

4. Following the final wash, completely decant the supernatant saline to ensure removal of all residual saline and a resultant “dry” red cell button.
5. Add two drops of NOVACLONE™ Anti-IgG to each tube containing a ‘dry’ button of washed red cells.
6. Mix tube gently, but thoroughly, to resuspend the red cells.
7. Centrifuge for:

a) 15 seconds at 900-1000 rcf.

b) or centrifugation of equivalent force†.
8. Gently resuspend the red cell button and examine for agglutination.
9. Grade and record results. *(A microscope or other optical aid, may be used to confirm weak or negative hemagglutination reactions).*
10. Negative or weak positive antiglobulin test results should be appropriately controlled by the addition of IgG sensitized reagent control cells (See CONTROLS).

INDIRECT ANTIGLOBULIN TEST PROCEDURE
For Antibody Detection, Identification or Compatibility Testing

NOTE: The following test method is recommended only as a guide. Modifications to the test procedure following accepted and well documented immunohematology practices (such as an increase in serum:cell ratio and/or incubation time) may be desirable to comply with the requirements of individual laboratories. However, the application of low ionic strength antibody enhancement or potentiating reagents requires strict adherence to the respective manufacturer's Directions for Use. Red cell phenotyping with specific Blood Grouping Reagents should be performed in accordance with the manufacturer's recommended Directions for Use. User-defined modifications to test procedures may require validation.

The following is one example of a commonly used test protocol that may be used for antibody detection, antibody identification or compatibility testing:

1. Appropriately label one test tube for each donor cell, Screening Cell or Panel Cell to be tested.
2. Dispense at least two drops of serum into each tube‡.
3. Add one drop of 2-5% donor cell or reagent red cell (Screening Cell or Panel Cell) suspension to the appropriate test tube.
4. Mix tube contents thoroughly.
5. Centrifuge for:

a) 15 seconds at 900-1000 rcf.

b) or centrifugation of equivalent force†.
6. Examine supernatant for visible hemolysis.
7. Gently resuspend the red cell button and examine macroscopically for agglutination. Grade and record results.
8. Mix tube contents again and incubate test tubes at 37°C (±1°C) for 15-60 minutes.†
9. Centrifuge for:

a) 15 seconds at 900-1000 rcf.

b) or centrifugation of equivalent force‡.
10. Examine supernatant for visible hemolysis.
11. Gently resuspend the red cell button and examine macroscopically for agglutination. Grade and record results.
12. Gently, but thoroughly, mix tube contents again.
13. Wash red cells three to four times with isotonic saline. Decant supernatant saline completely following each wash and ensure thorough resuspension and mixing of red cells with each new addition of saline for subsequent washes.
14. Following the final wash, completely decant the supernatant saline to ensure removal of all residual saline and a resultant ‘dry’ red cell button.
15. Add two drops of NOVACLONE™ Anti-IgG to each tube containing a ‘dry’ button of washed red cells.
16. Mix gently, but thoroughly, to resuspend red cells.
17. Centrifuge for:

a) 15 seconds at 900-1000 rcf.

b) or centrifugation of equivalent force‡.
18. Gently resuspend the red cell button and examine for agglutination. Grade and record results. *(A microscope, or other optical aid, may be used to confirm weak or negative hemagglutination reactions).*
19. Negative or weak positive antiglobulin test results should be appropriately controlled by the addition of IgG sensitized reagent control cells (See CONTROLS).

†No single centrifugation speed or time can be recommended for all types of available centrifuges or test applications. Each laboratory should calibrate their centrifuge equipment individually to determine the optimal centrifugation speed and time that produces the strongest agglutination reaction with antigen positive cells and allows complete and easy resuspension of negative reactions.

‡In other than low ionic test systems, it is a common and well-documented practice to increase the quantity of serum used in the test system (increase in serum:cell ratio). When using low ionic enhancement techniques, however, the low ionic reagents must be used exactly as outlined in the respective Directions for Use.

§In other than low ionic test systems, it is a common and well-documented practice to increase the incubation time beyond 15 minutes in order to increase the sensitivity of antibody detection/identification test procedures.

INTERPRETATION OF TEST RESULTS

POSITIVE: Agglutination of test red cells in the antiglobulin test phase of the direct or indirect antiglobulin test procedure constitutes a positive test result and, within the accepted limitations of the test procedure, indicates the presence of IgG on the red cells.

NEGATIVE: No agglutination of test red cells in the antiglobulin test phase of a direct or indirect antiglobulin test procedure constitutes a negative result and, within the accepted limitations of the test procedure, indicates the absence of serologically detectable IgG from the red cells.

STABILITY OF THE REACTION:

All test results should be read and interpreted immediately following centrifugation.

CONTROLS

Appropriate control tests are essential for all laboratory test procedures.

1. The application of IgG sensitized reagent control cells to aid in the confirmation of effective antiglobulin test results is an essential control test for antibody detection/identification procedures which include an indirect antiglobulin test phase (Refer to the relevant manufacturer's Directions for Use for IgG sensitized control cells).
2. The specificity and reactivity of Anti-Human Globulin may be verified by routine quality control procedures. Anti-IgG Anti-Human Globulin may be tested against red cells weakly sensitized with IgG and against red cells coated with C3 and C4 to verify the presence of active Anti-IgG and the absence of anti-C3 and anti-C4. Unsensitized red cells should be tested in parallel as a negative control.

LIMITATIONS OF THE TEST PROCEDURE

1. Anti-IgG should not be used as the sole antiglobulin reagent for direct antiglobulin tests. The exclusive use of anti-IgG for direct or indirect antiglobulin tests may result in false negative results.
2. Anti-IgG may be used for compatibility testing in place of Polyspecific Anti-Human Globulin. However, current scientific evidence indicates that the exclusive use of anti-IgG for this purpose may result in the failure to detect some blood group antibodies. Furthermore, the reactions of some complement-binding IgG antibodies may be enhanced when the Anti-Human Globulin reagent contains anti-complement in addition to anti-IgG.
3. A negative DAT result does not necessarily preclude clinical diagnosis of ABO Hemolytic Disease of the Fetus and Newborn (HDFN) or Autoimmune Hemolytic Anemia (AIHA).
4. Red cells with a positive DAT cannot be used in indirect antiglobulin test procedures.
5. Certain disease states and medication therapy may be associated with positive direct and/or indirect antiglobulin tests.
6. Weak or false negative antiglobulin test results may occur due to inactivation by residual human serum protein following inadequate wash procedures or dilution of the Anti-Human Globulin reagent associated with excess residual saline following wash procedures.
7. Overly vigorous or inappropriate resuspension of red cells in antiglobulin test procedures may result in weak or false negative results.
8. Procedural delays in antiglobulin test performance may result in weak or false negative results. Anti-IgG reactions, in particular, may be adversely affected by incubation and/or re-centrifugation.
9. Other variables such as improper technique, inappropriate centrifugation or incubation, improperly cleaned glassware, incorrect saline pH and/or contaminated materials may cause false negative or false positive results.
10. The use of green Anti-Human Globulin does not preclude the necessity for adequate control and confirmation of antiglobulin reactivity. *This dye provides a visual indication that the antiglobulin reagent has been added, but does not assure the reactivity of the reagent.*

SPECIFIC PERFORMANCE CHARACTERISTICS

Each lot of NOVACLONE™ Anti-IgG Anti-Human Globulin has been tested to ensure potency and specificity. Anti-IgG potency is assessed in serological tests with red cells coated specifically with IgG according to approved test procedures. Defined specificity is verified by tests against a variety of cells coated with different human proteins. The absence of anti-complement (anti-C3 and/or anti-C4) is verified in serological tests with cells coated with C3 and C4 according to approved test procedures. This reagent reacts specifically with red cells coated with human IgG when used in accordance with the recommended Directions for Use.

Deviation from the recommended Directions for Use may result in less than optimal product performance. User-defined modifications to test procedures may require validation.

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PRODUCT	PRODUCT CODES	
	10 mL vial	
NOVACLONE™ Anti-IgG Murine Monoclonal Blend	CLEAR	5461
	GREEN	5470

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