

# **Technical Data**

# SIM Motility Medium, Modified

# **M181F**

SIM Medium is recommended for determination of hydrogen sulphide production, indole formation and motility of enteric bacilli in accordance with FDA BAM.

### **Composition\*\***

Ingredients	Gms / Litre
Pancreatic digest of casein	20.000
Peptic digest of animal tissue	6.100
Ferrous ammonium sulfate	0.200
Sodium thiosulfate	0.200
Agar	3.500
Final pH ( at 25°C)	7.3±0.2

\*\*Formula adjusted, standardized to suit performance parameters

# **Directions**

Suspend 30.0 grams in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Dispense in tubes. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Allow the tubes to cool in an upright position.

# **Principle And Interpretation**

SIM Medium is recommended by FDA BAM, 1998 (1) to differentiate enteric bacilli particularly *Salmonella* and *Shigella* on the basis of sulphide production, indole formation and motility (2). Jordan and Victorson (3) reported that *Salmonella* Paratyphi A and Paratyphi B can be distinguished on the basis of H2S production using lead acetate. Sulkin and Willett (4) used Triple Sugar Iron Agar with 1% agar for motility along with H2S production and carbohydrate fermentation. Sosa (5) described a peptone medium with low agar for motility and indole determination.

Motility, indole and sulphide production tests are used to differentiate *Enterobacteriaceae* members. SIM Medium combines these three differential characteristics in a single medium. Peptonized iron and sodium thiosulphate are the indicators of H2S production. This H2S reacts with peptonized iron to form black precipitate of ferrous sulphide. *Salmonella* are H2S positive and *Shigella* are H2S negative. Motile organisms intensify the H2S reaction. Motile organisms grow away from line of inoculation showing diffused growth while non-motile organisms grow along the stabline. Motility detection is possible due to the semisolid nature of the medium. *Salmonella* is motile while *Shigella* are non motile. Tryptophan, from peptic digest of animal tissue, is degraded by specific bacteria to produce indole. The indole is detected by the addition of chemical reagents following the incubation period.

Inoculate fresh culture with a single stab using straight needle through the center of the medium. Following incubation, observe for motility (diffuse growth outward from the stabline or turbidity throughout the medium) and for H2S production (blackening of the medium). To detect indole production, add three or four drops of Kovacs reagent and observe for development of red color (positive reaction). Determine motility and H2S production prior to determination of indole production.

# **Quality Control**

Appearance

Cream to beige homogeneous free flowing powder **Gelling** Semisolid, comparable with 0.3% Agar gel. **Colour and Clarity of prepared medium** Medium amber coloured slightly opalescent gel forms in tubes as butts **Reaction** Reaction of 3.0% w/v aqueous solution at 25°C. pH : 7.3±0.2 **pH** 7.10-7.50

Please refer disclaimer Overleaf.

#### **Cultural Response**

Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

#### **Cultural Response**

Cultural Response						
Organism	Inoculum (CFU)	Growth	Motility	Indole production(on addition of Kovac's	H2S	
<b>Cultural Response</b> <i>Escherichia coli ATCC</i> 25922	50-100	luxuriant	positive, growth away from stabline causing turbidity	positive reaction, red ring at the interface of the medium	negative reaction	
Salmonella Typhimurium ATCC 14028	50-100	luxuriant	positive, growth away from stabline causing turbidity	negative reaction	positive reaction, blackening of medium	
Shigella flexneri ATCC 12022	50-100	luxuriant	negative, growth along the stabline, surrounding medium remains clear	negative reaction	negative reaction	
Salmonella Paratyphi A ATCC 9150	50-100	luxuriant	positive, growth away from stabline causing turbidity	negative reaction	Negative reaction	
Salmonella Paratyphi B ATCC 8739	50-100	luxuriant	positive, growth away from stabline causing turbidity	Negative reaction	Positive reaction, blackening of medium	
Klebsiella pneumoniae ATCC 13883	50-100	luxuriant	negative, growth along the stabline, surrounding medium remains clear	Negative reaction	Negative reaction	

#### **Storage and Shelf Life**

Store below 30°C in tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label.

#### Reference

1.FDA, U.S. 1998. Bacteriological Analytical Manual. 8 ed. Gaithersburg, MD: AOAC International.

2.MacFaddin, J. F. 1985. Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria vol. 1. Baltimore: Williams and Wilkins.

3.Jordan, E. O. and Victorson, R 1917. J. Inf. Dis, 21.

4.Sulkin, S. E. and Willett, J. C 1940. J. Lab. Clin. Med., 25.

5.Sosa, L 1943. Rev. Inst. Bacteriol, 11.

Revision : 2 / 2015

#### Disclaimer :

User must ensure suitability of the product(s) in their application prior to use. Products conform solely to the information contained in this and other related HiMedia<sup>™</sup> publications. The information contained in this publication is based on our research and development work and is to the best of our knowledge true and accurate. HiMedia<sup>™</sup> Laboratories Pvt Ltd reserves the right to make changes to specifications and information related to the products at any time. Products are not intended for human or animal or therapeutic use but for laboratory,diagnostic, research or further manufacturing use only, unless otherwise specified. Statements contained herein should not be considered as a warranty of any kind, expressed or implied, and no liability is accepted for infringement of any patents.

HiMedia Laboratories Pvt. Ltd. A-516, Swastik Disha Business Park, Via Vadhani Ind. Est., LBS Marg, Mumbai-400086, India. Customer care No.: 022-6147 1919 Email: techhelp@himedialabs.com Website: www.himedialabs.com