

Technical Data

Fraser Broth Base M1327

Intended use

Recommended, recommended as a primary as well as secondary enrichment medium, for the isolation and enumeration of *Listeria monocytogenes* from food and animal feeds. The composition and performance criteria of this media is as per the specification laid down in ISO 11290-1:2017 and ISO 11290-2:2017.

Composition**

ISO 11290 Specification - Half Fraser & Fraser		Fraser Broth: Half Fraser & Fraser broth			
Ingredients	Gms / Litre	Ingredients	Gms / Litre		
Enzymatic digest of animal tissues	5.000	Peptone #	5.000		
Enzymatic digest of casein	5.000	Tryptone \$	5.000		
Yeast extract	5.000	Yeast extract	5.000		
Meat extract	5.000	HM extract ##	5.000		
Sodium chloride	20.000	Sodium chloride	20.000		
Disodium hydrogen phosphate dihydrate	12.000	Disodium hydrogen phosphate dihydrate	12.000		
Potassium dihydrogen phosphate	1.350	Potassium dihydrogen phosphate	1.350		
Esculin	1.000	Esculin	1.000		
Lithium chloride	3.000	Lithium chloride	3.000		
Final pH (at 25°C)	7.2 ± 0.2	Final pH (at 25°C)	7.2 ± 0.2		

Supplements to be added after autoclaving

	Half fraser	Fraser		Half fraser	Fraser
	Gms / Litre	Gms / Litre		Gms / Litre	Gms / Litre
			<u>FD125I</u>	1 vial	2 vials
Acriflavin hydrochloride	0.0125	0.025	Acriflavin hydrochloride	0.0125	0.025
Nalidixic acid, sodium salt	0.01	0.02	Nalidixic acid, sodium salt	0.01	0.02
			FD141	2 vials	2 vials
Ammonium Iron citrate	0.5	0.50	Ammonium Iron citrate	0.5	0.50

^{**}Formula adjusted, standardized to suit performance parameters

Directions

Suspend 54.92 grams (the equivalent weight of dehydrated medium per litre) in 1000 ml purified / distilled water. Heat if necessary to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C and aseptically add rehydrated contents of 1 vial of Fraser Selective Supplement (FD125I) and 2 vials of Fraser Supplement (FD141) to 1000 ml medium for primary enrichment or 1 vial of each to 500 ml medium for secondary enrichment. Mix well and dispense in tubes or flasks as desired.

Principle And Interpretation

L.monocytogenes primarily causes meningitis, encephalitis or septicemia in humans (1,10). In pregnant women, L.monocytogenes often causes influenza like bacteremic illness that, if untreated, may leaded to ammionitis and infection of the fetus, resulting in abortion, still birth or premature birth. Contaminated foods are the primary vehicles of transmission (9). Fraser Broth Base is based on the formulation of Fraser and Sperber (11) is used for the detection of Listeria species in food products. Fraser Broth Base is formulated so as to provide optimum conditions for the growth of Listeria. This medium is recommended by ISO for primary and secondary enrichment of Listeria species.

^{# -} Equivalent to Enzymatic digest of animal tissues

^{\$ -} Equivalent to Enzymatic digest of casein

^{## -} Equivalent to Meat extract

Peptone, Tryptone, yeast extract, and HM extract make the media highly nutritive by providing essential nutrients including carbonaceous and nitrogenous substances. Phosphates maintain the buffering capacity of the medium. All Listeria species exhibit beta-glucosidase activity which is evident by the blackening of the media. *Listeria* species hydrolyze esculin (substituted glucoside) to glucose and esculetin. The latter combines with ferric ions of ferric ammonium citrate (FD141), resulting in the formation of 6-7 dihydroxycoumarin, a black brown complex. Ferric ammonium citrate also enhances the growth of *L.monocytogenes* (8). The high salt tolerance (of sodium chloride) of *Listeria* is used as means to inhibit the growth of Enterococci. Lithium chloride is also used to inhibit Enterococci, which also possess the ability to hydrolyze esculin. Growth of accompanying bacteria is largely inhibited by the addition of Nalidixic acid and Acriflavin hydrochloride (FD1251).

Type of specimen:

Food samples

Specimen Collection and Handling:

1. Initial suspension

This broth is used as an dilution fluid for the preparation of initial suspension

25grams/25 ml of sample to 225 ml of the medium (M1327 + 1 vial of FD125I + 2 vials of FD141)

2. Primary enrichment

The dilution prepared in Half Fraser broth is incubated at $30^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 24-26 hours.

The preenriched sample after incubation can be stored at 5°C for a maximum of 72 hours before transfer to Fraser Broth (secondary enrichment)

A black colouration can develop during incubation.

3. Secondary Enrichment

0.1 ml of culture from primary enrichment is added to 10 ml of Fraser Broth (secondary enrichment). It is incubated at $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 24 ± 2 hours.

Additional incubation of 24 hours for *Listeria* species other than *L.monocytogenes* is recommended to allow recovery of more species.

The sample from primary enrichment and secondary enrichment is then subcultured on L.mono Differential Agar Base (M1540) and on Listeria Oxford Medium Base (M1145) or Listeria Identification Agar Base (PALCAM) (M1064I). Incubate at 37 ± 1 °C for 24 ± 2 hours. Additional incubation at 37 ± 1 °C for 24 ± 2 hours is recommended for *Listeria* spp. other than *L.monocytogenes* for recovery of more species. (6,7)

Warning and Precautions:

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidleines should be followed while handling specimens. Saftey guidelines may be referred in individual safety data sheets

Limitations:

- 1. Presence of L.monocytogenes is often masked by other Listeria species like L.inocua and L.ivanovii.
- 2. Further subculture of organisms on selective media is required.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Cream to yellow homogeneous free flowing powder

Colour and Clarity of prepared medium

Basal medium: Yellow coloured clear solution with slight precipitate. After addition: Fluorescent yellow coloured clear solution with slight precipitate forms in tubes.

Reaction

Reaction of 5.49% w/v aqueous solution at 25°C. pH: 7.2±0.2

pН

7.00-7.40

Cultural l	Response
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Half Fraser (Primary Enrichment)

Organism	Inoculum	Growth	Esculin	Recovery	Colour of colony
	(CFU)		Hydrolysis	on M1540*	on M1540*

Productivity

Cultural characteristics observed on addition of FD125I and FD141 after an incubation at 30 ± 1 °C for 25 ± 1 hour. Further subculture is carried out on M1540 at 37 ± 1 °C for 48 ± 4 hours.

Listeria monocytogenes 1/2a ATCC 35152 (00109*) + Escherichia coli ATCC 25922 (00013*) + Enterococcus faecalis ATCC 29212 (00087*)	$50-100$ >= 10^4 >= 10^4	good-luxurian	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
Listeria monocytogenes 1/2a ATCC 35152 (00109*) + Escherichia coli ATCC 8739 (00012*) + Enterococcus faecalis ATCC 19433 (00009*)	$50-100$ $>=10^4$ $>=10^4$	good-luxuriant	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
Listeria monocytogenes 4b ATCC 13932 (00021*) + Escherichia coli ATCC 25922 (00013*) + Enterococcus faecalis ATCC 29212 (00087*)	$50-100$ $>=10^4$ $>=10^4$	good-luxuriant	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
Listeria monocytogenes 4b ATCC 13932 (00021*)+ Escherichia coli ATCC 8739 (00012*)+ Enterococcus faecalis ATCC 19433 (00009*)	$50-100$ $>=10^4$ $>=10^4$	good-luxuriant	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo

Selectivity

Cultural characteristics observed on addition of FD125I and FD141 after an incubation at $30 \pm 1^{\circ}$ C for 25 ± 1 hour. Further subculture is carried on Tryptone Soya Agar (M290) after an incubation at $37 \pm 1^{\circ}$ C for 48 ± 4 hours.

Escherichia coli ATCC 25922 (00013*)	>=104	inhibited	-	0
Escherichia coli ATCC 8739 (00012*)	>=104	inhibited	-	0
Enterococcus faecalis ATCC 29212 (00087*)	$C >= 10^4$	none-poor	-	<100 colonies
Enterococcus faecalis ATCO 19433 (00009*)	$C >= 10^4$	none-poor	-	<100 colonies

Fraser (Secondary Enrichment)

Organism	Inoculum	Growth	Esculin	Recovery	Colour of colony
	(CFU)		Hydrolysis	on M1540*	on M1540*

Productivity

Cultural characteristics observed on addition of FD125I and FD141 after an incubation at 37 ± 1 °C for 24 ± 2 hours. Further subculture is carried out on M1540 at 37 ± 1 °C for 48 ± 4 hours.

Listeria monocytogenes 1/2a ATCC 35152 (00109*) + Escherichia coli ATCC 25922 (00013*) + Enterococcus faecalis	50-100 >=10 ⁴	good-luxuriant	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
ATCC 29212 (00087*)	>=104				
Listeria monocytogenes 1/2a ATCC 35152 (00109*) +	50-100	good-luxuriant	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
Escherichia coli ATCC 8739 (00012*) +	>=104		medium		opaque naio
Enterococcus faecalis ATCC 19433 (00009*)	>=104				
Listeria monocytogenes 4b ATCC 13932 (00021*) +	50-100		positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
<i>Escherichia coli</i> ATCC 25922 (00013*) +	>=104				11
Enterococcus faecalis ATCC 29212 (00087*)	>=104				
Listeria monocytogenes 4b ATCC 13932 (00021*) +	50-100	good-luxuriant	positive reaction, blackening of medium	>10 colonies	Blue green colonies w/ opaque halo
Escherichia coli ATCC 8739 (00012*) +	>=104		medium		opaque naio
Enterococcus faecalis ATCC 19433 (00009*)	>=104				

Selectivity

Cultural characteristics observed on addition of FD125I and FD141 after an incubation at $30 \pm 1^{\circ}$ C for 25 ± 1 hour. Further subculture is carried on Tryptone Soya Agar (M290) after an incubation at $37 \pm 1^{\circ}$ C for 48 ± 4 hours.

Escherichia coli ATCC >=10 ⁴ 25922 (00013*)	inhibited	-	0
Escherichia coli ATCC >=10 ⁴ 8739 (00012*)	inhibited	-	0
Enterococcus faecalis ATCC>=10 ⁴ 29212 (00087*)	none-poor	-	<100 colonies
Enterococcus faecalis ATCC>=10 ⁴ 19433 (00009*)	none-poor	-	<100 colonies

Storage and Shelf Life

Store between $10\text{-}30^{\circ}\text{C}$ in a tightly closed container . Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle inorder to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Use before expiry date on the label. Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (4,5).

Reference

- 1. American Public Health Association, Standard Methods for the Examination of Dairy Products, 1978, 14th Ed., Washington D.C.
- 2. Cowart R. E. and Foster B. G., 1985, J. Infect. Dis.; 151:17
- 3. Fraser and Sperber, 1988, J. Food Prot., 51:762-76
- 4. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
- 5. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
- 6.Microbiology of the food chain Horizontal method for the detection and enumeration of *Listeria monocytogenes* and of *Listeria* spp. Part 1, Detection method; ISO 11290-1:2017
- 7. Microbiology of the food chain Horizontal method for the detection and enumeration of *Listeria monocytogenes* and of *Listeria* spp. Part 2, Detection method; ISO 11290-2:2017
- 8. Murray P. R., Baron E. J., Jorgensen J. H., Pfaller M. A., Yolken R. H., (Eds.), 8th Ed., 2003, Manual of Clinical Microbiology, ASM, Washington, D.C.
- 9. Nieman R. E., and Lorber B., 1980, Rev. Infect. Dis. 2: 207-2
- 10.Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
- 11 Schuchat A. B., Swaminathan and C. V. Broome, Clin. Microbiol., Rev. 4: 169-1
- 12. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.

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