

virotype[®] CSFV 2.0 RT-PCR Kit Handbook

For detection of RNA from *Classical Swine
Fever Virus (CSFV)*

Licensed in accordance with § 11 (2) of the German Animal Health Act
MA No.: FLI-C 113

REF 96 reactions (Cat. no. VT281825)



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Contents

Kit contents.....	3
Intended use.....	3
Symbols.....	4
Quality control.....	4
Storage.....	4
Safety information.....	5
Introduction.....	6
Principle.....	6
RNA extraction.....	7
Equipment and reagents to be supplied by user.....	9
Important notes.....	10
General precautions	10
Protocol: Real-time RT-PCR for detection of RNA from <i>Classical Swine Fever Virus</i>	12
Important points before starting.....	12
Things to do before starting.....	12
Procedure	13
Data analysis and interpretation.....	15
Interpretation of results.....	15
Change index.....	20

Kit contents

virotype CSFV 2.0 RT-PCR Kit	(96)
Cat. no.	VT281825
Number of reactions	96

Master Mix (tube with orange cap), includes primers, probes and enzymes	2 x 980 µl
Positive Control (tube with red cap)	1 x 150 µl
Negative Control (tube with blue cap)	1 x 150 µl
Handbook	1

Intended use

The virotype CSFV 2.0 RT-PCR Kit is intended for the detection of RNA from *Classical Swine Fever Virus* (CSFV) in blood, tissue, swabs, and samples in stabilizing transport media from pigs and wild boar.

The kit is approved by the Friedrich-Loeffler-Institut and licensed in accordance with § 11 (2) of the German Animal Health Act (FLI-C 113) for use in Germany for veterinary diagnostic procedures.

For veterinary use only.

Symbols



Legal manufacturer



Lot number



Use by date



Temperature limitations for storage



Handbook



Catalog number



Material number



Protect from light



For samples from pigs and wild boar

Quality control

In accordance with INDICAL's ISO-certified Quality Management System, each lot of virotype CSFV 2.0 RT-PCR Kit is tested against predetermined specifications to ensure consistent product quality.

Storage

The components of the virotype CSFV 2.0 RT-PCR Kit should be stored at -30°C to -15°C and are stable until the expiration date stated on the label. Avoid repeated thawing and freezing (> 3x), as this may reduce assay sensitivity. Freeze the components in aliquots if they will only be used intermittently.

Safety information

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, please consult the appropriate safety data sheets (SDSs). These are available from your local sales representative or by Email request under **compliance@indical.com**.

All sample residues and objects that have come into contact with samples must be decontaminated or disposed of as potentially infectious material.

Introduction

The virotype CSFV 2.0 RT-PCR Kit is a highly sensitive and specific solution for the detection of RNA from *Classical Swine Fever Virus* (CSFV) in samples from pigs and wild boar. Classical Swine Fever (CSF) is economically one of the most important viral infectious diseases of swine. CSF is widespread in domestic pig and wild boar populations. CSF is an internationally notifiable animal disease. The causative agent, *Classical Swine Fever Virus*, is a single-stranded RNA virus and a member of the genus *Pestivirus* which belongs to the *Flaviviridae* family like *Bovine Viral Diarrhea Virus* (BVDV) in cattle and *Border Disease Virus* (BDV) in sheep.

The high sensitivity of the virotype CSFV 2.0 RT-PCR Kit allows the early detection of the pathogen in individual or pooled blood samples, in individual or pooled tissue samples, as well as from swabs, and samples in stabilizing transport media from pigs and wild boar. The virotype CSFV 2.0 RT-PCR Kit detects all known CSFV-strains of various genotypes. In rare cases the test may detect CSF vaccine virus. Positive results of animals from areas where CSFV vaccination is performed should therefore be verified.

Principle

Polymerase chain reaction (PCR) is based on the amplification of specific regions of the pathogen genome. In real-time PCR, the amplified product is identified using fluorescent dyes. These are usually linked to oligonucleotide probes that bind specifically to the amplified product. Monitoring the fluorescence intensities during the PCR run (i.e., in real time) allows detection of the accumulating product without the need to re-open the reaction tubes afterward.

The virotype CSFV 2.0 RT-PCR Kit contains all of the necessary reagents for the detection of CSFV RNA, including a Positive and Negative Control. With this kit, both, reverse transcription and PCR are performed in one reaction tube, reducing the risk of contamination.

An internal control excludes the possibility of false-negative results. The kit uses two specific primer/ probe combinations:

- FAM™ fluorescence for RNA of CSFV
- JOE™ fluorescence for the endogenous Internal Control (EC; β -actin present within the sample)

A Positive Control serves to verify the functionality of the reaction mix for the amplification of the CSFV RNA target.

RNA extraction

The virotype CSFV RT-PCR Kit can be used for the detection of CSFV RNA from blood, tissue, swabs, and samples in stabilizing transport media from pigs and wild boar.

Due to the high sensitivity of the test individual or pooled samples can be tested. For domestic pigs, up to 20 individual serum, plasma, or EDTA-blood samples or up to 10 tissue samples can be used. Furthermore, for wild boar, pools can consist of up to 10 serum, plasma, EDTA-blood, or tissue samples. Also swab samples and samples in stabilizing transport media from pigs and wild boar can be tested.

Note: For use in Germany the specifications described in the „*Amtliche Methodensammlung*“ apply.

Prior to real-time RT-PCR, viral RNA must be extracted from the starting material. INDICAL offers a range of validated kits for the extraction of RNA from animal samples.

Extraction based on magnetic beads:

- **IndiMag® Pathogen Kit*** (SP947457)
- **IndiMag Pathogen Kit* w/o plastics** (SP947257)
- **IndiMag Pathogen IM48 Cartridge** (SP947654P608, SP947654P224)
- **IndiMag Pathogen KF96 Cartridge** (SP947855P196)

Extraction based on spin columns:

- **IndiSpin® Pathogen Kit*** (SP54104, SP54106)
- **IndiSpin QIAcube® HT Pathogen Kit** (SP54161)

* suitable for simultaneous extraction of CSFV RNA und ASFV DNA

Note: When using difficult sample material, it is recommended to use INDICAL's „Pretreatment T4 (phenol extraction)“.

If real-time RT-PCR is not performed immediately after extraction, store the RNA at -20°C or at -70°C for longer storage.

For further information on automated and manual extraction of CSFV RNA from different sample types, refer to the respective handbook or contact INDICAL Support at **support@indical.com**.

Equipment and reagents to be supplied by user

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

- Pipets
- Nuclease-free, aerosol-resistant pipet tips with filters
- Sterile 1.5 ml Eppendorf® tubes
- Nuclease-free (RNase/ DNase-free) consumables. Special care should be taken to avoid nuclease contamination of all reagents and consumables used to set up PCR for sensitive identification of viral nucleic acids
- Cooling device or ice
- Benchtop centrifuge with rotor for 1.5 ml tubes
- Real-time cycler with appropriate fluorescent channels
- Appropriate software for chosen real-time cycler
- Appropriate strip tubes and caps or 96-well optical microplate with optical sealing film or cover for chosen real-time cycler

Important notes

General precautions

The user should always pay attention to the following:

- Use nuclease-free pipet tips with filters.
- Store and extract positive materials (specimens, positive controls and amplicons) separately from all other reagents and add them to the reaction mix in a spatially separated facility.
- Thaw all components on ice before starting as assay.
- When thawed, mix the components by inverting and centrifuge briefly.
- Do not use components of the test kit past the expiration date.
- Keep samples and controls on ice or in a cooling block during the setup of reactions.

Negative control

At least one negative control reaction should be included in each PCR run, containing all the components of the reaction except for the pathogen template. This enables assessment of contamination in the reaction.

Positive control

When performing PCR on unknown samples, it is recommended to perform a positive control reaction in the PCR run, containing a sample that is known to include the targeted viral RNA. A positive control serves to prove the functionality of the pathogen assay, e.g., the correct setup of the reaction mix. Use 5 µl of the Positive Control provided with

the virotype CSFV 2.0 RT-PCR Kit to test for successful amplification of the target.

Extraction and amplification control

For increased process safety and convenience, one extraction and amplification control assay is included in the test kit.

An endogenous internal control (EC) detects the β -actin gene present within the sample. This allows extraction and amplification to be monitored.

Protocol: Real-time RT-PCR for detection of RNA from *Classical Swine Fever Virus*

Important points before starting

- Please read „Important notes“ on page 10 before starting.
- Include at least one positive control (Positive Control) and one negative control (Negative Control) per PCR run.
- Before beginning the procedure, read through the protocol and ensure that you are familiar with the operation of the chosen real-time PCR cyclers.
- RNA is unstable. Perform the protocol without interruption.

Things to do before starting

- Thaw all reagents on ice and protect from light.
- Before use, spin the reagents briefly.
- Maintain reagents on ice or in a cooling block during PCR setup.

Procedure

1. Before use, mix the Master Mix by inverting 5 times or until mixed thoroughly, then centrifuge briefly to collect the fluids.
2. Pipet 20 μl of the Master Mix into each reaction tube. Then add 5 μl of the sample RNA (Table 1).

Include positive and negative control reactions.

Positive Control: Use 5 μl of the positive control (Positive Control) instead of sample RNA.

Negative Control: Use 5 μl of the negative control (Negative Control) instead of sample RNA.

Table 1. Preparation of reaction mix

Component	Volume
Master Mix	20 μl
Sample	5 μl
Total volume	25 μl

3. Close the reaction tubes or seal the plate and invert 5 times or until mixed thoroughly. Then centrifuge briefly to collect the fluids.
4. Set the filters for the reporter dyes in the software of your thermal cycler according to Table 2.

Table 2. Filter settings for the reporter

Pathogen/ internal control	Reporter
CSFV	FAM
Endogenous internal control (EC)	HEX/ JOE™ ¹
Passive reference ²	ROX™

1 Use the option appropriate for your thermal cycler.

2 Internal reference for use with ABI PRISM® Sequence Detection Systems (Applied Biosystems®)

5. Run the real-time PCR protocol according to Table 3.

Table 3. Real-time RT-PCR protocol for CSFV 2.0

Step	Temperature	Time	Number of cycles
Reverse Transcription	50°C	10 min	1
Initial Activation	95°C	2 min	1
2-step cycling			
Denaturation	95°C	5 s	40
Annealing/ Extension*	60°C	30 s	

* Fluorescence data collection, approximate run time 72 min (CFX96, Bio-Rad™)

Note: The above protocol can also be used in combination with the virotype ASFV 2.0 PCR Kit.

Data analysis and interpretation

Interpretation of results

For the assay to be valid the Positive Control must give a signal in the FAM and HEX/ JOE channels with a $C_T^1 < 35$. The Negative Control must give no signal.

The following results are possible if working with unknown samples. The possible sample results are also summarized in Table 4 on page 17.

The sample is positive for CSFV, and the assay is valid, if the following criteria are met:

- The sample yields a signal in the FAM channel (regardless of any signal in the HEX/ JOE channel).
- The Positive Control yields a signal in the FAM and HEX/ JOE channel.
- The Negative Control does not yield a signal in the FAM and the HEX/ JOE channels.

Note that very high concentrations of CSFV RNA in the sample may lead to a reduced signal or no signal for the endogenous Internal Control (EC; HEX/ JOE channel) due to competition with the internal control.

¹ Threshold cycle (C_T) — cycle at which the amplification plot crosses the threshold, i.e., there is the first clearly detectable increase in fluorescence

The sample is negative for CSFV, and the assay is valid, if the following criteria are met:

- The sample **does not yield** any signal in the FAM channel.
- The sample yields a signal in the HEX/ JOE channel.
- The Positive Control yields a signal in the FAM and HEX/ JOE channels.
- The Negative Control does not yield a signal in the FAM and HEX/ JOE channels.

The sample results are inconclusive, and the assay is invalid, if the following criteria are met:

- The sample yields no signal in the FAM and HEX/ JOE channels.

If no signal is detected in the FAM (CSFV) and the HEX/ JOE (endogenous Internal Control, EC) channel, the result is inconclusive. The absence of a signal for the housekeeping gene indicates strong PCR inhibition and/or other malfunctions, e.g. during extraction.

To check for inhibition, we recommend 1:5 dilution of the sample RNA in nuclease-free water, to repeat the RNA extraction procedure, or repeat the whole test procedure starting with new sample material.

Check that there is a fluorescence signal in the FAM channel for the positive control reaction (Positive Control). Absence of a signal for the Positive Control indicates an error, which could be due to incorrect setup of the reaction mix or incorrect cycling conditions.

Table 4. Results interpretation table*

Sample result	FAM (CSFV)	HEX/ JOE (EC)
CSFV positive	X	X
CSFV strong positive	X	
CSFV negative		X
Inconclusive		

* Interpretation of sample results can be determined provided positive and negative control reactions are performed. The Positive Control must yield a signal in the FAM and HEX/ JOE channels. The Negative Control must yield no signal in any channel. For a complete explanation of possible sample results please refer to “Data analysis and interpretation” on page 15.

INDICAL offers a range of ELISA kits and real-time PCR and real-time RT-PCR kits for the detection of animal pathogens.

Visit www.indical.com for more information about afosa, bactotype, cador, cattletype, flocktype, pigtype, Svanovir and virotype products.

For up-to-date licensing information and product-specific disclaimers, see the respective INDICAL kit handbook or user manual.

Notes

Notes

Limited License Agreement for virotype CSFV 2.0 RT-PCR Kit

Use of this product signifies the agreement of any purchaser or user of the product to the following terms:

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Change index

Handbook	Version	Change
HB-2576-EN-001	July 2022	Product launch