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M627 | Lipovitellin Salt Mannitol Agar Base | Low risk | 20/12/2012 |
| DCM | M817 | Liquoid Broth | Low risk | 20/12/2012 |
| DCM | MV817 | Liquoid HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M569 | Listeria Enrichment Broth (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV569 | Listeria Enrichment HiVeg™ Broth (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV890A | Listeria Enrichment HiVeg™ Medium Base (UVM) | Low risk | 20/12/2012 |
| DCM | M890A | Listeria Enrichment Medium Base (UVM) | Low risk | 20/12/2012 |
| DCM | M1064 | Listeria Identification Agar Base (PALCAM) | Low risk | 22/04/2019 |
| DCM | M1090 | Listeria Identification Broth Base (PALCAM) | Low risk | 22/04/2019 |
| DCM | MV1064 | Listeria Identification HiVeg Agar Base (PALCAM) | Low risk | 22/04/2019 |
| DCM | MV1090 | Listeria Identification HiVeg Broth Base (PALCAM) | Low risk | 22/04/2019 |
| DCM | MCD1145 | Listeria Oxford HiCynth™ Medium Base | Low risk | 25/08/2016 |
| DCM | MV1145 | Listeria Oxford HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M1145R | Listeria Oxford Medium Base | Low risk | 25/08/2016 |

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| DCM | M1145 | Listeria Oxford Medium Base | Low risk | 20/12/2012 |
| DCM | M1781 | Listeria Oxford Medium Base, Modified | Low risk | 20/12/2012 |
| DCM | M567 | Listeria Selective Agar (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1474 | Listeria Selective Agar Base | Low risk | 20/12/2012 |
| DCM | M889 | Listeria Selective Broth Base | Low risk | 20/12/2012 |
| DCM | M1865 | Listeria Selective Enrichment Broth | Low risk | 22/04/2019 |
| DCM | MV567 | Listeria Selective HiVeg™ Agar (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV889 | Listeria Selective HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M507 | Litmus Lactose Bile Salt Agar (LLBSA) | Low risk | 10/11/2020 |
| DCM | MV507 | Litmus Lactose HiVeg™ Agar | Low risk | 10/11/2020 |
| DCM | M373 | Littman Bile Agar Base | Low risk | 20/12/2012 |
| DCM | M663 | Littman Bile Broth Base | Low risk | 20/12/2012 |
| DCM | MV373 | Littman HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV663 | Littman HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1001 | LM Agar | Low risk | 20/12/2012 |
| DCM | M1934 | LM Agar, Modified | Low risk | 20/12/2012 |
| DCM | MV537 | Loeffler HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M537 | Loeffler Medium Base | Low risk | 20/12/2012 |
| DCM | M1189 | Loeffler Serum Medium Base | Low risk | 20/12/2012 |
| DCM | MM162 | Lowenstein - Jensen Medium (L.J. Medium) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M162R | Lowenstein Jensen Medium Base, Modified (L J Medium Base, Modified) | Low risk | 25/08/2016 |
| DCM | M162 | Lowenstein Jensen Medium Base (L.J. Medium) | Low risk | 20/12/2012 |
| DCM | M1542 | Lowenstein Jensen Medium Base w/o Starch | Low risk | 20/12/2012 |
| DCM | M2032 | Lowenstein Jensen Medium Base, Modified | Low risk | 25/08/2016 |
| DCM | M176 | LV Agar (Liver Veal Agar) | Low risk | 10/11/2020 |
| DCM | M1977 | Lysine Indole Motility Medium, Modified | Low risk | 10/11/2020 |
| DCM | MH081 | MacConkey Agar | Low risk | 22/04/2019 |
| DCM | M1024 | MacConkey Agar Base | Low risk | 20/12/2012 |
| DCM | M1819 | MacConkey Agar II w/o CV | Low risk | 20/12/2012 |
| DCM | M008E | MacConkey Agar Medium | Low risk | 20/12/2012 |
| DCM | M081 | MacConkey Agar w/ 0.15% Bile Salts, CV and NaCl | Low risk | 20/12/2012 |
| DCM | M061 | MacConkey Agar w/ Bromo Thymol Blue | Low risk | 20/12/2012 |
| DCM | M1582 | MacConkey Agar w/ CV and w/o NaCl | Low risk | 20/12/2012 |
| DCM | M081A | MacConkey Agar w/ CV, NaCl, and 0.15% Bile Salts | Low risk | 20/12/2012 |
| DCM | M008 | MacConkey Agar w/o CV w/ 0.15% Bile Salts | Low risk | 20/12/2012 |
| DCM | M082A | MacConkey Agar w/o CV, NaCl w/ 0.5% Bile Salts | Low risk | 20/12/2012 |
| DCM | M082 | MacConkey Agar w/o CV, NaCl w/ 0.5% Sodium Taurocholate | Low risk | 20/12/2012 |
| DCM | M008A | MacConkey Agar w/o CV, w/ 0.5% Bile Salts | Low risk | 20/12/2012 |
| DCM | M008B | MacConkey Agar w/o CV, w/ 1.2% Agar | Low risk | 20/12/2012 |

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| DCM | M1785 | MacConkey Agar w/o CV, w/0.5% Sodium Taurocholate | Low risk | 20/12/2012 |
| DCM | M1702 | MacConkey Agar, RS | Low risk | 20/12/2012 |
| DCM | MH083 | MacConkey Broth | Low risk | 22/04/2019 |
| DCM | M083 | MacConkey Broth Purple w/BCP | Low risk | 22/04/2019 |
| DCM | MCD081 | MacConkey HiCynth™ Agar w/ 0.15% Bile Salts | Low risk | 25/08/2016 |
| DCM | MCD082 | MacConkey HiCynth™ Agar w/o CV, NaCl | Low risk | 25/08/2016 |
| DCM | MV1024 | MacConkey HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV061 | MacConkey HiVeg™ Agar w/ Bromo Thymol Blue | Low risk | 20/12/2012 |
| DCM | MV081 | MacConkey HiVeg™ Agar w/ CV, NaCl, 0.003% NR and 1.5% Agar | Low risk | 20/12/2012 |
| DCM | MV081A | MacConkey HiVeg™ Agar w/ CV, NaCl, 0.005% NR and 1.5% Agar | Low risk | 20/12/2012 |
| DCM | MV082 | MacConkey HiVeg™ Agar w/o CV and NaCl, w/ 0.004% NR and 2.0% Agar | Low risk | 20/12/2012 |
| DCM | MV082A | MacConkey HiVeg™ Agar w/o CV and NaCl, w/ 0.0075% NR and 1.2% Agar | Low risk | 20/12/2012 |
| DCM | MV008B | MacConkey HiVeg™ Agar w/o CV, w/ 0.003% NR and 1.2% Agar | Low risk | 20/12/2012 |
| DCM | MV008 | MacConkey HiVeg™ Agar w/o CV, w/ 0.003% NR and 1.5% Agar | Low risk | 20/12/2012 |
| DCM | MV008A | MacConkey HiVeg™ Agar w/o CV, w/ 0.0075% NR and 1.2% Agar | Low risk | 20/12/2012 |
| DCM | MV083 | MacConkey HiVeg™ Broth Purple w/ BCP | Low risk | 22/04/2019 |
| DCM | M298 | MacConkey Sorbitol Agar (Sorbitol Agar) | Low risk | 20/12/2012 |
| DCM | M298I | MacConkey Sorbitol Agar Base | Low risk | 20/12/2012 |
| DCM | M1727R | MacConkey Sorbitol Agar Base (w/ Rhamnose) | Low risk | 25/08/2016 |
| DCM | M1727 | MacConkey Sorbitol Agar Base w/ Rhamnose | Low risk | 20/12/2012 |
| DCM | MCD298 | MacConkey Sorbitol HiCynth™ Agar ( Sorbitol HiCynth™ Agar) | Low risk | 28/04/2017 |
| DCM | MV298 | MacConkey Sorbitol HiVeg™ Agar (Sorbitol HiVeg™Agar) | Low risk | 20/12/2012 |
| DCM | M2074 | MacConkey Sorbitol Rhamnose Selective Agar Base | Low risk | 16/12/2017 |
| DCM | M382 | Malonate Broth | Low risk | 25/08/2016 |
| DCM | M137 | Malt Extract Agar Base (w/ Mycological Peptone) | Low risk | 20/12/2012 |
| DCM | M995 | Malt Extract Agar Base, Modified as per Thom and Church | Low risk | 20/12/2012 |
| DCM | M255 | Malt Extract Broth Base | Low risk | 20/12/2012 |
| DCM | M1128 | Malt Extract Broth, Modified as per Thom and Church | Low risk | 20/12/2012 |
| DCM | MV137 | Malt Extract HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV995 | Malt Extract HiVeg™ Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | MV255 | Malt Extract HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1967 | Malt Yeast Agar | Low risk | 20/12/2012 |
| DCM | M1624 | Mannitol Agar w/Prilion | Low risk | 20/12/2012 |
| DCM | M1071 | Mannitol Lysine Agar | Low risk | 20/12/2012 |
| DCM | MCD1071 | Mannitol Lysine HiCynth™Agar | Low risk | 25/08/2016 |
| DCM | M1320 | Mannitol Motility Nitrate Medium | Low risk | 20/12/2012 |
| DCM | MV770 | Mannitol Motility Test HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M770 | Mannitol Motility Test Medium | Low risk | 20/12/2012 |
| DCM | MH118 | Mannitol Salt Agar | Low risk | 22/04/2019 |

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| DCM | M118 | Mannitol Salt Agar Base | Low risk | 20/12/2012 |
| DCM | M383 | Mannitol Salt Broth | Low risk | 20/12/2012 |
| DCM | MCD118 | Mannitol Salt HiCynth™ Agar Base | Low risk | 12/08/2015 |
| DCM | MV118 | Mannitol Salt HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV383 | Mannitol Salt HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1534 | Mannitol Selenite Broth (Selenite Mannitol Broth) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1537 | Mannitol Selenite Broth w/Brilliant Green (Twin Pack) | Low risk | 04/07/2018 |
| DCM | MV379 | Marine Oxidation Fermentation HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M379 | Marine Oxidation Fermentation Medium | Low risk | 20/12/2012 |
| DCM | M2085 | Martin Lewis Agar Base | Low risk | 22/04/2019 |
| DCM | M1030 | Maximum Recovery Diluent | Low risk | 22/04/2019 |
| DCM | MV1030 | Maximum Recovery Diluent HiVeg™ | Low risk | 22/04/2019 |
| DCM | M386 | McBride Listeria Agar Base | Low risk | 20/12/2012 |
| DCM | MV386 | McBride Listeria HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1354 | M-CP Agar Base | Low risk | 10/11/2020 |
| DCM | MV1354 | M-CP HiVeg™Agar Base | Low risk | 10/11/2020 |
| DCM | M1426 | M-E.coli Broth | Low risk | 22/04/2019 |
| DCM | M1594 | MeReSa Agar Base | Low risk | 20/12/2012 |
| DCM | M1974R | MeReSa Agar Base (HiCrome™ Rapid MRSA Agar) | Low risk | 25/08/2016 |
| DCM | M1812 | M-FC Basal Medium | Low risk | 10/11/2020 |
| DCM | M199 | Middlebrook 7H10 Agar Base | Low risk | 20/12/2012 |
| DCM | M196 | Middlebrook 7H10 Agar Base, Special | Low risk | 20/12/2012 |
| DCM | M511 | Middlebrook 7H11 Agar Base | Low risk | 20/12/2012 |
| DCM | M511A | Middlebrook 7H11 Agar Base w/o Malachite Green | Low risk | 20/12/2012 |
| DCM | MV511 | Middlebrook 7H11 HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M197 | Middlebrook 7H9 Agar Base | Low risk | 20/12/2012 |
| DCM | M198 | Middlebrook 7H9 Broth Base | Low risk | 20/12/2012 |
| DCM | M259 | Mitis Salivarius Agar Base | Low risk | 20/12/2012 |
| DCM | MV259 | Mitis Salivarius HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M5319 | Modified B.Q. Vaccine Medium | Low risk | 28/04/2017 |
| DCM | M1150 | Modified Bile Esculin Azide Agar | Low risk | 20/12/2012 |
| DCM | M892 | Modified Buffered Charcoal Agar Base | Low risk | 20/12/2012 |
| DCM | MV892 | Modified Buffered Charcoal HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1660 | Modified Cary-Blair Medium | Low risk | 20/12/2012 |
| DCM | MV460 | Modified CPLM HiVeg™ Medium Base (Trichomonas Modified CPLM HiVeg™ Medium Base) | Low risk | 20/12/2012 |
| DCM | M460 | Modified CPLM Medium Base (Trichomonas Modified CPLM Medium Base) | Low risk | 20/12/2012 |
| DCM | M1170 | Modified Czapek Dox Agar | Low risk | 20/12/2012 |
| DCM | M1285 | Modified EC Broth Base | Low risk | 20/12/2012 |
| DCM | M1445 | Modified Lactobacillus Agar | Low risk | 20/12/2012 |

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| DCM | M1643 | Modified Lauryl Sulphate Tryptose Broth Base | Low risk | 20/12/2012 |
| DCM | M1457R | Modified Listeria Lecithinase Agar Base | Low risk | 25/08/2016 |
| DCM | M1897 | Modified Listeria Oxford Agar Base | Low risk | 25/11/2017 |
| DCM | M891 | Modified McBride Listeria Agar Base | Low risk | 20/12/2012 |
| DCM | MV891 | Modified McBride Listeria HiVeg™™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1139 | Modified MYP Agar Base | Low risk | 20/12/2012 |
| DCM | MV1139 | Modified MYP HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1606 | Modified Protease Agar | Low risk | 20/12/2012 |
| DCM | M1681 | Modified Sabourauds Chloramphenicol Agar | Low risk | 20/12/2012 |
| DCM | M1068 | Modified Salt Broth | Low risk | 20/12/2012 |
| DCM | M2049 | Modified Shieh Agar (LMG Medium 215) | Low risk | 28/04/2017 |
| DCM | M1286I | Modified Soyabean Bile Broth Base | Low risk | 22/04/2019 |
| DCM | M795 | Modified Thayer Martin Medium Base (w/o Supplement) | Low risk | 20/12/2012 |
| DCM | M393 | Moeller Decarboxylase Broth Base (Decarboxylase Broth Base, Moeller) | Low risk | 25/08/2016 |
| DCM | MCD393 | Moeller Decarboxylase HiCynth™ Broth Bas | Low risk | 25/08/2016 |
| DCM | M246 | Mold Inhibitory Agar, Ulrich | Low risk | 20/12/2012 |
| DCM | M474 | Monsur Medium Base | Low risk | 20/12/2012 |
| DCM | M1927 | MRS Agar w/ Low pH | Low risk | 10/11/2020 |
| DCM | M1864 | MSM Broth Base | Low risk | 20/12/2012 |
| DCM | M173 | Mueller Hinton Agar | Low risk | 20/12/2012 |
| DCM | M1825 | Mueller Hinton Agar 2% Glucose w/ Methylene blue | Low risk | 20/12/2012 |
| DCM | M1825R | Mueller Hinton Agar Modified (As per CLSI) | Low risk | 25/08/2016 |
| DCM | M1084 | Mueller Hinton Agar No. 2 | Low risk | 20/12/2012 |
| DCM | M5389 | Mueller Hinton Agar w/ 2% NaCL | Low risk | 30/10/2018 |
| DCM | M391 | Mueller Hinton Broth | Low risk | 20/12/2012 |
| DCM | M1657 | Mueller Hinton Broth No. 2 Control Cations | Low risk | 20/12/2012 |
| DCM | MV173 | Mueller Hinton HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV1084 | Mueller Hinton HiVeg™ Agar No. 2 | Low risk | 20/12/2012 |
| DCM | MV391 | Mueller Hinton HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1202 | Mueller Tellurite Agar Base | Low risk | 20/12/2012 |
| DCM | M1373 | MUG EC O157 Agar | Low risk | 16/12/2017 |
| DCM | M1429 | MUG EC O157 Agar, Modified | Low risk | 20/12/2012 |
| DCM | M1080 | MUG MacConkey Agar | Low risk | 20/12/2012 |
| DCM | MV1080 | MUG MacConkey HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1205 | MUG Sorbitol Agar | Low risk | 20/12/2012 |
| DCM | M977 | Mutans-Sanguis Agar | Low risk | 20/12/2012 |
| DCM | M094 | Mycological Agar | Low risk | 20/12/2012 |
| DCM | M095 | Mycological Agar w/ Low pH | Low risk | 20/12/2012 |
| DCM | M1422 | Mycological Agar, Modified | Low risk | 20/12/2012 |

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| DCM | M264 | Mycological Broth | Low risk | 20/12/2012 |
| DCM | M265 | Mycological Broth w/ Low pH | Low risk | 20/12/2012 |
| DCM | M266 | Mycoplasma Agar Base (PPLO Agar Base) | Low risk | 20/12/2012 |
| DCM | M268 | Mycoplasma Broth Base w/ CV (PPLO Broth Base w/ CV) | Low risk | 20/12/2012 |
| DCM | M267 | Mycoplasma Broth Base w/o CV (PPLO Broth Base w/o CV) | Low risk | 20/12/2012 |
| DCM | M1498 | Mycoplasma Cultivation Broth Base | Low risk | 20/12/2012 |
| DCM | MV266 | Mycoplasma HiVeg™ Agar Base (PPLO HiVeg™ Agar Base) | Low risk | 20/12/2012 |
| DCM | MV268 | Mycoplasma HiVeg™ Broth Base w/ CV (PPLO HiVeg™ Broth Base w/ CV) | Low risk | 20/12/2012 |
| DCM | MV267 | Mycoplasma HiVeg™ Broth Base w/o CV (PPLO HiVeg™ Broth Base w/o CV) | Low risk | 20/12/2012 |
| DCM | MV624 | Mycoplasma Synoviae HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M624 | Mycoplasma Synoviae Medium Base | Low risk | 20/12/2012 |
| DCM | M1374 | Mycoplasma Urogenital Broth Base (Urogenital Mycoplasma Broth Base ) | Low risk | 20/12/2012 |
| DCM | M636 | MYP Agar Base (Phenol Red Egg Yolk Polymyxin Agar Base) | Low risk | 20/12/2012 |
| DCM | MCD636 | MYP HiCynth™ Agar Base (Phenol Red Egg Yolk Polymyxin HiCynth™ Agar Base | Low risk | 28/04/2017 |
| DCM | MV636 | MYP HiVeg™ Agar Base (Phenol Red Polymyxin HiVeg™ Agar Base) | Low risk | 20/12/2012 |
| DCM | MV217 | Nickerson HiVeg™ Medium (Bi.G.G.Y. HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | M217 | Nickerson Medium (Bi.G.G.Y. Agar) | Low risk | 20/12/2012 |
| DCM | M072 | Nitrate Agar | Low risk | 10/11/2020 |
| DCM | MV072 | Nitrate HiVeg™ Agar | Low risk | 10/11/2020 |
| DCM | M681 | NNN Modified Medium (Twin Pack) | Low risk | 10/11/2020 |
| DCM | M001 | Nutrient Agar | Low risk | 20/12/2012 |
| DCM | M001A | Nutrient Agar | Low risk | 20/12/2012 |
| DCM | M087 | Nutrient Agar 1.5% | Low risk | 20/12/2012 |
| DCM | M1269 | Nutrient Agar No.2 | Low risk | 20/12/2012 |
| DCM | M012 | Nutrient Agar w/ 1% Peptone | Low risk | 20/12/2012 |
| DCM | M561 | Nutrient Agar, pH 6.8 | Low risk | 20/12/2012 |
| DCM | M002 | Nutrient Broth | Low risk | 20/12/2012 |
| DCM | M1362 | Nutrient Broth No. 2 | Low risk | 20/12/2012 |
| DCM | M1902 | Nutrient Broth No.3 | Low risk | 20/12/2012 |
| DCM | M060 | Nutrient Gelatin | Low risk | 20/12/2012 |
| DCM | MCD001 | Nutrient HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | MCD002 | Nutrient HiCynth™ Broth | Low risk | 12/08/2015 |
| DCM | MV001 | Nutrient HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV087 | Nutrient HiVeg™ Agar 1.5% | Low risk | 20/12/2012 |
| DCM | MV1269 | Nutrient HiVeg™ Agar No.2 | Low risk | 20/12/2012 |
| DCM | MV012 | Nutrient HiVeg™ Agar w/ 1% HiVeg™ Peptone | Low risk | 20/12/2012 |
| DCM | MV561 | Nutrient HiVeg™ Agar, pH 6.8 | Low risk | 20/12/2012 |

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| DCM | MV002 | Nutrient HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1348 | NYC Agar Base | Low risk | 20/12/2012 |
| DCM | MCD395 | OF Basal HiCynth™ Medium | Low risk | 25/08/2016 |
| DCM | MV395 | OF Basal HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M395 | OF Basal Medium | Low risk | 20/12/2012 |
| DCM | M1811 | OFPBL Agar Base (Oxidation Fermentation Polymyxin Bacitracin Lactose Agar Base) | Low risk | 20/12/2012 |
| DCM | M1930 | ONPG BROTH | Low risk | 20/12/2012 |
| DCM | M933 | Orange Serum Agar | Low risk | 22/04/2019 |
| DCM | MV933 | Orange SerumHiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | M1454 | Oxacillin Resistance Screening Agar Base | Low risk | 20/12/2012 |
| DCM | M1390 | Pagano Levin Base | Low risk | 20/12/2012 |
| DCM | M867 | Peizer TB Medium Base | Low risk | 20/12/2012 |
| DCM | M1207 | Pepted M Broth | Low risk | 20/12/2012 |
| DCM | M028 | Peptone Water | Low risk | 20/12/2012 |
| DCM | MCD837 | Perfringens HiCynth™ Agar Base (T.S.C/S.F.P HiCynth™ Agar Base) | Low risk | 28/04/2017 |
| DCM | MV837 | Perfringens HiVeg™ Agar Base (T.S.C/S.F.P HiCynth™ Agar Base) | Low risk | 28/04/2017 |
| DCM | M269A | Phenylethanol Agar Base | Low risk | 20/12/2012 |
| DCM | M269 | Phenylethyl Alcohol Agar Base | Low risk | 20/12/2012 |
| DCM | MV269 | Phenylethyl Alcohol HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M540 | Phenylethyl Blood Agar Base (Anaerobic) | Low risk | 20/12/2012 |
| DCM | M1866 | Phosphate Buffered Saline (PBS) pH 7.4 | Low risk | 22/04/2019 |
| DCM | M519 | Pike Streptococcal Broth Base | Low risk | 20/12/2012 |
| DCM | MV519 | Pike Streptococcal HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M282 | PKU Test Agar Base | Low risk | 20/12/2012 |
| DCM | M398 | PKU Test Agar w/ Thienylalanine | Low risk | 20/12/2012 |
| DCM | M091 | Plate Count Agar (Standard Methods Agar) | Low risk | 28/04/2017 |
| DCM | MCD091 | Plate Count HiCynth™Agar (Standard Methods HiCynth™ Agar) | Low risk | 28/04/2017 |
| DCM | MV091 | Plate Count HiVeg™ Agar (Standard Methods HiVeg™ Agar) | Low risk | 28/04/2017 |
| DCM | M574 | Plesiomonas Differential Agar (Inositol Brilliant Green Bile Agar) | Low risk | 20/12/2012 |
| DCM | MV574 | Plesiomonas Differential HiVeg™ Agar (Inositol Brilliant Green HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | M1446 | PLET Agar Base | Low risk | 20/12/2012 |
| DCM | M1451 | PLET Agar Base, Modified | Low risk | 20/12/2012 |
| DCM | M835 | PNY Medium | Low risk | 20/12/2012 |
| DCM | MH096 | Potato Dextrose Agar | Low risk | 22/04/2019 |
| DCM | M096 | Potato Dextrose Agar | Low risk | 22/04/2019 |
| DCM | M5391 | PPLO Agar Base | Low risk | 30/10/2018 |
| DCM | M1586 | PPLO Modified Broth Base w/o CV | Low risk | 20/12/2012 |

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| DCM | M899 | Preston Enrichment Broth Base (Campylobacter Enrichment Broth Base) | Low risk | 20/12/2012 |
| DCM | MV899 | Preston Enrichment HiVeg™ Broth Base (Campylobacter Enrichment HiVeg™ Broth Base) | Low risk | 20/12/2012 |
| DCM | M956 | Propionibacter Isolation Agar Base | Low risk | 20/12/2012 |
| DCM | M1697 | Proskauer Beck medium | Low risk | 20/12/2012 |
| DCM | M085 | Pseudomonas Agar Base | Low risk | 22/04/2019 |
| DCM | MV085 | Pseudomonas HiVeg Agar Base | Low risk | 22/04/2019 |
| DCM | M406 | Pseudomonas Isolation Agar Base | Low risk | 20/12/2012 |
| DCM | MCD406 | Pseudomonas Isolation HiCynth™ Agar | Low risk | 25/08/2016 |
| DCM | MV406 | Pseudomonas Isolation HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1489 | PYR Agar | Low risk | 10/11/2020 |
| DCM | M1743 | R2A Agar, Modified | Low risk | 22/04/2019 |
| DCM | MV1078 | RajHans HiVeg™ Medium (Salmonella Differential HiVeg™ Agar) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1078 | RajHans Medium (Salmonella Differential Agar) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1453A | Rapid HiColiform Broth w/Tryptophan | Low risk | 22/04/2019 |
| DCM | MCD1465 | Rapid HiColiform HiCynth™ Agar | Low risk | 10/11/2020 |
| DCM | M1465 | Rapid HiColiform™ Agar | Low risk | 10/11/2020 |
| DCM | MV1465 | Rapid HiColiform™ HiVeg™ Agar | Low risk | 10/11/2020 |
| DCM | MCD1491 | Rappaport Vassiliadis HiCynth™ Broth | Low risk | 12/08/2015 |
| DCM | M1530 | Rappaport Vassiliadis R10 Medium | Low risk | 20/12/2012 |
| DCM | MH1491 | Rappaport Vassiliadis Salmonella Enrichment Broth | Low risk | 22/04/2019 |
| DCM | M1491 | Rappaport Vassiliadis Soya Broth (RVS Broth) | Low risk | 20/12/2012 |
| DCM | M1448 | Rappaport Vassiliadis Soyabean Meal Broth (RVSM) | Low risk | 20/12/2012 |
| DCM | MH443 | Reinforced Medium for Clostridia | Low risk | 22/04/2019 |
| DCM | M1626 | Reuter's Sorbic Acid Agar Base | Low risk | 20/12/2012 |
| DCM | M459 | Robinson Medium for Entamoeba (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M149 | Robinson'c Cooked M Medium (R.C. Medium) | Low risk | 16/12/2017 |
| DCM | M1899 | Rogosa Agar, Modified | Low risk | 20/12/2012 |
| DCM | M130 | Rogosa SL Agar | Low risk | 20/12/2012 |
| DCM | M958 | Rogosa SL Agar w/ 0.15% Bile | Low risk | 20/12/2012 |
| DCM | M407 | Rogosa SL Broth | Low risk | 20/12/2012 |
| DCM | MCD130 | Rogosa SL HiCynth™ Agar | Low risk | 28/04/2017 |
| DCM | MV130 | Rogosa SL HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV407 | Rogosa SL HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M842 | Rose Bengal Agar Base | Low risk | 20/12/2012 |
| DCM | M640 | Rose Bengal Chloramphenicol Agar | Low risk | 22/04/2019 |
| DCM | MV640 | Rose Bengal Chloramphenicol HiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | M1972 | RPMI 1640 Agar w/ MOPS & 2% Glucose w/o Sodium Bicarbonate ( Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV576 | RS HiVeg Medium Base | Low risk | 22/04/2019 |

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|---|---|---|---|---|
| DCM | M576 | RS Medium Base | Low risk | 22/04/2019 |
| DCM | M409 | SABHI Agar Base | Low risk | 20/12/2012 |
| DCM | MV409 | SABHI HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1744 | Sabouraud Agar Glucose 4% | Low risk | 20/12/2012 |
| DCM | M1067 | Sabouraud Chloramphenicol Agar | Low risk | 20/12/2012 |
| DCM | MV1067 | Sabouraud Chloramphenicol HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M664 | Sabouraud Cycloheximide Chloramphenicol Agar | Low risk | 20/12/2012 |
| DCM | MV664 | Sabouraud Cycloheximide Chloramphenicol HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MH063 | Sabouraud Dextrose Agar | Low risk | 22/04/2019 |
| DCM | M063 | Sabouraud Dextrose Agar | Low risk | 20/12/2012 |
| DCM | M286 | Sabouraud Dextrose Agar Base, Modified (Dextrose Agar Base, Emmons) | Low risk | 20/12/2012 |
| DCM | MH033 | Sabouraud Dextrose Broth | Low risk | 22/04/2019 |
| DCM | M033 | Sabouraud Dextrose Broth (Sabouraud Liquid Medium) | Low risk | 20/12/2012 |
| DCM | MCD063 | Sabouraud Dextrose HiCynth™ Agar | Low risk | 12/08/2015 |
| DCM | MCD033 | Sabouraud Dextrose HiCynth™ Broth | Low risk | 12/08/2015 |
| DCM | MV063 | Sabouraud Dextrose HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV286 | Sabouraud Dextrose HiVeg™ Agar Base, Modified (Dextrose HiVeg™ Agar Base, Emmons) | Low risk | 20/12/2012 |
| DCM | MV033 | Sabouraud Dextrose HiVeg™ Broth (Sabouraud Liquid HiVeg™ Medium) | Low risk | 20/12/2012 |
| DCM | M1313 | Sabouraud Dextrose Maltose Agar | Low risk | 20/12/2012 |
| DCM | M1460 | Sabouraud Dextrose Maltose Broth | Low risk | 20/12/2012 |
| DCM | MV1313 | Sabouraud Dextrose Maltose HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MCD013 | Sabouraud Fluid HiCynth™ Medium | Low risk | 12/08/2015 |
| DCM | M1472 | Sabouraud Glucose Agar Base w/ Antibiotics | Low risk | 20/12/2012 |
| DCM | M062 | Sabouraud Maltose Agar | Low risk | 20/12/2012 |
| DCM | M064 | Sabouraud Maltose Broth | Low risk | 20/12/2012 |
| DCM | MV062 | Sabouraud Maltose HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV064 | Sabouraud Maltose HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M844 | Saccharose Broth | Low risk | 20/12/2012 |
| DCM | M1619 | Sakazakii DHL Agar | Low risk | 20/12/2012 |
| DCM | M942 | Saline Agar | Low risk | 20/12/2012 |
| DCM | M1778 | Saline Lysine Decarboxylase Medium | Low risk | 20/12/2012 |
| DCM | M1633 | Salmonella Agar (HiCrome™ RajHans Medium) | Low risk | 20/12/2012 |
| DCM | M1634 | Salmonella Agar, Modified (HiCrome™ RajHans Medium, Modified) | Low risk | 20/12/2012 |
| DCM | M573 | Salmonella Agar, ONOZ | Low risk | 20/12/2012 |
| DCM | M1078 | Salmonella Differential Agar (Twin Pack) (RajHans Medium) | Low risk | 20/12/2012 |
| DCM | M1082 | Salmonella Differential Agar, Modified (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MCD1078 | Salmonella Differential HiCynth™ Agar (Twin Pack) | Low risk | 25/08/2016 |

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|---|---|---|---|---|
| DCM | MV1078 | Salmonella Differential HiVeg™ Agar (RajHans HiVeg™ Medium) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV1082 | Salmonella Differential HiVeg™ Agar, Modified (Twin Pack) | Low risk | 20/12/2012 |
| DCM | MV573 | Salmonella HiVeg™ Agar, ONOZ | Low risk | 20/12/2012 |
| DCM | M1767 | Salt Agar, Modified | Low risk | 20/12/2012 |
| DCM | M1290 | Salt Broth, Modified | Low risk | 20/12/2012 |
| DCM | M155 | Salt M Broth | Low risk | 20/12/2012 |
| DCM | M821 | Salt Polymyxin Broth Base | Low risk | 20/12/2012 |
| DCM | MV821 | Salt Polymyxin HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1276 | Sauton's Fluid Medium Base | Low risk | 20/12/2012 |
| DCM | M1535 | SBG Enrichment Broth (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M291 | Schaedler Agar | Low risk | 20/12/2012 |
| DCM | M292 | Schaedler Broth | Low risk | 20/12/2012 |
| DCM | MV291 | Schaedler HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV292 | Schaedler HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1882 | Selective Broth for MRSA | Low risk | 20/12/2012 |
| DCM | M052 | Selenite Broth (Selenite F Broth) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M970 | Selenite Broth Base w/o Biselenite | Low risk | 20/12/2012 |
| DCM | M1079 | Selenite Cystine Broth Base w/o Biselenite | Low risk | 20/12/2012 |
| DCM | M1536 | Selenite F Broth w/ Dulcitol (Dulcitol Selenite Broth) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1534 | Selenite Mannitol Broth (Mannitol Selenite Broth) (Twin Pack) | Low risk | 20/12/2012 |
| DCM | M1321 | Semisolid LM Medium | Low risk | 20/12/2012 |
| DCM | M1282 | Semisolid Rappaport Vassiliadis Medium, Modified | Low risk | 22/04/2019 |
| DCM | M1998 | Semisolid RV Medium w/ 0.9% Agar | Low risk | 25/08/2016 |
| DCM | MV296 | Sensitivity Test HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M296 | Sensitivity Test Medium | Low risk | 20/12/2012 |
| DCM | M1301 | Sheep Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M1739 | Shepard's Differential Agar Base (A7 Agar Base) | Low risk | 20/12/2012 |
| DCM | M411 | Simmons Agar Base | Low risk | 20/12/2012 |
| DCM | M099 | Simmons Citrate Agar | Low risk | 20/12/2012 |
| DCM | M099S | Simmons Citrate Agar | Low risk | 20/12/2012 |
| DCM | M612A | Slanetz and Bartley Medium w/o TTC | Low risk | 10/11/2020 |
| DCM | M5296 | SM Tryptone Glucose Glycerin Medium | Low risk | 25/11/2017 |
| DCM | M960 | Smibert's Semisolid Brucella Medium | Low risk | 20/12/2012 |
| DCM | M106 | Snyder Test Agar (B.C.G. - Dextrose Agar) | Low risk | 20/12/2012 |
| DCM | MV106 | Snyder Test HiVeg™ Agar (B.C.G. - Dextrose HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | M767 | Sodium Azide Crystal Violet Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M1079B | Sodium Biselenite | Low risk | 22/04/2019 |
| DCM | M298 | Sorbitol Agar (MacConkey Sorbitol Agar) | Low risk | 20/12/2012 |
| DCM | MV298 | Sorbitol HiVeg™Agar (MacConkey Sorbitol HiVeg™ Agar) | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| DCM | M299 | Sorbitol Iron Agar | Low risk | 20/12/2012 |
| DCM | MV299 | Sorbitol Iron HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M935 | Soya Peptone Yeast Extract Agar | Low risk | 20/12/2012 |
| DCM | M1286 | Soyabean Bile Broth Base | Low risk | 20/12/2012 |
| DCM | M290 | Soyabean Casein Digest Agar (Tryptone Soya Agar) | Low risk | 22/04/2019 |
| DCM | M109 | Soyabean Casein Digest Agar w/ Yeast Extract and Hemin (Tryptone Soya Agar w/ Yeast Extract and Hemin) | Low risk | 20/12/2012 |
| DCM | M011 | Soyabean Casein Digest Medium (Tryptone Soya Broth) | Low risk | 22/04/2019 |
| DCM | M323 | Soyabean Casein Digest Medium w/ 0.1% Agar (Tryptone Soya Broth w/ 0.1% Agar) | Low risk | 20/12/2012 |
| DCM | M207 | Soyabean Casein Digest Medium w/ Yeast Extract and Ferric pyrophosphate | Low risk | 20/12/2012 |
| DCM | M322 | Soyabean Casein Digest Medium w/o Dextrose (Tryptone SoyaBroth w/o Dextrose) | Low risk | 28/04/2017 |
| DCM | MV1286 | Soyabean HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | MV011 | Soyabean HiVeg™ Medium | Low risk | 22/04/2019 |
| DCM | MV323 | Soyabean HiVeg™ Medium w/ 0.1% Agar (Tryptone Soya HiVeg™ Broth w/ 0.1% Agar) | Low risk | 20/12/2012 |
| DCM | MV207 | Soyabean HiVeg™ Medium w/ Yeast Extract and Ferric pyrophosphate | Low risk | 20/12/2012 |
| DCM | MV290 | SoyabeanHiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | MH011 | Soybean Casein Digest Medium (Casein Soybean Digest Broth) | Low risk | 22/04/2019 |
| DCM | MH290 | Soybean-Casein Digest Agar (Casein Soyabean Digest Agar) | Low risk | 22/04/2019 |
| DCM | M211 | Special Infusion Agar (BHI Agar) | Low risk | 20/12/2012 |
| DCM | MV211 | Special Infusion Agar, HiVeg™ (BHI Agar, HiVeg™) | Low risk | 20/12/2012 |
| DCM | M1613 | Special YM Medium | Low risk | 20/12/2012 |
| DCM | M300 | Specimen Preservative Medium Base (SP Hajna) | Low risk | 20/12/2012 |
| DCM | M445 | Spirit Blue Agar | Low risk | 20/12/2012 |
| DCM | MV445 | Spirit Blue HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M412 | Spirolate Broth, OMATA | Low risk | 20/12/2012 |
| DCM | MV412 | Spirolate HiVeg™ Broth, OMATA | Low risk | 20/12/2012 |
| DCM | MCD108 | SS HiCynth™ Agar (Salmonella Shigella HiCynth™ Agar) | Low risk | 12/08/2015 |
| DCM | M108 | SS Agar (Salmonella Shigella Agar) | Low risk | 20/12/2012 |
| DCM | M108D | SS Agar (Salmonella Shigella Agar) | Low risk | 16/12/2017 |
| DCM | M1979R | SS Agar Modified (w/sucrose) | Low risk | 25/08/2016 |
| DCM | M1979 | SS Agar w/sucrose | Low risk | 20/12/2012 |
| DCM | M1032 | SS Agar, Modified | Low risk | 20/12/2012 |
| DCM | MV108 | SS HiVeg™ Agar (Salmonella Shigella HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | M1959 | SS Selective Agar, Improved | Low risk | 20/12/2012 |
| DCM | M1703 | SSDC agar | Low risk | 20/12/2012 |
| DCM | M1608 | ß-Streptococcus Selective Agar Base | Low risk | 20/12/2012 |
| DCM | M675 | Staib's Medium (Bird Seed Agar) | Low risk | 20/12/2012 |
| DCM | M883 | Standard Infusion Agar | Low risk | 20/12/2012 |

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| DCM | MV883 | Standard Infusion Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M116 | Standard Nutrient Broth (H.S. Vaccine Medium) | Low risk | 20/12/2012 |
| DCM | MV116 | Standard Nutrient HiVeg™ Broth (H.S. Vaccine HiVeg™ Medium) | Low risk | 20/12/2012 |
| DCM | M578 | Standard Staphylococcus Broth | Low risk | 20/12/2012 |
| DCM | MV578 | Standard Staphylococcus HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M156 | Staphylococcus Agar No. 110 w/ Azide | Low risk | 20/12/2012 |
| DCM | M521 | Staphylococcus Agar No.110 | Low risk | 20/12/2012 |
| DCM | MV521 | Staphylococcus HiVeg™ Agar No. 110 | Low risk | 20/12/2012 |
| DCM | M1965 | Stenotrophomonas Selective Agar Base | Low risk | 20/12/2012 |
| DCM | M1840R | Streptococcus Agalactiae Selective Agar Base (HiCrome™ Strep B Selective Agar Base) | Low risk | 30/10/2018 |
| DCM | M465 | Streptococcus Enrichment Broth (SE Broth) | Low risk | 20/12/2012 |
| DCM | MV465 | Streptococcus Enrichment HiVeg™ Broth (SE HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M304 | Streptococcus Selection Agar | Low risk | 20/12/2012 |
| DCM | M303 | Streptococcus Selection Broth | Low risk | 20/12/2012 |
| DCM | MV304 | Streptococcus Selection HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV303 | Streptococcus Selection HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M1735 | Stuart Medium w/o Methylene Blue with Charcoal | Low risk | 20/12/2012 |
| DCM | M306 | Stuart Transport Medium (Transport Medium, Stuart) | Low risk | 20/12/2012 |
| DCM | M1131 | Stuart Transport Medium w/o Methylene Blue | Low risk | 20/12/2012 |
| DCM | M1203 | Stuart Transport Medium w/o Sodium Glycerophosphate | Low risk | 20/12/2012 |
| DCM | M308 | Sulpha Sensitivity Test Agar | Low risk | 20/12/2012 |
| DCM | MV837 | T.S.C./S.F.P. HiVeg™ Agar Base (Perfringens HiVeg™ Agar Base) | Low risk | 20/12/2012 |
| DCM | M100 | TB Broth Base | Low risk | 20/12/2012 |
| DCM | M034 | TB Broth Base w/o Tween 80 | Low risk | 20/12/2012 |
| DCM | MV100 | TB HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | MV034 | TB HiVeg™ Broth Base w/o Tween 80 | Low risk | 20/12/2012 |
| DCM | M189 | TCBS Agar | Low risk | 20/12/2012 |
| DCM | M870 | TCBS Agar (Selective) | Low risk | 20/12/2012 |
| DCM | M870A | TCBS Agar, Modified | Low risk | 20/12/2012 |
| DCM | MCD870 | TCBS HiCynth™ Agar (Selective) | Low risk | 25/08/2016 |
| DCM | MV189 | TCBS HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV870 | TCBS HiVeg™ Agar (Selective) | Low risk | 20/12/2012 |
| DCM | M529 | Teepol Broth (Twin Pack) | Low risk | 10/11/2020 |
| DCM | MV529 | Teepol HiVeg™ Broth (Twin Pack) | Low risk | 10/11/2020 |
| DCM | M1260 | Tellurite Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M448 | Tellurite Glycine Agar Base | Low risk | 20/12/2012 |
| DCM | M616 | Tergitol-7 Agar Base | Low risk | 20/12/2012 |
| DCM | M850 | Tergitol-7 Agar H | Low risk | 20/12/2012 |
| DCM | M851 | Tergitol-7 Broth | Low risk | 20/12/2012 |

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| DCM | MV616 | Tergitol-7 HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV850 | Tergitol-7 HiVeg™ Agar H | Low risk | 20/12/2012 |
| DCM | MV851 | Tergitol-7 HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M032 | Tetrathionate Broth Base (w/o Iodine and BG) (Fluid Tetrathionate Medium w/o Iodine and BG) | Low risk | 20/12/2012 |
| DCM | MV032 | Tetrathionate HiVeg™ Broth Base (w/o Iodine and BG) (Fluid Tetrathionate HiVeg™ Medium w/o Iodine and BG) | Low risk | 20/12/2012 |
| DCM | MV413 | Thayer Martin HiVeg™ Medium Base | Low risk | 20/12/2012 |
| DCM | M413 | Thayer Martin Medium Base | Low risk | 20/12/2012 |
| DCM | M610 | Thiogel Medium | Low risk | 20/12/2012 |
| DCM | M608 | Thioglycollate Agar | Low risk | 20/12/2012 |
| DCM | M010 | Thioglycollate Broth, Alternative ( Alternative Thioglycollate Medium)(NIH Thioglycollate Broth) | Low risk | 20/12/2012 |
| DCM | MCD010 | Thioglycollate HiCynth™ Broth, Alternative ( Alternative Thioglycollate HiCynth™ Medium)(NIH Thioglycollate HiCynth™ Broth) | Low risk | 12/08/2015 |
| DCM | MV608 | Thioglycollate HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | MV010 | Thioglycollate HiVeg™ Broth, Alternative ( Alternative Thioglycollate HiVeg™ Medium)(NIH HiVeg™ Thioglycollate Broth) | Low risk | 20/12/2012 |
| DCM | MV195 | Thioglycollate HiVeg™ Medium, Linden (Brewer Thioglycollate HiVeg™ Medium, Modified) | Low risk | 20/12/2012 |
| DCM | M979 | Thioglycollate Medium w/ Hemin and Vitamin K | Low risk | 20/12/2012 |
| DCM | M195 | Thioglycollate Medium, Linden (Brewer Thioglycollate Medium, Modified) | Low risk | 20/12/2012 |
| DCM | M853 | Thiol Broth | Low risk | 20/12/2012 |
| DCM | MV853 | Thiol HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | MV852 | Thiol HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M852 | Thiol Medium | Low risk | 20/12/2012 |
| DCM | M314 | Tinsdale Agar Base | Low risk | 20/12/2012 |
| DCM | MV314 | Tinsdale HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M313 | Todd Hewitt Broth | Low risk | 20/12/2012 |
| DCM | MV313 | Todd Hewitt HiVeg™ Broth | Low risk | 20/12/2012 |
| DCM | M2127 | Todd Hewitt Broth w/colistin & Nalidixic Acid | Low risk | 17/06/2021 |
| DCM | M879 | Tomato Juice Agar, Special | Low risk | 20/12/2012 |
| DCM | MV879 | Tomato Juice HiVeg™ Agar, Special | Low risk | 20/12/2012 |
| DCM | M1149 | Transgrow Medium Base | Low risk | 20/12/2012 |
| DCM | M315 | Transport Charcoal Medium | Low risk | 20/12/2012 |
| DCM | M1487 | Transport Liquid Medium | Low risk | 20/12/2012 |
| DCM | M306 | Transport Medium Stuart (Stuart Transport Medium) | Low risk | 20/12/2012 |
| DCM | M202 | Transport Medium w/o Charcoal (Cary - Blair Medium Base) | Low risk | 20/12/2012 |
| DCM | M684 | Transport Medium, Amies w/o Charcoal | Low risk | 20/12/2012 |
| DCM | M665 | Trichomonas Agar Base | Low risk | 20/12/2012 |
| DCM | M1204 | Trichomonas Broth Base No. 2 | Low risk | 20/12/2012 |

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| DCM | M305 | Trichomonas Broth Base, Kupferberg (Kupferberg Trichomonas Broth Base) | Low risk | 20/12/2012 |
| DCM | MV665 | Trichomonas HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV305 | Trichomonas HiVeg™ Broth Base, Kupferberg (Kupferberg Trichomonas HiVeg™ Broth Base) | Low risk | 20/12/2012 |
| DCM | MV460 | Trichomonas Modified CPLM HiVeg™ Medium Base (Modified CPLM HiVeg™ Medium Base) | Low risk | 20/12/2012 |
| DCM | M460 | Trichomonas Modified CPLM Medium Base (Modified CPLM Medium Base) | Low risk | 20/12/2012 |
| DCM | M531 | Trichophyton Agar-1 | Low risk | 20/12/2012 |
| DCM | M532 | Trichophyton Agar-2 | Low risk | 20/12/2012 |
| DCM | M533 | Trichophyton Agar-3 | Low risk | 20/12/2012 |
| DCM | M534 | Trichophyton Agar-4 | Low risk | 20/12/2012 |
| DCM | M535 | Trichophyton Agar-5 | Low risk | 20/12/2012 |
| DCM | M536 | Trichophyton Agar-6 | Low risk | 20/12/2012 |
| DCM | M152 | Trichophyton Agar-7 | Low risk | 20/12/2012 |
| DCM | MV531 | Trichophyton HiVeg™ Agar-1 | Low risk | 20/12/2012 |
| DCM | MV532 | Trichophyton HiVeg™ Agar-2 | Low risk | 20/12/2012 |
| DCM | MV533 | Trichophyton HiVeg™ Agar-3 | Low risk | 20/12/2012 |
| DCM | MV534 | Trichophyton HiVeg™ Agar-4 | Low risk | 20/12/2012 |
| DCM | MV535 | Trichophyton HiVeg™ Agar-5 | Low risk | 20/12/2012 |
| DCM | M021 | Triple Sugar Iron Agar | Low risk | 22/04/2019 |
| DCM | MV021 | Triple Sugar Iron HiVeg™ Agar | Low risk | 22/04/2019 |
| DCM | M1028 | Tryptic Digest Broth(Field's Tryptic Digest Broth) | Low risk | 20/12/2012 |
| DCM | MV1028 | Tryptic Digest Broth, HiVeg™ (Field's Tryptic Digest Broth, HiVeg™) | Low risk | 20/12/2012 |
| DCM | M1591 | Tryptone Bile Glucuronic Agar (TBX Agar) | Low risk | 22/04/2019 |
| DCM | M463 | Tryptone Broth (Tryptone Water) | Low risk | 22/04/2019 |
| DCM | MV364 | Tryptone Nitrate HiVeg™ Medium (Indole Nitrate HiVeg™ Medium) | Low risk | 20/12/2012 |
| DCM | M364 | Tryptone Nitrate Medium (Indole Nitrate Medium) | Low risk | 20/12/2012 |
| DCM | M969 | Tryptone Peptone Glucose Yeast Extract Broth Base w/o Trypsin | Low risk | 20/12/2012 |
| DCM | MV969 | Tryptone Peptone Glucose Yeast Extract HiVeg™ Broth Base w/o Trypsin | Low risk | 20/12/2012 |
| DCM | M323 | Tryptone Soya Broth w/ 0.1% Agar (Soyabean Casein Digest Medium w/ 0.1% Agar) | Low risk | 20/12/2012 |
| DCM | MV323 | Tryptone Soya HiVeg™ Broth w/ 0.1% Agar (Soyabean HiVeg™ Medium w/ 0.1% Agar) | Low risk | 20/12/2012 |
| DCM | M1948 | Tryptone Soya Serum Bacitracin Vancomycin Agar (TSBV) | Low risk | 08/12/2017 |
| DCM | M1217 | Tryptone Sucrose Tetrazolium Agar Base (TSTA) | Low risk | 20/12/2012 |
| DCM | M1056 | Tryptone Tellurite Agar Base | Low risk | 20/12/2012 |
| DCM | MV463 | Tryptone Water, HiVeg™ (Tryptone Broth,HiVeg™) | Low risk | 22/04/2019 |
| DCM | M1975 | Tryptone yeast extract cystine w/sucrose and w/O bacitracin agar (TYCSB) | Low risk | 20/12/2012 |
| DCM | M2046I | Tryptone Yeast Sodium Sulphite Agar Base | Low risk | 10/11/2020 |

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|---|---|---|---|---|
| DCM | M538 | Tryptose Agar | Low risk | 20/12/2012 |
| DCM | M996 | Tryptose Agar w/ Thiamine HCl | Low risk | 20/12/2012 |
| DCM | MV996 | Tryptose Agar w/ Thiamine HCl, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MV538 | Tryptose Agar, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M097 | Tryptose Blood Agar Base | Low risk | 20/12/2012 |
| DCM | M450 | Tryptose Blood Agar Base w/ Yeast Extract | Low risk | 20/12/2012 |
| DCM | MV450 | Tryptose Blood Agar Base w/ Yeast Extract, HiVeg™ | Low risk | 20/12/2012 |
| DCM | MV097 | Tryptose Blood Agar Base, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M177 | Tryptose Broth | Low risk | 20/12/2012 |
| DCM | M997 | Tryptose Broth w/ Thiamine HCl | Low risk | 20/12/2012 |
| DCM | MV177 | Tryptose Broth, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M5393 | Tryptose Phosphate Broth | Low risk | 30/10/2018 |
| DCM | M093 | Tryptose Phosphate Broth | Low risk | 20/12/2012 |
| DCM | MV093 | Tryptose Phosphate Broth, HiVeg™ | Low risk | 20/12/2012 |
| DCM | M1532 | Tryptose Phosphate Broth, Modified | Low risk | 20/12/2012 |
| DCM | M093G | Tryptose Phosphate Broth, Sterile | Low risk | 22/04/2019 |
| DCM | M2060 | Tryptose Serum Agar Base | Low risk | 10/11/2020 |
| DCM | M2019 | Tryptose Serum Broth Base(Modified Newin | Low risk | 25/08/2016 |
| DCM | M837 | Tryptose Sulphite Cycloserine (T.S.C. / S.F.P.) Agar Base (Perfringens Agar Base) | Low risk | 20/12/2012 |
| DCM | M1780 | TS Saline Agar (Triple Sugar Saline Iron Agar) | Low risk | 20/12/2012 |
| DCM | M2016 | TSB w/6.5% NaCL | Low risk | 25/08/2016 |
| DCM | M1220 | TTC Broth Base (Triclosan Ticarcillin Chlorate Broth) | Low risk | 20/12/2012 |
| DCM | MV1220 | TTC HiVeg™ Broth Base | Low risk | 20/12/2012 |
| DCM | M1912 | Tween Esterase Test Agar Base | Low risk | 20/12/2012 |
| DCM | M1817 | Universal Fastidious Culture Agar | Low risk | 20/12/2012 |
| DCM | M1818 | Universal Fastidious Culture Broth | Low risk | 10/11/2020 |
| DCM | M112S | Urea Agar Base (Christensen) | Low risk | 20/12/2012 |
| DCM | M112 | Urea Agar Base (Christensen) (Autoclavable) | Low risk | 20/12/2012 |
| DCM | M112A | Urea Agar Base (Filter Sterilizable) (w/o Agar) | Low risk | 20/12/2012 |
| DCM | M112I | Urea Agar Base, Christensen | Low risk | 20/12/2012 |
| DCM | M111A | Urea Broth (Filter Sterilizable) | Low risk | 20/12/2012 |
| DCM | M111 | Urea Broth Base (Diagnostic Stuart's Urea Broth Base) | Low risk | 20/12/2012 |
| DCM | MV112 | Urea HiVeg™ Agar Base (Christensen) (Autoclavable) | Low risk | 20/12/2012 |
| DCM | M1784I | Urea Indole Broth, Modified | Low risk | 20/12/2012 |
| DCM | M1784 | Urea Indole Medium | Low risk | 20/12/2012 |
| DCM | M328 | V Infusion Agar | Low risk | 20/12/2012 |
| DCM | M329 | V Infusion Broth | Low risk | 20/12/2012 |
| DCM | M1057 | Vaginalis Agar Base | Low risk | 20/12/2012 |
| DCM | M1763 | Vancomycin Resistant Enterococci (VRE) Agar Base | Low risk | 20/12/2012 |

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| DCM | M1762 | Vancomycin Resistant Enterococci (VRE) Broth Base | Low risk | 20/12/2012 |
| DCM | M416 | Veillonella Agar Base | Low risk | 20/12/2012 |
| DCM | MV416 | Veillonella HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M820 | Vibrio Agar | Low risk | 20/12/2012 |
| DCM | MV820 | Vibrio HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M049 | Violet Red Bile Agar | Low risk | 28/04/2017 |
| DCM | M049A | Violet Red Bile Agar | Low risk | 16/12/2017 |
| DCM | M1684 | Violet Red Bile Agar w/ Glucose and Lactose | Low risk | 22/04/2019 |
| DCM | MH581 | Violet Red Bile Glucose Agar | Low risk | 22/04/2019 |
| DCM | M581 | Violet Red Bile Glucose Agar w/o Lactose | Low risk | 25/11/2017 |
| DCM | MCD581 | Violet Red Bile Glucose HiCynth™ Agar w/o Lactose | Low risk | 04/07/2018 |
| DCM | MV581 | Violet Red Bile Glucose HiVeg™ Agar w/o Lactose | Low risk | 04/07/2018 |
| DCM | MCD049 | Violet Red Bile HiCynth™ Agar | Low risk | 28/04/2017 |
| DCM | MV049 | Violet Red HiVeg™ Agar | Low risk | 28/04/2017 |
| DCM | MCD023 | Vogel Johnson HiCynth™ Agar Base w/o Tellurite (V.J. HiCynth™ Agar) | Low risk | 12/08/2015 |
| DCM | M023 | Vogel-Johnson Agar Base w/o Tellurite (V.J. Agar) | Low risk | 20/12/2012 |
| DCM | MU023 | Vogel-Johnson Agar Medium | Low risk | 20/12/2012 |
| DCM | MV023 | Vogel-Johnson HiVeg™ Agar Base w/o Tellurite (V. J. HiVeg™ Agar) | Low risk | 20/12/2012 |
| DCM | MV662 | VP HiVeg™ Medium | Low risk | 20/12/2012 |
| DCM | M662 | VP Medium | Low risk | 20/12/2012 |
| DCM | M626 | Wagatsuma Agar Base | Low risk | 20/12/2012 |
| DCM | MV626 | Wagatsuma HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M1059 | Wayne Sulphatase Agar Base | Low risk | 20/12/2012 |
| DCM | M832 | Wilkins Chalgren Anaerobic Agar Base | Low risk | 20/12/2012 |
| DCM | M863 | Wilkins Chalgren Anaerobic Broth Base | Low risk | 20/12/2012 |
| DCM | MV832 | Wilkins Chalgren Anaerobic HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV863 | Wilkins Chalgren Anaerobic HiVeg™ Broth Base | Low risk | 25/08/2016 |
| DCM | M331 | Wilson Blair Agar Base | Low risk | 20/12/2012 |
| DCM | M332 | Wilson Blair Agar w/ BG | Low risk | 20/12/2012 |
| DCM | MV331 | Wilson Blair HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | MV332 | Wilson Blair HiVeg™ Agar w/ BG | Low risk | 20/12/2012 |
| DCM | MV031 | XLD HiVeg™ Agar | Low risk | 20/12/2012 |
| DCM | M1147 | XLT4 Agar Base | Low risk | 20/12/2012 |
| DCM | MV1147 | XLT4 HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | M336 | Xylose Lysine Agar Base | Low risk | 20/12/2012 |
| DCM | M031 | Xylose Lysine Deoxycholate Agar (XLD Agar) | Low risk | 20/12/2012 |
| DCM | MCD031 | Xylose Lysine Deoxycholate HiCynth™ Agar (XLD HiCynth™ Agar) | Low risk | 12/08/2015 |
| DCM | MH031 | Xylose-Lysine-Deoxycholate Agar | Low risk | 22/04/2019 |

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|---|---|---|---|---|
| DCM | M424 | Yeast Malt Agar (YM Agar) (ISP Medium No. 2) | Low risk | 22/04/2019 |
| DCM | M425 | Yeast Malt Broth (YM Broth) | Low risk | 20/12/2012 |
| DCM | MV424 | Yeast Malt HiVeg™ Agar (YM HiVeg™ Agar) | Low risk | 22/04/2019 |
| DCM | MV425 | Yeast Malt HiVeg™ Broth (YM HiVeg™ Broth) | Low risk | 20/12/2012 |
| DCM | M1421 | YEP Agar | Low risk | 30/10/2018 |
| DCM | M1823 | YEP Agar, Modified | Low risk | 10/11/2020 |
| DCM | M1367 | Yersinia Enrichment Broth Base | Low risk | 20/12/2012 |
| DCM | M843 | Yersinia Selective Agar Base | Low risk | 20/12/2012 |
| DCM | M1861 | Yersinia Selective Broth Base | Low risk | 20/12/2012 |
| DCM | MV843 | Yersinia Selective HiVeg™ Agar Base | Low risk | 20/12/2012 |
| DCM | EC211CR | BHI Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC210CR | BHI Broth (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC073DR | Blood Agar Base (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1297ACR | HiCrome™ Candida Differential Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1297ARDR | HiCrome™ Candida Differential Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1297ADR | HiCrome™ Candida Differential Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1674CCLR | HiCrome™ MeReSa Agar Base (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1353CCLR | HiCrome™ UTI Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1353CR | HiCrome™ UTI Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1353DR | HiCrome™ UTI Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC211CCL | HiEncap™ BHI Agar (HiEncap™ Special Infusion Agar) | Low risk | 12/08/2015 |
| DCM | EC210D | HiEncap™ BHI Broth | Low risk | 12/08/2015 |
| DCM | EC210CCL | HiEncap™ BHI Broth | Low risk | 12/08/2015 |
| DCM | EC073D | HiEncap™ Blood Agar Base | Low risk | 12/08/2015 |
| DCM | EC073CCL | HiEncap™ Blood Agar Base | Low risk | 12/08/2015 |
| DCM | EC081CCL | HiEncap™ MacConkey Agar w/0.15% Bile Salt | Low risk | 12/08/2015 |
| DCM | EC082ACCL | HiEncap™ MacConkey Agar w/o CV, NaCl w/Bile Salts | Low risk | 12/08/2015 |
| DCM | EC173CCL | HiEncap™ Mueller Hinton Agar | Low risk | 12/08/2015 |
| DCM | EC173D | HiEncap™ Mueller Hinton Agar | Low risk | 12/08/2015 |
| DCM | EC1084CCL | HiEncap™ Mueller Hinton Agar No.2 | Low risk | 12/08/2015 |
| DCM | EC1084D | HiEncap™ Mueller Hinton Agar No.2 | Low risk | 12/08/2015 |
| DCM | EC391CCL | HiEncap™ Mueller Hinton Broth | Low risk | 12/08/2015 |
| DCM | EC391D | HiEncap™ Mueller Hinton Broth | Low risk | 12/08/2015 |
| DCM | EC001DR | HiEncap™ Nutrient Agar | Low risk | 25/08/2016 |
| DCM | EC001CCL | HiEncap™ Nutrient Agar | Low risk | 12/08/2015 |
| DCM | EC001D | HiEncap™ Nutrient Agar | Low risk | 12/08/2015 |
| DCM | EC002CCL | HiEncap™ Nutrient Broth | Low risk | 12/08/2015 |
| DCM | EC002D | HiEncap™ Nutrient Broth | Low risk | 12/08/2015 |

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|---|---|---|---|---|
| DCM | EC002M | HiEncap™ Nutrient Broth | Low risk | 12/08/2015 |
| DCM | EC091D | HiEncap™ Plate Count Agar | Low risk | 16/12/2017 |
| DCM | EC091CCL | HiEncap™ Plate Count Agar | Low risk | 16/12/2017 |
| DCM | EC063CCL | HiEncap™ Sabouraud Dextrose Agar | Low risk | 12/08/2015 |
| DCM | EC033CCL | HiEncap™ Sabouraud Dextrose Broth | Low risk | 12/08/2015 |
| DCM | EC033D | HiEncap™ Sabouraud Dextrose Broth | Low risk | 12/08/2015 |
| DCM | EC173DR | Mueller Hinton Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC1084DR | Mueller Hinton Agar No.2 (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC391CR | Mueller Hinton Broth (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC002CR | Nutrient Broth (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC063CCLR | Sabouraud Dextrose Agar (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC033CR | Sabouraud Dextrose Broth (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | EC031CCLR | Xylose Deoxycholate Agar (XLD Agar) (HiEncap™ water-soluble capsule) | Low risk | 25/08/2016 |
| DCM | GM618 | Alkaline Peptone Water, Granulated | Low Risk | 12/08/2015 |
| DCM | GM491 | Anaerobic Agar (Brewer) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM672 | Aspargine Broth (Coccidiodin and Histoplasmin Broth) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM043 | Baird Parker Agar Base,Granulated | Low Risk | 12/08/2015 |
| DCM | GM1091 | Baird Staphylococcus Enrichment Broth Base, Granulated | Low risk | 10/11/2020 |
| DCM | GM211 | BHI Agar (Special Infusion Agar) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM210 | BHI Broth, Granulated | Low Risk | 12/08/2015 |
| DCM | GM217 | Bi.G.G.Y. Agar (Nickerson Medium) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM027 | Bismuth Sulphite Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM073 | Blood Agar Base (Infusion Agar) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM073R | Blood Agar Base (Infusion Agar) w/o Blood, Granulated | Low risk | 25/08/2016 |
| DCM | GM834A | Blood Agar Base No. 2 w/ 1.2% Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM016A | Brilliant Green Agar Base w/ 1.2% Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM971 | Brilliant Green Agar Base w/ Phosphates, Granulated | Low risk | 20/12/2012 |
| DCM | GM074 | Brucella Agar Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM614 | Buffered Peptone Water , Granulated | Low risk | 22/04/2019 |
| DCM | GM1275 | Buffered Peptone Water w/ NaCl, Granulated | Low Risk | 12/08/2015 |
| DCM | GMH1275 | Buffered Sodium Chloride-Peptone Solution pH 7.0 , Granulated | Low risk | 22/04/2019 |
| DCM | GM792 | C.L.E.D. Agar w/ Bromo Thymol Blue, Granulated | Low Risk | 12/08/2015 |
| DCM | GMH024 | Cetrimide Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM024 | Cetrimide Agar Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM497 | Clostridial Agar, Granulated | Low risk | 25/08/2016 |
| DCM | GMH144 | Columbia Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM144 | Columbia Blood Agar Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM188 | D.T.M. Agar Base (Dermatophyte Test Agar Base) , Granulated | Low Risk | 12/08/2015 |

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|---|---|---|---|---|
| DCM | GM030 | Deoxycholate Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM065 | Deoxycholate Citrate Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM286 | Dextrose Agar Base, Emmons (Sabouraud Dextrose Agar Base, Modified) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM1129 | Dichloran Glycerol Medium Base , Granulated | Low risk | 22/04/2019 |
| DCM | GM1603 | Differential Reinforced Clostridial Agar,Granulated | Low risk | 10/11/2020 |
| DCM | GM127 | EC Broth, Granulated | Low Risk | 12/08/2015 |
| DCM | GM317 | EMB Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM022 | EMB Agar, Levine, Granulated | Low Risk | 12/08/2015 |
| DCM | GM029 | Endo Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM029R | Endo Agar, Special | Low risk | 25/08/2016 |
| DCM | GM1075 | Endo Agar,Modified, Granulated | Low Risk | 12/08/2015 |
| DCM | GMH287 | Enterobacteria Enrichment Broth, Mossel , Granulated | Low risk | 22/04/2019 |
| DCM | GM013 | Fluid Sabouraud Medium (Sabouraud Medium,Fluid) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM025 | Fluid Selenite Cystine Medium (Selenite Cystine Broth) (Twin Pack) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM032 | Fluid Tetrathionate Medium w/o Iodine and BG (Tetrathionate Broth Base w/o Iodine and BG) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM009 | Fluid Thioglycollate medium (Thioglycollate medium Fluid) , Granulated | Low risk | 22/04/2019 |
| DCM | GM434 | GC Agar Base,Granulated | Low Risk | 04/07/2018 |
| DCM | GM070 | Glucose Phosphate Broth (Buffered Glucose Broth) , Granulated | Low risk | 12/08/2015 |
| DCM | GM070R | Glucose Phosphate Broth (Buffered Glucose Broth) , Granulated | Low risk | 04/07/2018 |
| DCM | GMV070 | Glucose Phosphate HiVeg™ Broth (Buffered Glucose HiVeg™ Broth) , Granulated | Low risk | 20/12/2012 |
| DCM | GM242 | GN Broth, Hajna, Granulated | Low Risk | 12/08/2015 |
| DCM | GM467 | Hektoen Enteric Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1297A | HiCrome™ Candida Differential Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1353 | HiCrome™ UTI Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1007 | KF Streptococcus Agar Base w/ BCP, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1232 | Kimmig Fungi Agar Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1543 | King's Medium A Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM078 | Kligler Iron Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM641 | Lactobacillus MRS Agar (MRS Agar) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM369 | Lactobacillus MRS Broth (MRS Broth) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM1003 | Lactose Broth , Granulated | Low risk | 22/04/2019 |
| DCM | GM080 | Lauryl Sulphate Broth (Lauryl Tryptose Broth) , Granulated | Low risk | 22/04/2019 |
| DCM | GM1380 | Leifson Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM890A | Listeria Enrichment Medium Base (UVM) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM1064 | Listeria Identification Agar Base (PALCAM) , Granulated | Low risk | 22/04/2019 |
| DCM | GM1090 | Listeria Identification Broth Base (PALCAM) , Granulated | Low risk | 22/04/2019 |

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| DCM | GM1145 | Listeria Oxford Medium Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM889 | Listeria Selective Broth Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1865 | Listeria Selective Enrichment Broth , Granulated | Low risk | 22/04/2019 |
| DCM | GM1001 | LM Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM162 | Lowenstein Jensen Medium Base (L.J. Medium) , Granulated | Low Risk | 12/08/2015 |
| DCM | GMH081 | MacConkey Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM081 | MacConkey Agar w/0.15% Bile Salts,CV and NaCL, Granulated | Low Risk | 12/08/2015 |
| DCM | GM082A | MacConkey Agar w/o CV,NaCL w/0.5% Bile Salts, Granulated | Low Risk | 12/08/2015 |
| DCM | GM082 | MacConkey Agar w/o CV,NaCLw/0.5% Sodium Taurocholate, Granulated | Low Risk | 12/08/2015 |
| DCM | GMH083 | MacConkey Broth , Granulated | Low risk | 22/04/2019 |
| DCM | GM083 | MacConkey Broth Purple w/BCP , Granulated | Low risk | 22/04/2019 |
| DCM | GM137 | Malt Extract Agar Base (w/ Mycological Peptone) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM255 | Malt Extract Broth Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GMH118 | Mannitol Salt Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM118 | Mannitol Salt Agar Base , Granulated | Low risk | 12/08/2015 |
| DCM | GM1030 | Maximum Recovery Diluent , Granulated | Low risk | 22/04/2019 |
| DCM | GM1170 | Modified Czapek Dox Agar, Granulated | Low risk | 25/08/2016 |
| DCM | GM1285 | Modified EC Broth Base, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1286I | Modified Soyabean Bile Broth Base , Granulated | Low risk | 22/04/2019 |
| DCM | GM1084 | Mueller Hinton Agar No. 2, Granulated | Low Risk | 12/08/2015 |
| DCM | GM173 | Mueller Hinton Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM391 | Mueller Hinton Broth, Granulated | Low Risk | 12/08/2015 |
| DCM | GM636 | MYP Agar Base (Phenol Red Egg Yolk Polymyxin Agar Base) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM1269 | Nutrient Agar No.2 , Granulated | Low risk | 12/08/2015 |
| DCM | GM001 | Nutrient Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM002 | Nutrient Broth, Granulated | Low Risk | 12/08/2015 |
| DCM | GM395 | OF Basal Medium, Granulated | Low Risk | 12/08/2015 |
| DCM | GM933 | Orange Serum Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM028 | Peptone Water, Granulated | Low Risk | 04/07/2018 |
| DCM | GM837 | Perfringens Agar Base ( Tryptose Sulphite Cycloserine Agar Base) (T.S.C./S.F.P. Agar Base) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM091 | Plate Count Agar (Standard Methods Agar),Granulated | Low Risk | 28/04/2017 |
| DCM | GMH096 | Potato Dextrose Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM096 | Potato Dextrose Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM085 | Pseudomonas Agar Base , Granulated | Low risk | 22/04/2019 |
| DCM | GM085 | Pseudomonas Agar Base, Granulated | Low Risk | 22/04/2019 |
| DCM | GMH1491 | Rappaport Vassiliadis Salmonella Enrichment Broth , Granulated | Low risk | 22/04/2019 |
| DCM | GM1491 | Rappaport Vassiliadis Soya Broth (RVS Broth) , Granulated | Low Risk | 12/08/2015 |
| DCM | GMH443 | Reinforced Medium for Clostridia , Granulated | Low risk | 22/04/2019 |

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| DCM | GM149 | Robinson's Cooked M Medium (R.C. Medium), Granulated | Low Risk | 16/12/2017 |
| DCM | GM130 | Rogosa SL Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM842 | Rose Bengal Agar Base, Granulated | Low risk | 12/08/2015 |
| DCM | GM1067 | Sabouraud Chloramphenicol Agar, Granulated | Low risk | 25/08/2016 |
| DCM | GM063 | Sabouraud Dextrose Agar , Granulated | Low Risk | 12/08/2015 |
| DCM | GMH063 | Sabouraud Dextrose Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM033 | Sabouraud Dextrose Broth (Sabouraud Liquid Medium) , Granulated | Low Risk | 12/08/2015 |
| DCM | GMH033 | Sabouraud Dextrose Broth , Granulated | Low risk | 22/04/2019 |
| DCM | GMV033 | Sabouraud Dextrose HiVeg™ Broth (Sabouraud Liquid HiVeg™ Medium) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM1313 | Sabouraud Dextrose Maltose Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM062 | Sabouraud Maltose Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1619 | Sakazakii DHL Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM1078 | Salmonella Differential Agar (Twin Pack), Raj Hans Medium (Twin Pack) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM052 | Selenite Broth (Selenite F Broth) (Twin Pack) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM612A | Slanetz and Bartley Medium w/o TTC, Granulated | Low risk | 10/11/2020 |
| DCM | GM298R | Sorbitol Agar (Sorbitol MacConkey Agar) | Low risk | 25/08/2016 |
| DCM | GM290 | Soyabean Casein Digest Agar (Tryptone Soya Agar) , Granulated | Low risk | 22/04/2019 |
| DCM | GM011 | Soyabean Casein Digest Medium (Tryptone Soya Broth) , Granulated | Low risk | 22/04/2019 |
| DCM | GMH011 | Soybean Casein Digest Medium (Casein Soybean Digest Broth) , Granulated | Low risk | 22/04/2019 |
| DCM | GMH290 | Soybean-Casein Digest Agar (Casein Soyabean Digest Agar) , Granulated | Low risk | 22/04/2019 |
| DCM | GM108 | SS Agar (Salmonella Shigella Agar) , Granulated | Low Risk | 12/08/2015 |
| DCM | GM189 | TCBS Agar, Granulated | Low Risk | 12/08/2015 |
| DCM | GM010 | Thioglycollate Broth, Alternative ( Alternative Thioglycollate Medium)(NIH Thioglycollate Broth) , Granulated | Low risk | 12/08/2015 |
| DCM | GM021 | Triple Sugar Iron Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM463 | Tryptone Broth (Tryptone Water) , Granulated | Low risk | 22/04/2019 |
| DCM | GM177 | Tryptose Broth, Granulated | Low Risk | 12/08/2015 |
| DCM | GM112 | Urea Agar Base (Christensen) (Autoclavable) | Low Risk | 30/10/2018 |
| DCM | GM112A | Urea Agar Base (Filter sterilizable), Granulated | Low risk | 25/08/2016 |
| DCM | GM111A | Urea Broth (Filter sterilizable), Granulated | Low risk | 25/08/2016 |
| DCM | GM049 | Violet Red Bile Agar, Granulated | Low risk | 28/04/2017 |
| DCM | GMH581 | Violet Red Bile Glucose Agar , Granulated | Low risk | 22/04/2019 |
| DCM | GM581 | Violet Red Bile Glucose Agar w/o Lactose, Granulated | Low risk | 04/07/2018 |
| DCM | GM031 | Xylose Lysine Deoxycholate Agar (XLD Agar) , Granulated | Low Risk | 12/08/2015 |
| DCM | GMH031 | Xylose-Lysine-Deoxycholate Agar , Granulated | Low risk | 22/04/2019 |

| Product group | Type/ Model / Ref number | Device Name | Risk Class | Date of CE compliance |
|---|---|---|---|---|
| **Dehydrated Culture Media -Supplements** | | | | |
| DCM-S | FD001 | Non Spore Anaerobic Supplement | Low risk | 20/12/2012 |
| DCM-S | FD002 | G.N. Spore Anaerobic Supplement | Low risk | 20/12/2012 |
| DCM-S | FD003 | Polymyxin B Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD003B | Polymyxin B Selective Supplement | Low risk | 04/07/2018 |
| DCM-S | FD004 | Bordetella Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD005 | Brucella Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD006 | Campylobacter Supplement-I (Blaser-Wang) | Low risk | 20/12/2012 |
| DCM-S | FD007 | Campylobacter Supplement - II (Butzler) | Low risk | 20/12/2012 |
| DCM-S | FD008 | Campylobacter Supplement- III (Skirrow) | Low risk | 20/12/2012 |
| DCM-S | FD009 | Campylobacter Growth Supplement | Low risk | 20/12/2012 |
| DCM-S | FD010 | Clostridium Difficile Supplement | Low risk | 20/12/2012 |
| DCM-S | FD013 | S.F.P. Supplement (Perfringens S.F.P. Supplement) | Low risk | 20/12/2012 |
| DCM-S | FD014 | T.S.C. Supplement (Perfringens T.S.C. Supplement) | Low risk | 20/12/2012 |
| DCM-S | FD015 | Dermato Supplement | Low risk | 20/12/2012 |
| DCM-S | FD017 | Legionella Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD018 | Middlebrook OADC Growth Supplement | Low risk | 20/12/2012 |
| DCM-S | FD019 | Middlebrook ADC Growth Supplement | Low risk | 20/12/2012 |
| DCM-S | FD019R | Middlebrook ADC Growth Supplement | Low risk | 10/11/2020 |
| DCM-S | FD020 | Oleic Albumin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD021 | GC Supplement w/ Antibiotics | Low risk | 20/12/2012 |
| DCM-S | FD022 | Haemoglobin Powder | Low risk | 20/12/2012 |
| DCM-S | FD023 | V.C.N. Supplement | Low risk | 20/12/2012 |
| DCM-S | FD024 | V.C.N.T. Supplement | Low risk | 20/12/2012 |
| DCM-S | FD025 | Vitamino Growth Supplement (Twin Pack) | Low risk | 20/12/2012 |
| DCM-S | FD025R | Vitamino Growth Supplement (Twin Pack) | Low risk | 10/11/2020 |
| DCM-S | FD026 | Linco T Supplement | Low risk | 20/12/2012 |
| DCM-S | FD026R | Linco T Supplement | Low risk | 10/11/2020 |
| DCM-S | FD027 | Yeast Autolysate Supplement | Low risk | 20/12/2012 |
| DCM-S | FD028 | Vanclo T Supplement | Low risk | 20/12/2012 |
| DCM-S | FD029 | Cetrinix Supplement | Low risk | 22/04/2019 |
| DCM-S | FD030 | Staph-Strepto Supplement | Low risk | 20/12/2012 |
| DCM-S | FD031 | Strepto supplement | Low risk | 25/08/2016 |
| DCM-S | FD033 | Chloramphenicol Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD034 | Yersinia Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD035 | CC Supplement | Low risk | 20/12/2012 |
| DCM-S | FD036 | CFC Supplement | Low risk | 22/04/2019 |
| DCM-S | FD037 | Legionella Selective Supplement II | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| DCM-S | FD038 | Legionella Selective Supplement III | Low risk | 20/12/2012 |
| DCM-S | FD039 | Aeromonas Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD040 | Legionella Selective Supplement IV (MWY) | Low risk | 20/12/2012 |
| DCM-S | FD041A | Legionella Supplement (Twin Pack) | Low risk | 20/12/2012 |
| DCM-S | FD041AR | Legionella Growth Supplement (Legionella Supplement) (Twin Pack) | Low risk | 25/08/2016 |
| DCM-S | FD042 | Campylobacter Selective Supplement IV (Preston Selective Supplement) | Low risk | 20/12/2012 |
| DCM-S | FD043 | Doyle's Antibiotic Supplement | Low risk | 20/12/2012 |
| DCM-S | FD045 | Egg Yolk Emulsion (100 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD045B | Egg Yolk Emulsion | Low risk | 04/07/2018 |
| DCM-S | FD045L | Egg Yolk Emulsion (50ml per vial) | Low risk | 04/07/2018 |
| DCM-S | FD045R | Egg Yolk Emulsion (100 ml per vial) | Low risk | 25/08/2016 |
| DCM-S | FD045RC | Egg Yolk Emulsion (100 ml per vial) | Low risk | 10/11/2020 |
| DCM-S | FD046 | Egg Yolk Tellurite Emulsion ( 100 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD046B | Egg Yolk Tellurite Emulsion | Low risk | 04/07/2018 |
| DCM-S | FD046L | Egg Yolk Tellurite Emulsion (50ml per vial) | Low risk | 04/07/2018 |
| DCM-S | FD046N | Egg Yolk Tellurite Emulsion, Modified | Low risk | 04/07/2018 |
| DCM-S | FD046NL | Egg Yolk Tellurite Emulsion, Modified | Low risk | 04/07/2018 |
| DCM-S | FD046R | Egg Yolk Tellurite Emulsion | Low risk | 10/11/2020 |
| DCM-S | FD047 | Potassium Tellurite 3.5% (1 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD048 | Urea 40% (5 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD049 | C.B.I. Supplement | Low risk | 20/12/2012 |
| DCM-S | FD052 | Potassium Tellurite 1% (1 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD053 | Gruft Mycobacterial Supplement | Low risk | 20/12/2012 |
| DCM-S | FD054 | GBS Supplement | Low risk | 20/12/2012 |
| DCM-S | FD056 | G. Vaginalis Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD057 | TTC Solution 1% (10 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD059 | Basic Fuchsin (6.0 gm per vial) | Low risk | 20/12/2012 |
| DCM-S | FD061 | Listeria Selective Supplement (PALCAM) | Low risk | 22/04/2019 |
| DCM-S | FD061R | Listeria Selective Supplement (PALCAM) | Low risk | 04/07/2018 |
| DCM-S | FD062 | Bacteroides Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD063 | Listeria Selective Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD063I | Listeria Selective Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD066 | Leptospira Enrichment | Low risk | 20/12/2012 |
| DCM-S | FD068 | Sulpha Supplement | Low risk | 20/12/2012 |
| DCM-S | FD069 | B P Sulpha Supplement | Low risk | 20/12/2012 |
| DCM-S | FD070 | McBride Listeria Supplement | Low risk | 20/12/2012 |
| DCM-S | FD071 | Oxford Listeria Supplement | Low risk | 20/12/2012 |
| DCM-S | FD072 | KL Virulence Enrichment (20 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD072D | KL Virulence Enrichment (500 ml) | Low risk | 10/11/2020 |

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|---|---|---|---|---|
| DCM-S | FD072M | KL Virulence Enrichment (1000 ml) | Low risk | 10/11/2020 |
| DCM-S | FD073 | Diphtheria Virulence Supplement (Part A & B) | Low risk | 20/12/2012 |
| DCM-S | FD075 | Mycoplasma Enrichment Supplement | Low risk | 20/12/2012 |
| DCM-S | FD075R | Mycoplasma Enrichment Supplement | Low risk | 10/11/2020 |
| DCM-S | FD078 | Campylobacter Selective Supplement (Karmali) | Low risk | 10/11/2020 |
| DCM-S | FD082 | Ampicillin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD090 | Campylobacter Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD091 | Bromo Thymol Blue Supplement (20 mg per vial) | Low risk | 20/12/2012 |
| DCM-S | FD092 | MUG Supplement (50 mg per vial) | Low risk | 10/11/2020 |
| DCM-S | FD093 | Bromo Cresol Purple | Low risk | 22/04/2019 |
| DCM-S | FD094 | Trichomonas Selective Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD095 | 10% Lactic Acid Solution (10 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD096 | Novobiocin Supplement | Low risk | 22/04/2019 |
| DCM-S | FD099 | Trichomonas Selective Supplement I | Low risk | 20/12/2012 |
| DCM-S | FD100 | Mueller Tellurite Serum (25 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD102 | Ticarcillin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD103 | Potassium Chlorate Supplement | Low risk | 20/12/2012 |
| DCM-S | FD105 | Park and Sanders Selective Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD106 | Campylobacter Supplement VI (Butzler) | Low risk | 20/12/2012 |
| DCM-S | FD111 | Kimmig Selective Supplement (Twin Pack) | Low risk | 20/12/2012 |
| DCM-S | FD112 | George Kimmig Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD114 | Vitamin K1 Supplement | Low risk | 20/12/2012 |
| DCM-S | FD117 | Haemophilus Growth Supplement | Low risk | 20/12/2012 |
| DCM-S | FD118 | Mucasol | Low risk | 20/12/2012 |
| DCM-S | FD119 | Streptococcus Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD120 | Chlortetracycline Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD126 | Listeria Moxalactam Supplement | Low risk | 20/12/2012 |
| DCM-S | FD130 | Nalidixic Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD132 | Campylobacter Selective Supplement w/ Hemin (Karmali) | Low risk | 10/11/2020 |
| DCM-S | FD135 | CCDA Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD136 | Listeria UVM Supplement I | Low risk | 20/12/2012 |
| DCM-S | FD137 | Listeria UVM Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD142 | Legionella Growth Supplement (BCYE) | Low risk | 10/11/2020 |
| DCM-S | FD142X | Legionella Growth Supplement | Low risk | 10/11/2020 |
| DCM-S | FD143 | Legionella (GVPC) Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD144 | Legionella BMPA Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD147 | Tellurite - Cefixime Supplement | Low risk | 20/12/2012 |
| DCM-S | FD149 | Neomycin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD150 | NYC Supplement | Low risk | 20/12/2012 |
| DCM-S | FD152 | XLT4 Supplement | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| DCM-S | FD153 | M-CP Selective Supplement - I | Low risk | 10/11/2020 |
| DCM-S | FD154 | M-CP Selective Supplement - II | Low risk | 10/11/2020 |
| DCM-S | FD154A | M-CP Selective Supplement, Modified | Low risk | 10/11/2020 |
| DCM-S | FD157 | Urea 5% (5 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD158 | Campylobacter Selective Supplement IV (Preston), Modified | Low risk | 20/12/2012 |
| DCM-S | FD159 | Doyle'S Antibiotic Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD160 | Legionella (GVPA) Selective Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD161 | Brucella Selective Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD163 | Listeria Selective Supplement II, Modified | Low risk | 04/07/2018 |
| DCM-S | FD164 | Park and Sanders Selective Supplement II, Modified | Low risk | 04/07/2018 |
| DCM-S | FD165 | Campylobacter Supplement -II (Butzler), Modified | Low risk | 04/07/2018 |
| DCM-S | FD169 | CC Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD171 | McBride Listeria Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD172 | Oxford Listeria Supplement, Modified | Low risk | 28/04/2017 |
| DCM-S | FD172R | Oxford Listeria Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD173 | Mycoprep (for 2 tests) | Low risk | 16/12/2017 |
| DCM-S | FD173B | Mycoprep (for 10 tests) | Low risk | 16/12/2017 |
| DCM-S | FD175 | Mycoplasma Urogenital Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD176 | Dermato Supplement, Modified | Low risk | 04/07/2018 |
| DCM-S | FD179 | Antibiotic Mixture for Borrelia (100 X) (5 ml per vial) | Low risk | 20/12/2012 |
| DCM-S | FD180 | Rabbit serum | Low risk | 25/08/2016 |
| DCM-S | FD181 | HiCrome™ Listeria Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD183 | Legionella Selective Supplement II, Modified | Low risk | 04/07/2018 |
| DCM-S | FD185 | Anthracis Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD187 | HiCrome™ EC O157 : H7 Selective Supplement | Low risk | 05/11/2020 |
| DCM-S | FD190 | HiCrome°Hicrome ECC Selective Supplement | Low risk | 22/04/2019 |
| DCM-S | FD191 | Oxacillin Resistance Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD192 | HiCrome™ Candida Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD195 | Fibrinogen Plasma Trypsin Inhibitor Supplement | Low risk | 20/12/2012 |
| DCM-S | FD196 | Tetracycline Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD198 | Mycoplasma Cultivation Supplement | Low risk | 20/12/2012 |
| DCM-S | FD201 | Albumin Glucose Supplement | Low risk | 20/12/2012 |
| DCM-S | FD206 | Legionella Growth Supplement w/o L-Cysteine | Low risk | 05/11/2020 |
| DCM-S | FD206R | Legionella Growth Supplement w/o L-Cysteine | Low risk | 10/11/2020 |
| DCM-S | FD212 | L. mono Selective Supplement I | Low risk | 20/12/2012 |
| DCM-S | FD212A | OA Listeria Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD212B | L. mono Selective Supplement I | Low risk | 04/07/2018 |
| DCM-S | FD213 | L. mono Selective Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD214 | L. mono Enrichment Supplement I | Low risk | 20/12/2012 |
| DCM-S | FD215 | Vitamino Growth Supplement, Modified (Twin Pack) | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| DCM-S | FD215B | Vitamino Growth Supplement, Modified | Low risk | 22/04/2019 |
| DCM-S | FD225 | Klebsiella Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD226 | Enterococcus faecium Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD227 | L. mono Enrichment Supplement II | Low risk | 20/12/2012 |
| DCM-S | FD229 | MeReSa Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD230 | HiCrome EC 0157: H7 Selective Supplement | Low risk | 22/04/2019 |
| DCM-S | FD232 | Burkholderia Cepacia Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD233 | Vancomycin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD236 | Sorbic Acid Supplement | Low risk | 20/12/2012 |
| DCM-S | FD241 | Poctri supplement | Low risk | 25/08/2016 |
| DCM-S | FD242 | Legionella Selective Supplement(GVPN) | Low risk | 20/12/2012 |
| DCM-S | FD243 | Clostridium Difficile Supplement | Low risk | 20/12/2012 |
| DCM-S | FD245 | HiCrome™ Nickels & Leesment Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD246 | Cefixime Supplement | Low risk | 20/12/2012 |
| DCM-S | FD247 | ECO157:H7 Selective Supplement | Low risk | 22/04/2019 |
| DCM-S | FD248 | Coagulase Plasma | Low risk | 04/07/2018 |
| DCM-S | FD248A | Coagulase Plasma w/ EDTA (From Rabbit) | Low risk | 22/04/2019 |
| DCM-S | FD248B | Rabbit plasma with EDTA and 15% NaCl | Low risk | 22/04/2019 |
| DCM-S | FD248R | Coagulase Supplement for Staphilococci | Low risk | 22/04/2019 |
| DCM-S | FD252 | Gentamycin Selective Supplement | Low risk | 22/04/2019 |
| DCM-S | FD253 | Urea Solution | Low risk | 20/12/2012 |
| DCM-S | FD254 | Ureaplasma Selective Supplement | Low risk | 04/07/2018 |
| DCM-S | FD255 | Ureaplasma Growth Supplement | Low risk | 20/12/2012 |
| DCM-S | FD259 | Cefoxitin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD261 | Vancomycin Supplement | Low risk | 20/12/2012 |
| DCM-S | FD266 | Listeria Moxalactam Supplement Modified | Low risk | 20/12/2012 |
| DCM-S | FD269 | OFPBL Selective Supplement | Low risk | 25/11/2017 |
| DCM-S | FD270 | Chromogenic Supplement | Low risk | 10/11/2020 |
| DCM-S | FD271 | MDR Acinetobacter Selective Supplement | Low risk | 25/08/2016 |
| DCM-S | FD274 | HiCrome™ Selective Salmonella Agar Supplement | Low risk | 22/04/2019 |
| DCM-S | FD277 | HiCrome™ VRE Agar supplement | Low risk | 20/12/2012 |
| DCM-S | FD278 | HiCrome™ ESBL Agar Supplement | Low risk | 20/12/2012 |
| DCM-S | FD279 | HiCrome™ KPC Agar Supplement | Low risk | 20/12/2012 |
| DCM-S | FD280 | Sterile Charcoal Supplement for Legionella Agar | Low risk | 10/11/2020 |
| DCM-S | FD283R | HiCrome™ Candida Differential Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD284 | Acriflavin-Cefsulodin-Vancomycin Supplement (ACV Supplement) | Low risk | 20/12/2012 |
| DCM-S | FD285 | Bifidobacterium Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD286 | Yersinia Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD287 | Growth Supplement I for MSM | Low risk | 20/12/2012 |
| DCM-S | FD288 | Growth Supplement II for MSM | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| DCM-S | FD290 | Novobiocin Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD295 | HiCrome™ ECO157:H7 Selective Supplement Modified | Low risk | 22/04/2019 |
| DCM-S | FD299 | Selective Supplement for MRSA | Low risk | 20/12/2012 |
| DCM-S | FD300 | Hayflick Supplement | Low risk | 20/12/2012 |
| DCM-S | FD302 | Group A Selective Supplement | Low risk | 20/12/2012 |
| DCM-S | FD304 | Arcobacter Selective Supplement | Low risk | 05/11/2020 |
| DCM-S | FD306 | Modified Listeria Oxford Selective Supplement | Low risk | 22/04/2019 |
| DCM-S | FD309 | Monensin Selective Supplement | Low risk | 22/04/2019 |
| DCM-S | FD312 | VIA Supplement | Low risk | 20/12/2012 |
| DCM-S | FD319 | MRSA Supplement | Low risk | 25/08/2016 |
| DCM-S | FD319R | MeReSa Selective Supplement (MRSA Selective Supplement) | Low risk | 25/08/2016 |
| DCM-S | FD320 | Clostridium difficle  Selective Supplement | Low risk | 25/08/2016 |
| DCM-S | FD321 | TVCSB Supplement | Low risk | 25/08/2016 |
| DCM-S | FD322 | Middlebrook ADC Growth Supplement, Modified | Low risk | 25/08/2016 |
| DCM-S | FD323 | TSBV Supplement | Low risk | 25/08/2016 |
| DCM-S | FD324 | Bacillus Selective Supplement | Low risk | 25/08/2016 |
| DCM-S | FD327 | NAD Supplement | Low risk | 25/08/2016 |
| DCM-S | FD329 | Middlebrook OADC Enrichment Supplement | Low risk | 25/08/2016 |
| DCM-S | FD332 | Lecithin solution | Low risk | 10/11/2020 |
| DCM-S | FD333 | Modified L.mono Selective supplement | Low risk | 10/11/2020 |
| DCM-S | FD334 | Mycoplasma selective supplement | Low risk | 25/08/2016 |
| DCM-S | FD335 | Leeds Acinetobacter selective supplement | Low risk | 25/08/2016 |
| DCM-S | FD335R | MDR Acinetobacter Selective Supplement | Low risk | 25/08/2016 |
| DCM-S | FD338 | LCN Supplement | Low risk | 25/08/2016 |
| DCM-S | FD340 | PACT Supplement | Low risk | 28/04/2017 |
| DCM-S | FD342 | Rapid Listeria Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD343 | Growth Supplement for Fastidious Organism | Low risk | 16/12/2017 |
| DCM-S | FD344 | ECC Selective Supplement Modified | Low risk | 22/04/2019 |
| DCM-S | FD345 | Ciprofloxacin Supplement | Low risk | 10/11/2020 |
| DCM-S | FD347 | PCP Supplement | Low risk | 04/07/2018 |
| DCM-S | FD347B | PCP Supplement | Low risk | 10/11/2020 |
| DCM-S | FD348 | OADS Supplement | Low risk | 16/12/2017 |
| DCM-S | FD349 | Vancomycin Polymyxin B Supplement | Low risk | 04/07/2018 |
| DCM-S | FD352 | Acinetobacter Selective Supplement | Low risk | 30/10/2018 |
| DCM-S | FD353 | VCAT Supplement | Low risk | 30/10/2018 |
| DCM-S | FD354 | STEC Selective Supplement | Low risk | 30/10/2018 |
| DCM-S | FD355 | HiCrome™ Colistin Resistant Selective Supplement | Low risk | 30/10/2018 |
| DCM-S | FD356 | Diphenyl supplement | Low risk | 22/04/2019 |
| DCM-S | FD357 | Carba Selective Supplement | Low risk | 10/11/2020 |
| DCM-S | FD360 | C.auris Selective Supplement | Low risk | 05/11/2019 |
| DCM-S | FD361 | BCSA Selective Supplement | Low risk | 05/11/2019 |

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|---|---|---|---|---|
| DCM-S | FD362 | Coagulase Supplement (for M2126) | Low risk | 10/11/2020 |
| DCM-S | FD363 | HiMRSA Selective Supplement | Low risk | 17/06/2021 |
| DCM-S | FD725R | Mycoprep (Modified,Bulk powder) | Low risk | 25/08/2016 |
| DCM-S | FD726 | Mycoprep (Modified, powder for 1000ml) | Low risk | 25/08/2016 |
| DCM-S | FD743R | Bifido Selective Supplement C | Low risk | 25/08/2016 |
| DCM-S | FD744R | Bifido Selective Supplement D | Low risk | 25/08/2016 |
| DCM-S | FD745R | Bifido Selective Supplement E | Low risk | 25/08/2016 |
| DCM-S | FD749 | Supplement for HiCrome™ Candida Agar | Low risk | 04/07/2018 |
| DCM-S | FD750 | L. J. Media Supplement w/ Capreomycin | Low risk | 25/08/2016 |
| DCM-S | FD751 | L. J. Medium Supplement w/ Clarithromycin | Low risk | 25/08/2016 |
| DCM-S | FD752 | L. J. Media Supplement w/ D-Cycloserine | Low risk | 25/08/2016 |
| DCM-S | FD753 | L. J. Media Supplement w/ Ethambutol | Low risk | 25/08/2016 |
| DCM-S | FD754 | L. J. Media Supplement w/ Ethionamide | Low risk | 25/08/2016 |
| DCM-S | FD755 | L. J. Medium Supplement w/ Gatifloxacin | Low risk | 25/08/2016 |
| DCM-S | FD756 | L. J. Media Supplement w/ Isoniazide | Low risk | 25/08/2016 |
| DCM-S | FD757 | L. J. Media Supplement w/ Kanamycin | Low risk | 25/08/2016 |
| DCM-S | FD758 | L. J. Medium Supplement w/ Levofloxacin | Low risk | 25/08/2016 |
| DCM-S | FD759 | L. J. Medium Supplement w/ Lomefloxacin | Low risk | 04/07/2018 |
| DCM-S | FD760 | L. J. Medium Supplement w/ Ofloxacin | Low risk | 04/07/2018 |
| DCM-S | FD761 | L. J. Medium Supplement w/ p-Aminosalicylic acid | Low risk | 04/07/2018 |
| DCM-S | FD762 | L. J. Medium Supplement w/ Pyrazinamide | Low risk | 04/07/2018 |
| DCM-S | FD763 | L.J.Medium Supplementw/Rifabutin | Low risk | 04/07/2018 |
| DCM-S | FD764 | L.J.Medium Supplementw/Rifampicin | Low risk | 04/07/2018 |
| DCM-S | FD765 | L.J.Medium Supplementw/Sodium Salicylate | Low risk | 04/07/2018 |
| DCM-S | FD766 | L.J.Medium Supplementw/Streptomycin | Low risk | 04/07/2018 |
| DCM-S | FD767 | L.J.Medium Supplementw/TCH | Low risk | 04/07/2018 |
| DCM-S | FD768 | Chloramphenicol Supplement | Low risk | 04/07/2018 |
| DCM-S | FD772 | L.J. Media Supplement w/Amikacin | Low risk | 04/07/2018 |
| DCM-S | FD775 | L.J. Media Supplement w/ p-Nitrobenzoic acid | Low risk | 04/07/2018 |
| DCM-S | FD780 | L.J. Media Supplement w/Moxifloxacin | Low risk | 04/07/2018 |
| DCM-S | FD804 | Enriched growth Supplement for Mycobacteria | Low risk | 04/07/2018 |
| DCM-S | FD805 | Growth Supplement for Anaerobic cultures | Low risk | 10/11/2020 |
| DCM-S | FD808 | Supplement for GC Agar Base | Low risk | 10/11/2020 |
| DCM-S | FD812 | Selective Supplement for Gram positive bacteria (Clostridium, Staphylococcus spp. etc.) | Low risk | 10/11/2020 |
| DCM-S | FD814 | PANTA Supplement | Low risk | 10/11/2020 |
| DCM-S | FD815B | Selective Supplement for SS Agar | Low risk | 10/11/2020 |
| DCM-S | FD816 | Selective supplement for Enterobacteriaceae | Low risk | 10/11/2020 |
| DCM-S | FD817 | Selective Supplement for Staphylococcus | Low risk | 10/11/2020 |
| DCM-S | FD820 | Selective Supplement for SS Agar | Low risk | 10/11/2020 |

| Product group | Type/ Model / Ref number | Device Name | Risk Class | Date of CE compliance |
|---|---|---|---|---|
| **Ready Prepared Media** | | | Low risk | 10/06/2021 |
| RPM - Ready Prepared Plates | HB001 | HiCombi™ Nutrient - MacConkey Agar Plate | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB003 | HiCombi™ CLED - MacConkey Agar Plate | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB004 | HiCombi™ XLD - MacConkey Agar Plate | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB005 | HiCombi™ Cetrimide - MacConkey Agar Plate | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB006 | HiCombi™ Blood- MacConkey Agar Plate | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB007 | HiCombi™ MacConkey-Mannitol Salt Agar | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB008 | HiCombi™ Blood -Chocolate Agar | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB009 | HiCombi™ Blood -Mannitol Salt Agar | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB010 | HiCombi™ Chocolate - MacConkey Agar Plate | Low risk | 20/12/2012 |
| RPM - Ready Prepared Plates | HB017 | HiCombi™ Sabouraud Dextrose-Sheep Blood Agar Plate | Low risk | 17/06/2021 |
| RPM- HiDip Slides | HD001 | HiDip™ Cled-Cetri-Mac Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD002 | HiDip™ Mac-Cled-Sab Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD003 | HiDip™ Mac-Cled-Bile Esculin Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD004 | HiDip™ Cled-Mac Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD005 | HiDip™ Cled-MUG Mac Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD006 | HiDip™ Cled-HiCrome™ UTI Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD007 | HiDip™ Mac-HiCrome™ UTI Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD007R | HiDip™ Mac-HiCrome™ UTI Medium | Low risk | 10/11/2020 |
| RPM- HiDip Slides | HD018 | HiDip™ TSA-CLED Agar w/ B.T.B Indicator Medium | Low Risk | 20/12/2012 |
| RPM- HiDip Slides | HD020 | HiDip™ Pseudomonas Agar - MacConkey Agar Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD021 | HiDip™ PCA - MacConkey Agar Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD024 | HiDip™ Modified Rogosa Medium-Modified Rogosa Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD025 | HiDip™ Modified Nickerson Medium-Modified Nickerson Medium | Low risk | 20/12/2012 |
| RPM- HiDip Slides | HD041 | HiDip HiCrome™ Universal Agar-PCA | Low risk | 28/04/2017 |
| RPM- HiDip Slides | HD042 | HiDip HiCrome™ UTI Agar - Dey Engley Neutralzing agar | Low risk | 28/04/2017 |
| RPM- HiDip Slides | HD046 | HiDip TSA-TCBS | Low risk | 30/10/2018 |
| RPM- HiDip Slides | HD047 | HiDip TSA-MRS | Low risk | 30/10/2018 |
| RPM- HiSafe Blood Culturing System | LQ003 | BHI | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ003A | BHI | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ004 | BHI - Supplemented w/ 0.05% SPS | Low risk | 20/12/2012 |

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| RPM- HiSafe Blood Culturing System | LQ004R | BHI - Supplemented w/ 0.05% SPS | Low risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ004A | BHI - Supplemented w/ 0.05% SPS | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ004AR | BHI - Supplemented w/ 0.05% SPS | Low risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ005 | TSB - Tryptone Soya Broth w/ 10% Sucrose | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ005A | TSB - Tryptone Soya Broth w/ 10% Sucrose | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ006 | Columbia Broth | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ006A | Columbia Broth | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ007 | Thioglycollate Broth | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ007R | Thioglycollate Broth | Low Risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ007A | Thioglycollate Broth | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ007AR | Thioglycollate Broth | Low Risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ008 | Schaedler Broth | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ008A | Schaedler Broth | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ009 | TSB - Tryptone Soya Broth | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ009A | TSB - Tryptone Soya Broth | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ010 | Glucose Broth Supplemented w/ 0.05% SPS | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ010A | Glucose Broth Supplemented w/ 0.05% SPS | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ010AR | Glucose Broth Supplemented w/ 0.05% SPS | Low risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ010V | Glucose Broth supplemented w/0.05% SPS | Low risk | 22/04/2019 |
| RPM- HiSafe Blood Culturing System | LQ011 | TSB - Tryptone Soya Broth Supplemented w/ 0.05% SPS | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ011A | TSB - Tryptone Soya Broth Supplemented w/ 0.05% SPS | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ011AR | TSB - Tryptone Soya Broth Supplemented w/ 0.05% SPS | Low risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ013V | Hartley Broth | Low risk | 22/04/2019 |
| RPM- HiSafe Blood Culturing System | LQ014 | Modified Wilkins Chalgren Broth | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ014A | Modified Wilkins Chalgren Broth | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ012 | HiCombi™ Dual Performance Medium | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ012R | HiCombi™ Dual Performance Medium | Low Risk | 10/11/2020 |

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| RPM- HiSafe Blood Culturing System | LQ013 | Hartley Broth | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ013A | Hartley Broth | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ023 | Fluid thioglycollate Medium w/0.05% SPS | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ023A | Fluid thioglycollate Medium w/0.05% SPS | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ029 | HiCombi™ Dual Performance Salmonella Medium - SS | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ029A | HiCombi™ Dual Performance Salmonella Medium - SS | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ029AR | HiCombi™ Dual Performance Salmonella Medium - SS | Low Risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ030 | HiCombi™ Dual Performance Salmonella Medium - XLD | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ030A | HiCombi™ Dual Performance Salmonella Medium - XLD | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ031 | HiCombi™ Dual Performance Salmonella Medium - DCA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ031A | HiCombi™ Dual Performance Salmonella Medium - DCA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ031AR | HiCombi™ Dual Performance Salmonella Medium - DCA | Low risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ032 | HiCombi™ Dual Performance Salmonella Medium - HEA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ032A | HiCombi™ Dual Performance Salmonella Medium - HEA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ033 | HiCombi™ Dual Performance Medium | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ033R | HiCombi™ Dual Performance Medium | Low risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ034 | HiCombi™ Dual Performance Fungal Medium Kit | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ034R | HiCombi™ Dual Performance Fungal Medium Kit | Low Risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ034A | HiCombi™ Dual Performance Fungal Medium Kit | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ034AR | HiCombi™ Dual Performance Fungal Medium Kit | Low Risk | 10/11/2020 |

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| RPM- Ready Prepared Dual Media | LQ035 | HiCombi™ Dual Performance Selective Medium - HEA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ035A | HiCombi™ Dual Performance Selective Medium - HEA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ035AR | HiCombi™ Dual Performance Selective Medium - HEA | Low risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ036 | HiCombi™ Dual Performance Selective Medium - SS | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ036R | HiCombi™ Dual Performance Selective Medium - SS | Low risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ036A | HiCombi™ Dual Performance Selective Medium - SS | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ036AR | HiCombi™ Dual Performance Selective Medium - SS | Low risk | 10/11/2020 |
| RPM- Ready Prepared Dual Media | LQ037 | HiCombi™ Dual Performance Selective Medium - HEA | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ038 | HiCombi™ Dual Performance Selective Medium - SS | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ038A | HiCombi™ Dual Performance Selective Medium - SS | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ004Al | BHI-Supplemented w/0.05% SPS | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ004AL | BHI-Supplemented w/0.05% SPS | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ004V | BHI - Supplemented w/ 0.05% SPS | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ0151 | Medium 11. GN Broth | Low risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ069 | Alkaline Peptone Water | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ069R | Enrichment Medium For Vibrio | Low Risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ070 | Selenite Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ070V | Selenite Broth | Low Risk | 25/08/2016 |

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| RPM- Ready Prepared Liquid Medium | LQ077 | BHI Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ077V | BHI Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ077R | Enrichment Medium | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ079 | Bile Broth | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ079V | Bile Broth | Low Risk | 22/04/2019 |
| RPM- Ready Prepared Liquid Medium | LQ080 | Cooked M Medium | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ080C | Cooked M Medium | Low Risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ080V | Cooked M Medium | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ088 | Tetrathionate Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ089 | Peptone Water | Low risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ089X | Peptone Water | Low risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ093 | Cooked M Medium w/ Glucose, Hemin & Vitamin K | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ095 | Hartley Broth w/ 0.05% SPS | Low Risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ095A | Hartley Broth w/ 0.05% SPS | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ104 | Rappaport Vassiliadis Salmonella Enrichment Broth | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ104C | Rappaport Vassiliadis Salmonella Enrichment Broth | Low Risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ104V | Rappaport Vassiliadis Salmonella Enrichment Broth | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ104XX | Rappaport Vassiliadis Salmonella Enrichment Broth | Low Risk | 28/04/2017 |
| RPM- Ready Prepared Liquid Medium | LQ105 | Kirchner Medium Base | Low Risk | 20/12/2012 |

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| RPM- Ready Prepared Dual Media | LQ109 | HiCombi™ Dual Performance Trans Isolate Medium | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ109R | HiCombi™ Dual Performance Trans Isolate Medium | Low risk | 10/11/2020 |
| RPM- Ready Prepared Liquid Medium | LQ126 | Urea Indole Medium | Low risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ129 | Sabouraud's Dextrose Broth | Low risk | 04/07/2018 |
| RPM- Ready Prepared Liquid Medium | LQ129V | Sabouraud's Dextrose Broth | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ132 | Campylo Thioglycollate Broth w/Selective Supplement | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ134 | L Broth | Low Risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ146 | Mannitol Selenite Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ157 | GN Broth, Hajna | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ159 | Hayflick Medium | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ170 | Selective Enrichment Medium For Group B | Low risk | 25/08/2016 |
| RPM- Ready Prepared Liquid Medium | LQ180V | Brucella Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ181V | Mannitol Salt Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ182V | Mueller Hinton Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ296X | Hugh Leifson Medium | Low risk | 10/11/2020 |
| RPM- HiSafe Blood Culturing System | LQ188 | HiCombi™ Dual Performance Fungal Medium, Modified | Low risk | 20/12/2012 |
| RPM- HiSafe Blood Culturing System | LQ208 | Eugonic LT100 Broth | Low Risk | 22/04/2019 |
| RPM- Ready Prepared Liquid Medium | LQ208L | Eugonic LT100 Broth | Low risk | 28/04/2017 |
| RPM- Ready Prepared Liquid Medium | LQ208CCL | Eugonic LT100 Broth | Low risk | 28/04/2017 |

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| RPM- Ready Prepared Liquid Medium | LQ210C | BHI Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Liquid Medium | LQ210D | BHI Broth | Low risk | 20/12/2012 |
| RPM- Ready Prepared Dual Media | LQ241 | HiCombi Trans Isolate Medium | Low risk | 28/04/2017 |
| RPM- Ready Prepared Liquid Medium | LQ246CCL | Sauton's Fluid Medium Base | Low risk | 16/12/2017 |
| RPM- Ready Prepared Liquid Medium | LQ314II | HiMiC™ Diluent | Low risk | 10/11/2020 |
| RPM- Ready Prepared Liquid Medium | LQ319V | Thioglycollate Medium with Hemin & Vitamin K | Low risk | 17/06/2021 |
| RPM- Ready Prepared Liquid Medium | LQ319VIII | Thioglycollate Medium with Hemin & Vitamin K | Low risk | 17/06/2021 |
| RPM- Ready Prepared Liquid Medium | LQ089CCLR | Peptone Water | Low risk | 17/06/2021 |
| RPM -Ready Prepared Plates | MP001 | Nutrient Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP001L | Nutrient Agar Plate (150mm plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP015 | Hoyles Media Plate with supplements. | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP016 | Brilliant Green Agar, Modified Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP022 | EMB Agar, Levine Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP023 | Vogel Johnson Agar Plate (V.J. Agar Plate) | Low Risk | 22/04/2019 |
| RPM -Ready Prepared Plates | MP024 | Cetrimide Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP029 | Endo Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP031 | Xylose Lysine Deoxycholate Agar (XLD Agar) Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP043 | Baird Parker Agar Plate | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP043L | Baird Parker Agar Plate | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP043M | Baird Parker Agar Plate (150mm) | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP049 | Violet Red Bile Agar Plate | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP063 | Sabouraud Dextrose Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP063L | Sabouraud Dextrose Agar Plate (150 mm plate) | Low Risk | 20/12/2012 |

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| RPM -Ready Prepared Plates | MP063M | Sabouraud Dextrose Agar Plate (120 mm plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP065 | Deoxycholate Citrate Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP073 | Blood Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP074 | Brucella Agar Plate | Low Risk | 22/04/2019 |
| RPM -Ready Prepared Plates | MP081 | MacConkey Agar w/ 0.15% Bile Salts, CV and NaCl Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP081XL | MacConkey Agar w/ 0.15% Bile Salts, CV and NaCl Plate (200mm plate) | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP082 | MacConkey Agar w/o CV, NaCl w/ 0.5% Sodium Taurocholate Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP091 | Plate Count Agar Plate | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP103 | Chocolate Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP108 | SS Agar (Salmonella Shigella Agar) Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1032 | SS Agar Plate, Modified (Salmonella Shigella Agar Plate, Modified) | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1039 | Brucella Agar Plate with Hemin & Vitamin K1 | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1057 | G. vaginilis Selective Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1067 | Sabouraud Chloramphenicol Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1084 | Mueller Hinton Agar No. 2 Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1084HB | Mueller Hinton Agar No.2 Plate w/ Horse Blood | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1084SB | Mueller Hinton Agar No.2 Plate w/ Sheep Blood | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1139 | Modified MYP Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP118 | Mannitol Salt Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1259 | Haemophilus Test Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1260 | Tellurite Blood Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1295 | HiCrome™ E.coli Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1297A | HiCrome™TM Candida Differential Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1301 | Sheep Blood Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1301C | Sheep Blood Agar Plate (Individually Packed) | Low risk | 28/04/2017 |
| RPM -Ready Prepared Plates | MP1301M | Sheep Blood Agar Plate | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP1345 | Anaerobic Blood Agar Plate w/Neomycin | Low risk | 30/10/2018 |

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| RPM -Ready Prepared Plates | MP1353 | HiCrome™ UTI Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP137 | Malt Extract Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP144 | Columbia 5% Sheep Blood Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1418 | HiCrome™ UTI Agar Plate, Modified | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1454 | Oxacillin Resistant Screening Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1540I | HiCrome™ Listeria Ottaviani Agosti Agar Plate | Low Risk | 17/06/2021 |
| RPM -Ready Prepared Plates | MP1548 | Chocolate No. 2 Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1594 | MeReSa Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP160 | DCLS Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1600 | HiCrome™ Universal Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1640 | Burkholderia Cepacia Agar Plate | Low Risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP1674 | HiCrome™ MeReSa Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1682 | HiCrome™ Vibrio Agar Plate | Low Risk | 17/06/2021 |
| RPM -Ready Prepared Plates | MP1702 | MacConkey Agar RS Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP173 | Mueller Hinton Agar Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP173C | Mueller Hinton Agar Plate (100 mm Plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP173L | Mueller Hinton Agar Plate (150mm plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP173M | Mueller Hinton Agar Plate (120mm plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP173XL | Mueller Hinton Agar Plate (200mm plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP173SP | Mueller Hinton Agar Plate (150 mm scored plate) | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP175 | Bordet Gengou Agar Plate w/15% Sheep blood | Low Risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP175SB | Bordet Gengou Agar Plate with 25% Sheep Blood | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1763 | VRE Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP180 | Lead Acetate Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1806 | Mueller Hinton Agar plate w/ 5% Sheep Blood | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1806M | Mueller Hinton Agar plate w/ 5% Sheep Blood | Low Risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP1811 | OFBL Agar Plate (Oxidation Fermentation Polymyxin Bacitracin Lactose Agar Plate) | Low risk | 10/11/2020 |

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| RPM -Ready Prepared Plates | MP1825 | Mueller Hinton Agar Plate with 2% Glucose w/Methylene Blue | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1829 | HiCrome™ ESBL Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1831 | HiCrome™ KPC Agar Plate | Low Risk | 28/04/2017 |
| RPM -Ready Prepared Plates | MP1837 | HiCrome™ Staph Agar Plate, Modified | Low Risk | 28/04/2017 |
| RPM -Ready Prepared Plates | MP1858 | Bifidobacterium Agar Modified Plate | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP188 | D.T.M Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1832 | HiCrome™ Coliform Agar Plate, Modified | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP1925 | HiCrome™ VRE Agar Plate | Low Risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1938 | HiCrome™ Acinetobacter Agar Plate | Low Risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP1947 | Enriched Tryptone Soya Agar Plate (ETSA) | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1948 | Tryptone Soya Serum Bacitracin Vancomycin Agar (TSBV) | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1949 | Tryptone Soya Agar w/ Hemin & Menadione | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP1966 | HiCrome™ Strep B Selective Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP1974 | HiCrome™ Rapid MRSA Agar Plate | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP2062I | HiCrome™ Cronobacter Isolation Agar Plate (CCI Agar Plate) | Low risk | 17/06/2021 |
| RPM -Ready Prepared Plates | MP2085 | Martin Lewin Agar | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP2089 | Burkholderia cepacia Selective Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP211 | BHI Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP2116 | HiCrome™ Salmoconfirm Selective Agar Plate | Low risk | 17/06/2021 |
| RPM -Ready Prepared Plates | MP217 | Bi.G.G.Y. Agar Plate (Nickerson Agar Plate) | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP291 | Schaedler Agar Plate | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP1296 | HiCrome™ Salmonella Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP298 | MacConkey Sorbitol Agar Plate | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP317 | EMB Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP406 | Pseudomonas Isolation Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP413 | Thayer Martin Agar Plate w/VCNT | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP467 | Hektoen Enteric Agar Plate | Low risk | 04/07/2018 |

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| RPM -Ready Prepared Plates | MP491 | Anaerobic Agar (Brewer) Plate | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP540 | Phenylethyl Blood Agar Plate w/ 5% Sheep Blood | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP5269 | Modified Nickerson Medium | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5208 | CNA Agar Plate with 5% Sheep Blood | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP5304 | Blood agar Plate w/5mg/l Gentamicin | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP5316 | Chocolate Agar Plate w/ 5% Sheep Blood | Low risk | 28/04/2017 |
| RPM -Ready Prepared Plates | MP5332 | Sabouraud Dextrose Agar Plate w/Chloramphenicol & gentamicin | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP5333 | Chocolate Agar Plate w/ Bacitracin | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP5334 | Sabouraud Dextrose Agar plate w/Penicillin & Streptomycin | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP5339 | Regan Lowe Agar Plate (Charcoal Blood Plate w/Cephalexin) | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP5340 | Bordet Gengou Blood Agar Plate w/Cephalexin | Low risk | 16/12/2017 |
| RPM -Ready Prepared Plates | MP5380 | BHI Agar Plate w/ Blood | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5381 | BHI Agar Plate w/ Vancomycin | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5382 | BHI Blood agar plate w/ Vancomycin | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5383 | BCYE Selective Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5384 | GBS Agar | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5386 | Sabouraud Dextrose Agar Plate w/Gentamicin | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5387 | Sabouraud Dextrose Agar Plate w/ Cycloheximide | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5389 | Mueller Hinton Agar Plate w/ 2% NaCL | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5390 | Helicobacter Pylori Selective Agar | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP5476 | Mucormycosis Selective Agar Plate | Low risk | 10/06/2021 |
| RPM -Ready Prepared Plates | MP5477 | Candida Selective Agar Plate | Low risk | 10/06/2021 |
| RPM -Ready Prepared Plates | MP511 | Middlebrook 7H11 Agar w/TCH | Low risk | 04/07/2018 |
| RPM -Ready Prepared Plates | MP5426 | Middlebrook 7H11 Agar w/ PANTA supplement | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MP616 | Tergitol-7 Agar Plate | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP641 | MRS Agar Plate | Low risk | 28/04/2017 |
| RPM -Ready Prepared Plates | MP636C | MYP Agar Plate (100mm plate) | Low risk | 10/11/2020 |

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| RPM -Ready Prepared Plates | MP641-I | MRS Agar w/ 10 ppm cycloheximide | Low risk | 28/04/2017 |
| RPM -Ready Prepared Plates | MP664 | Sabouraud Dextrose Agar Plate w/Chloramphenicol (50mg/L) and Cycloheximide 500mg/L | Low Risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP792 | CLED Agar w/ Bromothymol Blue Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MP805 | Bacteroides Bile Esculin Agar Plate | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP813I | BCYE Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP843 | Yersinia Selective Agar Plate | Low risk | 30/10/2018 |
| RPM -Ready Prepared Plates | MP870 | TCBS Agar Plate | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP975A | Anaerobic Blood Agar Plate | Low risk | 25/08/2016 |
| RPM -Ready Prepared Plates | MP994 | Campylobacter Agar Plate | Low risk | 20/12/2012 |
| RPM -Ready Prepared Plates | MPV081 | MacConkey HiVeg™ Agar Plate w/ CV, Nacl, 0.003% NR and 1.5% Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | MPV173 | Mueller Hinton HiVeg™ Agar Plate | Low risk | 10/11/2020 |
| RPM -Ready Prepared Plates | QP001 | Middlebrooke 7H11 Agar Plate | Low risk | 04/07/2018 |
| RPM- Transport Medium w/ swabs | MQ651P | HiCulture™ Transport Swabs w/ Amies Medium w/ Charcoal | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MQ5203P | HiCulture™ Transport Swab w/ Enteric Pathogen Transport Medium | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MQ306P | HiCulture™ Transport Swabs w/ Stuart Transport Medium | Low risk | 20/12/2012 |
| RPM- Viral Transport Medium w/ swabs | AL167 | HiViral Transport Medium | Low risk | 20/12/2012 |
| RPM- Viral Transport Medium w/ swabs | MS052A | HiCulture™ Transport Swabs w/Selenite Medium (A) | Low risk | 25/08/2016 |
| RPM- Viral Transport Medium w/ swabs | MS316 | HiCulture™ Transport Swabs w/CVTR Medium | Low risk | 20/12/2012 |
| RPM- Viral Transport Medium w/ swabs | MS316S | HiCulture™ Transport Swabs w/CVTR Medium | Low risk | 10/11/2020 |
| RPM- Viral Transport Medium w/ swabs | MS316SR | HiCulture™ Transport Swabs w/CVTR Medium w/metal stick | Low risk | 10/11/2020 |
| RPM- Viral Transport Medium w/ swabs | MS316A | HiCulture™ Transport Swabs w/CVTR Medium,Modified | Low risk | 25/08/2016 |
| RPM- Viral Transport Medium w/ swabs | MS1145 | HiCulture™ Listeria Isolation and Transport Swabs | Low risk | 20/12/2012 |
| RPM- Viral Transport Medium w/ swabs | MS1145R | HiCulture™ Listeria Isolation and Transport Swabs | Low risk | 10/11/2020 |

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| RPM- Viral Transport Medium w/ swabs | MS1145S | HiCulture™ Listeria Isolation and Transport Swabs with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS1514 | Hiculture™ Transport swabs w/Modified Campylobacter Thioglycollate Medium | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS1514R | Hiculture™ Transport swabs w/Modified Campylobacter Thioglycollate Medium in polystyrene tube | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS1557 | Hiculture™ Transport swabs w/BHI broth for H.pylori | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS1759 | Hiculture™ Transport swabs | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS2016A | HiCulture™ Transport Swabs w/ Soyabean Casein Digest Medium w/6.5% NaCL | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS2016B | HiCulture™ Transport Swabs w/ Soyabean Casein Digest Medium w/6.5% NaCL | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS202 | HiCulture™ Transport Swabs w/ Cary Blair Medium | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS202A | HiCulture™ Transport Swabs w/ Cary Blair Medium (A) | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS202R | HiCulture™ Transport Swabs w/ Cary Blair Medium in polystyrene tube | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS202S | HiCulture™ Transport Swabs w/ Cary Blair Medium with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS2055 | HiCulture™ Transport Medium for Helicobacter pylori | Low risk | 28/04/2017 |
| RPM- Transport Medium w/ swabs | MS2127 | HiCulture™ Transport Swab w/ Todd Hewitt Broth w/Colistin & Nalidixic Acid | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS306 | HiCulture™ Transport Swabs w/ Stuart Transport Medium | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS306R | HiCulture™ Transport Swabs w/ Stuart Transport Medium | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS306S | HiCulture™ Transport Swabs w/ Stuart Transport Medium with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS5002 | HiCulture™ Transport Swabs w/ 0.85% Sodium chloride and 0.1% Buffered Ppetone Water in polystyrene tube | Low risk | 04/07/2018 |
| RPM- Transport Medium w/ swabs | MS5215 | HiViral™ Transport Medium for Cloacal Samples | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS5296 | HiCulture™ Skim Milk Tryptone Glucose Glycerin Medium swabs | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS5321 | HiCulture Sterile swabs w/ 0.9% Saline | Low risk | 22/04/2019 |
| RPM- Transport Medium w/ swabs | MS651 | HiCulture™ Transport Swabs w/ Amies Medium w/ Charcoal | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS651R | HiCulture™ Transport Swabs w/ Amies Medium w/ Charcoal in polystyrene tube | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS651S | HiCulture™ Transport Swabs w/ Amies Medium w/ Charcoal with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS651SR | HiCulture™ Transport Swabs w/ Amies Medium w/ Charcoal with metal stick | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS684 | HiCulture™ Transport Swabs w/ Amies Medium w/o Charcoal | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS684R | HiCulture™ Transport Swabs w/ Amies Medium w/o Charcoal in polystyrene tube | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS684A | HiCulture™ Transport Swabs w/ Amies Medium (A) | Low risk | 25/08/2016 |

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| RPM- Transport Medium w/ swabs | MS684B | HiCulture™ Transport Swabs w/ Amies Medium (B) | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS684C | HiCulture™ Transport Swabs w/ Amies Medium (C) | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS684D | HiCulture™ Transport Swabs w/ Amies Medium (D) | Low risk | 25/08/2016 |
| RPM- Transport Medium w/ swabs | MS684S | HiCulture™ Transport Swabs w/ Amies Medium w/o Charcoal with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS010 | HiCulture™ Transport Swabs w/ Alternative Thioglycollate Medium | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS010R | HiCulture™ Transport Swabs w/ Alternative Thioglycollate Medium in polystyrene tube | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS010S | HiCulture™ Transport Swabs w/ Alternative Thioglycollate Medium with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS113 | HiCulture™ Transport Swabs w/ Chlamydospore Medium | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS113R | HiCulture™ Transport Swabs w/ Chlamydospore Medium in polystyrene tube | Low risk | 10/11/2020 |
| RPM- Transport Medium w/ swabs | MS113S | HiCulture™ Transport Swabs w/ Chlamydospore Medium with metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS198S | HiCulture™ Transport Swab w/ Middlebrook 7H9 Broth w/metal stick | Low risk | 20/12/2012 |
| RPM- Transport Medium w/ swabs | MS5478 | HiFungal Transport medium w/ Swab | Low risk | 10/06/2021 |
| RPM- Ready Prepared Medium | MT001 | Modified Middlebrook 7H9 Broth with Indicator | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001 | L.J. Medium Slant | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001H | L.J. Medium in glass bottle | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL001B | L.J. Medium Slant | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001L | L.J. Medium Slant in long tube | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001LD | L.J. Medium Slant (in long tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001M | L.J.Medium Slant (In Medium Length tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001T | L.J. Medium Slant in thick glass bottles | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL001X | L.J. Medium Slant | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL002 | L.J.Medium Kit | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL003 | L.J.Medium Plus Kit | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL004 | L.J.Medium w/ Pyruvate | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL004L | L.J.Medium w/ Pyruvate (0.2%) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL005 | L.J.Medium w/ Streptomycin (4 mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL005L | L.J.Medium w/Streptomycin (4 mcg / ml) | Low risk | 20/12/2012 |

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| RPM- L.J.Medium Slants | SL006 | L.J.Medium w/ INH | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL007 | L.J.Medium Slant w/ Rifampicin (40µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL008 | Acid Egg Medium Slant | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL009 | Acid Egg Medium Slant w/ pyruvate | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL010 | Modified L. J. Medium Plus Kit | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL011 | L.J. Medium Slant w/ Ciprofloxacin | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL011L | L.J. Medium Slant w/ Ciprofloxacin (12.5 mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL012 | L.J. Medium Slant w/ Amikacin | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL013 | L.J. Medium Slant w/ Clarithromycin | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL014 | L.J. Medium Slant w/Ethionamide (20µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL015 | L.J. Medium Slant w/Rifabutin (0.5 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL016 | L. J. Medium Plus Kit w/ kanamycin µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL017 | L.J. Medium Slant w/ D-Cycloserine (30 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL017L | L.J. Medium Slant w/ D-Cycloserine (30 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL018 | L.J.Medium w/Pyrazinamide of pH 5.5 | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL018L | L.J. Medium Slant w/ Pyrazinamide pH 5.5 | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL019 | L.M. Slant (Loeffler Medium) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL020 | L.J. Medium w/TCH | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL021 | L.J. Medium Slant w/ p-Nitrobenzoic acid (500 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL021L | L.J. Medium Slant w/ p-Nitrobenzoic acid | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL022 | L.J. Medium Slant w/o Glycerol | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL023 | Tuberculosis First Line Kit (Total 7 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL023L | Tuberculosis First Line Kit (Total 7 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL023LD | Tuberculosis First Line Kit (Total 7slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL023R | Tuberculosis First Line Kit (Total 7 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL024 | Tuberculosis Second Line Kit (Total 10 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL024L | Tuberculosis Second Line Kit (Total 10 slants) | Low risk | 20/12/2012 |

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| RPM- L.J.Medium Slants | SL024LD | Tuberculosis Second Line Kit (Total 10 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL024R | Tuberculosis Second Line Kit (Total 8 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL025 | Dorset Egg Medium Slant | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL026 | L.J. Medium Slant w/Streptomycin (5mcg) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL027 | L.J. Medium Slant w/Ethambutol (2mcg) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL029 | L.J. Medium Slant w/P-Amino Salicylic acid | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL029L | L.J. Medium Slant w/ p-Aminosalicylic acid | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL031 | Dermatophyte Test Medium Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL032 | Kligler Iron Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL033 | Motility Indole Lysine Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL034 | Simmons Citrate Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL034T | Simmon Citrate Agar Slant in long tubes | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL035 | Urea Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL035T | Urea Agar Slant in Tube | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL036 | Sabouraud Dextrose Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL036L | Sabouraud Dextrose Agar Slant | Low risk | 08/12/2017 |
| RPM- L.J.Medium Slants | SL037 | Tuberculosis First Line Plus Kit (Total 9 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL037R | Tuberculosis First Line Plus Kit (9 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL038 | Tuberculosis Second Line Plus Kit (Total 11 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL038R | Tuberculosis Second Line Plus Kit (11 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL038U | Lowenstein - Jensen Medium Slant with tu | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL040 | L.J. Medium Slant w/ Moxifloxacin | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL041 | Gelatin Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL042 | MIU Medium Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL043 | Nitrate Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL044 | Phenyl Alanine Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL045 | Triple Sugar Iron Agar Slant | Low risk | 20/12/2012 |

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| RPM- Ready Prepared Slants | SL045T | Triple Sugar Iron Agar Slant in Tube | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL047 | L.J.Medium Slant w/ Ethambutol (2µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL049 | L.J. Medium Slant w/ Ofloxacin (2 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL049L | L.J Medium Slant w/ Ofloxacin (2µg/ml) (long tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL055L | L.J.Medium Slant w/ Isoniazide (0.2 mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL061 | L.J. Medium Slant w/ Pyrazinamide (50 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL066 | L.J. Medium Slant w/ Capreomycin (20 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL067 | L.J. Medium Slant w/ Capreomycin (40 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL067L | L.J Medium Slant w/ Capreomycin (40 µg/ml) (long tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL067X | L.J Medium Slant w/ Capreomycin (40 µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL070 | L.J. Medium Slant w/ D-Cycloserine (40 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL071 | L.J. Medium Slant w/ Ethambutol (4 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL072 | L.J. Medium Slant w/ Ethambutol (5 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL076 | L.J. Medium Slant w/Ethionamide (40µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL078 | L.J. Medium Slant w/ Isoniazide (0.2 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL079 | L.J. Medium Slant w/ Isoniazide (5 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL081 | L.J. Medium Slant w/ Kanamycin (20 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL081L | L.J. Medium Slant w/ Kanamycin (20µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL082 | L.J. Medium Slant w/ Kanamycin (30 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL091 | L.J. Medium Slant w/ p-Aminosalicyclic acid (0.25 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL092 | L.J. Medium Slant w/ p-Aminosalicyclic acid (0.5 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL092L | L.J. Medium Slant w/ p-Aminosalicyclic acid | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL094 | L.J. Medium Slant w/ Ciprofloxacin 2µg/ml | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL098 | L.J. Medium Slant w/ Pyruvate (0.2%) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL099 | L.J.Medium Slants w/ Isoniazid (0.2 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL105L | L.J.Medium Slant w/ Rifampicin (20mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL107 | L.J. Medium Slant w/ Rifampicin (50 µg/ml) | Low risk | 20/12/2012 |

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| RPM- L.J.Medium Slants | SL109 | L.J. Medium Slant w/ Streptomycin (8 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL110 | L.J. Medium Slant w/ Streptomycin (25 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL115 | L.J. Medium Slant w/ Pyrazinamide of pH 5.5 | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL115L | L.J. Medium Slant w/ Pyrazinamide pH 5.5 | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL116 | Rapid UTI Diagnostic Slants | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL120L | L.J. Medium Slant pH 5.5 | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL121 | HiPyrazide glass tube w/ PYZ agar | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL121R | HiPyrazide glass tube w/ PYZ agar | Low risk | 10/11/2020 |
| RPM- L.J.Medium Slants | SL122 | HiCatalase glass tubes w/ 5ml of L.J. medium | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL122R | HiCatalase glass tubes w/ 5ml of L.J. medium | Low risk | 10/11/2020 |
| RPM- L.J.Medium Slants | SL123 | Tuberculosis first line plus kit (Modified) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL124 | L.J. Medium slant (Tubes with Aluminium caps) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL125 | L.J. Medium w/Isoniazid (1.0µg/ml) (Tube | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL125L | L.J.Medium Slant w/ Isoniazide (1mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL125M | L.J.Medium Slant w/ Isoniazid in Maccart | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL125X | L. J. Medium Slant w/ Isoniazide (1µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL126 | L.J. Medium w/Rifampicin (40.0µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL126L | L.J.Medium Slant w/ Rifampicin (40mcg/ml | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL126M | L.J.Medium Slant w/ Rifampicin in Maccar | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL126X | L. J. Medium Slant w/ Rifampicin (40.0 µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL127 | L.J. Medium w/Ethambutol (2.0µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL127L | L.J. Medium Slant w/Ethambutol (2µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL127X | L. J. Medium Slant w/ Ethambutol (2.0 µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL128 | L.J. Medium w/Streptomycin (10.0µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL128L | L.J.Medium w/Streptomycin - 10mcg / Ml | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL129 | L.J. Medium w/Ethionamide (30.0µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL130 | L.J. Medium w/Kanamycin (30.0µg/ml | Low risk | 20/12/2012 |

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| RPM- L.J.Medium Slants | SL130L | L.J. Medium Slant w/ Kanamycin (30µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL131 | L.J. Medium w/Ofloxacin (2.0µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL132 | L.J. Medium w/Capreomycin (30.0µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL133 | L.J. Medium w/P-aminosalicylic acid (1.0 | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL136L | Tuberculosis Second Line Kit, Modified | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL141 | Modified L. J. Medium Plus Kit | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL142 | Cystine Tryptone Agar with 1% Sugars | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL143 | Tuberculosis First Line Kit, Modified | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL144 | L.J Medium slant w/ Amikacin (1.0 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL147 | L.J. Medium Slant w/Rifampicin (64µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL148 | L.J. Medium Slant w/Ethambutol (6µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL149 | L.J. Medium Slant w/Streptomycin (16µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL150 | L.J Medium slant w/ Streptomycin (32 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL151 | TB Five Antitubercular Kit w/o Control | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL152 | Kit for Mycobiograme in Lowenstein Jensen | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL153 | Tuberculosis First Line Plus Kit (Modified) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL154 | L.J. Medium Plus Kit (total 9 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL155L | L.J. Medium Slant w/ TCH | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL156 | L.J.Medium Slant w/Rifampicin (128 µg /ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL157 | L.J.Medium Slant w/Pyrazinamide pH 5.5 | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL158 | L.J.Medium Slant w/Ethambutol (8 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL159 | L.J. Medium Slant w/Ethambutol (16 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL160L | Tuberculosis kit with antitubercular Age | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL161 | L.J. Medium Slant w/ Ciprofloxacin (16 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL162 | L.J. Medium Slant w/ Ciprofloxacin | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL163 | L.J. Medium Slant w/Amikacin (20 µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL164 | L.J. Medium Slant w/Amikacin (700 µg/ml) | Low risk | 20/12/2012 |

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| RPM- L.J.Medium Slants | SL165L | L.J.Medium w/ Pyruvate (0.48%) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL166L | Tuberculosis kit with antitubercular Agent | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL167 | L.J. Medium slants w/ Augmentin(20µg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL168 | L.J.Medium Slant w/ Ofloxacin (40µg/ml) | Low risk | 25/08/2016 |
| RPM- L.J.Medium Slants | SL168L | L.J.Medium Slant w/ Ofloxacin (40µg/ml) (long tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL169 | L.J.Medium Slant w/ Ethionamide (20µg/ml | Low risk | 25/08/2016 |
| RPM- L.J.Medium Slants | SL169L | L.J.Medium Slant w/ Ethionamide (20µg/ml) (long tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL170L | L.J Medium Slant w/ Ethionamide (40µg/ml) (long tube) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL170X | L.J Medium Slant w/ Ethionamide (40 µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL171L | L.J Medium Slant w/ p-Amino salicylic acid (1µg/ml) (long tube) | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL172 | Chocolate Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL173 | Nutrient Agar Slant | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL174 | B.C.G.-Dextrose Agar Butt (Synder Test Agar) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL175L | L.J.Medium Slant w/ Amikacin (30mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL175X | L. J. Medium Slant w/ Amikacin (30 mcg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL176L | L.J.Medium Slant w/ Ofloxacin (4mcg/ml) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL177 | Tuberculosis First Line Kit, Modified (Total 5 slants) | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL179 | L.J.Slopes for BCG Vaccines | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL180 | BHI Agar Slant w/5% Sheep Blood | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL181 | BHI Agar Slant w/10 % Sheep Blood,Chloramphenicol and Gentamicin | Low risk | 20/12/2012 |
| RPM- Ready Prepared Slants | SL182 | BHI CC Agar Slant w/10 % Sheep Blood and Gentamicin | Low risk | 20/12/2012 |
| RPM- L.J.Medium Slants | SL187 | L.J.Medium slants w/ LCN Supplement | Low risk | 25/08/2016 |
| RPM- L.J.Medium Slants | SL188L | L.J.Medium Slant w/ Levofloxacin (2 mg/ml) | Low risk | 16/12/2017 |
| RPM- L.J.Medium Slants | SL188X | L.J Medium Slant w/ Levofloxacin (2 µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL189L | L.J.Medium Slant w/ Levofloxacin (2.5 mg/ml) | Low risk | 16/12/2017 |
| RPM- L.J.Medium Slants | SL189X | L.J Medium Slant w/ Moxifloxacin (2.5 µg/ml) | Low risk | 17/06/2021 |
| RPM- L.J.Medium Slants | SL190 | L.J.Medium Slant w/ Rifampicin (20mcg/ml) | Low risk | 04/07/2018 |

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| RPM- L.J.Medium Slants | SL191 | L.J.Medium Slant w/ Amikacin (8mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL192 | L.J.Medium Slant w/ Ofloxacin (5mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL193 | L.J.Medium Slant w/ Levofloxacin (5 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL194 | L.J.Medium Slant w/ Ethionamide (5 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL195 | L.J.Medium Slant w/ Ethionamide (25 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL196 | L.J.Medium Slant w/ Prothionamide (5 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL197 | L.J.Medium Slant w/ Prothionamide (25 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL198 | L.J.Medium Slant w/ Linezolid (30 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL199 | L.J.Medium Slant w/ Clofazimine (1 mcg/ml) | Low risk | 04/07/2018 |
| RPM- L.J.Medium Slants | SL202 | Middlebrook 7H10 Agar Slant | Low risk | 30/10/2018 |
| RPM- L.J.Medium Slants | SL204 | L.J. Medium Slant w/ Prothionamide (40 mcg/ml) | Low risk | 22/04/2019 |
| RPM- L.J.Medium Slants | SL205 | L.J. Medium Slant w/ Amikacin (40 mcg/ml) | Low risk | 22/04/2019 |
| RPM- L.J.Medium Slants | SL211 | BHI Agar Slant | Low risk | 30/10/2018 |
| RPM- L.J.Medium Slants | SL1067L | Sabouraus Chloramphenicol Agar Slant | Low risk | 16/12/2017 |
| RPM- Ready Prepared Solid Medium | SM001 | Nutrient Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM001CCL | Nutrient Agar | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM001D | Nutrient Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM016C | Brilliant Green Agar, Modified | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM027C | Bismuth Sulphite Agar | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM049C | Violet Red Bile Agar | Low risk | 17/06/2021 |
| RPM- Ready Prepared Solid Medium | SM049D | Violet Red Bile Agar | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM063 | Sabouraud Dextrose Agar | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM063D | Sabouraud Dextrose Agar | Low risk | 25/08/2016 |

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| RPM- Ready Prepared Solid Medium | SM078 | Kligler Iron Agar | Low risk | 25/08/2016 |
| RPM- Ready Prepared Solid Medium | SM081 | MacConkey Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM081D | MacConkey Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM082 | MacConkey Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM082D | MacConkey Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM091 | Plate Count Agar | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM091D | Plate Count Agar | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM091DCC | Plate Count Agar | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM091M | Plate Count Agar | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM103A | Modified Chocolate Agar Kit w/o Selective | Low risk | 25/08/2016 |
| RPM- Ready Prepared Solid Medium | SM103AR | Modified Chocolate Agar kit w/osupplement | Low risk | 10/11/2020 |
| RPM- Ready Prepared Solid Medium | SM103H | Modified Chocolate Agar kit | Low risk | 25/08/2016 |
| RPM- Ready Prepared Solid Medium | SM103HR | Modified Chocolate Agar kit w/supplement | Low risk | 10/11/2020 |
| RPM- Ready Prepared Solid Medium | SM1067 | Sabouraud Chloramphenicol Agar Plate | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM1067C | Sabouraud Chloramphenicol Agar Plate | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM1067D | Sabouraud Chloramphenicol Agar | Low risk | 17/06/2021 |
| RPM- Ready Prepared Solid Medium | SM1067CCL | Sabouraud Chloramphenicol Agar Plate | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM1296D | HiCrome™ Salmonella Agar | Low risk | 04/07/2018 |

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| RPM- Ready Prepared Solid Medium | SM1297A | HiCrome™ Candida Differential Agar | Low risk | 25/08/2016 |
| RPM- Ready Prepared Solid Medium | SM1353 | HiCrome™ UTI Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM1353CC | HiCrome™ UTI Agar | Low risk | 04/07/2018 |
| RPM- Ready Prepared Solid Medium | SM154D | Reinforced Clostridial Agar | Low risk | 10/11/2020 |
| RPM- Ready Prepared Solid Medium | SM173 | Mueller Hinton Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM173CCL | Mueller Hinton Agar | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM173D | Mueller Hinton Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM211 | BHI Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM211D | BHI Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM331C | Wilson Blair Agar | Low risk | 22/04/2019 |
| RPM- Ready Prepared Solid Medium | SM434 | GC Agar | Low risk | 25/08/2016 |
| RPM- Ready Prepared Solid Medium | SM434R | Modified GC Agar Kit | Low risk | 10/11/2020 |
| RPM- Ready Prepared Solid Medium | SM434H | GC Agar,Modified | Low risk | 25/08/2016 |
| RPM- Ready Prepared Solid Medium | SM467 | Hektoen Enteric Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM467D | Hektoen Enteric Agar | Low risk | 20/12/2012 |
| RPM- Ready Prepared Solid Medium | SM792 | C.L.E.D. Agar w/ Bromothymol Blue | Low risk | 30/10/2018 |
| RPM- Ready Prepared Solid Medium | SM837 | Tryptose Sulphite Cycloserine(T.S.C) Agar | Low risk | 17/06/2021 |
| RPM- Ready Prepared Solid Medium | SM933D | Orange Serum Agar | Low risk | 22/04/2019 |

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| RPM- Ready Prepared UTI Diagnostic Kits | K041 | Rapid UTI ABST Kit | Low Risk | 20/12/2012 |
| RPM- Ready Prepared UTI Diagnostic Kits | K084A | Ecopathology Uro Kit-1 | Low risk | 20/12/2012 |
| RPM- Ready Prepared UTI Diagnostic Kits | K084B | Ecopathology Uro Kit-1, Modified | Low risk | 30/10/2018 |
| RPM- Ready Prepared UTI Diagnostic Kits | K085A | Ecopathology Uro Kit-2 | Low risk | 20/12/2012 |
| RPM- Ready Prepared UTI Diagnostic Kits | K089 | Ecopathology Uro Kit-3 | Low risk | 20/12/2012 |
| RPM- Ready Prepared UTI Diagnostic Kits | K090 | Ecopathology Uro Kit-4 | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K043 | Nitrate Reduction Test Kit for Mycobacteria | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K044 | Catalase Test Kit for Mycobacteria | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K044R | Catalase Test Kit for Mycobacteria | Low risk | 10/11/2020 |
| RPM- Biochemical Kits for Mycobacteria | K045 | Pyrazinmidase Test Kit for Mycobacteria | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K045R | Pyrazinmidase Test Kit for Mycobacteria ( | Low risk | 10/11/2020 |
| RPM- Biochemical Kits for Mycobacteria | K046 | Thiopene Carboxylic Hydrazide Test Kit for Mycobacteria | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K047 | Niacin Detection Kit w/ syringe | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K048 | Niacin Detection Kit Modified w/o syringe | Low risk | 20/12/2012 |
| RPM- Biochemical Kits for Mycobacteria | K050 | Kit for Selective Isolation of M.tuberculosis | Low risk | 20/12/2012 |
| RPM-MRSA Kits | K058S | MRSA Alert kit (w/swabs) | Low risk | 25/08/2016 |
| RPM-MRSA Kits | K058SR | MeReSa Agar Base,MRSA Alert Kit (w/swabs) | Low risk | 25/08/2016 |
| RPM-MRSA Kits | K086R | Enterococcus Presumptive Broth (VRE Alert) | Low risk | 25/08/2016 |
| RPM- Ready Prepared Diagnostic Kits | K144 | Mucormycosis Detection Kit | Low risk | 10/06/2021 |
| RPM- Biochemical Identification Kits | KB001 | HiIMViC™ Biochemical Test Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB001R | HiIMViC Biochemical Test Kit | Low risk | 25/08/2016 |

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| RPM- Biochemical Identification Kits | KB002 | HiAssorted™ Biochemical Test Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB002R | HiAssorted Biochemical Test Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB003 | Hi25™ Enterobacteriaceae Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB003R | Hi25 Enterobacteriaceae Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB004 | HiStaph™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB004R | HiStaph Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB005A | HiStrep™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB005AR | HiStrep Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB006 | HiCandida™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB006R | HiCandida Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB007 | HiVibrio™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB007R | HiVibrio Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB008 | HiNeisseria™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB008R | HiNeisseria Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB009 | HiCarbo™ Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB009R | HiCarbo Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB009A | HiCarbo™ Kit- Part A | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB009AR | HiCarbo Kit- Part A | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB009B1 | HiCarbo™ Kit- Part B | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB009C | HiCarbo™ Kit- Part C | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB010 | HiE. coli™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB010R | HiE.coli™ Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB011 | HiSalmonella™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB011R | HiSalmonella Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB012A | HiListeria™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB012AR | HiListeria Identification Kit | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB013 | HiBacillus™ Identification Kit | Low risk | 20/12/2012 |

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| RPM- Biochemical Identification Kits | KB013R | HiCarbo Kit (HiBacillus Identification Kit) | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB014 | HiAcinetobacter™ Identification Kit | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KB014R | HiCarbo Kit (HiAcinetobacter Identification Kit) | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KB015 | HiCorynebacteria Identification Kit | Low risk | 12/08/2015 |
| RPM- Biochemical Identification Kits | KB016 | Hi24™ Enterobacteriaceae Identification Kit,Modified | Low risk | 12/08/2015 |
| RPM- Biochemical Identification Kits | KB019 | Hi24™ Nonfermenters Identification Kit | Low risk | 28/04/2017 |
| RPM- Biochemical Identification Kits | KB020 | HiLacto Identification Kit | Low risk | 28/04/2017 |
| RPM- Biochemical Identification Kits | KB021 | HiBifido Identification Kit | Low risk | 28/04/2017 |
| RPM- Biochemical Identification Kits | KBM001 | HiMotility™ Biochemical Kit for E.coli | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KBM001R | HiMotility Biochemical Kit for E.coli | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KBM002 | HiMotility™ Biochemical Kit for Salmonella | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KBM002R | HiMotility™ Biochemical Kit for Salmonella | Low risk | 25/08/2016 |
| RPM- Biochemical Identification Kits | KBM003A | HiMotility™ Biochemical Kit for Listeria | Low risk | 20/12/2012 |
| RPM- Biochemical Identification Kits | KBM003AR | HiMotility™ Biochemical Kit for Listeria | Low risk | 25/08/2016 |

| **Product group** | **Type/ Model / Ref number** | **Device Name** | **Risk Class** | **Date of CE compliance** |
|---|---|---|---|---|
| **Epidemeology Screening Kit** | | | | |
| ESK- Hi Aureus Confirmation Kits | K053AD | Hiaureus Coagulase Confrimation Kit (w/o swabs) | Low risk | 07/02/2012 |
| ESK- Hi Aureus Confirmation Kits | K053ADS | Hiaureus Coagulase Confrimation Kit (w/ swabs) | Low risk | 07/02/2012 |

| Product group | Type/ Model / Ref number | Device Name | Risk Class | Date of CE compliance |
|---|---|---|---|---|
| **Bacteriological Differentiation Aids** | | | | |
| BDA- HiDtect Rapid Identification Discs | DT001 | HiDtect™ UTI Identification Disc | Low risk | 20/12/2012 |
| BDA- HiDtect Rapid Identification Discs | DT003 | HiDtect™ Pseudomonas Identification Disc | Low risk | 20/12/2012 |
| BDA- HiDtect Rapid Identification Discs | DT015 | HiDtect™ Universal Enviro Identification Disc | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I001 | Andrade's Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I002 | Bromocresol Green Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I003 | Bromocresol Purple Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I004 | Bromophenol Blue Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I005 | Bromothymol Blue Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I006 | Methyl Orange Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I007 | Methyl Red Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I008 | Neutral Red Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I009 | Phenolphthalein, 0.1% w/v | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I010 | Phenol Red Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I011 | Thymol Blue Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I012 | Thymolphthalein Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I013 | Universal Indicator | Low risk | 20/12/2012 |
| BDA- Readymade Indicators in Liquid | I014 | Mixed Indicator Solution (25X) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K001 | Gram Stains - Kit (contains S012, S032, S013 and S027 or S038) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K001CCL | Gram Stains - Kit (contains S012, S032, S013 and S027 or S038) | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | K001D | Gram Staining Kit | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | K001L | Gram Stains - Kit (contains S012, S032, S013 and S027 or S038) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K001M | Gram Stains - Kit | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | K002 | Albert`s Metachromatic Stains - Kit | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K002L | Albert`s Metachromatic Stains - Kit | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K003 | Neisser's Metachromatic Stains - Kit (contains S013, S023 and S037) | Low risk | 20/12/2012 |

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| BDA- Readymade Stains in Liquid | K003L | Neisser's Metachromatic Stains - Kit (contains S013, S023 and S037) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K004 | Capsule Stains - Kit (contains S021, S025 and S047) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K004L | Capsule Stains - Kit (contains S021, S025 and S047) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K005 | ZN Acid Fast Stains - Kit (contains S033,S005 and S022) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K005CCL | ZN Acid Fast Stains - Kit (contains S033,S005 and S022) | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | K005D | ZN Acid Fast Stains - Kit | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | K005L | ZN Acid Fast Stains - Kit (contains S033, S005 & S022) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K005M | ZN Acid Fast Stains - Kit (contains S033, S005 & S022) | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | K006 | Schaeffer & Fulton's Spore Stains - Kit (contains S028 and S029) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K006L | Schaeffer & Fulton's Spore Stains - Kit (contains S028 and S029) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K011 | Malarial Parasite - Kit (contains S008 and S009) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K011L | Malarial Parasite - Kit (contains S008 and S009) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K021 | Fluorescent Stains - Kit for Mycobacteria (contains S042, S043 and S044) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K021L | Fluorescent Stains - Kit for Mycobacteria (contains S042, S043 and S044) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K021R | Fluorescent Stains Kit for Mycobacteria (conatins S054,S055,S056) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K021Y | Fluorescent Stains Kit for Mycobacteria (conatins S042Y,S043Y,S044Y) | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | K049 | Malarial Parasite - Kit (contains S008 and S009) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K061 | HiFluo-Phenol Free Stain - kit for Mycobacteria [Kit contains 200ml each of Auramine – Rhodamine solution (Phenol free)-S082, Decolourizer-S099 (2 x200), Potassium Permanganate Solution-S083] | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K062 | HiCold Stain TB - Kit for Mycobacteria [Kit contains 500ml each of Carbol Fuchsin Solution-S080, Decolourizer-S099, Counter Stain (Loeffler's Methylene Blue)-S081] | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K062S | HiCold Stain TB - Kit for Mycobacteria [Kit contains 100ml each of Carbol Fuchsin Solution-S080, Decolourizer-S099, Counter Stain (Loeffler's Methylene Blue) S081] | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | K063 | Modified Neisser's Metachromatic Stains - Kit (1 minute staining) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R001 | Barium Chloride Solution, 10% w/v | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R002 | Benedict's Qualitative Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R003 | Benedict's Quantitative Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R004 | C.S.F. Diluting Fluid | Low risk | 20/12/2012 |

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| BDA - Readymade Reagents in Liquid | R005 | Ehrlich's Aldehyde Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R006 | Folin & Wu's Alkaline Copper Solution | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R007 | Folin & Wu's Phosphate, Molybdate Solution | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R008 | Kovacs' Indole Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R009 | a-Naphthylamine solution | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R010 | Nessler's Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R011 | Potassium Chromate, 5% w/v | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R012 | Potassium Oxalate, 5% w/v | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R013 | R.B.C. Diluting Fluid (Hayemis) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R014 | Sodium Citrate, 3.8% w/v | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R015 | Sulphanilic acid, 0.8% | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R016 | W.B.C. Diluting Fluid | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R017 | Nessler's Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R018 | Fouchet's Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R019 | E.D.T.A. (di-sodium) 5% | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R020 | Sulphosalicylic Acid 3% | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R021 | Topfer Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R022 | o-Toluidine reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R023 | R.B.C. Diluting Fluid (Grower's) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R024 | o-Toluidine Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R026 | Gordon-McLeod Reagent (Oxidase reagent) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R027 | Gaby-Hadley Reagent A | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R028 | Gaby-Hadley Reagent B | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R029 | Barritt Reagent A (for VP test) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R030 | Barritt Reagent B (for VP test) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R031 | O'Meara Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R035 | DMACA Reagent | Low risk | 20/12/2012 |

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| BDA - Readymade Reagents in Liquid | R036 | TDA Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R037 | Fehling Solution No. 1 | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R038 | Fehling Solution No. 2 | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R043 | PYR Reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R044 | Iodine Solution | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R075 | 10X RBC Lysis Buffer Solution | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R083 | Thrombocount reagent | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R084 | HiDecal (mild decalcifying solution) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R085 | HiDecal (strong decalcifying solution) | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R092 | McFarland Standard Tube | Low risk | 20/12/2012 |
| BDA - Readymade Reagents in Liquid | R092A | Mcfarland standard 0.5 | Low risk | 22/04/2019 |
| BDA - Readymade Reagents in Liquid | R092B | Mcfarland standard 1 | Low risk | 22/04/2019 |
| BDA - Readymade Reagents in Liquid | R092C | Mcfarland standard 2 | Low risk | 22/04/2019 |
| BDA - Readymade Reagents in Liquid | R092D | Mcfarland standard 3 | Low risk | 22/04/2019 |
| BDA - Readymade Reagents in Liquid | R092E | Mcfarland standard 4 | Low risk | 22/04/2019 |
| BDA - Readymade Reagents in Liquid | R092R | Test Tubes (McFarland Standard Tube) | Low risk | 25/08/2016 |
| BDA - Readymade Reagents in Liquid | R092S | McFarland Standard Set (0.5,1,2) | Low risk | 04/07/2018 |
| BDA - Readymade Reagents in Liquid | R097 | Millons reagent | Low risk | 28/04/2017 |
| BDA- Readymade Stains in Liquid | S001 | Albert's Stain A | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S002 | Albert's Stain B | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S003 | Borax Carmine (Grenacher's), Alcoholic Stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S004 | Borax Carmine (Grenacher's), Aqueous Stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S005 | Carbol Fuchsin (ZN,Strong) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S005D | Carbol Fuchsin (ZN,Strong) | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S005M | Carbol Fuchsin (ZN,Strong) | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S006 | Carbol Fuchsin (ZN, Dilute) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S007 | Eosin, 2% w/v | Low risk | 20/12/2012 |

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| BDA- Readymade Stains in Liquid | S008 | Field's Stain A | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S009 | Field's Stain B | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S010 | Gentian Violet | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S011 | Giemsa's Stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S012 | Gram's Crystal Violet | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S012D | Gram's Crystal Violet | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S012M | Gram's Crystal Violet | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S013 | Gram's Iodine | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S013D | Gram's Iodine | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S013M | Gram's Iodine | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S014 | Haematoxylin (Delafield's) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S015 | Lactophenol | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S016 | Lactophenol Cotton Blue | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S017 | Lactophenol Picric Acid | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S018 | Leishman's Stain (Twin Pack) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S018S | Leishman's Stain Solution | Low risk | 25/11/2017 |
| BDA- Readymade Stains in Liquid | S019 | Lugol's Iodine | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S020 | Malachite Green, 1% w/v | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S021 | Methylene Blue (Aqueous) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S022 | Methylene Blue (Loeffler's) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S022D | Methylene Blue (Loeffler's) | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S022M | Methylene Blue (Loeffler's) | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S023 | Neisser's Methylene Blue | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S024 | Newman's Stain, Modified | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S025 | Nigrosin Stain,10% w/v | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S026 | Picric Acid (Saturated, Aqueous) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S027 | Safranin, 0.5% w/v | Low risk | 20/12/2012 |

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| BDA- Readymade Stains in Liquid | S027D | Safranin, 0.5% w/v | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S027M | Safranin, 0.5% w/v | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S028 | Schaeffer & Fulton's Spore Stain A | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S029 | Schaeffer & Fulton's Spore Stain B | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S030 | Wright's Stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S031 | Mayer's Mucicarmine Stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S032 | Gram's Decolourizer | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S032D | Gram's Decolourizer | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S032M | Gram's Decolourizer | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S033 | Acid Fast Decolourizer | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S033D | Acid Fast Decolourizer | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S033M | Acid Fast Decolourizer | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S034 | Haematoxylin (Harris) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S035 | Papanicolaou-OG-6 | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S036 | Papanicolaou-EA-36 | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S037 | Neutral Red Solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S038 | Basic Fuchsin 0.1% w/v | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S038D | Basic Fuchsin 0.1% w/v | Low risk | 04/07/2018 |
| BDA- Readymade Stains in Liquid | S038M | Basic Fuchsin 0.1% w/v | Low risk | 22/04/2019 |
| BDA- Readymade Stains in Liquid | S039 | May-Grunwald's Stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S041 | FA Rhodamine Counterstain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S042 | Phenolic auramine | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S042Y | Phenolic auramine O | Low risk | 08/12/2017 |
| BDA- Readymade Stains in Liquid | S043 | Mycobacteria decolourizer | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S043Y | Mycobacteria decolourizer | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S044 | Potassium permanganate | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S044Y | Potassium permanganate | Low risk | 08/12/2017 |

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| BDA- Readymade Stains in Liquid | S047 | M'Fadyean Stain (Polychrome Methylene Blue) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S054 | Fluorochrome Solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S055 | Decolourising Solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S056 | Background Solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S057 | Grams Iodine, Stabilized | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S058 | Haematoxylin (Mayer) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S059 | Haematoxylin (Ehrlich) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S062 | Fixing solution, for fixing Haematological samples | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S066 | Brilliant Cresyl Blue Solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S067 | Congo red (1% aqueous)Solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S068 | Papanicolaou-EA-50 | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S070 | Schiff's fuchsin-sulphite reagent | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S073 | Periodic Acid Solution (PAS) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S074 | Schiff's Reagent | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S076 | Haematoxylin (Gill No.3) | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S102 | Fixative, for fixing cytological or histological samples | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S109 | Fixative (Buffered Formalin fixative) for fixing cytological or histological samples | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S118 | Fixative, for rapid fixing of haematological samples | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S119 | Fixative (BFA), for Rapid fixing of haematological samples | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S125 | Romanowsky-Giemsa (RG) stain | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S126 | Shorr's Stain solution | Low risk | 20/12/2012 |
| BDA- Readymade Stains in Liquid | S127 | Gabbett Counterstaining Solution | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S128 | HiGrams Stain Crystal Violet | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S129 | HiGrams Iodine | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S130 | HiGrams Decolouriser | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S131 | HiGrams Counter Stain | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S132 | HiCarbol Fuchsin | Low risk | 16/12/2017 |

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|---|---|---|---|---|
| BDA- Readymade Stains in Liquid | S133 | HiAcid Fast Decolouriser | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S134 | HiAcid Fast Counter Stain | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S135 | Solution for Leishman's Stain L (Twin Pack) | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S136 | Solution for Leishman's Stain R (Twin Pack) | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S137 | Solution for Leishman's Stain HP (Twin Pack) | Low risk | 16/12/2017 |
| BDA- Readymade Stains in Liquid | S138 | Gentian Violet 1 % Solution | Low risk | 22/04/2019 |
| BDA- Differentiation Discs | DD015 | Bacitracin | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD024 | Bile Esculin | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD040 | DMACA Indole | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD035 | Hippurate hydrolysis | Low risk | 20/12/2012 |
| BDA- Differentiation Strips | DD034 | Lead Acetate Paper strips | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD041 | Nitrate Discs | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD042 | Nitrate Reagent Discs | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD008 | ONPG | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD009 | Optochin | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD009R | Optochin (5mcg) | Low risk | 25/08/2016 |
| BDA- Differentiation Discs | DD018 | Oxidase | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD020 | X factor | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD022 | X+V Factor | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD021 | V Factor | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD047 | Vibrio 0129 Differential Disc (10 mcg) | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD048 | Vibrio 0129 Differential Disc (150 mcg) | Low risk | 20/12/2012 |
| BDA- Differentiation Discs | DD055 | Bacitracin B | Low risk | 25/08/2016 |
| BDA- Differentiation Discs | DD056 | Sodium Biselenite Disc | Low risk | 04/07/2018 |
| BDA- Differentiation Discs | DB001 | Sodium Biselenite Bud | Low risk | 04/07/2018 |

| Product group | Type/ Model / Ref number | Device Name | Risk Class | Date of CE compliance |
|---|---|---|---|---|
| **Antimicrobial Susceptibility Systems** | | | | |
| ASS- Sensitivity Discs (Multi Discs) | DE001 | Dodeca Universal-I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE002 | Dodeca G-I-Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE003 | Dodeca G-I-Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE004 | Dodeca UTI-I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE005 | Dodeca UTI-II | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE006 | Dodeca UTI-III | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE007 | Dodeca Universal-II | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE008 | Dodeca Universal-III | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE009 | Dodeca G-II-Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE010 | Dodeca G-II-Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE011 | Dodeca UTI-IV | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE012 | Dodeca Universal-IV | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE013 | Dodeca Universal-V | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE014 | Dodeca Universal-VI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE015 | Dodeca Universal-VII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE016 | Dodeca Universal III | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE017 | Dodeca Universal-IX | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE018 | Dodeca G-III-Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE019 | Dodeca G-III-Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE020 | Dodeca Pseudo-I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE021 | Dodeca UTI-V | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE022 | Dodeca Universal X | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE023 | Dodeca G-IV Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE024 | Dodeca G-IV minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE025 | Dodeca UTI-VI | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| ASS- Sensitivity Discs (Multi Discs) | DE026 | Dodeca Universal -XI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE027 | Dodeca Universal -XII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE028 | Dodeca Universal -XIII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE029 | Dodeca G-V minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE030 | Dodeca UTI-VII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE031 | Dodeca G-VI minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE032 | Dodeca G-V Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE033 | Dodeca G-VII Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE034 | Dodeca UTI-VIII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE035 | Dodeca Universal XIV | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE036 | Dodeca G-VI Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE037 | Dodeca G-VIII Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE038 | Dodeca G-VII Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE039 | Dodeca G-IX Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE040 | Dodeca UTI-IX | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE041 | Dodeca Pseudo-II | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE042 | Dodeca Universal XV | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE043 | Dodeca G-X Minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE044 | Dodeca - G-VIII Plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE045 | Dodeca G-XI Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE046 | Dodeca G-XII Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE047 | Dodeca G-IX Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE048 | Dodeca Staphylococci - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE049 | Dodeca Staphylococci - 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE050 | Dodeca Enterococcus -1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE051 | Dodeca Pseudomonas -1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE052 | Dodeca Pseudomonas 2 | Low risk | 25/08/2016 |

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|---|---|---|---|---|
| ASS- Sensitivity Discs (Multi Discs) | DE053 | Dodeca Enterobacteriaceae - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE054 | Dodeca Enterobacteriaceae - 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | DE700 | Dodeca Staphylococci - 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE701 | Dodeca Staphylococci - 2 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE702 | Dodeca Enterococcus - 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE703 | Dodeca Pseudomonas - 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE704 | Dodeca Pseudomonas - 2 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE705 | Dodeca Enterobacteriaceae - 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE706 | Dodeca Enterobacteriaceae - 2 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE707 | Dodeca Universal - 16 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE708 | Dodeca UTI - 10 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE709 | Dodeca G-Minus 13 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE710 | Dodeca G-Plus 10 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE711 | Dodeca G minus XIV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE712 | Dodeca G minus XV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE713 | Dodeca G minus 16 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE714 | Dodeca G minus 17 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE715 | Dodeca G minus 18 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE716 | Dodeca G plus 11 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE717 | Dodeca G plus 12 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE718 | Dodeca G minus 19 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE719 | Dodeca UTI 10 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE720 | Dodeca UTI 11 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE721 | Dodeca Universal 16 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE722 | Dodeca Universal 17 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE723 | Dodeca G Plus 13 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE724 | Dodeca UTI-12 | Low risk | 25/08/2016 |

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|---|---|---|---|---|
| ASS- Sensitivity Discs (Multi Discs) | DE725 | Dodeca Universal-18 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE726 | Dodeca UTI - 13 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE727 | Dodeca G-minus 20 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE728 | Dodeca UTI 14 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE729 | Dodeca G-Plus 14 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE730 | Dodeca G-Minus 21 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE731 | Dodeca G-Minus 22 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE732 | Dodeca Universal 19 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE733 | Dodeca Universal 20 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE734 | Dodeca Universal 21 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE735 | Dodeca Universal 22 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE736 | Dodeca G-Plus 15 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE737 | Dodeca G-Minus 23 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE738 | Dodeca G-Minus 24 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE739 | Dodeca UTI 13 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE740 | Dodeca G-Plus 16 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE741 | Dodeca G-Minus 25 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE742 | Dodeca Pseudomonas -3 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | DE743 | Dodeca G-Minus 26 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | DE744 | Dodeca UTI 15 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | DE745 | Dodeca Pseudomonas -4 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | DE746 | Dodeca G-Plus 17 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | DE747 | Dodeca G-Minus 27 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | DE748 | Dodeca UTI 16 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | DE749 | Dodeca G-Plus 18 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE750 | Dodeca G-Plus 19 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE751 | Dodeca G-Minus 28 | Low risk | 10/11/2020 |

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| ASS- Sensitivity Discs (Multi Discs) | DE752 | Dodeca G-Minus 29 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE753 | Dodeca G-Plus 20 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE754 | Dodeca G-Minus 30 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE755 | Dodeca Pseudomonas -5 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE756 | Dodeca G-Minus 31 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | DE757 | Dodeca G-Plus 21 | Low risk | 10/11/2020 |
| ASS- Ezy MIC Strips | EM001 | Amikacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM002 | Amoxycillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM003 | Amoxyclav (2:1) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM004 | Azithromycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM006 | Aztreonam | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM008 | Cefazolin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM009 | Cefdinir | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM011 | Cefpirome | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM012 | Ceftazidime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM013 | Ceftriaxone | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM016 | Chloramphenicol | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM017 | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM018 | Clarithromycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM019 | Clindamycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM020 | Colistin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM020S | Colistin | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM021 | Co-Trimoxazole (1:19) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM021S | Co-Trimoxazole (1:19) | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM022 | Erythromycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM023 | Fusidic Acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM024 | Gatifloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM025 | Gentamicin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM026 | Kanamycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM027 | Levofloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM029 | Linezolid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM032 | Minocycline | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM033 | Moxifloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM035 | Nalidixic acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM037 | Nitrofurantoin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM038 | Norfloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM039 | Ofloxacin | Low risk | 20/12/2012 |

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| ASS- Ezy MIC Strips | EM041 | Piperacillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM042 | Piperacillin/Tazobactam | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM043 | Polymixin B | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM044 | Pristinomycin (Quinupristin/Dalfopristin) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM045 | Rifampicin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM046 | Roxithromycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM047 | Sparfloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM048 | Streptomycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM055 | Teicoplanin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM055S | Teicoplanin | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM056 | Tetracycline | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM057 | Ticarcillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM058 | Tobramycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM059 | Trimethoprim | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM060 | Vancomycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM060S | Vancomycin | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM061 | Gentamicin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM062 | Penicillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM063 | Oxacillin - Vancomycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM064 | Cefotaxime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM065 | Oxacillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM066 | Ceftriaxone | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM066S | Ceftriaxone | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM068 | Ampicillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM070 | Cefepime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM071 | Amphotericin-B | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM072 | Fluconazole | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM073 | Itraconazole | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM074 | Ketoconazole | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM076 | Gemifloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM077 | Vancomycin -Cefoxitin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM078 | Imipenem w&w/o EDTA | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM079A | Triple ESBL detection Strip | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM080 | Meropenem | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM081A | ESBL & AmpC Detection Strip | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM082 | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM083 | Co-Trimoxazole (1:19) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM084 | Penicillin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM085 | Ertapenem | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM086 | Voriconazole | Low risk | 20/12/2012 |

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| ASS- Ezy MIC Strips | EM087 | Mupirocin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM088 | Daptomycin (Supplemented with Calcium ions) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM089 | Tigecycline | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM090 | Doripenem | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM091 | Faropenem | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM092 | Meropenem with & without EDTA | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM093 | Cefepime/Tazobactam (2:1) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM094 | Cefoperazone/Sulbactam (2:1) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM095 | Netilmicin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM097 | Ceftriaxone/Sulbactam (2:1) | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM098 | Ceftazidime / Ceftazidime+ Clavulanic acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM099 | Cefotaxime / Cefotaxime + Clavulanic acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM100 | Cefotaxime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM101 | Cefoxitin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM102 | Cefuroxime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM103 | Doxycycline | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM104 | Imipenem | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM105 | Cefotetan | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM106 | Cephalothin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM107 | Cefaclor | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM108 | Fosfomycin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM109 | Ampicillin/Sulbactam | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM110 | Cefixime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM111 | Vancomycin - Teicoplanin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM112 | Cefoperazone | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM113 | Cefonicid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM114 | Cefmetazole | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM115 | Enrofloxacin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM116 | Cefepime / cefepime + Clavulanic acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM117 | Ceftriaxone / Ceftriaxone + Clavulanic acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM118 | Flucytosine | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM119 | Caspofungin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM120 | Posaconazole | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM121 | Micafungin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM122 | Anidulafungin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM123 | Ceftizoxime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM124 | Mecillinam | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM125 | Ticarcillin/Clavulanic Acid | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM126 | Bacitracin | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM127 | Cefotetan / Cefotetan + Cloxacillin | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| ASS- Ezy MIC Strips | EM128 | Metronidazole | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM129 | Cefpodoxime | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM130 | Cefprozil | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM131 | Sulbactam | Low risk | 20/12/2012 |
| ASS- Ezy MIC Strips | EM132 | Improved ESBL Detection Ezy MIC Strip (Mix+/Mix) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM133 | Improved AmpC Detection Ezy MIC Strip (Mix+/Mix) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM134 | MBL Plus ESBL Detection Ezy MIC Strip (ESBL+/ESBL) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM135 | MBL Plus AmpC Detection Ezy MIC Strip (AmpC+/Amp) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM136 | ESBL-AmpC Coexistence Detection Ezy MIC Kit | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM137 | MBL-ESBL-AmpC Co-existence Detection Ezy MIC Kit | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM138 | Cefpodoxime/Clavulanic Acid Ezy MIC Strip | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM139 | Amoxyclav Ezy MIC Strip (AUG) (0.016-256 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM140 | Ampicillin/Sulbactam Ezy MIC Strip (SAM) (4 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM141 | Ertapenem/Ertapenem + Boronic acid Ezy MIC Strip (ETP+/ETP) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM142 | Terbinafine Ezy MIC Strip(TRB) (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM143 | Griseofulvin EZY MIC Strip (GRI) (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM144 | Clotrimazole EZY MIC Strip (CLO) (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM145 | Terbinafine Ezy MIC Strip(TRB) (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM146 | Miconazole EZY MIC Strip (MIC) (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM147 | Flucloxacillin Ezy MIC Stripr (FLC) (0.016-256 mcg/ml) | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM148 | Cefepime/Clavulanic acid Ezy MIC Stripr (FIC) (0.016-256 mcg/ml) | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM149 | Ceftazidime /Tazobactum Ezy MIC Strip (CAT) (0.016-256 mcg/ml) | Low risk | 04/07/2018 |
| ASS- Ezy MIC Strips | EM150 | Natamycin Ezy MIC Strip (NAT) (0.016-256 mcg/ml) | Low risk | 22/04/2019 |
| ASS- Ezy MIC Strips | EM151 | Cefpirome/Sulbactam Ezy MIC™ Strip | Low risk | 22/04/2019 |
| ASS- Ezy MIC Strips | EM152 | Ceftizoxime/Sulbactam Ezy MIC™ Strip | Low risk | 10/11/2020 |
| ASS- Ezy MIC Strips | EM153 | Ceftazidime/Avibactam Ezy MIC™ Strip | Low risk | 10/11/2020 |
| ASS- Ezy MIC Strips | EM154 | Faropenem/Clavulanic acid Ezy MIC™ strip (FAC) | Low risk | 01/11/2020 |
| ASS- Ezy MIC Strips | EM155 | Cefuroxime/Clavulanic acid Ezy MIC™ strip (CXC) | Low risk | 01/11/2020 |
| ASS- Ezy MIC Strips | EM701 | Xylomonas Ezy MIC Strip (0.016-256mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM702 | Arbekacin Ezy MIC Strip (ABK) (0.016-256 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM703 | Garenoxacin EZY MIC Strip (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM705 | Biapenem EZY MIC Strip (BPM) (0.002-32 mcg/ml) | Low risk | 25/08/2016 |
| ASS- Ezy MIC Strips | EM706 | Reinvexin EZY MIC Strip (PB) (0.016-256 mcg/ml) | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | HX001 | Hexa G-plus 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX002 | Hexa G-plus 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX003 | Hexa G-plus 3 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX004 | Hexa G-plus 4 | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | HX005 | Hexa G-plus 5 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX006 | Hexa G-minus 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX007 | Hexa G-minus 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX008 | Hexa G-minus 3 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX009 | Hexa G-minus 4 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX010 | Hexa G-minus 5 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX011 | Hexa Pseudo 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX012 | Hexa Pseudo 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX013 | Hexa Pseudo 3 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX014 | Hexa UTI-1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX015 | Hexa UTI-2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX016 | Hexa Haemophilus 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX017 | Hexa Haemophilus 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX018 | Hexa Haemophilus 3 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX019 | Hexa Pneumococci 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX020 | Hexa Pneumococci 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX021 | Hexa Anaerobic 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX022 | Hexa G-plus 6 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX023 | Hexa G-plus 7 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX024 | Hexa G-plus 8 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX025 | Hexa G-Minus 6 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX026 | Hexa Pseudo 4 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX027 | Hexa G-Plus 9 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX028 | Hexa G-minus 7 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX029 | Hexa Pseudo 5 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX030 | Hexa G-Minus 8 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX031 | Hexa G-Plus 10 | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | HX032 | Hexa Universal - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX033 | Hexa UTI 3 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX034 | Hexa G-plus11 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX035 | Hexa G-minus 9 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX036 | Hexa G-minus 29 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX037 | Hexa UTI 4 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX038 | Hexa Universal-2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX039 | Hexa G-plus 12 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX040 | Hexa G-plus 13 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX041 | Hexa Pneumococci - 3 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX042 | Hexa Pneumococci-4 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX043 | Hexa Pneumococci - 5 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX044 | Hexa Pneumococci - 6 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX045 | Hexa Pneumococci-7 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX046 | Hexa Pneumococci-8 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX047 | Hexa G-plus 25 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX048 | Hexa G-plus 26 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX049 | Hexa G-plus 27 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX050 | Hexa Pseudo 6 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX051 | Hexa Pseudo 7 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX052 | Hexa Pseudo 8 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX053 | Hexa Pseudo 9 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX054 | Hexa Pseudo 10 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX055 | Hexa Pseudo 11 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX056 | Hexa G-minus 26 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX057 | Hexa G-minus 27 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX058 | Hexa G-minus 28 | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | HX059 | Hexa G-minus 10 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX060 | Hexa G-minus 11 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX061 | Hexa G-Minus 12 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX062 | Hexa G-minus 13 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX063 | Hexa G-minus 14 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX064 | Hexa G-minus 15 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX065 | Hexa G-Minus 16 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX066 | Hexa G-minus 17 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX067 | Hexa G-minus 18 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX068 | Hexa G-minus 19 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX069 | Hexa G-minus 20 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX070 | Hexa G-minus 21 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX071 | Hexa G-Minus 22 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX072 | Hexa UTI 4 (Modified) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX073 | Hexa UTI 5 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX074 | Hexa UTI 6 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX075 | Hexa UTI 7 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX076 | Hexa UTI 8 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX077 | Hexa UTI 9 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX078 | Hexa UTI 10 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX079 | Hexa UTI 11 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX080 | Hexa G-plus 14 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX081 | Hexa G-plus 15 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX082 | Hexa G-Plus 16 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX083 | Hexa G-plus 17 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX084 | Hexa Haemophilus 4 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX085 | Hexa Haemophilus 5 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | HX086 | Hexa Haemophilus 6 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX087 | Hexa Haemophilus 7 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX088 | Hexa Haemophilus 8 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX089 | Hexa Haemophilus 9 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX090 | Hexa G-plus 18 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX091 | Hexa G-plus 19 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX092 | Hexa G-plus 20 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX093 | Hexa G-plus 21 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX094 | Hexa G-Plus 22 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX095 | Hexa G-minus 23 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX096 | Hexa G-minus 24 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX097 | Hexa Universal-2 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX098 | Hexa Universal-3 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX099 | Hexa UTI 12 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX100 | Hexa G-Plus 23 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX101 | Hexa G-plus 24 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX102 | Hexa G-minus 25 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX103 | Hexa Pseudo 12 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX104 | Hexa Antimyco-01 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | HX700 | Hexa G-Plus 25 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX701 | Hexa G-Minus 26 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX703 | Hexa Pseudo-13 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX704 | Hexa G-Minus 27 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX705 | Hexa Anaerobic 2 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX706 | Hexa UTI 14 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX707 | Hexa G-Plus 26 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX708 | Hexa G-Minus 28 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | HX709 | Hexa Pseudo 14 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX710 | Hexa UTI-15 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX711 | Hexa G-Plus 27 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX712 | Hexa Pseudo 15 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX713 | Hexa Anaerobic-3 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX714 | Hexa Combi 1 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX715 | Hexa Universal 4 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX716 | Hexa Universal 5 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX717 | Hexa Combi 2 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX718 | Hexa Combi 3 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX719 | Hexa Combi 4 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX720 | Hexa Combi 5 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX721 | Hexa Combi 6 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX722 | Hexa Combi 7 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX723 | Hexa Combi 8 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | HX724 | Hexa Combi 9 | Low risk | 28/04/2017 |
| ASS- Sensitivity Discs (Multi Discs) | HX725 | Hexa Combi 10 | Low risk | 28/04/2017 |
| ASS- Sensitivity Discs (Multi Discs) | HX726 | Hexa Combi 11 | Low risk | 28/04/2017 |
| ASS- Sensitivity Discs (Multi Discs) | IC001 | Icosa Universal - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC002 | Icosa G-I-Plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC003 | Icosa G-I-Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC004 | Icosa UTI - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC005 | Icosa Pseudo - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC006 | Icosa Universal - 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC007 | Icosa Pseudo - 2 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC008 | Icosa G-II-Minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | IC701 | Icosa Universal - 3 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | IC702 | Icosa Universal - 4 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | IC703 | Icosa Universal 5 | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD001 | Amikacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD002 | Amoxycillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD003 | Amoxyclav (Amoxycillin/ Clavulanic acid) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD004 | Azithromycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD005 | Azlocillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD006 | Aztreonam | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD007 | Carbenicillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD008 | Cefazolin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD009 | Cefdinir | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD010 | Cefepime | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD011 | Cefpirome | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD012 | Ceftazidime | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD013 | Ceftriaxone | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD014 | Cefalexin (Cephalexin) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD015 | Cefotaxime (Cephotaxime) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD016 | Chloramphenicol | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD017 | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD018 | Clarithromycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD019 | Clindamycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD020 | Colistin (Methane Sulphonate) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD021 | Co-Trimoxazole (Sulpha/Trimethoprim) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD022 | Erythromycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD023 | Fusidic Acid | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD024 | Gatifloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD025 | Gentamicin | Low risk | 20/12/2012 |

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| ASS-HiComb MIC Strips | MD026 | Kanamycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD027 | Levofloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD028 | Lincomycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD029 | Linezolid | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD030 | Lomefloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD031 | Methicillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD032 | Minocycline | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD033 | Moxifloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD034 | Mupirocin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD035 | Nalidixic Acid | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD036 | Neomycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD037 | Nitrofurantoin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD038 | Norfloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD039 | Ofloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD040 | Pefloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD041 | Piperacillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD042 | Piperacillin/Tazobactam | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD043 | Polymyxin-B | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD044 | Pristinomycin (Quinupristin/Dalfopristin) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD045 | Rifampicin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD046 | Roxithromycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD047 | Sparfloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD048 | Streptomycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD049 | Sulfasomidine | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD050 | Sulphadiazine | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD051 | Sulphafurazole (Sulfisoxazole) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD052 | Sulphamethizole | Low risk | 20/12/2012 |

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| ASS-HiComb MIC Strips | MD053 | Sulphamethoxypyridazine | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD054 | Sulphaphenazole | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD055 | Teicoplanin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD056 | Tetracycline | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD057 | Ticarcillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD058 | Tobramycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD059 | Trimethoprim | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD060 | Vancomycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD061 | Gentamicin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD062 | Benzyl Penicillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD063 | Vancomycin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD064 | Cefotaxime (Cephotaxime) | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD065 | Oxacillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD066 | Ceftriaxone | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD067 | Amikacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD068 | Ampicillin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD069 | Ceftazidime | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD070 | Cefepime | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD071 | Amphotericin B | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD072 | Fluconazole | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD073 | Itraconazole | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD074 | Ketoconazole | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD076 | Gemifloxacin | Low risk | 20/12/2012 |
| ASS-HiComb MIC Strips | MD701 | Cefepime/Tazobactam | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD702 | Ceftazidime/Tazobactum | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD704 | Nadifloxacin | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD706 | Cefoperazone/Tazobactam CST | Low risk | 25/08/2016 |

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|---|---|---|---|---|
| ASS-HiComb MIC Strips | MD707 | Balofloxacin | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD708 | Cefuroxime CXM | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD709 | Cefpodaxime CPD | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD710 | Cefpodaxime / Clavulanic acid (2:1) | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD711 | Netilimicin NET | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD712 | Cefixime CFM | Low risk | 25/08/2016 |
| ASS-HiComb MIC Strips | MD713 | Pazufloxacin | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD001 | G-I-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD001R | G-I-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD002 | G-II-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD002R | G-II-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD003 | G-III-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD0032R | Combi I | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD003R | G-III-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD004 | G-IV-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD004R | G-IV-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD005 | G-I-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD005R | G-I-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD006 | G-II-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD006R | G-II-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD007 | G-III-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD007R | G-III-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD008 | Pseudo | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD008R | Pseudo | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD009 | UTI-I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD009R | UTI-I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD010 | UTI-II | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| ASS- Sensitivity Discs (Multi Discs) | OD010R | UTI-II | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD011 | G-X-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD011R | G-X-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD012 | G-IX-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD012R | G-IX-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD014 | G-IV-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD014R | G-IV-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD015 | G-V-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD015R | G-V-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD016 | UTI-IV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD016R | UTI-IV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD017 | UTI-VI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD017R | UTI-VI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD018 | UTI-VII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD018R | UTI-VII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD019 | UTI-V | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD019R | UTI-V | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD020 | Combi I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD020R | Combi I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD021 | Combi II | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD021R | Combi II | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD022 | Combi III | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD022R | Combi III | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD023 | Combi IV | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD023R | Combi IV | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD024 | Combi V | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD024R | Combi V | Low risk | 25/08/2016 |

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|---|---|---|---|---|
| ASS- Sensitivity Discs (Multi Discs) | OD025 | Combi VI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD025R | Combi VI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD026 | Combi VII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD026R | Combi VII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD027 | Combi VIII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD027R | Combi VIII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD028 | Combi IX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD028R | Combi IX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD029 | Combi X | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD029R | Combi X | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD030 | Combi XI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD030R | Combi XI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD031 | Combi XII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD031R | Combi XII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD032 | Combi XIII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD032R | Combi XIII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD033 | G-V-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD033R | G-V-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD034 | G-VI-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD034R | G-VI-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD035 | UTI III | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD035R | UTI III | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD036 | Pseudo I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD036R | Pseudo I | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD037 | G-VII-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD037R | G-VII-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD038 | G-VIII-plus | Low risk | 20/12/2012 |

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|---|---|---|---|---|
| ASS- Sensitivity Discs (Multi Discs) | OD038R | G-VIII-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD039 | G-XI-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD039R | G-XI-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD040 | UTI-VIII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD040R | UTI-VIII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD041 | G-XII-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD041R | G-XII-plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD042 | G-VI-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD042R | G-VI-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD043 | G-VII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD043R | G-VII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD044 | G-VIII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD044R | G-VIII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD045 | G-IX-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD045R | G-IX-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD046 | G-X-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD046R | G-X-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD047 | G-XI-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD047R | G-XI-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD048 | UTI-IX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD048R | UTI-IX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD049 | G-XIII- plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD049R | G-XIII- plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD050 | G-XIV- plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD050R | G-XIV- plus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD051 | UTI-X | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD051R | UTI-X | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | OD052 | UTI-XI | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD052R | UTI-XI | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD053 | G-XII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD053R | G-XII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD054 | UTI-XII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD054R | UTI-XII | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD055 | G-XIII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD055R | G-XIII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD056 | Combi 59 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD056R | Combi 59 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD057 | G-XVIII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD057R | G-XVIII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD058 | G-XIX-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD058R | G-XIX-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD059 | G-XX-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD059R | G-XX-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD060 | G-XXI-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD060R | G-XXI-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD061 | G-XXII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD061R | G-XXII-minus | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD062 | G-XXIII-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD062R | G-XXIII-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD063 | Pseudo V | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD063R | Pseudo V | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD064 | Combi-XIV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD064R | Combi XIV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD065 | UTI-XVII | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD065R | UTI-XVII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD066 | Combi 82 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD066R | Combi 82 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD067 | Combi 83 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD067R | Combi 83 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD202 | Comb XXI | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD209 | Combi 28 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD211 | Combi 30 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD211R | Combi 30 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD212 | Combi 31 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD215 | Combi 34 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD215R | Combi 34 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD216 | Combi 35 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD216R | Combi 35 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD220 | Combi 39 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD221 | Combi 40 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD223 | G XIV minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD224 | G XV plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD225 | UTI XIII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD226 | Combi 41 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD227 | UTI-XIV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD228 | Pseudo II | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD229 | G-XV-minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD230 | G-XVI-plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD231 | Combi -42 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD232 | Combi 43 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD233 | Combi 44 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD233R | Combi 44 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD234 | Combi 45 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD234R | Combi 45 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD241 | Combi 49 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD241R | Combi 49 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD243 | G XVII minus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD244 | G XIX plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD248 | Combi-53 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD249 | Combi-54 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD250 | Pseudo - III for Pseudomonas | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD251 | GXX plus | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD253 | Combi 56 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD256 | Combi 59 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD256R | Combi 59 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD257 | Combi 60 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD257R | Combi 60 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD258 | Combi 61 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD258R | Combi 61 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD259 | Combi 62 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD259R | Combi 62 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD260 | UTI-XIII | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD261 | UTI-XIV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD262 | UTI-E | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD263 | UTI-XV | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD264 | Pseudo II | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD265 | Combi 63 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD266 | Combi 64 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD267 | Combi 65 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD268 | Combi 66 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD269 | Combi 67 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD269R | Combi 67 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD270 | Combi 68 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD270R | Combi 68 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD271 | Combi 69 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD271R | Combi 69 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD272 | Combi 70 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD272R | Combi 70 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD273 | Combi 71 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD273R | Combi 71 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD274 | Combi 72 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD274R | Combi 72 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD275 | Combi 73 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD275R | Combi 73 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD275RS | Combi 60 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD275S | Combi 60 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD276 | Combi 84 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD276R | Combi 84 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD277 | Combi 77 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD277R | Combi 77 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD278 | Combi 78 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD278R | Combi 78 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD279 | Combi 79 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD279R | Combi 79 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD280 | Combi 80 | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | OD280R | Combi 80 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD281 | Combi 85 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD281R | Combi 85 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD282 | Combi 505 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD282R | Combi 505 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD283 | Combi 506 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD283R | Combi 506 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD284 | Combi 508 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD284R | Combi 508 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD285 | Combi 509 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD285R | Combi 509 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD286 | Combi 510 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD286R | Combi 510 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD287 | Combi 511 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD287R | Combi 511 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD288 | Combi 512 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD288R | Combi 512 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD289 | Combi 513 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD289R | Combi 513 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD290 | Combi 514 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD290R | Combi 514 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD291 | Combi 90 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD291R | Combi 90 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD292 | Combi 91 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD292R | Combi 91 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD293 | Combi 92 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD293R | Combi 92 | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | OD294 | Combi 93 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD294R | Combi 93 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD295 | Combi 516 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD295R | Combi 516 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD296 | Combi 517 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD296R | Combi 517 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD297 | Combi 518 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD297R | Combi 518 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD298 | Combi 94 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD298R | Combi 94 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD299 | Combi 95 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD299R | Combi 95 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD300 | Combi 96 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD300R | Combi 96 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD301 | G minus-24 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD301R | G minus-24 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD302 | G minus-25 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD302R | G minus-25 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD303 | G Plus-15 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD303R | G Plus-15 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD304 | G Plus-16 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD304R | G Plus-16 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD305 | G Plus-17 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD305R | G Plus-17 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD306 | UTI-18 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD306R | UTI-18 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD307 | Pseudo VI | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Multi Discs) | OD307R | Pseudo VI | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD308 | Universal - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD308R | Universal - 1 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD309 | G Plus-18 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD309R | G Plus-18 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD310 | G minus-26 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD310R | G minus-26 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD311 | G minus-27 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD311R | G minus-27 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD312 | G Minus - 28 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD312R | G Minus - 28 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD313 | G Minus - 29 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD313R | G Minus - 29 | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Multi Discs) | OD704 | Combi 77 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD705 | Combi 78 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD706 | Combi 79 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD707 | Combi 80 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD708 | Combi 81 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD709 | Combi 85 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD710 | Combi 86 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD711 | Combi 501 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD712 | Combi 502 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD713 | Combi 503 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD714 | Combi 504 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD715 | Combi 505 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD716 | Combi 506 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD717 | Octodiscs-A | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD718 | Octodiscs-B | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD719 | Octodiscs-C | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD720 | Octodiscs-D | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD721 | Octodiscs-E | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD722 | Octodiscs-F | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD723 | Octodiscs-G | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD724 | Combi 507 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD725 | Combi 508 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD726 | Combi 509 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD727 | Combi 510 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD728 | Combi 511 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD729 | Combi 512 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD730 | Combi 513 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD731 | Combi 514 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD732 | Combi 515 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD733 | Combi 516 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD734 | Combi 517 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD735 | Combi 518 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD736R | Combi 519 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD737 | Combi 520 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD737R | Combi 520 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD738 | Combi 521 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD738R | Combi 521 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD739 | Combi 522 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD739R | Combi 522 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD740 | Combi 523 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD740R | Combi 523 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD741 | Combi 524 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD741R | Combi 524 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD742 | Combi 525 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD742R | Combi 525 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD743 | Combi 526 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD743R | Combi 526 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD744 | Combi 527 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD744R | Combi 527 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD745 | Combi 528 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD745R | Combi 528 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD746 | Combi 529 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD746R | Combi 529 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD747 | Combi 530 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD747R | Combi 530 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD748 | Combi 531 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD748R | Combi 531 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD749 | Combi 532 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD749R | Combi 532 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD750 | Combi 533 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD750R | Combi 533 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD751 | Combi 534 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD751R | Combi 534 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD752 | Combi 535 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD752R | Combi 535 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD753 | Combi 536 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD753R | Combi 536 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD754 | Combi 537 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD754R | Combi 537 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD755 | Combi 538 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD755R | Combi 538 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD756 | Combi 539 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD756R | Combi 539 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD757 | Combi 540 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD757R | Combi 540 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD758 | Combi 541 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD758R | Combi 541 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD759 | Combi 542 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD759R | Combi 542 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD760 | Combi 543 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD760R | Combi 543 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD761 | Combi 544 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD761R | Combi 544 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD762 | Combi 545 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD762R | Combi 545 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD763 | Combi 546 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD763R | Combi 546 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD764 | Combi 547 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD764R | Combi 547 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD765 | Combi 548 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD765R | Combi 548 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD766 | Combi 549 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD766R | Combi 549 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD767 | Combi 550 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD767R | Combi 550 | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Multi Discs) | OD768 | Combi 551 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD768R | Combi 551 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD769 | Combi 552 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD769R | Combi 552 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD770 | Combi 553 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD770R | Combi 553 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD771 | Combi 554 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD771R | Combi 554 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Multi Discs) | OD772 | Combi 555 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD772R | Combi 555 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD773 | Combi 556 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD773R | Combi 556 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD774 | Combi 557 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD774R | Combi 557 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD775 | Combi 558 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD775R | Combi 558 | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Multi Discs) | OD776 | Combi 559 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | OD776R | Combi 559 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | OD777 | Combi 560 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | OD777R | Combi 560 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | OD778 | Combi 561 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | OD778R | Combi 561 | Low risk | 04/07/2018 |
| ASS- Sensitivity Discs (Multi Discs) | OD779 | Combi 562 | Low risk | 22/04/2019 |
| ASS- Sensitivity Discs (Multi Discs) | OD779R | Combi 562 | Low risk | 22/04/2019 |
| ASS- Sensitivity Discs (Multi Discs) | OD780 | Combi 563 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD780R | Combi 563 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD781 | Combi 564 | Low risk | 10/11/2020 |

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| ASS- Sensitivity Discs (Multi Discs) | OD781R | Combi 564 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD782 | Combi 565 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD782R | Combi 565 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD783 | Combi 566 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD783R | Combi 566 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD784 | Combi 567 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Multi Discs) | OD784R | Combi 567 | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Single Discs) | SD001 | Amoxycillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD002 | Ampicillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD002A | Ampicillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD003 | Bacitracin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD004 | Carbenicillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD005 | Cefaloridine (Cephaloridine) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD006 | Chloramphenicol | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD006B | Chloramphenicol(2 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD007 | Chlortetracycline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD008 | Cloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD009 | Colistin (Methane Sulphonate) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD010 | Co-Trimoxazole (Sulpha/Trimethoprim) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD012 | Doxycycline Hydrochloride | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD013 | Erythromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD014 | Framycetin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD015 | Furazolidone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD016 | Gentamicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD017 | Kanamycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD018 | Lincomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD019 | Methicillin | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD020 | Metronidazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD021 | Nalidixic Acid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD022 | Neomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD023 | Nitrofurantoin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD023A | Nitrofurantoin | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD024 | Nitrofurazone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD025 | Nystatin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD026 | Oleandomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD027 | Oxytetracycline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD028 | Penicillin-G | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD029 | Polymyxin-B | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD030 | Rifampicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD031 | Streptomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD032 | Sulphafurazole (Sulfisoxazole) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD033 | Sulphamethizole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD034 | Sulphadiazine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD035 | Amikacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD036 | Sulphaphenazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD037 | Tetracycline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD038 | Triple Sulphas | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD039 | Trimethoprim | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD040 | Cefotaxime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD040A | Cefotaxime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD041 | Cefoxitin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD042 | Furoxone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD043 | Oxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD044 | Tobramycin | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD045 | Vancomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD046 | Netillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD047 | Cefazolin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD048 | Cefalexin(Cephalexin) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD049 | Cycloserine | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD050 | Cephalothin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD051 | Clindamycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD052 | Dicloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD053 | Novobiocin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD054 | Spiramycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD055 | Sulphamethoxypyridazine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD056 | Sulfasomidine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD056A | Sulphamethoxazole | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD057 | Norfloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD058 | Co-Trimazine (Vet.) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD059 | Sisomicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD060 | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD060A | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD061 | Cefuroxime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD062 | Ceftazidime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD062A | Ceftazidime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD063 | Amoxyclav (Amoxycillin/Clavulanic acid) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD063A | Augmentine | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD064 | Azlocillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD065 | Ceftriaxone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD066 | Piperacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD066A | Piperacillin | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD067 | Sterile Discs | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD068 | Methanamine Mandalate | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD069 | Ofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD070 | Pefloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD071 | Co-Trimazine (Human) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD072 | Cefoperazone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD073 | Imipenem | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD074 | Ticarcillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD075 | Cloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD076 | Amoxycillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD077 | Ampicillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD078 | Amoxyclav | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD079 | Cefaloridine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD080 | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD081 | Chloramphenicol | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD082 | Amikacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD083 | Erythromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD084 | Lincomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD085 | Netillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD086 | Nitrofurantoin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD087 | Ofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD088 | Oxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD089 | Penicillin-G | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD090 | Nitrofurantoin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD091 | Streptomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD092 | Sulphadiazine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD093 | Trimethoprim | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD094 | Azlocillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD096 | Rifampicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD097 | Colistin (Methane Sulphonate) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD098 | Lincomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD099 | Metronidazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD101 | Spiramycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD102 | Penicillin-G (1.5 units) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD103 | Nitrofurantoin NIT | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD104 | Neomycin N | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD105 | Bacitracin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD106 | Polymyxin-B | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD107 | Metronidazole | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD108 | Colistin (Methane Sulphonate) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD109 | Ceftriaxone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD110 | Ceftizoxime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD111 | Amphotericin-B | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD112 | Ampicillin/Sulbactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD113 | Ampicillin/Cloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD114 | Fluconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD115 | Clotrimazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD116 | Cefadroxil (Cephadroxil) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD117 | Bacitracin (0.1 units) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD118 | Bacitracin (2 units) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD119 | Bacitracin (1 unit) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD120 | Doxycycline Hydrochloride | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD121 | Novobiocin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD123 | Tetracycline T | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Single Discs) | SD124 | Azithromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD125 | Lomefloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD126 | Roxithromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD127 | Rifampicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD128 | Rifampicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD129 | Amoxycillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD130 | Cephaloridine | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD131 | Chloramphenicol | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD132 | Piperacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD133 | Tetracycline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD134 | Tobramycin TB | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD135 | Trimethoprim | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD136 | Methicillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD137 | Methicillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD138 | Erythromycin | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD139 | Polymyxin-B | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD140 | Floxidin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD141 | Floxidin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD142 | Ciprofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD143 | Cloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD144 | Penicillin-G | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD145 | Penicillin-G | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD147 | Tetracycline | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD148 | Trimethoprim | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD149 | Trimethoprim | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD150 | Enrofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD152 | Penicillin-G | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Single Discs) | SD153 | Chloramphenicol | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD154 | Tobramycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD155 | Vancomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD156 | Enrofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD157 | Cefaclor | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD158 | Minocycline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD159 | Cephradine | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD160 | Cefradine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD161 | Trimethoprim | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD162 | Sparfloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD163 | Vancomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD164 | Clindamycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD165 | Cloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD166 | Gentamicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD167 | Penicillin-G | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD168 | Ceftriaxone Ci | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD169 | Fusidic Acid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD170 | Gentamicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD171 | Fusidic Acid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD174 | Polymyxin-B Pb | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD175 | Pipemidic Acid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD176 | Mecillinam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD177 | Mecillinam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD178 | Pristinomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD179 | Fosfomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD180 | Oxolinic Acid (10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD181 | Spectinomycin | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD182 | Virginamycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD184 | Norfloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD185 | Pipemidic Acid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD186 | Oxolinic Acid (2 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD187 | Flumequine (2 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD188 | Dibekacine (10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD189 | Oxolinic Acid (5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD190 | Flumequine (5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD191 | Kanamycin (1 mcg) (K1) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD192 | Clarithromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD195 | Gentamicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD196 | Nitroxoline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD197 | Furazolidone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD198 | Flumequine | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD199 | Tylosine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD200 | Cefamandole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD201 | Ticarcillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD203 | Cefoperazone | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD204 | Azithromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD205 | Fosfomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD206 | Lomefloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD207 | Ceftazidime /Clavulanic acid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD209 | Cefprozil | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD210 | Piperacillin/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD211 | Cefixime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD212 | Aztreonam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD213 | Teicoplanin | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD214 | Isepamicin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD215 | Linezolid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD216 | Levofloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD217 | Moxifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD218 | Cefdinir | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD219 | Cefepime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD220 | Moxalactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD221 | Itraconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD222 | Erythromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD223 | Kanamycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD224 | Ketoconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD225 | Mezlocillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD231 | Cefoperazone :Sulbactum (30mcg:10mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD232 | Fluconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD233 | Amphotericin B | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD234 | Cefepime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD235 | Cefpirome | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD236 | Streptomycin For detection of HLAR Strains. | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD237 | Enoxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD238 | Kit I for ESBL Identification, Cefotaxime (Cephotaxime) Kit contains 6 cartridges (6CT): 3CT of SD040 Cefotaxime (Cephotaxime) 30 mcg, 3CT of SD724 Cefotaxime (Cephotaxime)/Clavulanic acid 30/10 mcg | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD239 | Kit II for ESBL Identification, Cefepime Kit contains 6 cartridges (6CT): 3CT of SD219 Cefepime 30 mcg, 3CT of SD234 Cefepime /Clavulanic acid 30/10 mcg | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD240 | Kit III for ESBL Identification, Ceftazidime Kit contains 6 cartridges (6CT): 3CT of SD062 Ceftazidime 30 mcg, 3CT of SD207 Ceftazidime /Clavulanic acid 30/10 mcg | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD241 | Kit IV for ESBL Identification, Cefpirome Kit contains 6 cartridges (6CT): 3CT of SD738 Cefpirome 30 mcg, 3CT of SD235 Cefpirome /Clavulanic acid 30/7.5 mcg | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD242 | Kit V for ESBL identif | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD243 | Amoxyclav (Amoxycillin / Clavulanicacid) | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Single Discs) | SD244 | Cefmetazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD245 | Cinoxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD246 | Nafcillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD247 | Cefepime/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD248 | Cefonicid | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD249 | Cefotetan | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD250 | Gemifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD251 | Ceftriaxone/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD252 | Ceftazidime/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD253 | Cefoperazone/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD254 | Cefoperazone/ | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD255 | Cefpodoxime/ Clavulanic acid | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD256 | Ceftriaxone/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD257 | Cefepime/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD258 | Nadifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD259 | Cefoperazone/Sulbactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD260 | Lomefloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD261 | Ceftriaxone/ Sulbactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD262 | Cefepime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD263 | Aztreonam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD264 | Amoxycillin/ | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD265 | Imipenem/Cilastin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD266 | Cefixime/ | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD267 | Prulifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD268 | Prulifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD269 | Ceftazidime/Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD270 | Amphotericin B | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD271 | Nystatin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD272 | Miconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD273 | Miconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD274 | Ketoconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD275 | Ketoconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD276 | Itraconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD277 | Voriconazole | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD278 | Tigecycline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD279 | Faropenem | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD280 | Ertapenem | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD281 | Amoxyclav | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD282 | Imipenem-EDTA | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD283 | Doripenem | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD284 | Cloxacillin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD285 | Cefoxitin- | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD286 | Amoxycillin/Sulbactam | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD287 | Ampicillin/Sulbactam | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD288 | Cefotaxime CTX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD289 | Ceftriaxone CTR | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD290E | Ceftaroline | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD291E | Telithromycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD292E | Piperacillin / Tazobactam | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD293E | Mupirocin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD294E | Ceftibuten | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD295E | Cefotaxime CTX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD296E | Linezolid LZ | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD297 | Colistin Sulphate | Low risk | 17/06/2021 |

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| ASS- Sensitivity Discs (Single Discs) | SD298 | Caspofungin | Low risk | 17/06/2021 |
| ASS- Sensitivity Discs (Single Discs) | SD701 | Carbenicilline | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD704 | Cefradine | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD705 | Amoxycillin (2 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD709 | Novobiocin ( 5mcg ) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD712 | Oleandomycin (5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD715 | Fluconazole ( 25 mcg ) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD722 | Penicillin-G (2mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD723 | Ampicillin (20mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD724 | Cefotaxime/Clavulanic acid (30/10 mcg) | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD725 | Cefpodoxime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD726 | Ceftazimide/Clavulinic ( 3/10 mcg ) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD727 | Meropenem | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD730 | Metronidazole (50 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD731 | Neomycin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD732 | Novobiocin (5mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD736 | Bacitracin B 0.05 units /disc | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD737 | Gatifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD738 | Cefpirome (Cfp) (30mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD740 | Gatifloxacin | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD741 | Cephotaxime/Sulbactam (30/15 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD744 | Ofloxacin Of 30 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD745 | Norfloxacin (30mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD746 | Gentamicin (200mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD748 | Mupirocin MU 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD751 | Cefpodoxime/ Clavulanic acid (10/1 MCG) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD753 | Gatifloxacin | Low risk | 20/12/2012 |

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| ASS- Sensitivity Discs (Single Discs) | SD755 | Ceftiofur (0.2mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD756 | Ceftiaxone (30 mcg) / Sulbactam (15 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD761 | Sparfloxacin Sc (10mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD764 | Ceftriaxone/ Tazobactam (80/10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD765 | Gemifloxacin (GEM) 5mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD767 | Ceftazidime-Tazobactam (CaT) (30/10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD768 | Cefoperazone-tazobactam (75/10mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD769 | Cefoperazone-Sulbactam (Cfs) (75/10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD770 | Cefepime/Tazobactam (30/10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD771 | Cefpodoxime / Clavulanic acid (10/5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD773 | Piperacillin / Sulbactam (100/10 mcg ) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD774 | Faropenem (5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD775 | Ceftriaxone (30 mcg) / Tazobactam (10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD776 | Cefepime (80 mcg) / Tazobactam (10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD777 | Nadifloxacin (5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD779 | Cefoperazone / Sulbactam (50 / 50 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD780 | Lomefloxacin Lo (15 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD781 | Cefixime/Clavulanic acid Cmc (200/125 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD782 | Cefepime Cpm (50 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD783 | Aztreonam Ao (50 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD784 | Amoxycillin/Sulbactam Ams (30/15 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD785 | Imipenem/Cilastatin Ic (10/10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD786 | Cefixime / Clavulanic acid Cmc (5/10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD787 | Prulifloxacin Pr (10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD788 | Prulifloxacin Pr (5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD789 | Ceftriaxone / Sulbactam (500/250 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD790 | Ceftriaxone / Sulbactam (1000/500 mcg) | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Single Discs) | SD791 | Piperacillin + Tazobactam (80:10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD792 | Pazufloxacin (PZ) (25 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD793 | Cefditoren (10 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD794 | Cefpodoxime/Clavulanic acid (10/6.25mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD795 | Cefipime / Amikacin (30 / 7.5 mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD796 | Cefepime / Sulbactam (30/15 mcg) CPS | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD797 | Ceftazidime / Sulbactam (30/15 mcg) CAS | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD798 | Ceftriaxone/Tobramycin (30/5.4 mcg) CTB | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD799 | Ceftriaxone/Vancomycin (30/15 mcg) CVA | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD800 | Cefpirome / Sulbactam (30/15 mcg) CRS | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD801 | Cefaperazone/Sulbactum (70/35mcg)(CSB) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD802 | Ceftazidime Tobramycin (30+3.6 mcg) CFT | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD803 | Amoxycillin/Clavulanic acid AC 50/10 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD804 | Cefpodoxime / Clavulanic acid (24:15mcg) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD805 | Cefixime : Ofloxacin COF 5:5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD806 | Balofloxacin BF 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD807 | Tigecycline TGC 20 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD808 | Ampicillin / Cloxacillin 128/128µg Ax | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD809 | Amoxycillin/Cloxacillin 128/128µg ACX | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD810 | Gentamicin GEN 128µg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD811 | Enrofloxacin EX 8µg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD812 | Ciprofloxacin CIP 8µg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD813 | Tetracyclin TE 128µg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD814 | Chloramphenicol C 8 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD815 | Streptomycin/Penicillin SPN 128/128mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD816 | Ceftazidime/Tobramycin CFT 30/10mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD816V | Ceftazidime/Tobramycin CFT (30:10) | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Single Discs) | SD817 | Cefepime / Amikacin CPA 30/10mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD818 | Balofloxacin BF 10mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD819 | Oxacillin Ox 10mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD820 | Cefixime | Low risk | 20/12/2012 |
| ASS- Sensitivity Discs (Single Discs) | SD821 | Cefpodoxim CPD 30mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD822 | Garenoxacin GRN 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD823 | Sitafloxacin STX 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD824 | Tosufloxacin TOS 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD825 | Biapenem BPM 10 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD826 | Cefepime Amikacin 58.8:14.6.mcg CPA | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD827 | Florfenikol FLO 30mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD828 | Cefpodoxime:Levofloxacin 10:5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD829 | Meropenem/Sulbactam MRS 10:5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD830V | Ceftriazone Vancomycin CVA (30:30) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD831 | Ampicillin/Sulbactum (A/S) 20:10 | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD832 | Cefixime : Azithromycin CFA 5:15 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD833 | Cefquinome CEQ 30mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD834 | Ceftriaxone CTR 128 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD835 | Sulphatrimethoprim STM 128/128 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD836 | Erythromycin E 60 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD837 | Kanmycin K 1000 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD838 | Quninupristin/Dalfopristin RP 15/15 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD839 | Levofloxacin/Cefpodoxime LEC 250 : 200 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD840 | Ampicillin/Sulbactam A/S 20/12.5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD841 | Garenoxacin GRN 1mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD842 | Garenoxacin GRN 5mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD843 | Mipenem (Meropenem) MIP 10 mcg | Low risk | 25/08/2016 |

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| ASS- Sensitivity Discs (Single Discs) | SD844 | Ranicef (Cefdinir) RNF 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD845 | Clavamox (Amoxycillin / Clavulanic acid) | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD846 | Ciprotab (Ciprofloxacin) CPT 5 mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD847 | Ciprotab (Ciprofloxacin) CPT 10mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD848 | Meropenem/Sulbactam MRS 2/200mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD849 | Flucloxacillin FCO 30mcg | Low risk | 25/08/2016 |
| ASS- Sensitivity Discs (Single Discs) | SD850 | Cefuroxime/Clavulanic acid CCV 30/7.5mcg | Low risk | 28/04/2017 |
| ASS- Sensitivity Discs (Single Discs) | SD851 | Cefixime/Dicloxacillin CDC 5/12.5mcg | Low risk | 28/04/2017 |
| ASS- Sensitivity Discs (Single Discs) | SD852 | Cefpodoxime / Clavulanic acid CCL 10/5mcg | Low risk | 16/12/2017 |
| ASS- Sensitivity Discs (Single Discs) | SD853 | Nafithromycin NFT 15mcg | Low risk | 30/10/2018 |
| ASS- Sensitivity Discs (Single Discs) | SD854 | Levonadifloxacin LND 10mcg | Low risk | 30/10/2018 |
| ASS- Sensitivity Discs (Single Discs) | SD855 | Dicrysticin-S DCR 50mcg | Low risk | 22/04/2019 |
| ASS- Sensitivity Discs (Single Discs) | SD856 | Garenoxacin GRN 10mcg | Low risk | 22/04/2019 |
| ASS- Sensitivity Discs (Single Discs) | SD857 | Cefepime / sulbactam | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Single Discs) | SD858 | Cefotaxime / Sulbactam | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Single Discs) | SD859 | Ceftizoxime / Sulbactam | Low risk | 10/11/2020 |
| ASS- Sensitivity Discs (Single Discs) | SD860 | Meropenem / EDTA | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM001 | Amikacin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM002 | Amoxicillin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM003 | Amoxyclav HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM068 | Ampicillin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM109 | Ampicillin /Sulbactam HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM070 | Cefepime HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM064 | Cefotaxime HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM101 | Cefoxitin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM012 | Ceftazidime HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM066 | Ceftriaxone HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM016 | Chloramphenicol HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM017 | Ciprofloxacin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |

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| ASS-HiComb™ MIC Strip, Modified | MDM020 | Colistin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM108 | Fosfmycin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM025 | Gentamicin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM080 | Meropenem HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM065 | Oxacillin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM084 | Penicillin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM043 | Polymyxin B HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM055 | Teicoplanin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM056 | Tetracycline HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM089 | Tigecycline HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM059 | Trimethoprim HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM060 | Vancomycin HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM071 | Amphotericin B HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM072 | Fluconazole HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiComb™ MIC Strip, Modified | MDM086 | Voriconazole HiComb™ MIC Strip, Modified | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK001 | Amikacin HiMIC™ Plate Kit (contains HMP001,LQ314II,PW1378,R-MPK001) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK068 | Ampicillin HiMIC™ Plate Kit (contains HMP068,LQ314II,PW1378,R-MPK068) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK109 | Ampicillin/Sulbactam HiMIC™ Plate Kit (contains HMP109,LQ314II,PW1378,R-MPK109) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK071 | Amphotericin B HiMIC™ Plate Kit (contains HMP071,LQ314I,PW1378,R-MPK071) | Low risk | 17/06/2021 |
| ASS-HiMIC™ Plate Kit | MPK070 | Cefepime HiMIC™ Plate Kit (contains HMP070,LQ314I,PW1378,R-MPK070) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK101 | Cefoxitin HiMIC™ Plate Kit (contains HMP101,LQ314II,PW1378,R-MPK101) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK012 | Ceftazidime HiMIC™ Plate Kit (contains HMP012,LQ314II,PW1378,R-MPK012) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK016 | Chloramphenicol HiMIC™ Plate Kit (contains HMP016,LQ314II,PW1378,R-MPK016) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK017 | Ciprofloxacin HiMIC™ Plate Kit (contains HMP017,LQ314II,PW1378,R-MPK017) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK019 | Clindamycin HiMIC™ Plate Kit (contains HMP019,LQ314II,PW1378,R-MPK019) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK020 | Colistin HiMIC™ Plate Kit (contains HMP020,LQ314II,PW1378,R-MPK020) | Low risk | 10/11/2020 |

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| ASS-HiMIC™ Plate Kit | MPK085 | Ertapenem HiMIC™ Plate Kit (contains HMP085,LQ314II,PW1378,R-MPK085) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK025 | Gentamicin HiMIC™ Plate Kit (contains HMP025,LQ314II,PW1378,R-MPK025) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK104 | Imipenem HiMIC™ Plate Kit (contains HMP104,LQ314II,PW1378,R-MPK104) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK156 | Isavuconazole HiMIC™ Plate Kit (contains HMP156,LQ314I,PW1378,R-MPK156) | Low risk | 17/06/2021 |
| ASS-HiMIC™ Plate Kit | MPK073 | Itraconazole HiMIC™ Plate Kit (contains HMP073,LQ314I,PW1378,R-MPK073) | Low risk | 17/06/2021 |
| ASS-HiMIC™ Plate Kit | MPK080 | Meropenem HiMIC™ Plate Kit (contains HMP080,LQ314I,PW1378,R-MPK080) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK084 | Penicillin HiMIC™ Plate Kit (contains HMP084,LQ314II,PW1378,R-MPK084) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK042 | Piperacillin/Tazobactam HiMIC™ Plate Kit (contains HMP042,LQ314I,PW1378,R-MPK042) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK043 | Polymyxin B HiMIC™ Plate Kit (contains HMP043,LQ314II,PW1378,R-MPK043) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK120 | Posaconazole HiMIC™ Plate Kit (contains HMP120,LQ314II,PW1378,R-MPK120) | Low risk | 17/06/2021 |
| ASS-HiMIC™ Plate Kit | MPK055 | Teicoplanin HiMIC™ Plate Kit (contains HMP055,LQ314II,PW1378,R-MPK055) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK089 | Tigecycline HiMIC™ Plate Kit (contains HMP089,LQ314II,PW1378,R-MPK089) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK060 | Vancomycin HiMIC™ Plate Kit (contains HMP060,LQ314II,PW1378,R-MPK060) | Low risk | 10/11/2020 |
| ASS-HiMIC™ Plate Kit | MPK086 | Variconazole HiMIC™ Plate Kit (contains HMP086,LQ314II,PW1378,R-MPK086) | Low risk | 17/06/2021 |

# Amoxicillin Ezy MIC™ Strip (AMX) (0.016-256 mcg/ml) EM002

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use

It is a unique MIC determination paper strip which is coated with Amoxicillin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016 mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for 2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, *Bacteroides* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar, supplemented with 5% sterile, defibrinated sheep blood for fastidious organism such as streptococci.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Amoxicillin and other members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin, Carbapenems always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values "in-between" two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Amoxicillin showing reading of 0.38 mcg/ml should be rounded up to next concentration ie. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL :**

**Interpretation**

**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *S.pneumoniae* (non meningitis) | 35-37°C for 20-24hrs at 5% $CO_2$ | 2 | 4 | 8 |

**Quality Control**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures recommended by CLSI on suitable medium incubated appropriately.

**Table 2:** Following are the reference MIC values (mcg/ml) range for Amoxicillin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *K. pneumoniae* ATCC 700603 | Mueller Hinton Agar | 35-37°C for 18 hrs. | >128 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.03 – 0.06 – 0.12 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen, J. H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; 32nd Edition. M100-Ed32, Vol.42, No.2, Jan-2022.

**Packing:**
Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Amoxicillin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

Revision : 04/2022

CE

On receipt store at -20°C

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Do not use if package is damaged

# Chloramphenicol Ezy MIC™ Strip (CHL) (0.016-256 mcg/ml) EM016

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use

It is a unique MIC determination paper strip which is coated with Chloramphenicol in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).

Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland. This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

Please refer disclaimer Overleaf

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood is recommended. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117). For *B. fragilis*, Brucella Agar (M074) with Hemin and Vitamin K1 supplemented with 5 % v/v sterile defibrinated sheep blood is to be recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bacteriostatic drugs such Chloramphenicol, Tetracycline, Azithromycin, Fluconazole, Linezolid and Trimethoprim/ sulphamethoxazole, read MICs at 80% inhibition for homogenously sensitive strains such as QC control strains.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two-fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Chloramphenicol showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.

8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL:**

**Interpretation**
**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae*, *Enterococcus* spp., *Staphylococcus* spp., *S.maltophila*, *B.cepacia*, other non-*Enterobacteriaceae* | 35-37ºC for 18hrs | 8 | 16 | 32 |
| *Haemophillus* spp., *N.meningitidis* | 35-37ºCfor 20-24 hrs with 5% $CO_2$ | 2 | 4 | 8 |
| *Streptococcus* spp. Beta haemolytic groups, *Streptococcus* spp Viridans group | 35-37ºCfor 20-24 hrs with 5% $CO_2$ | 4 | 8 | 16 |
| *S.pneumoniae* | 35-37ºCfor 20-24 hrs with 5% $CO_2$ | 4 | - | 8 |
| Anerobes | 35-37ºC for 24 -48 hrs under anaerobic condition | 8 | 16 | 32 |

**Quality Control**
Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.

**Table 2:** Following are the reference MIC values (mcg/ml) range for Chloramphenicol.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S. aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 2.0 – 4.0 – 8.0 -16.0 |
| *E. faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 4.0 – 8.0 – 16.0 |
| *E. coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 2.0 – 4.0 – 8.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24 hrs at 5% $CO_2$ | 2.0 – 4.0 – 8.0 |
| *H. influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24 hrs at 5% $CO_2$ | 0.25 - 0.5 – 1.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen, J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; 32nd Edition. M100-Ed32, Vol.42, No.2, Jan-2022.


**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Chloramphenicol Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

Revision : 04/2022

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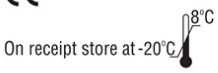


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Do not use if package is damaged

# Gentamicin Ezy MIC™ Strip (GEN) (0.016-256 mcg/ml) EM025

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use

It is a unique MIC determination paper strip which is coated with Gentamicin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is

recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Gentamicin, Vancomycin, Amikacin and members of β-lactams class of drugs, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values "in-between" two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Gentamicin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strips are at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL:**

**Interpretation**

**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae, Pseudomonas* spp, *Acinetobacter* spp, *Staphylococcus* spp other Non- *Enterobacteriaceae* | 35-37°C for 18 hrs. | 4 | 8 | 16 |

**Quality control**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.

**Table 2:** Following are the reference MIC values (mcg/ml) range for Gentamicin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.12 - 0.25 - 0.5 - 1.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 4.0 – 8.0 – 16.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.25 - 0.5 - 1.0 |
| *P.aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 – 1.0 – 2.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions .

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; 32nd Edition. M100-Ed32, Vol.42, No.2, Jan-2022.

**Packing:**
Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Gentamicin Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

Revision : 04/2022

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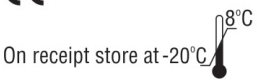


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# Linezolid Ezy MIC™ Strip (LNZ) (0.016-256 mcg/ml) EM029

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use

It is a unique MIC determination paper strip which is coated with Linezolid in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**CLSI RECOMMENDATION FOR LINEZOLID SENSITIVITY TEST**

- For reading results of Linezolid, the plates should be held up to light source (i.e. results are to be read with transmitted light).The zone margin is to be considered as the area showing no obvious, visible growth that can be detected with the unaided eye. Ignore the faint growth of tiny colonies that can be detected with the help of magnifying lens only, at the edge of the zone of inhibition.
- Any discernable growth observed within the zone of inhibition is indicative of resistance.
- Organisms with resistant results by disc diffusion should be confirmed with MIC method.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).

Please refer disclaimer Overleaf

Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood is recommended. For *B. fragilis*, Brucella Agar (M074) with Hemin and Vitamin K1 supplemented with 5 % v/v sterile defibrinated sheep blood is to be recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate pre-spread with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bacteriostatic drugs such Chloramphenicol, Tetracycline, Azithromycin, Fluconazole, Linezolid and Trimethoprim/ sulphamethoxazole, read MICs at 80% inhibition for homogenously sensitive strains such as QC control strains.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Linezolid showing reading of 0.75 mcg/ml should be rounded up to next concentration i.e. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.

8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL :**

**Interpretation:**
**Table 1:** Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Staphylococcus* spp. | 35-37°C for 18 hrs. | 4 | - | 8 |
| *Enterococcus* spp. | 35-37°C for 18 hrs. | 2 | 4 | 8 |
| *S.pneumoniae, Streptococcus* spp. Beta haemolytic group, *Streptococcus* spp. Viridans group | 35-37°C for 20-24hrs at 5% $CO_2$ | 2 | - | - |

**Quality control:**
Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.

**Table 2:** Following are the reference MIC values (mcg/ml) range for Linezolid.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 – 2.0 – 4.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 – 2.0 – 4.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 –0.5 – 1.0 – 2.0 |
| *B. fragilis* ATCC 25285 | Brucella Agar with Hemin and Vitamin K1, supplemented with 5 % v/v defibrinated sterile sheep blood | 35-37°C for 24-48 hrs under strict anaerobic condition | 2.0–4.0–8.0 |

**Storage & Shelf Life:**
1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the

clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; 32nd Edition. M100-Ed32, Vol.42, No.2, Jan-2022.

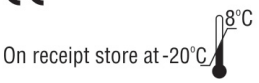

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Linezolid Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

Revision : 04/2022

CE




IVD In vitro diagnostic medical device

Plot No. C-40, Road No. 21Y,
Wagale Industrial Area,
Thane (W) - 400604,
Maharashtra, India

EC REP CEpartner4U,
ESDOORNLAAN 13,
3951DB MAARN, NL
www.cepartner 4u.eu

Do not use if package is damaged

# Moxifloxacin Ezy MIC™ Strip (MXF) (0.002 - 32 mcg/ml) EM033

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use

It is a unique MIC determination paper strip which is coated with Moxifloxacin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used. For *B. fragilis*, Brucella Agar (M074) with Hemin and Vitamin K1 supplemented with 5 % v/v sterile defibrinated sheep blood is to be recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Moxifloxacin and other members of quinolones class of drugs, Amikacin, Vancomycin, Gentamicin and members of β-lactams class of drugs always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Moxifloxacin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strips is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL:**

**Interpretation:**

**Table 1:** Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Staphylococcus* spp. | 35-37°C for 18 hrs. | 0.5 | 1 | 2 |
| *Haemophilus* spp. | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | - | - |
| *S. pneumoniae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |
| Anaerobe | 35-37°C for 20-24 hrs under anaerobic condition | 2 | 4 | 8 |

**Quality control**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
**Table 2:** Following are the reference MIC values (mcg/ml) range for Moxifloxacin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.016 - 0.03 - 0.06 - 0.12 |
| *E. faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.06 -0.12 -0.25 -0.5 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.008 - 0.016 - 0.03 - 0.06 |
| *P. aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 -2.0 - 4.0 -8.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.06 -0.12 - 0.25 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.008 - 0.016 - 0.03 |
| *B.fragilis* ATCC 25285 | Brucella Agar with Hemin and Vitamin K1, supplemented with 5 % v/v defibrinated sterile sheep blood | 35-37°C for 24-48 hrs under strict anaerobic condition | 0.125 - 0.25- 0.5 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; 32nd Edition. M100-Ed32, Vol.42, No.2, Jan-2022.


**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Moxifloxacin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

Revision : 04/2022

CE

On receipt store at -20°C / 8°C

IVD In vitro diagnostic medical device

Plot No. C-40, Road No. 21Y,
Wagale Industrial Area,
Thane (W) - 400604,
Maharashtra, India

EC REP CEpartner4U,
ESDOORNLAAN 13,
3951DB MAARN, NL
www.cepartner 4u.eu

Do not use if package is damaged

# Rifampicin Ezy MIC™ Strip (RIF) (0.002-32 mcg/ml) EM045

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Rifampicin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

Please refer disclaimer Overleaf

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Rifampicin and members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin, Carbapenemes always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Rifampicin showing reading of 0.38 mcg/ml should be rounded up to next concentration ie. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL (As per CLSI Guidelines):**

**Interpretation**

**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Staphylococcus* spp, *Enterococcus* spp | 35-37°C for 18 hrs. | 1 | 2 | 4 |
| *Haemophilus* spp., *S.pneumoniae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |
| *N.meningitidis* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | 1 | 2 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
**Table 2:** Following are the reference MIC values (mcg/ml) range for Rifampicin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.004 - 0.008 - 0.016 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 - 1.0 - 2.0 - 4.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 4.0 - 8.0 - 16.0 |
| *Ps.aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 16.0 - 32.0 - 64.0* |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.016 - 0.03 - 0.06 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 - 0.5 - 1.0 |

* Quality control test of *P. aeruginosa* ATCC 27853 may or may not show MIC value, as highest concentration is 32 mcg/ml

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Rifampicin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Streptomycin Ezy MIC™ Strip (STR) (0.016-256 mcg/ml) EM048

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Streptomycin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Streptomycin, Amikacin, Vancomycin, Gentamicin and other members of β-lactams class of drugs always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Streptomycin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| Enterococci (HLAR) | 35-37°C for 18 hrs. | - | - | 1000 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures on suitable medium incubated appropriately.

Following are the reference MIC values (mcg/ml) range for Streptomycin.*

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 2.0 – 4.0 – 8.0 |
| *P. aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 8.0 – 16.0 – 32.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 64.0 – 128.0 – 256.0 |

*: Ranges not as per CLSI guidelines.

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C. For prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *in vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Streptomycin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Teicoplanin Ezy MIC™ Strip (TEI) (0.016-256 mcg/ml) EM055

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Teicoplanin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for 2-8 hours until light to

moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).

Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland. This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, and streptococci for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Teicoplanin Amikacin, Vancomycin, Gentamicin and members of β-lactams class of drugs, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Teicoplanin showing reading of 0.75 mcg/ml should be rounded up to next concentration i.e. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as > the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.

7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Staphylococcus* spp, *Enterococcus* spp | 35-37°C for 18 hrs. | 8 | 16 | 32 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.

Following are the reference MIC values (mcg/ml) range for Teicoplanin:

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.25 - 0.5 - 1.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.25 - 0.5 - 1.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenb
3. Erg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
4. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
5. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Teicoplanin Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Vancomycin Ezy MIC™ Strip (VAN) (0.016-256 mcg/ml) EM060

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Vancomycin in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**CLSI RECOMMENDATION FOR VANCOMYCIN SENSITIVITY TEST**

High molecular weight antibiotics such as Vancomycin, polymyxin B and colistin do not diffuse in concentration gradient manner while diffusing through the agar medium when the disc susceptibility test is employed. The Antimicrobial Susceptibility Testing using disc diffusion test does not differentiate vancomycin-susceptible isolates of *S.aureus* from Vancomycin intermediate isolates, nor does the test differentiates among Vancomycin–susceptible, intermediate, and resistant isolates of coagulase-negative staphylococci, all of which may give similar size zones of inhibition.

**CLSI therefore recommends that MIC test should be performed to determine the susceptibility of all isolates of staphylococci to Vancomycin .[1]**

**Usefulness of Vancomycin Ezy MIC™ strip**

1) Besides obtaining accurate MIC values for Gram- positive cultures, VISA (Vancomycin Intermediate *Staphylococcus aureus*) can be detected when isolated colonies appear within the zone of inhibition of Vancomycin particularly when 1.0 McFarland inoculum is used and MIC is read on full 48 hrs incubation. The sensitivity of the method can be further enhanced for better detection of VISA/ VRSA (Vancomycin Resistant *Staphylococcus aureus* / hVISA (Hetro Vancomycin Intermediate *Staphylococcus aureus*) using BHI agar with higher inoculum and 48 hr incubation.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm yields $10^5$ - $10^6$ cells/ml).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, and streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar supplemented with 5% sterile, defibrinated blood is recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such Vancomycin, Gentamicin, Amikacin, and members of β-lactams class of drugs, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Vancomycin showing reading of 0.75 mcg/ml should be rounded up to next concentration i.e. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip ( does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strips is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL (As per CLSI Guidelines):**

**Interpretation**

**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| **When testing** | **Interpretative Criteria** | | |
|---|---|---|---|
| | **≤ S** | **I** | **≥ R** |
| *Staphylococcus* spp | 2 | 4-8 | 16 |
| Coagulase negative *Staphylococci* spps. and *Enterococcus* | 4 | 8-16 | 32 |
| *S.pneumoniae, Streptococcus* spps. Beta haemolytic group, *Streptococcus* spps. Viridans group | 1 | - | - |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Vancomycin.

| **Organism** | **Medium used** | **Incubation** | **Std. Quality Control limits (mcg/ml)** |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 – 1.0 – 2.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 – 2.0 – 4.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.12– 0.25 – 0.5 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**
Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.
1) Vancomycin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Cefotaxime Ezy MIC™ Strip (CTR) (0.016-256 mcg/ml) EM064

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Cefotaxime on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, *Bacteroides* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented

with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used. For *B. fragilis*, Brucella Agar (M074) with Hemin and Vitamin K1 supplemented with 5 % v/v sterile defibrinated sheep blood is to be recommended.

2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Cefotaxime and other members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Cefotaxime showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* | 35-37°C for 18 hrs. | 1 | 2 | 4 |
| Other non-*Enterobacteriaceae*, *Acinetobacter* spp, *Staphylococcus* spp | 35-37°C for 18 hrs. | 8 | 16-32 | 64 |
| *Haemophilus* spp | 35-37°C for 20-24hrs at 5% $CO_2$ | 2 | - | - |
| *N.meningitidis* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.12 | - | - |
| *Streptococcus spp.* Beta haemolytic group, *N.gonorrhoeae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | - | - |
| *S.pneumoniae*(meningitis) | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | 1 | 2 |
| *S.pneumoniae* (non meningitis), *Streptococcus* spp. Viridans group | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |
| Anaerobes | 35-37°C for 24-48 hrs under anaerobic condition | 16 | 32 | 64 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Cefotaxime.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 - 2.0 - 4.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.03 – 0.06 – 0.12 |
| *P. aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 8.0 - 16.0 - 32.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.03 – 0.06 – 0.12 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.12 - 0.25 - 0.5 |
| *N. gonorrhoeae* ATCC 49226 | GC Agar Base (M434) with 1% defined growth supplement (FD025) | 35-37°C for 24 – 48 hrs at 5% $CO_2$ | 0.016 – 0.03 – 0.06 |
| *B. fragilis* ATCC 25285 | Brucella Agar with Hemin and Vitamin K1, supplemented with 5 % v/v defibrinated sterile sheep blood | 35-37°C for 24-48 hrs under strict anaerobic condition | 8.0 – 16.0 – 32.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at room temperature and Ezy MIC™ Strip container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Second Informational Supplement. M100-S22, Vol. 32, No.3, Jan 2012.
5. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in a sealed glass vial with a desiccator capsule.

1) Cefotaxime Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Oxacillin Ezy MIC™ Strip (OXA) (0.016-256 mcg/ml) EM065

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Oxacillin in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

## CLSI RECOMMENDATION FOR DETECTION OF OXACILLIN REASISTANCE

- Of the penicillinase-stable penicillins, Oxacillin is preferred for *in vitro* testing. Oxacillin is more resistant to degradation in storage and is more likely to detect heteroresistant staphylococcal strains. Oxacillin susceptibility results can be applied to the other penicillinase-stable penicillins like Cloxacillin, Dicloxacillin, Methicillin, Flucloxacillin and Naficillin.
- Addition of 2% NaCl is required for dilution testing of Oxacillin to improve the detection of heteroresistant MRSA.
- The use of direct colony suspension method for preparation of inoculum is necessary.
- Incubate tests to detect MRS for full 24 hours at 35± 2°C when using Oxacillin (testing at temperature above 35°C may not detect MRS).

## METHOD AND USE OF EZY MIC™ STRIPS

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter.

Please refer disclaimer Overleaf

Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, and streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar with added 2% NaCl for detection of ORSA or ORSE.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15-30 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate pre-spread with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Oxacillin and other members of β-lactams class of drugs Carbapenems, Amikacin, Vancomycin, Gentamicin, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Oxacillin showing reading of 0.75 mcg/ml should be rounded up to next concentration i.e. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip),
report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.

4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Interpretative Criteria | | |
|---|---|---|---|
| | **≤ S** | **I** | **≥ R** |
| *S. aureus* and *S. lugdunensis* | 2 | - | 4 |
| Coagulase-negative Staphylococci except *S.lugdunesis* | 0.25 | - | 0.5 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Oxacillin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.125 – 0.25 – 0.5 |
| *E. faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 8.0 – 16.0 – 32.0 |
| *S.aureus* ATCC 43300 (MRSA) | Mueller Hinton Agar + 2 % NaCl | 35-37°C for 24 hrs. | 16.0 -32.0 – 64.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**

Each Pack contains following material packed in sealed glass vial with desiccator capsule.

1) Oxacillin Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Ceftriaxone Ezy MIC™ Strip (CTR) (0.016-256 mcg/ml) EM066

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Ceftriaxone on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Ceftriaxone and other members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Ceftriaxone showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**Interpretation:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* | 35-37°C for 18 hrs. | 1 | 2 | 4 |
| *Staphylococcus* spp, other non-*Enterobacteriaceae*, *Acinetobacter* spp | 35-37°C for 18 hrs. | 8 | 16-32 | 64 |
| *Haemophilus* spp | 35-37°C for 20-24hrs at 5% $CO_2$ | 2 | - | - |
| *N.gonorhoeae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | - | - |
| *N.meningitidis* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.12 | - | - |
| *Streptococcus* spp. Beta haemolytic group | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | - | - |
| *S.pneumoniae* (meningitis) | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | 1 | 2 |
| *S.pneumoniae* (non meningitis), *Streptococcus* spps. Viridians group | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |
| Anaerobes | 35-37°C for 20-24hrs under anaerobic condition | 16 | 32 | 64 |

**QUALITY CONTROL:**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Ceftriaxone.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 - 2.0 - 4.0 - 8.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.03 – 0.06 – 0.12 |
| *E.coli* ATCC 35218* | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.06 – 0.12 – 0.25 |
| *P. aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 8.0 - 16.0 - 32.0 - 64.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.03 – 0.06 – 0.12 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.06 - 0.12 - 0.25 |

*Not as per CLSI

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen, J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Ceftriaxone Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Ampicillin Ezy MIC™ Strip (AMP) (0.016-256 mcg/ml) EM068

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Ampicillin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Amikacin, Vancomycin, Gentamicin and members of β-lactams class of drugs, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Ampicillin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL (As per CLSI Guidelines):**

**Interpretation**

**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* | 35-37°C for 18 hrs. | 8 | 16 | 32 |
| *Staphylococcus* spp | 35-37°C for 18 hrs. | 0.25 | - | 0.5 |
| *Enterococcus* spp | 35-37°C for 18 hrs. | 8 | - | 16 |
| *Haemophilus* spp | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |
| *N.meningitidis* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.12 | 0.25-1 | 2 |
| *Streptococcus spps.* Beta haemolytic group | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | - | - |
| *Streptococcus* spps. Viridians group | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | 0.5-4 | 8 |
| Anaerobes | 35-37°C for 18 hrs. | 0.5 | 1 | 2 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
**Table 2:** Following are the reference MIC values (mcg/ml) range for Ampicillin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 - 1.0 - 2.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 - 1.0 - 2.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 2.0 - 4.0 - 8.0 |
| *E.coli* ATCC 35218 | Mueller Hinton Agar | 35-37°C for 18 hrs. | > 32.0 |
| *K.pneumoniae* ATCC 700603 | Mueller Hinton Agar | 35-37°C for 18 hrs. | > 128.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.06 - 0.12 - 0.25 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 2.0 - 4.0 - 8.0 |
| *B.fragilis* ATCC 25285 | Brucella Agar with Hemin and Vitamin K1, supplemented with 5 % v/v defibrinated sterile sheep blood | 35-37°C for 24-48 hrs under strict anaerobic condition | 16.0 - 32.0 - 64.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**
Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Ampicillin Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Cefepime Ezy MIC™ Strip (CPM) (0.016-256 mcg/ml) EM070

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Cefepime on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016 mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1,3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1,3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, and streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Cefepime and other members of β-lactams class of drugs, Carbapenems, Amikacin, Vancomycin, Gentamicin, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, micro colonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Cefepime showing reading of 0.38 mcg/ml should be rounded up to next concentration i.e. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION & QUALITY CONTROL (As per CLSI Guidelines):**

**Interpretation**

**Table 1:** Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* | 35-37°C for 18 hrs. | 2 | 4-8 | 16 |
| other non *Enterobacteriaceae*, *Pseudomonas aeruginosa*, *Acinetobacter* spp, *Staphylococcus* spp. | 35-37°C for 18 hrs. | 8 | 16 | 32 |
| *Haemophilus* spp | 35-37°C for 20-24hrs at 5% $CO_2$ | 2 | - | - |
| *Streptococcus spps.* Beta haemolytic group | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | - | - |
| *Neiserria gonorrhoea* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | - | - |
| *S. pneumoniae* (meningitis) | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | 1 | 2 |
| *S. pneumoniae* (Non meningitis) *Streptococcus* spps. Viridans group | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
**Table 2:** Following are the reference MIC values (mcg/ml) range for Cefepime:

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 - 2.0 - 4.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.016 – 0.03 – 0.06 - 0.12 |
| *P.aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 - 1.0 - 2.0 - 4.0 |
| *E.coli* ATCC 35218 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.008 - 0.016 – 0.03 – 0.06 |
| *K.pneumoniae* ATCC 700603 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5 – 1.0 – 2.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.03 - 0.06 - 0.12 - 0.25 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 05- 1.0 - 2.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Second Informational Supplement. M100-S22, Vol. 32, No.3, Jan 2012.
5. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Cefepime Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Amphotericin B Ezy MIC™ Strip (AP) (0.002-32 mcg/ml) EM071

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Amphotericin B on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Amphotericin B showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar + 2% Glucose + 0.5mcg/ml Methylene Blue incubated at 35-37°C for 24 - 48 hrs.
Following are the reference MIC values (mcg/ml) range for Amphotericin B.

| **Organism** | **Std. Quality Control limits (mcg/ml)** |
|---|---|
| *C.albicans* ATCC 90028 | 0.5 - 1.0 - 2.0 |
| *C.albicans* ATCC 24433 | 0.25 – 0.5 – 1.0 |
| *C.parapsilosis* ATCC 22019 | 0.25 - 0.5 - 1.0 |
| *C. parapsilosis* ATCC 90018 | 0.5 - 1.0 - 2.0 |
| *C.tropicalis* ATCC 750 | 0.5 – 1.0 – 2.0 |
| *C. krusei* ATCC 6258 | 0.25 – 0.5 – 1.0 –2.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *in vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Amphotericin B Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Fluconazole Ezy MIC™ Strip (FLC) (0.016-256 mcg/ml) EM072

Antimicrobial Susceptibility Testing

For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Fluconazole on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) as mentioned above
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Fluconazole showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.

7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | S-DD* | ≥ R |
| *Candida albicans* | 35-37°C for 24 hrs. | 2 | 4 | 8 |
| *Candida glabrata* | 35-37°C for 24 hrs. | - | ≤ 32 | 64 |
| *Candida parapsilosis* | 35-37°C for 24 hrs. | 2 | 4 | 8 |
| *Candida tropicalis* | 35-37°C for 24 hrs. | 2 | 4 | 8 |

* S-DD - Susceptible - Dose Dependent.
#: Isolates of *C.krusei* are assumed to be intrinsically resistant to Fluconazole. The results of Fluconazole susceptibility testing should not be interpreted using this criterion for this species.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar + 2% Glucose + 0.5mcg/ml Methylene Blue incubated at 35-37°C for 24 - 48 hrs.
Following are the reference MIC values (mcg/ml) range for Fluconazole.

| Organism | Std. Quality Control limits (mcg/ml) |
|---|---|
| *C.albicans* ATCC 90028 | 0.25 -0.5 – 1.0 |
| *C.albicans* ATCC 24433 | 0.25 -0.5 – 1.0 |
| *C.tropicalis* ATCC 750 | 1.0 - 2.0- 4.0 |
| *C.parapsilosis* ATCC 90018 | 0.25 – 0.5 – 1.0 |
| *C.parapsilosis* ATCC 22019* | 1.0 – 2.0 – 4.0 – 8.0 |
| *C.krusei* ATCC 6258 | 16.0 – 32.0 – 64.0 |
| *C.albicans* ATCC 10231* | 0.5 - 1.0 - 2.0 - 4.0 - 8.0 |

* Limits may not match with CLSI guidelines

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Fluconazole Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Itraconazole Ezy MIC™ Strip (ITR) (0.002- 32 mcg/ml) EM073

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Itraconazole on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of fungal cultures to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.

2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Itraconazole showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | S-DD* | ≥ R |
| *Candida* spps.# | 35-37°C for 24 hrs. | 0.125 | 0.25-0.5 | 1 |

* S-DD - Susceptible - Dose Dependent.

#: For Itraconazole, the data are based entirely on experience with mucosal infections, and data supporting breakpoints for invasive infections due to *Candida* spps. is not available.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar + 2% Glucose + 0.5mcg/ml Methylene Blue incubated at 35-37°C for 24 hrs.
Following are the reference MIC values (mcg/ml) range for Itraconazole.

| Organism | Std. Quality Control limits (mcg/ml) |
|---|---|
| *C.parapsilosis* ATCC 22019 | 0.06 - 0.12- 0.25- 0.5 |
| *C.krusei* ATCC 6258 | 0. 12 – 0.25 - 0.5 – 1.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.

6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Itraconazole Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Meropenem Ezy MIC™ Strip (MRP) (0.002 - 32 mcg/ml) EM080

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Meropenem on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, Streptococci and for testing Staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used. For *B. fragilis*, Brucella Agar (M074) with Hemin and Vitamin K1 supplemented with 5 % v/v sterile defibrinated sheep blood is to be recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Carbapenems, Amikacin, Vancomycin, Gentamicin and members of β-lactams class of drugs, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Meropenem showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strips is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* | 35-37°C for 18 hrs. | 1 | 2 | 4 |
| *P.aeruginosa, Acinetobacter* spp | 35-37°C for 18 hrs. | 2 | 4 | 8 |
| *B.cepecia*, other non-*Enterobacteriaceae*, *Staphylococcus* spp. | 35-37°C for 18 hrs. | 4 | 8 | 16 |
| *Haemophilus* spp, *Streptococcus* spp. Beta haemolytic group, *Streptococcus* spp. Viridians group. | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.5 | - | - |
| *N.meningitidis* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | - | - |
| *S.pneumoniae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | 0.5 | 1 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Meropenem.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.03 - 0.06 - 0.12 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 2.0 - 4.0 - 8.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.008 - 0.016 - 0.032 -0.06 |
| *P.aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.12 - 0.25 - 0.5 -1 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.03- 0.06 - 0.12 - 0.25 |
| *H.influenzae* ATCC 49766 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.03 - 0.06 - 0.12 |
| *B. fragilis* ATCC 25285 | Brucella Agar with Hemin and Vitamin K1, supplemented with 5 % v/v defibrinated sterile sheep blood | 35-37°C for 24-48 hrs under strict anaerobic condition | 0.03 – 0.06 – 0.12 – 0.25 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Second Informational Supplement. M100-S22, Vol. 32, No.3, Jan 2012.
5. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Meropenem Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Ciprofloxacin Ezy MIC™ Strip (CPH) (0.016 - 256 mcg/ml) EM082

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Ciprofloxacin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016 to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

In recent past, resistant strains for antibiotics like Ciprofloxacin have emerged. The high level Ciprofloxacin Ezy MIC™ strips helps in detection of these resistant cultures.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**
Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0. 2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm yields $10^5$ - $10^6$ cells/ml).

Please refer disclaimer Overleaf

Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, Streptococci and for testing Staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For *Haemophilus* Spp, Haemophillus Test Agar Base (M1259) with added supplement (FD117) and For *Neisseria gonorrhoeae*, GC Agar Base (M434) with 1% defined growth supplement (FD025) is recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Remove necessary number of Ezy MIC™ strips on clean dry surface preferably on paper.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate pre-spread with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such Ciprofloxacin, Amikacin, Vancomycin, Gentamicin and members of β-lactams class of drugs, always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Ciprofloxacin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.

7. Before using Ezy MIC™ Strips, ensure that the strips is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**Interpretation:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* other than *S.*Typhi and extraintestinal *Salmonella* spp | 35-37°C for 18 hrs. | 0.25 | 0.5 | 1 |
| Other non-*Enterobacteriaceae, Acinetobacter* spp, *Enterococcus* spp, *Staphylococcus* spp | 35-37°C for 18 hrs. | 1 | 2 | 4 |
| *P.aeruginosa* | 35-37°C for 18 hrs. | 0.5 | 1 | 2 |
| *For S.*Typhi and extraintestinal *Salmonella* spp | 35-37°C for 18 hrs. | 0.06 | 0.12-0.5 | 1 |
| *Haemophilus* spp | 35°C ± 2°C at 5% $CO_2$ for 20-24 hours | 1 | - | - |
| *N.gonorrhoeae* | 35°C ± 2°C at 5% $CO_2$ for 20-24 hours | 0.06 | 0.12-5 | 1 |
| *N.meningitidis* | 35°C ± 2°C at 5% $CO_2$ for 20-24 hours | 0.03 | 0.06 | 0.12 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Ciprofloxacin.*

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.12- 0.25 -0.5 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.25 - 0.5 – 1.0 – 2.0 |
| *P. aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.12 - 0.25 - 0.5 -1.0 |

* Quality Control test for *E.coli* ATCC 25922, *H.influenzae* ATCC 49247 and *N. gonorrhoeae ATCC* 49226 cannot be performed using this strip as the QC limits of these cultures are below the lowest concentration of the strip.

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *in vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Ciprofloxacin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Penicillin Ezy MIC™ Strip (PEN) (0.002-32 mcg/ml) EM084

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Penicillin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

Please refer disclaimer Overleaf

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood and for *Neisseria gonorrhoeae*, GC Agar Base (M434) with 1% defined growth supplement (FD025) is recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Penicillin and other members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin, always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Penicillin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Staphylococcus* spp | 35-37°C for 18 hrs. | 0.12 | - | 0.25 |
| *Enterococcus* spp | 35-37°C for 18 hrs. | 8 | - | 16 |
| *S.pneumoniae* (Non Meningitis) | 35-37°C for 20-24hrs with 5% $CO_2$ | 2 | 4 | 8 |
| *S.pneumoniae* (Meningitis) | 35-37°C for 20-24hrs with 5% $CO_2$ | 0.06 | - | 0.12 |
| *Streptococcus* spps. Beta haemolytic group | 35-37°C for 20-24hrs with 5% $CO_2$ | 0.12 | - | - |
| *Streptococcus* spps. Viridans group | 35-37°C for 20-24hrs with 5% $CO_2$ | 0.12 | 0.25-2 | 4 |
| *N. gonorrhoeae* | 35-37°C for 20-24hrs with 5% $CO_2$ | 0.06 | 0.12-1 | 2 |
| *N. meningitidis* | 35-37°C for 20-24hrs with 5% $CO_2$ | 0.06 | 0.12-0.25 | 0.5 |
| Anaerobes | 35-37°C for 24-48hrs under anaerobic condition. | 0.5 | 1 | 2 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures recommended by CLSI on suitable medium incubated appropriately.

Following are the reference MIC values (mcg/ml) range for Penicillin

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S. aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.25-0.5-1.0-2.0 |
| *E. faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 1.0 - 2.0 - 4.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 - 0.5 - 1.0 |
| *Neisseria gonorrhoeae* ATCC 49226 | GC Agar Base (M434) with 1% defined growth supplement (FD025) | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 - 0.5 - 1.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**


1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.


**Packing:**

Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Penicillin Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Voriconazole Ezy MIC™ Strip (VRC) (0.002- 32 mcg/ml) EM086

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Voriconazole on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of fungal cultures to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Voriconazole showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | S-DD* | ≥ R |
| *Candida* parapsilosis | 35-37°C for 24-48 hrs. | 0.12 | 0.25-0.5 | 1 |
| *Candida krusei* | 35-37°C for 24-48 hrs. | 0.5 | 1 | 2 |
| *Candida albicans* | 35-37°C for 24-48 hrs. | 0.12 | 0.25-0.5 | 1 |
| *Candida tropicalis* | 35-37°C for 24-48 hrs. | 0.12 | 0.25-0.5 | 1 |

* S-DD - Susceptible - Dose Dependent.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar + 2% Glucose + 0.5mcg/ml Methylene Blue incubated at 35-37°C for 24-48 hrs.
Following are the reference MIC values (mcg/ml) range for Voriconazole.

| Organism | Std. Quality Control limits (mcg/ml) |
|---|---|
| *C.parapsilosis* ATCC 22019 | 0.016 – 0.032 - 0.064 – 0.12 |
| *C.krusei* ATCC 6258 | 0.06 – 0. 12 – 0.25 - 0.5 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Voriconazole Ezy MIC™ strips (30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Cefixime Ezy MIC™ Strip (FIX) (0.016-256 mcg/ml) EM110

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Cefixime on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.016 mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

**Please refer disclaimer Overleaf**

- **Test Procedure**

1. Prepare plates with suitable make of Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as Cefixime and members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin, Carbapenemes always read the MIC at the point of complete inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Cefixime showing reading of 0.38 mcg/ml should be rounded up to next concentration ie. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae* | 35-37°C for 18 hrs. | 1 | 2 | 4 |
| *Haemophilus* spp. | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | - | - |
| *N.gonorrhoeae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | - | - |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Cefixime.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 8.0 - 16.0 - 32.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.25 - 0.5 - 1.0 |
| *H. influenzae* ATCC 49766 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.12 - 0.25 - 0.5 - 1.0 |
| *N.gonorrhoeae* ATCC 49226 | GC Agar Base (M434) with 1% defined growth supplement (FD025) | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.004 - 0.008 - 0.016 – 0.03 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**

Each Pack contains following material packed in sealed glass vial with a desiccator capsule.

1) Cefixime Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Caspofungin Ezy MIC™ Strip (CAS) (0.002-32 mcg/ml) EM119

Antifungal Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Caspofungin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of fungi to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, 2% glucose with methylene blue (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow it to solidify.
**Note : Surface of the medium should be dry when inoculated.**

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, 2% glucose with methylene blue (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
   **Note: Once the plate has been swabbed with the test culture, the plate should be dry for at least 1 to 1½ hour before placing the Ezy MIC Strip.**
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. Ensure that strip is completely adsorbed to the agar surface, Press if required.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Caspofungin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I* | ≥ R |
| *Candida albicans* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida glabrata* | 35-37°C for 24 hrs. | 0.12 | 0.25 | 0.5 |
| *Candida tropicalis* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida krusei* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida parapsilosis* | 35-37°C for 24-48 hrs. | 2.0 | 4.0 | 8.0 |
| *Candida guilliermondii* | 35-37°C for 24 hrs. | 2.0 | 4.0 | 8.0 |

*: the ability to successfully treat infections with isolates for which the MIC results are in the intermediate category. The available data do not permit the MIC results to be clearly categorized as either "susceptible" or "resistant"
.

**QUALITY CONTROL:**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar, 2% Glucose with 0.5 mcg/ml Methylene Blue incubated at 35-37°C for 24 - 48 hrs.
Following are the reference MIC values (mcg/ml) range for Caspofungin.

| Organism | Std. Quality Control limits (mcg/ml) |
|---|---|
| *C.parapsilosis* ATCC 22019 | 0.25 – 0.5 – 1.0 |
| *C. krusei* ATCC 6258 | 0.12 – 0.25 – 0.5 – 1.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Caspofungin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Posaconazole Ezy MIC™ Strip (POS) (0.002-32 mcg/ml) EM120

Antifungal Susceptibility Testing

For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Posaconazole on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of yeast and fungi to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, 2% glucose with methylene blue (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow it to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, 2% glucose with methylene blue (M1825) as mentioned above.

Please refer disclaimer Overleaf

2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Posaconazole showing reading of 0.38 mcg/ml should be rounded up to next concentration ie. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar, 2% Glucose with mcg/ml Methylene Blue incubated at 35-37°C for 24 - 48 hrs.
Following are the reference MIC values (mcg/ml) range for Posaconazole.

| Organism | Std. Quality Control limits (mcg/ml) |
|---|---|
| *C.parapsilosis* ATCC 22019 | 0.03 – 0.06 – 0.12 – 0.25 |
| *C. krusei* ATCC 6258 | 0.06 – 0.12 – 0.25 – 0.5 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Posaconazole Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Micafungin Ezy MIC™ Strip (MYC) (0.002-32 mcg/ml) EM121

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Micafungin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of yeast and fungi to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, 2% glucose with methylene blue (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

Please refer disclaimer Overleaf

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, 2% glucose with methylene blue (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Micafungin showing reading of 0.38 mcg/ml should be rounded up to next concentration ie. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I* | ≥ R |
| *Candida albicans* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida glabrata* | 35-37°C for 24 hrs. | 0.06 | 0.12 | 0.25 |
| *Candida tropicalis* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida krusei* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida parapsilosis* | 35-37°C for 24-48 hrs. | 2.0 | 4.0 | 8.0 |
| *Candida guilliermondii* | 35-37°C for 24 hrs. | 2.0 | 4.0 | 8.0 |

*: the ability to successfully treat infections with isolates for which the MIC results are in the intermediate category. The available data do not permit the MIC results to be clearly categorized as either "susceptible" or "resistant"

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures recommended by CLSI on suitable medium incubated appropriately.

Following are the reference MIC values (mcg/ml) range for Micafungin.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *C. parapsilosis* ATCC 22019 | Mueller Hinton Agar, 2% Glucose with 0.5 mg/ml Methylene Blue | 35-37°C for 24-48 hrs. | 0.5 – 1.0 – 2.0 – 4.0 |
| *C. krusei* ATCC 6258 | Mueller Hinton Agar, 2% Glucose with 0.5 mg/ml Methylene Blue | 35-37°C for 24 hrs. | 0.12 – 0.25 – 0.5 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Micafungin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Anidulafungin Ezy MIC™ Strip (AND) (0.002-32 mcg/ml) EM122

Antifungal Susceptibility Testing

For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Anidulafungin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of yeast and fungi to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, 2% glucose with methylene blue (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow it to solidify.

**Note: Surface of the medium should be dry when inoculated.**

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, 2% glucose with methylene blue (M1825) as mentioned above.

Please refer disclaimer Overleaf

2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Anidulafungin showing reading of 0.38 mcg/ml should be rounded up to next concentration ie. 0.5 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strips are at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**INTERPRETATION:**

Use following interpretive criteria for susceptibility categorization.

| When testing | Incubation | Interpretive Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I* | ≥ R |
| *Candida albicans* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida glabrata* | 35-37°C for 24 hrs. | 0.12 | 0.25 | 0.5 |
| *Candida tropicalis* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida krusei* | 35-37°C for 24 hrs. | 0.25 | 0.5 | 1.0 |
| *Candida parapsilosis* | 35-37°C for 24-48 hrs. | 2.0 | 4.0 | 8.0 |
| *Candida guilliermondii* | 35-37°C for 24 hrs. | 2.0 | 4.0 | 8.0 |

*: the ability to successfully treat infections with isolates for which the MIC results are in the intermediate category. The available data do not permit the MIC results to be clearly categorized as either "susceptible" or "resistant"

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures on Mueller Hinton Agar, 2% Glucose with 0.5 mcg/ml Methylene Blue incubated at 35-37°C for 24 - 48 hrs.
Following are the reference MIC values (mcg/ml) range for Anidulafungin.

| Organism | Std. Quality Control limits (mcg/ml) |
|---|---|
| *C.parapsilosis* ATCC 22019 | 0.25 – 0.5 – 1 – 2 |
| *C. krusei* ATCC 6258 | 0.03 – 0.06 – 0.09 – 0.12 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Anidulafungin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Nystatin Ezy MIC™ Strip (AP) (0.002-32 mcg/ml) EM145

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Nystatin on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

Please refer disclaimer Overleaf

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Nystatin showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures. Following are the reference MIC values (mcg/ml) range for Nystatin.

| Organism | Medium Used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *C.parapsilosis* ATCC 22019 | Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) Or Mueller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue | 35-37°C for 24 - 48 hrs. | 0.25 – 0.5 – 1.0 |
| *C.tropicalis* ATCC 750 | | | 0.125 – 0.25 – 0.5 – 1.0 |
| *C. krusei* ATCC 6258 | | | 0.5 – 1.0 – 2.0 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *in vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.
7. Arikan, S., Ostrosky-Zeichner, L., Lozano-Chiu, Mario., Paetznick, V., Gordon, D., Wallace, T., Rex, J.H. (2002). In Vitro Activity of Nystatin Compared with Those of Liposomal Nystatin, Amphotericin B, and Fluconazole against Clinical *Candida* Isolates. *J.Clin.Microbiol*. Apr. 2002, p. 1406–1412.

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Nystatin Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

# Miconazole Ezy MIC™ Strip (MIC) (0.002-32 mcg/ml) EM146

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Miconazole on a single paper strip in a concentration gradient manner, capable of showing MICs in the range of 0.002 mcg/ml to 32 mcg/ml, on testing against the test organism.

**Introduction:**
Ezy MIC™ strip is useful for quantitative determination of susceptibility of yeast and fungi to antifungal agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antifungal agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar or Mueller Hinton Agar, 2% glucose with methylene blue is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.
7. **The strips give reproducible MIC values that are equivalent to the standard reference MIC obtained by Broth dilution performed as per guidelines with less efforts.**

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) from dehydrated powder according to the directions specified on the label. Alternately, prepare Mueller Hinton Agar with added 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (this could be added pre or post sterilization). Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

Please refer disclaimer Overleaf

- **Test Procedure**

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye i.e. Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) as mentioned above.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate swabbed with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniform and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation
4. Isolated colonies, pinpoint microcolonies and hazes may appear within the zone of inhibition frequently and they should be ignored. In such cases, consider reading for MIC determination at a point on the scale at which prominent reduction of growth is seen.
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Miconazole showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC cultures.
Following are the reference MIC values (mcg/ml) range for Miconazole*.

| Organism | Medium Used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *C. albican* ATCC 24433 | Mueller Hinton Agar, Modified (as per CLSI for antifungal) (M1825) Or Mueller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue | 35-37°C for 24 - 48 hrs. | 0.06 – 0.125 – 0.25 |
| *C.tropicalis* ATCC 750 | | | 0.125 – 0.25 – 0.5 – 1.0 |
| *C. glabrata* ATCC 2001 | | | 0.125 – 0.25 – 0.5 |

*Not as per CLSI guidelines

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strip container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
5. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Third Edition. Vol.28 No.14, April- 2008 CLSI document M27-S3.
6. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts; Fourth Informational Supplement. Vol.32 No.17, December 2012 CLSI document M27-S4.
7. Microbiology Review, Division of Special Pathogen and Transplant Products. Center for Drug Evaluation Research. NDA 22-404 (SDN 9 – Original).

**Packing:**

Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.

1) Miconazole Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert

## Fluconazole FLC 25 mcg SD232

Fluconazole FLC 25mcg discs are used for antimicrobial susceptibility testing of fungal cultures

## Composition

| *Ingredients | Concentration |
|---|---|
| Fluconazole | 25mcg/disc |

## Susceptibility Test Procedure:

Preparation of Inoculum:

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

Test Procedure:

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (M1825) for carrying out susceptibility of antifungal discs. The medium in the plates should be sterile and have a depth of about 4 mm.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Apply the discs using aseptic technique. Deposit the discs with centers at least 24 mm apart.
4. Invert the plates and place in an incubator set to 35 ± 2°C within 15 minutes after the discs are applied.
5. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniformly circular and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation.

## Principle:

Antimicrobial susceptibility testing (AST) of bacterial and fungal isolates is a common and important technique in most clinical laboratories. The results of these tests are used for selection of the most appropriate antimicrobial agent(s) for treatment against the infectious organisms. Till the 1950s, laboratories were lacking in the methodologies and equipments for the accurate determination of in vitro responses of organisms to antimicrobial agents. Bauer et al (1) began the development of standardized methods for antimicrobial susceptibility testing, using disc diffusion system. However the susceptibility results may not always correlate with the patient's response to therapy. The response of an infected patient to antimicrobial agent(s) is a complex interrelationship of host responses, drug dynamics and microbial activity. Antimicrobial susceptibility tests are either quantitative or qualitative. Disc diffusion test is a qualitative test method. The National Committee for Clinical Laboratory Standards (NCCLS), now known as Clinical Laboratory Standards Institute (CLSI) has published comprehensive documents regarding the disc diffusion systems. The agar disc diffusion test is the most convenient and widely used method for routine antimicrobial susceptibility testing. In subsequent and current practice, antimicrobial impregnated paper discs are applied onto the agar surface. Based on the Bauer-Kirby Method, standardized reference procedures for the disc systems were published by WHO and FDA and are periodically updated by the CLSI (formerly NCCLS)(2). For any antimicrobial testing, Quality control or clinical testing, the method to be followed is the same as mentioned above.

However few precautions are to be maintained while handling of the Sensitivity discs,

- On receipt the discs are to be immediately stored at the recommended temperature.
- Medium preparation, Inoculum preparation and incubation to be done as specified.

## Interpretation:

Use following interpretive criteria for susceptibility categorization.

| Antimicrobial agent | Interpretative criteria for | Resistant mm or less | S-DD# mm | S mm or more |
|---|---|---|---|---|
| Fluconazole 25mcg | *Candida* spps | 14 | 15-18 | 19 |

# S-DD - Susceptible - Dose dependent

## Quality Control:

**Appearance:** Filter paper discs of 6mm diameter with printed "FLC 25" on centre of each side of the disc.

**Cultural response**: Average diameter of zone of inhibition observed on Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye after 24-48 hours incubation at 35-37°C for standard cultures.

| Organisms (ATCC) | Std. zone of diameter (mm) |
|---|---|
| *C.albicans* (90028)* | 28-39 |
| *C.parapsilosis* (22019)* | 22-33 |
| *C.tropicalis* (750)* | 26-37 |
| *C.albicans*(10231) | 25-30 |

* = Q.C. Strains recommended by CLSI

## Storage and Shelf-life:

Discs should always be stored at -20°C to +8°C under dry conditions, along with the dessicator provided in individual pack. Use before expiry date on the label.

## References:

1. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
2. Zone Diameter Interpretive Standards, Corresponding Minimal Inhibitory Concentration (MIC) Interpretive Breakpoints, and Quality Control Limits for Antifungal Disk Diffusion Susceptibility Testing Of Yeasts, Third International Supplement CLSI document - M44-S3.

*** Not for Medicinal Use**

Revision : 1 / 2012

CE

## Voriconazole VRC 1 mcg SD277

Voriconazole VRC 1 mcg discs are used for antimicrobial susceptibility testing of fungal cultures

## Composition

| *Ingredients | Concentration |
|---|---|
| Voriconazole | 1mcg/disc |

## Susceptibility Test Procedure:

Preparation of Inoculum:

1. Inoculum is prepared by picking five distinct colonies of approximately 1mm from 24 hours old culture grown on Sabouraud Dextrose Agar (M063) and incubated at 35 ± 2°C. Colonies are suspended in 5ml of sterile 0.85% Saline.
2. Vortex the resulting suspension and adjust the turbidity to yield 1 x $10^6$ - 5 x $10^6$ cells /ml (i.e. 0.5 McFarland standard).

Test Procedure:

1. Prepare plates with Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye (M1825) for carrying out susceptibility of antifungal discs. The medium in the plates should be sterile and have a depth of about 4 mm.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum (turbidity so adjusted, as to obtain semi confluent growth on the petri plate) and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
3. Apply the discs using aseptic technique. Deposit the discs with centers at least 24 mm apart.
4. Invert the plates and place in an incubator set to 35 ± 2°C within 15 minutes after the discs are applied.
5. Examine each plate after 20 - 24 hours of incubation. If plate was satisfactorily streaked the resulting zones of inhibition will be uniformly circular and there will be a semi-confluent lawn of growth. Read at 48 hours only when insufficient growth is observed after 24 hours incubation.

## Principle:

Antimicrobial susceptibility testing (AST) of bacterial and fungal isolates is a common and important technique in most clinical laboratories. The results of these tests are used for selection of the most appropriate antimicrobial agent(s) for treatment against the infectious organisms. Till the 1950s, laboratories were lacking in the methodologies and equipments for the accurate determination of in vitro responses of organisms to antimicrobial agents. Bauer et al (1) began the development of standardized methods for antimicrobial susceptibility testing, using disc diffusion system. However the susceptibility results may not always correlate with the patient's response to therapy. The response of an infected patient to antimicrobial agent(s) is a complex interrelationship of host responses, drug dynamics and microbial activity. Antimicrobial susceptibility tests are either quantitative or qualitative. Disc diffusion test is a qualitative test method. The National Committee for Clinical Laboratory Standards (NCCLS), now known as Clinical Laboratory Standards Institute (CLSI) has published comprehensive documents regarding the disc diffusion systems. The agar disc diffusion test is the most convenient and widely used method for routine antimicrobial susceptibility testing. In subsequent and current practice, antimicrobial impregnated paper discs are applied onto the agar surface. Based on the Bauer-Kirby Method, standardized reference procedures for the disc systems were published by WHO and FDA and are periodically updated by the CLSI (formerly NCCLS)(2). For any antimicrobial testing, Quality control or clinical testing, the method to be followed is the same as mentioned above.

However few precautions are to be maintained while handling of the Sensitivity discs,

- On receipt the discs are to be immediately stored at the recommended temperature.
- Medium preparation, Inoculum preparation and incubation to be done as specified.

## Interpretation:

Use following interpretive criteria for susceptibility categorization.

| Antimicrobial agent | Interpretative criteria for | Resistant mm or less | S-DD# mm | S mm or more |
|---|---|---|---|---|
| Voriconazole 1 mcg | *Candida* spps | 13 | 14-16 | 17 |

# S-DD - Susceptible - Dose dependent

## Quality Control:

**Appearance:** Filter paper discs of 6mm diameter with printed "VRC 1" on centre of each side of the disc.

**Cultural response**: Average diameter of zone of inhibition observed on Muller Hinton Agar + 2% Glucose + 0.5 mcg/ml Methylene Blue Dye after 24-48 hours incubation at 35-37°C for standard cultures.

| Organisms (ATCC) | Std. zone of diameter (mm) |
|---|---|
| *C.albicans* (90028)* | 31-42 |
| *C.parapsilosis* (22019)* | 28-37 |
| *C.krusei* (6528)* | 16-25 |
| *C.albicans*(10231) | 30-40 |
| *S.cerevesiae* (9763) | 29-38 |

* = Q.C. Strains recommended by CLSI

## Storage and Shelf-life:

Discs should always be stored at -20°C to +8°C under dry conditions, along with the dessicator provided in individual pack. Use before expiry date on the label.

## References:

1. Method for Antifungal Disk Diffusion Susceptibility Testing of Yeasts; Approved Guidelines-Second edition Vol.29 No.17, August- 2009 CLSI document M44-A2. For more details refer to this volume
2. Zone Diameter Interpretive Standards, Corresponding Minimal Inhibitory Concentration (MIC) Interpretive Breakpoints, and Quality Control Limits for Antifungal Disk Diffusion Susceptibility Testing Of Yeasts, Third International Supplement CLSI document - M44-S3.

*** Not for Medicinal Use**

Revision : 1 / 2012

CE

# Tetracycline Ezy MIC™ Strip (TET) (0.016-256 mcg/ml) EM056

Antimicrobial Susceptibility Testing
For *In Vitro* Diagnostic use Not for Medicinal Use

It is a unique MIC determination paper strip which is coated with Tetracycline in a concentration gradient manner, capable of showing MICs in the range of 0.016mcg/ml to 256 mcg/ml, on testing against the test organism.

**Introduction**

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

**Ezy MIC™ Strip FEATURES AND ADVANTAGES**

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material.
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

**METHOD AND USE OF EZY MIC™ STRIPS**

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow to solidify. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm).
Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland .This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with Mueller Hinton Agar for rapidly growing aerobic organisms as mentioned above. For fastidious organisms such as Streptococci, Mueller Hinton Agar is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) is to be used. For *Neisseria gonorrhoeae*, GC Agar Base (M434) with 1% defined growth supplement (FD025) is recommended. For *B. fragilis*, Brucella Agar (M074) with Hemin and Vitamin K1 supplemented with 5 % v/v sterile defibrinated sheep blood is to be recommended.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove Ezy MIC™ strip container from cold and keep it at room temperature for 15 minutes before opening.
4. Remove one applicator from the self sealing bag stored at room temperature.
5. Hold the applicator in the middle and gently press its broader sticky side on the centre of Ezy MIC™ strip.
6. Lift the applicator along with attached Ezy MIC™ strip.
7. Place the strip at a desired position on agar plate pre-spread with test culture. Gently turn the applicator clockwise with fingers. With this action, the applicator will detach from the strip.
8. DO NOT PRESS EZY MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
9. Ezy MIC™ strip should not be repositioned or adjusted once placed.
10. Transfer plates in the incubator under appropriate conditions.

**MIC Reading:**

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bacteriostatic drugs such Tetracycline, Chloramphenicol, Tetracycline, Azithromycin, Fluconazole and Trimethoprim/ Sulphamethoxazole, read MICs at 80% inhibition for homogenously sensitive strains such as QC control strains.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for MIC determination at a point on the scale above which no resistant colonies are observed close to MIC strip (within 1-3 mm distance from the strip).
5. Since Ezy MIC™ strip has continuous gradient, MIC values “in-between” two fold dilutions can be obtained.
6. Always round up these values to the next two-fold dilution before categorization. For example: Tetracycline showing reading of 0.75 mcg/ml should be rounded up to next concentration ie. 1.0 mcg/ml.
7. If the ellipse intersects the strip in between 2 dilutions, read the MIC as the value which is nearest to the intersection.
8. When growth occurs along the entire strip, report the MIC as $\geq$ the highest values on the MIC strip. When the inhibition ellipse is below the strip (does not intersect the strip), report the MIC < the lowest value on the MIC scale.

**Warning and Precautions:**

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strip is at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

**Interpretation**

Use following interpretive criteria for susceptibility categorization as per CLSI.

| When testing | Incubation | Interpretative Criteria | | |
|---|---|---|---|---|
| | | ≤ S | I | ≥ R |
| *Enterobacteriaceae*, other non-*Enterobacteriaceae*, *Staphylococcus* spp, *Acinetobacter* spp, *Enterococcus* spp | 35-37°C for 18 hrs. | 4 | 8 | 16 |
| *Haemophilus* spp, *Streptococcus* spp. Beta haemolytic group, *Streptococcus* spp. Viridans group | 35-37°C for 20-24hrs at 5% $CO_2$ | 2 | 4 | 8 |
| *S.pneumoniae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 1 | 2 | 4 |
| *N.gonorrhoeae* | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.25 | 0.5-1 | 2 |
| Anaerobes | 35-37°C for 20-24hrs under anaerobic condition | 4 | 8 | 16 |

**QUALITY CONTROL**

Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.
Following are the reference MIC values (mcg/ml) range for Tetracycline.

| Organism | Medium used | Incubation | Std. Quality Control limits (mcg/ml) |
|---|---|---|---|
| *S.aureus* ATCC 29213 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.12-0.25-0.5-1.0 |
| *E.faecalis* ATCC 29212 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 8.0-16.0-32.0 |
| *E.coli* ATCC 25922 | Mueller Hinton Agar | 35-37°C for 18 hrs. | 0.5-1.0-2.0 |
| *P. aeruginosa* ATCC 27853 | Mueller Hinton Agar | 35-37°C for 18 hrs | 8.0-16.0-32.0 |
| *S. pneumoniae* ATCC 49619 | Mueller Hinton Agar w/ 5% Sheep Blood | 35-37°C for 20-24hrs at 5% $CO_2$ | 0.06 - 0.12-0.25-0.5 |
| *H.influenzae* ATCC 49247 | Haemophilus Test Medium | 35-37°C for 20-24hrs at 5% $CO_2$ | 4.0-8.0-16.0-32.0 |
| *N. gonorrhoeae* ATCC 49226 | GC Agar Base (M434) with 1% defined growth supplement (FD025) | 35-37°C for 24 – 48 hrs at 5% $CO_2$ | 0.25-0.5-1 |
| *B.fragilis* ATCC 25285 | Brucella Agar with Hemin and Vitamin K1, supplemented with 5 % v/v defibrinated sterile sheep blood | 35-37°C for 24-48 hrs under strict anaerobic condition | 0.125-0.25-0.5 |

**Storage & Shelf Life:**

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at 2-8°C, for prolonged use store below -20°C.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

**Disposal:**
After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

**Limitation of Test**

Ezy MIC™ Strips provides *In vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

**References:**

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen,J.H., Pfaller , M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance standards of Antimicrobial Susceptibility Testing; Twenty Ninth Informational Supplement. M100-S29, Vol. 39, No.1, Jan 2019.

**Packing:**
Each Pack contains following material packed in air-tight plastic container with a desiccator capsule.
1) Tetracycline Ezy MIC™ strips (10/30/60/90/120/150 Strips per pack)
2) Applicator sticks
3) Package insert