

General Information

This diagnostic kit is designed to detect antibodies directed against the internal nucleocapsid of the Influenza A virus.

It can be used with birds, horse or swine serum or plasma, but also with porcine oral fluid or chicken egg yolk samples.

Description and Principle

Wells are coated with Influenza A nucleoprotein (NP).

Specimens to be tested and controls are added to the microwells. Anti-NP antibodies, if present, form an antibody-antigen complex which masks the NP epitopes.

An anti-NP-peroxidase (HRP) conjugate is added to the microwells. It fixes to the remaining free NP epitopes, forming an antigen-conjugate-HRP complex.

After washing in order to eliminate the excess conjugate, the substrate solution (TMB) is added.

The resulting coloration depends on the quantity of specific antibodies present in the specimen to be tested:

- in the absence of antibodies, a blue coloration appears which becomes yellow after addition of the stop solution.
- in the presence of antibodies, no coloration appears.

The microplate is read at 450 nm.

Kit Components

Reagents*
Microplates coated with the NP
Concentrated Conjugate (10X)
Positive Control
Negative Control
Dilution Buffer 3
Dilution Buffer 2
Wash Concentrate (20X)
Substrate Solution
Stop Solution (0.5 M)

* Quantities supplied are indicated on the kit label.

1. The conjugate, the controls and the substrate solution must be stored at 5°C (± 3°C).
2. The other reagents can be stored between +2°C and +26°C.
3. For detailed storage conditions of opened and/or diluted components, please refer to www.innovative-diagnostics.com/storage-conditions
4. Wash and stop solutions can be used for the entire IDvet product range. Substrate solutions and dilution buffers with same batch numbers are interchangeable.

Materials required but not provided

1. Mono or multi-channel micropipettes capable of delivering volumes of 10 µL, 100 µL, and 500 µL.
2. Disposable tips.
3. 96 well pre-dilution plate
4. Distilled or deionized water.
5. Manual or automatic wash system.
6. 96-well microplate reader.

Precautions

1. Do not pipette by mouth.
2. Contains components that can be harmful to the skin and eyes and may cause sensitisation by skin contact. Avoid contact with skin and eyes. Use protective lab coat, one-way gloves and safety

glasses. The stop solution (0.5 M acid) may be harmful if swallowed.

3. Do not expose the substrate solution to bright light nor to oxidizing agents.
4. All waste should be properly decontaminated prior to disposal. Dispose in accordance with local regulations.

Please refer to the Material Safety Data Sheet, available upon request at info@innovative-diagnostics.com, for more detailed information

Wash Solution Preparation

If necessary, bring the **Wash Concentrate (20X)** to room temperature and mix thoroughly to ensure that the Wash Concentrate is completely solubilized.

Prepare the **Wash Solution (1X)** by diluting the **Wash Concentrate (20X)** to 1:20 in distilled/deionized water.

The quality of the wash step may influence results. Ensure that wells are completely empty between washes. If using an automatic washer, it is extremely important to correctly parameter the machine (mode, type of aspiration, aspiration height). For more information, please consult the "IDvet Washing Guide", available upon request.

Testing Procedure

Allow all the reagents to come to room temperature (21°C ± 5°C) before use. Homogenize all reagents by inversion or vortexing.

Note: Protocols for chicken, turkey, quail, guinea fowl, horse, duck, ostrich, and swine samples are proposed. For use in other species, please contact us.

Samples from different species require different dilution factors.

SERUMS OR PLASMAS:

For chicken, turkey, quail, guinea fowl, duck, geese, or ostrich samples

1. In the ELISA microplate, add:
 - 40 µL of **Dilution Buffer 2** in each well.
 - 10 µL of the **Positive Control** to wells A1 and B1.
 - 10 µL of the **Negative Control** to wells C1 and D1.
 - 10 µL of each sample to be tested to the remaining wells.

For horse samples

1. In the ELISA microplate, add:
 - 90 µL of **Dilution Buffer 2** in each well.
 - 10 µL of the **Positive Control** to wells A1 and B1.
 - 10 µL of the **Negative Control** to wells C1 and D1.
 - 10 µL of **each sample to be tested** to the remaining wells.

For swine samples

1. In the ELISA microplate, add:
 - 90 µL of **Dilution Buffer 2** and 10µL of the **Positive Control** to wells A1 and B1.
 - 90 µL of **Dilution Buffer 2** and 10µL of the **Negative Control** to wells C1 and D1.
 - 200 µL of **Dilution Buffer 2** and 5 µL of each sample to be tested in the remaining wells.

ORAL FLUIDS:

For swine samples

1. In the ELISA microplate, add:
 - 90 µL of **Dilution Buffer 2** and 10µL of the **Positive Control** to wells A1 and B1.
 - 90 µL of **Dilution Buffer 2** and 10µL of the **Negative Control** to wells C1 and D1.
 - 50 µL of **Dilution Buffer 2** and 50 µL of **each sample to be tested** in the remaining wells.

EGG YOLKS:

For chicken samples

1. a) Pre-dilute the samples 1:5 in **Wash Solution 1X** as follows:
 - Mix 80 µL of **Wash Solution 1X** with 20 µL of **egg yolk**
 - Mix well before testing
2. Note: These 1:5 diluted egg yolk samples can be stored at -20°C for future test runs. Not more than 3 freeze-thaw cycles.

1. b) In the ELISA microplate, add:
 - 90 µL of **Dilution Buffer 2** and 10µL of the **Positive Control** to wells A1 and B1.
 - 90 µL of **Dilution Buffer 2** and 10µL of the **Negative Control** to wells C1 and D1.
 - 100 µL of **each diluted egg yolk sample** to be tested to the remaining wells.

FOR ALL TYPES OF SAMPLES:

2. Cover the plate and incubate **60 min ± 6 min** at **37°C (± 2°C)**.
3. Empty the wells. Wash each well **5 times** with at least 300 µL of **Wash Solution**. Avoid drying of the wells between washes.
4. Prepare the **Conjugate 1X** by diluting the **Concentrated Conjugate 10X** to 1:10 in **Dilution Buffer 3**.
5. Add 50 µL of the **Conjugate 1X** to each well.
6. Cover the plate and incubate **30 min ± 3 min** at **21°C (± 5°C)**.
7. Empty the wells. Wash each well 3 times with at least 300 µL of **Wash Solution**. Avoid drying of the wells between washes.
8. Add 50 µL of the **Substrate Solution** to each well.
9. Cover the plate and incubate **10 min ± 1 min** at **21°C (± 5°C)** in the dark.
10. Add 50 µL of the **Stop Solution** to each well in the same order as in step No. 8 to stop the reaction.
11. Read and record the O.D. at 450 nm.

Validation

The test is validated if:

✓ the mean value of the Negative Control O.D. (OD_{NC}) is greater than 0.700.

$$\text{OD}_{\text{NC}} > 0.700$$

✓ the mean value of the Positive Control (OD_{PC}) is less than 30 % of the OD_{NC}.

$$\text{OD}_{\text{PC}} / \text{OD}_{\text{NC}} < 0.3$$

Interpretation

For each sample, calculate the competition percentage (S/N%):

$$\text{S/N\%} = \frac{\text{OD}_{\text{sample}}}{\text{OD}_{\text{NC}}} \times 100$$

Samples presenting a S/N%:

- greater than or equal to 50% are considered negative.
- between 45% and 50% are considered doubtful.
- less than or equal to 45% are considered positive.

Result	Status
S/N% ≤ 45%	POSITIVE
45% < S/N% < 50%	DOUBTFUL
S/N% ≥ 50%	NEGATIVE

Note: The IDSoft™ data analysis program is available free-of-charge. For more information, please contact support.software@innovative-diagnostics.com.

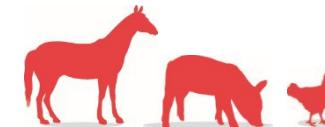
This software program can calculate many parameters (validation criteria, S/P or S/N values, titers, vaccination age, groups) and offers a graphic representation of the serological profiles of the animals tested).



Certified
management
system



ID Screen® Influenza A Antibody Competition Multi-Species



Competitive ELISA for the detection of antibodies against the nucleoprotein of the Influenza A virus in avian, porcine, or equine serum, plasma, in porcine oral fluid or in chicken egg yolk samples.

For *in vitro* use

September 2021

Protocol for egg yolk samples now included in the insert
(while previously provided separately)

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